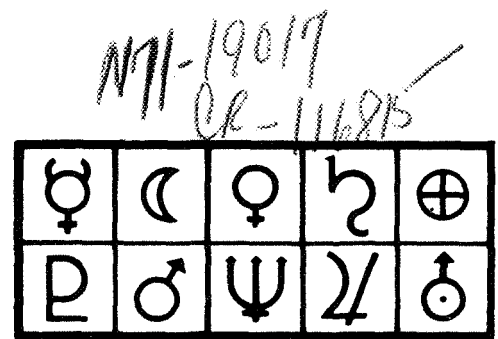




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AN APPROACH TO COMPUTERIZED BACTERIAL IDENTIFICATION



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Program Completed -- November 1970

ABSTRACT

This report describes the analysis and design of a general approach to bacterial identification which readily lends itself to computerization. This approach has been applied to the microbial sampling data of four Apollo spacecraft. These data were supplied by the PHS Spacecraft Bioassay Laboratory who obtained them in connection with NASA's lunar planetary quarantine responsibility. The approach described here was found to agree quite well with the PHS identifications assigned in the laboratory.

The computer program used in this application is described in detail, and some analysis of the results of the use of this computer program with the Apollo data is given. Other possible applications of the general approach to bacterial identification described here are mentioned.

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AN APPROACH TO COMPUTERIZED BACTERIAL IDENTIFICATION

Introduction

Interest in the use of a computer to identify microorganisms arose during microbial sampling of Apollo spacecraft. This sampling was done in response to the lunar planetary quarantine responsibility of NASA which requires that an inventory of probable post landing biological contamination levels at each manned and automated landing site (1) be maintained, and that microbial life be identified, quantified, and insofar as possible located on the lunar surface (2). The U.S. Public Health Service (PHS) collected, cultured, identified and stored for future reference the microbial samples obtained from the spacecraft. A Lunar Information System, designed by Sandia Laboratories (3), is now being used to store, categorize and analyze the data collected by the Public Health Service so that the above information is continuously available to the NASA Planetary Quarantine Officer (4).

Inasmuch as the data collected by the PHS, as well as the results of their identification, were already being stored in the Lunar Information System, it was questioned whether it was feasible for the Lunar Information System to "standardize" the identification process so that it could be accomplished within the information system. In fact, this possibility was earlier considered in the Lunar Information System concept (3). The feasibility of computerized bacterial identification was demonstrated earlier (5) using the Apollo 10, 11, and 12 data that were then available in the Lunar Information System. Feasibility here was based upon agreement with the PHS identification, and it was found that a gross computerized identification scheme agreed with PHS identification in excess of 87 percent of the time. Based on these results, it was proposed that a program be incorporated into the Lunar Information System which would utilize the laboratory data provided by the PHS and, in fact, make the identification. Such a program has been written and its efficiency has been determined by use of previous data from Apollo missions 10, 11, 12, and 13. The data are shown in Figure 1. Some of these data have previously been published (6). It was felt that a total of 7,049 samples would provide a good data base for comparison of the two approaches.

		Apollo			
		10	11	12	13
A. Catalase positive, gram positive, aerobic cocci					
001 Staphylococcus, Subgroup I		142	28	1	0
002 " " II		261	355	334	152
003 " " III		38	6	29	2
004 " " IV		131	242	228	51
005 " " V		163	215	152	34
006 " " VI		134	154	216	48
007 Micrococcus, Subgroup 1		142	207	160	124
008 " " 2		85	103	82	80
009 " " 3		68	36	77	75
010 " " 4				1	0
011 " " 5		20	26	20	4
012 " " 6				3	0
013 " " 7		94	104	283	220
014 " " 8		4	1		0
Totals Scheme A		1282	1477	1566	790
B. Catalase negative, gram positive, aerobic cocci					
015 Entrococcus		2			
016 Lactic					
017 Viridans		15	5		1
018 Pyogenic		3			
Totals Scheme B		20	5	0	1
C. Gram positive, spore forming, aerobic rods					
019 B. alvei		3		2	
020 B. badius			2	5	
021 B. brevis		1		1	
022 B. cereus		2	4	4	4
023 B. circulans		13	1	8	5
024 B. coagulans			2	7	4
025 B. firmus		2	4	3	4
026 B. laterosporus		1			
027 B. lentus		7	2	1	4
028 B. licheniformis		6	8	2	1
029 B. macerans		3	1		
030 B. megaterium		3	1	1	
031 B. pathothenicus		1	20	3	3
032 B. polymyxa		2	12	4	2
033 B. pulvificiens		2	1	3	2
034 B. pumilus		1	7		0
035 B. sphaericus		1	1	4	2
036 B. subtilis				3	1
Totals Scheme C		45	66	51	32
D. Aerobic and anaerobic non-spore forming gram positive rods, anaerobic gram negative cocci and anerobic gram negative rods: Test reactions only.					
037 Aerobic, non-sporeforming gram positive rods		160	183	195	159
038 Anaerobic, non-sporeforming, gram positive rods					
039 Anaerobic gram negative cocci					
040 Anaerobic gram negative rods					
Totals Scheme D		160	183	195	159
E. Aerobic gram negative cocci:					
041 Neisseria, spp.					
Totals Scheme E		0	0	0	0
F. Aerobic gram negative rods:					
042 Achromobacter, spp.			2		
043 Alcaligenes, spp.		3	9	2	
044 Flavobacterium, spp.		1	1	7	
045 Pseudomonas, spp.		1			
Totals Scheme F		5	12	9	0

Figure 1. PHS Identifications for Apollo 10, 11, 12, and 13

		Apollo			
		10	11	12	13
G.	046 Gram positive, catalase negative, anaerobic cocci				
	Totals Scheme G	0	0	0	0
H.	047 Gram positive, catalase negative anaerobic cocci				
	Totals Scheme H	0	0	0	0
I.	Anaerobic sporeforming gram positive rods				
	048 C. acetobutylicum				
	049 C. aerofotidum				
	050 C. amyolyticum				
	051 C. amylosaccharobutylprophylicum				
	052 C. aurantibutyricum				
	053 C. bejerinckii				
	054 C. bifermentans				
	055 C. botulinum				
	056 C. butylicum				
	057 C. butyricum				
	058 C. capitoale				
	059 C. caproicum				
	060 C. carnis				
	061 C. cochlearium				
	062 C. difficile				
	063 C. fallax				
	064 C. felsineum				
	065 C. festeri				
	066 C. flavum				
	067 C. haemolyticum				
	068 C. histolyticum				
	069 C. innocuum				
	070 C. iodophilum				
	071 C. lactoacetophilum				
	072 C. lentoputrescens				
	073 C. novyi				
	074 C. parabolulinum				
	075 C. paraputrificum				
	076 C. pasteurianum				
	077 C. pectinovorum				
	078 C. perfringens				
	079 C. roseum				
	080 C. septicum				
	081 C. sordelli				
	082 C. phenoides				
	083 C. sporogenes				
	084 C. subterminale				
	085 C. tertium				
	086 C. tetani				
	087 C. tetanomorphum				
	Totals Scheme I	0	0	0	0
J.	088 Actinomycetes	2	7	1	2
	Totals Scheme J	2	7	1	2
K.	089 Streptomycetes				1
	Totals Scheme K	0	0	0	1
L.	090 Yeasts	5	8	13	18
	Totals Scheme L	5	8	13	18
M.	091 Molds	37	34	36	14
	Totals Scheme M	37	34	36	14
	092 Atypical cocci	9	57	22	2
	093 Aerobacter	1	0	0	0
	094 Atypical bacillus	17	13	2	0
	095 Nogro Differential Media	409	173	76	1*
	Total Apollo	1992	2020	1997	1020
	*Lost in process	Total = 7049 samples			

Figure 1. (Cont.)

When discussing the "identification" of bacteria, the meaning of the word must be agreed upon as accurately as possible. It would be nice if there were a well defined set containing all bacteria, each of which could be characterized by the results of some standard tests. Then any unknown could be subjected to the tests and the test outcomes compared to those describing the above categories until a match was obtained yielding an identification. Unfortunately, such an idealistic view of the identification of microbes is, to date, impractical. The existing knowledge about the set of all microorganisms is too incomplete, and a theoretical or practical basis for completely subdividing the set of those we know about has serious limitations. Some of the difficulties are as follows:

1. A precise workable definition of bacteria is not available. For example, a recent text (7) uses a modified definition due to Cohn, 1872, which is totally inadequate in view of recent knowledge.
2. Even if well-defined, the class of all bacteria would be open ended. Naturally occurring or artificially induced mutations are being formed constantly, many with newly observed properties.
3. The existing classification used in the U.S., Bergey's Manual (8), is based at least in part on old categories which were devised by means of old lists, some of which are now regarded as inconclusive or, at best, subjective.
4. The use of all "comprehensive" keys, including Bergey's, for identification requires the frequent interpretation of statements qualified as: rarely, frequently, some, and usually - which leads to overlapping categories.

The above difficulties reflect those differences between the mechanical way a computer operates and the manner in which people function. The computer requires well-defined categories and tests with a set of rules to relate them. It is difficult to include subjective judgments such as: "since these other tests came out that way and this is variable, its importance is minimal since it looks like _____ anyway". It might seem that a mechanized (or standardized) identification would be difficult, if not impossible, which is probably a fair conclusion if one operates within the above traditional framework. However, simple efficient schemes, capable of computerization, can be devised for special purposes. It should be noted that an identification scheme need only be adequate for the task at hand. There should be no objection to

having such a scheme suitable for the PHS Apollo task, another for clinical diagnostic work, another for work in the problem of bacterial classification or taxonomy, and yet others suitable for research with particular strains.

For NASA's Apollo Program, the U. S. PHS had devised an identification scheme tailored to that particular need (9). Categories appropriate to this need were defined in terms of test outcomes without mandatory reference to traditional categories of bacteria. Appendix 1 contains the current Data Reporting Form and list of categories being used. There was a backup inasmuch as cultures of the sampled microbes were to be stored for future reference. In addition, all laboratory data and the associated identification were stored and available to a computer as part of the Lunar Information System. Thus, it was possible to simulate the identification process and conveniently compare results with those of the PHS. Moreover, the large amount of data available in the NASA Lunar Information System provided an excellent basis for comparison of the two methods.

Every effort was made during the development of the system to retain as much flexibility as possible in the hope that it would have wider applications than the Apollo system for which it was designed. It was felt, in particular, that it should be constructed so that it could easily be modified by a microbiologist without reprogramming. This was accomplished by arranging the program so that it performed operations only on the data subject to the particular identification scheme which it had read. That is, not only were the data input, but the identification process itself became input. The form of the PHS identification scheme suggested a manner in which this could easily be done. The process that the PHS uses in order to identify a sample with the data they have collected was abstracted so that the computer would make the same (or similar) decisions. Thus, with some limitations, almost any scheme to be used can be handled by the program. The program will be described first by using this scheme as an example and then by showing the results and potential applications.

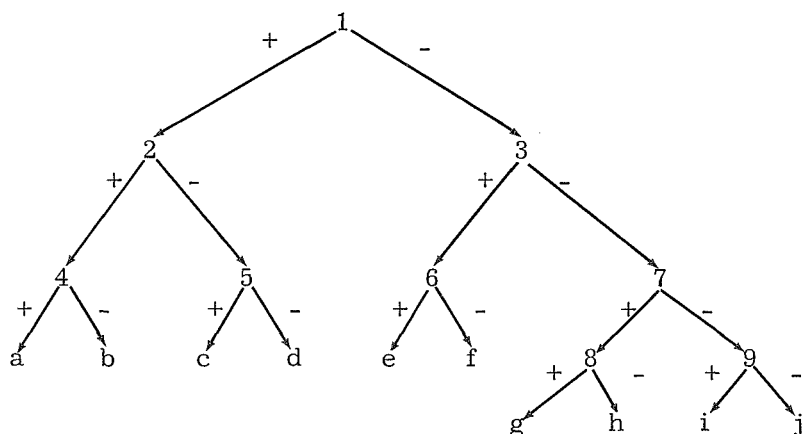
The Abstract Approach

Ideally, a microbiological identification system would consist of a (closed) class, M , of microbiological categories and a set, T , of tests which are sufficient to span M , that is, a set of tests with the property that for every pair of categories in M there is

at least one test which distinguishes between these. Then the problem of identification of a particular sample would reduce to taking the sequence of test results arising from the sample and comparing those with the test results associated with each category in M until an "identification" is made. Indeed, something approaching this ideal is necessary if one is to think in terms of a computerized identification scheme. However, as indicated in the Introduction, the class of all microorganisms as traditionally defined is not closed and, in fact, is continually changing, as is the set of tests that are used. Furthermore, there is no well-defined and generally accepted manner by which one can proceed from a specified series of test outcomes to a definite microorganism category. Rather, there are numerous different approaches to the problem of microbial identification. Some of these have been developed for specific groups of microorganisms, for particular application, and, in some cases, more as a method of research documentation. Some of the various types of approaches will be discussed.

Every approach to identification of organisms in some class M relies on an underlying classification of the organisms belonging to M. A classification is merely a specification of the abstract relationship between organism categories (in M) and test results. There are two basic means of presenting such a classification scheme, both of which from an informational point of view, are essentially equivalent, but which have totally different implications to practicing microbiologists when used for identification purposes.

First, there is the more traditional tree form of a classification scheme, [as exemplified in Bergey's Manual (8), discussed earlier]. In this approach, the tests occur as nodes of a tree, the end branches of which represent organism categories. An abstract example of this is shown below:



In this figure, there are organism categories a, b, c, d, e, f, g, h, i, and j. In the tree, nine tests are needed to distinguish among these. Although this tree indicates each test has only one of two outcomes (+ or -), this need not be the case in general classification schemes [e. g., Bergey's Manual (8)]. When used for identification purposes, this approach to classification has the implied disadvantage that the performance of tests must be done in a sequential manner with time. This results in running the fewest number of tests, but not necessarily in obtaining identification in the least amount of time, or indeed, with the least man-time in repeated routine identification programs.

The second approach to the presentation of classification schemes is through the use of matrices relating test results to organism categories. An example of such a matrix is shown in the figure below:

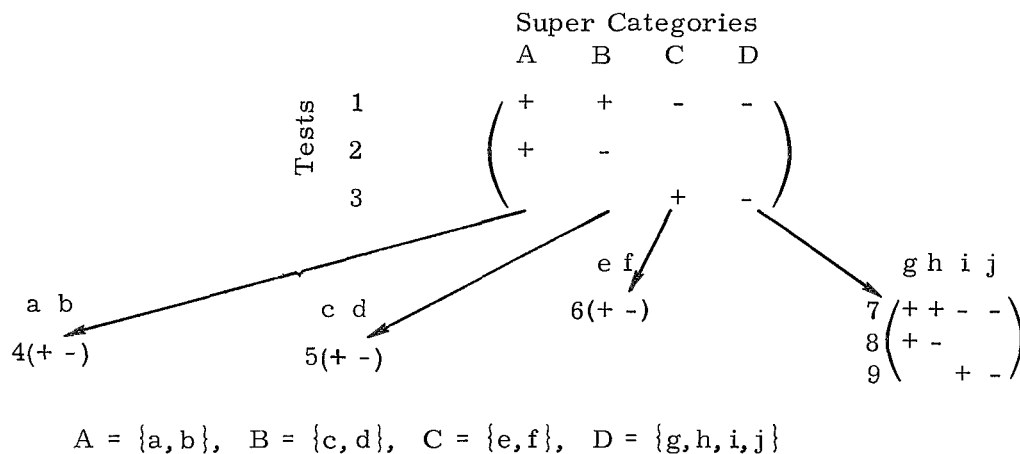
	Organism Categories									
	a	b	c	d	e	f	g	h	i	j
Tests	1	+	+	+	+	-	-	-	-	-
	2	+	+	-	-					
	3				+	+				
	4	+								
	5		+							
	6				+					
	7						+	+		
	8						+	-		
	9								+	

Here the rows represent tests, the columns represent organism categories, and an entry in the matrix represents the test outcome for the row (in which it occurs) and for the organism category of the column in which it occurs. Thus, for example, the (5, c) entry indicates that test 5 yields a + result when run on organisms in category c. The matrix form of a classification scheme implies in effect that one must run all tests prior to attempting an identification. This is not to say that all are needed a posteriori, but they must be known since the organism being identified is not known. This has the disadvantage of frequently taking much unnecessary data and requiring unnecessary time, but has the advantage that the scheme itself imposes no sequential time constraints like those found with a tree.

It was asserted earlier that these two approaches (trees and matrices) have essentially the same abstract informational value. This may be seen by comparing the above tree and matrix. They both use precisely the same test results to characterize each organism category. For example, in both cases, to characterize organism g, one needs only specify that tests 1 and 3 be negative and that tests 7 and 8 be positive. Indeed, any column of the matrix in which entries occur is obtained from the path of the tree leading to the organism category corresponding to that column.

From the point of view of taking laboratory data in identification situations, neither the tree nor the matrix approach has an abstract clear-cut advantage as regards "efficiency". Any such advantage would depend highly upon the categories and tests. For example, if some tests occur in all paths of the tree (that is, their results must be known in identifying all the organism categories), then these tests might just as well be represented by a matrix.

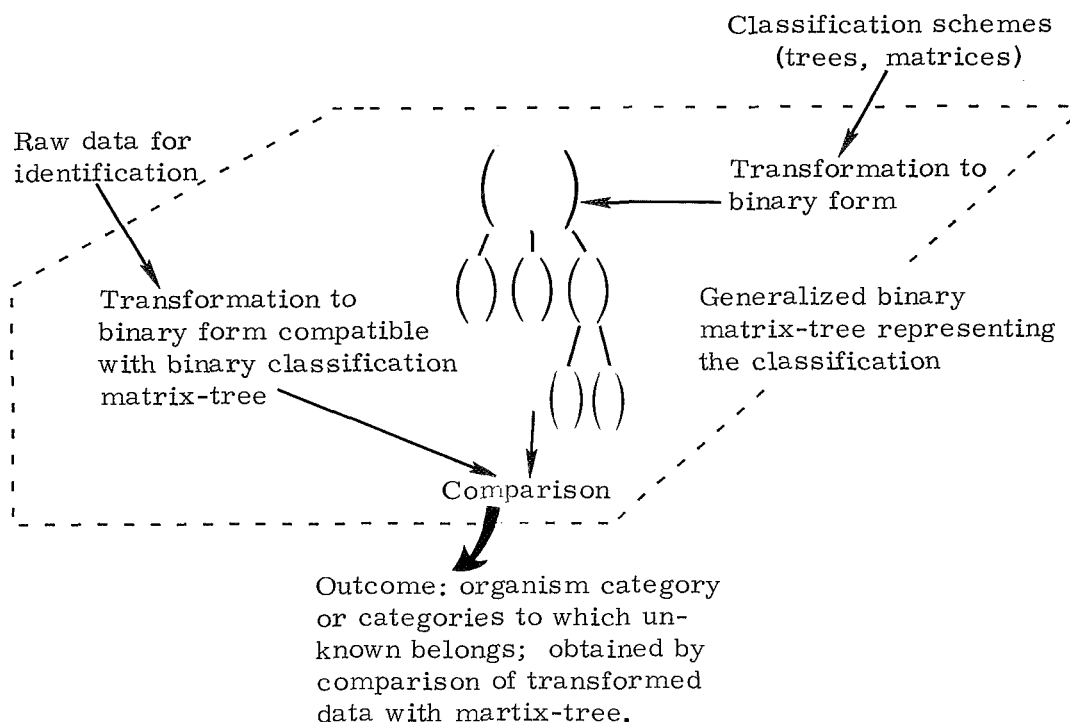
From such considerations, and the consideration of the PHS Apollo scheme, a very general approach to classification has been devised which possesses the advantages of both the matrix and tree approaches and minimizes their practical disadvantages. In simple terms this is accomplished by generalizing the notion of a tree in which nodes represent tests, to a tree in which nodes represent matrices. The "categories" of the matrices at the upper levels of the tree are supersets of the original categories (i. e., if a, b are two original categories, one possible outcome for a matrix at the upper levels of the tree is $A = \{a, b\}$, meaning A is a or b). To see how this concept might work, we will consider the above examples of a tree and matrix. This may be redrawn as follows:



The practical implications of this figure are that tests 1, 2 and 3 will be run simultaneously (or as nearly so as possible) at the outset. The unknown will then belong to supercategory A, B, C or D. From this knowledge, the appropriate test or tests needed to determine the category a, b, c, d, e, f, g, h, i or j will then be run. This represents a compromise between a strict sequential running of tests and running all tests concurrently. This is particularly advantageous if, for example, tests 1, 2 and 3 are particularly easy to run.

There is another advantage to this general approach when computerization is contemplated. All "decisions" are made using matrices. This is advantageous for two reasons. First, this represents a particularly simple computer operation; and second, all operations are of exactly the same type and hence may be regarded as essentially the same computer operation, leading to considerable computer program efficiency (as will be seen later).

One other point in abstracting classification and identification schemes needs to be addressed. In the above examples, the test results were all binary in character (+, -). As stated earlier, this is not always the case. At the same time there are several reasons why it is best that the system be binary for computerization purposes. First, if multiple outcomes occur in a matrix, then the general utility of the program is somewhat lessened since the program and its inputs become considerably more complex. Additionally, decisions about identification become complicated (as will be seen later) due to the increased possibility of "multiple" identifications. Since, as a general precept, we may, through analysis, always regard multiple outcomes in one scheme as binary, in another (by redesigning the former) it has been elected to consider classification and identification as preferably a binary process. This necessitates the introduction of the concept of "transformation" whereby both classification schemes and test data may be transformed from multiple test outcomes to essentially binary outcomes. This is not precisely so since "variable" outcomes are allowed as a third possibility. To explain this, "transformations" of data are distinguished from "transformations" of classification schemes. Abstractly, a description of these transformations is complex. Because of this, only their existence and desirability are discussed here. Later, specific examples will be given in profusion. Thus, the total abstract approach may be summarized in the following diagram:



The part within the dashed lines is performed in the computer.

In application, it was relatively simple to convert the trees in use by the Public Health Service into matrices. Next, these matrices are particularly easy to deal with on modern computers. This choice provided a uniform format which made possible an exceedingly simple computer program. Furthermore, selecting this option made it easy to distinguish between the variability of the tests used and the judgment or decisions of a microbiologist in recording the data he had obtained in the PHS program. The utilization of matrices as the standard identification system permitted the establishment of normal (or standardized) entries in the matrices through transformations that will be described later. This general approach permitted the particular identification scheme (tree, key, etc.) or transformation to be included, not as part of the program, but as input to the program which the microbiologist or other user could change at will, since any use of computers to assist microbiologists in the interpretation of data to make an identification should allow easy modification of both test interpretation and the actual identification scheme itself. To achieve the desired flexibility the computer program should be quite independent of the particular identification scheme in use. It was possible to accomplish this with the PHS problem by analyzing the entire process and separating it into a number of separate operations.

These operations then might be categorized as (1) those associated with the "standard" general identification process (or form of identification), (2) those operations particular to the identification scheme in use, and finally (3) those operations which reflect the decisions or judgments of a microbiologist. To show how the preceding abstract approach was conceived, and as a base for describing the precise computerization of the PHS problem, a description of the latter as it was first seen, is presented below.

It was noted earlier that a microbial identification scheme need only be adequate for some particular purpose. In keeping with this, categories of interest for the Apollo program were established. To classify or identify samples taken from a spacecraft as belonging to some of these categories, appropriate sets of tests were used. Hence, a collection of test sets both necessary and sufficient to classify a sample in the appropriate category was developed. Hopefully such a test set is also nearly minimal in the sense of requiring the least number of tests for this purpose. The identification system used for the Apollo program was developed by the U.S. PHS (9). This system will be described in some detail as a basis for the approach to incorporating it in a computer. At first it would appear that their identification system consists of two parts. The first, or general scheme, takes the form of a flow diagram or tree which uses the results of five tests to enter the appropriate place in the second part. This flow diagram is reproduced as Figure 2.

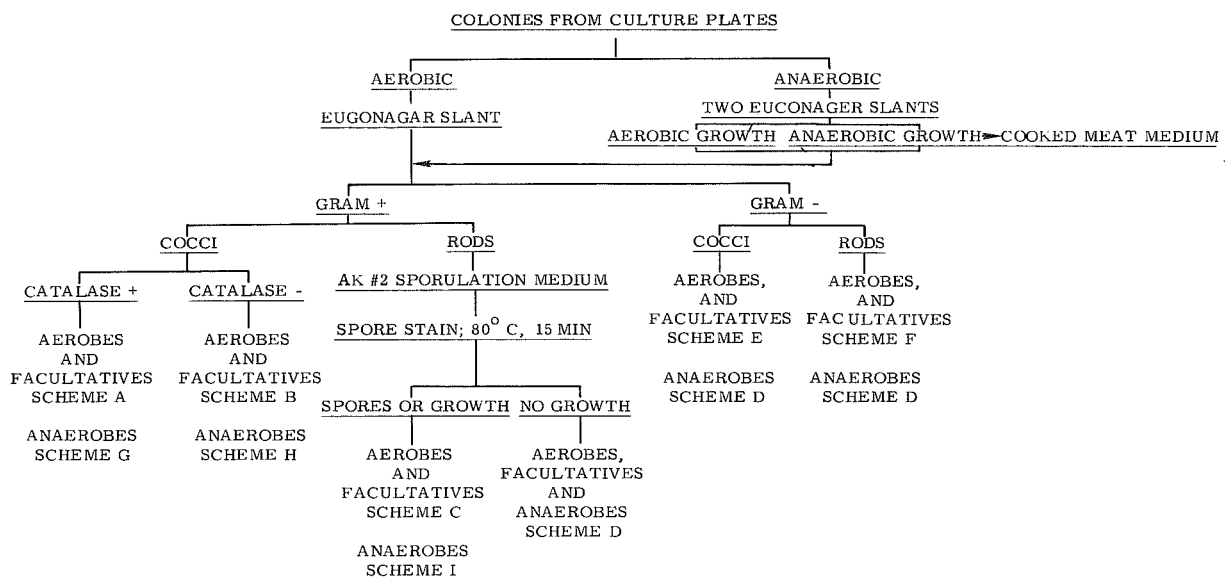


Figure 2. Flow Diagram of General Scheme for PHS Identification

The second part of the system consists of a number of individual schemes (in Figure 2 - A through I) which are reached after stepping through the first part of the identification system. In general, the schemes of this second part allow an identification to be made (i. e., assignment to a category) on the basis of certain specified tests. This relationship of test to microorganism category is usually presented in matrix form. For example, the key for Scheme A of Figure 2 is given as the following matrix.

Subgroup:	Group I Staphylococcus Rosenbach						Group II Micrococcus Cohn							
	I	II	III	IV	V	VI	1	2	3	4	5	6	7	8
Pink Pigment	-	-	-	-	-	-	-	-	-	-	-	-	-	+
Acid from glucose:														
(1) aerobic	+	+	+	+	+	+	+	+	+	+	+	+	±	±
(2) anaerobic	+	+	+	+	+	+	-	-	-	-	-	-	-	-
Coagulase	+	-	-	-	-	-	-	-	-	-	-	-	-	-
Phosphatase	+	+	+	-	-	-	-	-	-	-	-	+	-	-
Acetyl-methyl-carbinol	+	+	-	+	+	+	+	+	+	+	-	-	-	-
Acid from:														
(1) arabinose	-	-	-	-	-	-	-	-	-	+	v	+	-	-
(2) lactose	+	+	v	-	+	v	-	+	v	+	+	+	-	-
(3) maltose	+	+	-	v	+	v	v	+	+	+	+	+	-	±
(4) Mannitol	+	-	-	-	-	+	-	-	+	+	+	+	-	-

± = weak or negative, v = variable

Note: Staphylococcus subgroup I corresponds to S. aureus and subgroups II-VI to S. epidermidis; Micrococcus subgroup 7 corresponds to M. luteus and subgroup 8 to M. roseus.

Figure 3. Key for Scheme A (Baird-Parker)

The key for scheme A may be regarded as typical although there are keys which are more complicated, i. e. consisting of two matrices (one indicating entry to another) as well as keys which admit only a single identification or classification. Scheme A is taken from Baird-Parker (10) and may be regarded as typical of the key approach to identification. This particular key will be discussed in more detail later.

In practice, then, the system consists of a list of microbial categories and a set of tests with rules or procedures relating them. For example, five (or fewer) test results, in conjunction with Figure 2, indicate a scheme to be used. The key for this scheme specifies a number of tests to be performed whose results will permit classification of the sample into a category. A computer is to be programmed so that, given the data, it can simulate the process of identification; that is, it can duplicate the steps the microbiologist takes after the data is collected.

These steps include first interpreting each test result associated with an unknown as positive, negative, or inconclusive. Then the interpreted results are compared with those specified by the key (or matrix) for the scheme being used in order to determine if a suitably close match can be obtained for an identification. The system, and hence the computer, must simulate these steps. In addition, it must allow for differing degrees of variability in the interpretation of the data. The operation might best be seen, in a general way, in terms of the matrix for Scheme A above.

In the case of Scheme A the operation is that the results of a number of tests are compared with the entries in a matrix to obtain a match. This, for example, is done with a matrix constructed to represent the initial tree in order to determine the appropriate final scheme. Then appropriate data is compared with the entries of the matrix for that scheme to determine the sample identification. The fact that the same operations are repeatedly performed with various matrices allows for an extremely simple program. This suggests that the matrices and appropriate indices could be provided as input in the program rather than being part of the program itself. This allows the particular identification scheme being used to be easily changed at any time. There is, however, still the problem of variable or indeterminate test results referred to above.

The way this difficulty was addressed was to incorporate a set of transformations for each of the matrices which would translate the data into a standard form. These transformations are based upon previous experience and are easily modified as required.

The indeterminate nature of the test is taken care of at two levels, thus allowing more flexibility. That indeterminacy which is largely inherent in the scheme is incorporated in the corresponding matrix or matrices, while the indeterminacy due to interpretation of the test is considered in the data transformation. Hopefully this

separation permits the computer to more closely simulate microbiological practice. The precise manner in which this is accomplished will be described in the detailed system description.

In summary, the objective was to write a program (as general as possible) which would be able to make an identification on the basis of recorded laboratory data. This was accomplished by analyzing the PHS identification system and abstracting from this the operations that are performed in making an identification, the schemes used as a basis for the data and identification, and finally the data interpretations that are made by the microbiologist (subject to the system that he is using). This was done in as simple a manner as could be accomplished so that the program could easily be modified to accommodate modifications in the process used by the PHS or even in entirely new identification schemes. There are only a few subroutines which need to be changed for a completely new identification system. The program will be described in detail in the next section.

Identification by Computer

As indicated in the preceding section, bacterial identification can be simple. One takes the results of a number of tests and compares these with those attributed to various categories of bacteria. When the observed results agree with the known results, an identification is made. Practically, there are a number of difficulties as noted before. For computerized identification it is important to approach the above ideal as closely as possible. This was one of the major motivations for the particular approach taken, namely transforming the data so that they could be compared with matrices representing individual schemes in a uniform manner. Through this device, which has other advantages, a relatively simple program can be written since similar operations are iteratively performed. That is, different transformed test results are compared with a matrix representing a different scheme, but the same operations are repeatedly performed.

This is accomplished by representing each scheme by a matrix having as entries only +1, -1, and 99 which represent respectively a positive, negative, or inconclusive test result. Thus, to make a comparison, transformations must be

specified which will convert reported test results into this form. In setting up these transformations it is possible to incorporate some of the judgment that a microbiologist used in making an identification at the bench. In conjunction with the interpretation of test results as used in the matrices, considerable flexibility is allowed. To make the above concepts specific the PHS identification system will be used in some detail. In the preceding section it was noted that the identification procedure begins with a flow diagram which uses the results of five tests to determine which of nine schemes to enter. These latter, in general, were given in a matrix form that related test results to organism categories.

The first part of the Public Health Service system, namely the flow diagram (Figure 2), does not at first appear to fit the above description of a binomial tree. However, during the feasibility analysis, as has been previously mentioned, it was demonstrated that the flow diagram could be regarded as a binomial tree which used the outcomes of five tests to determine the appropriate scheme to enter. This tree is reproduced as Figure 4.

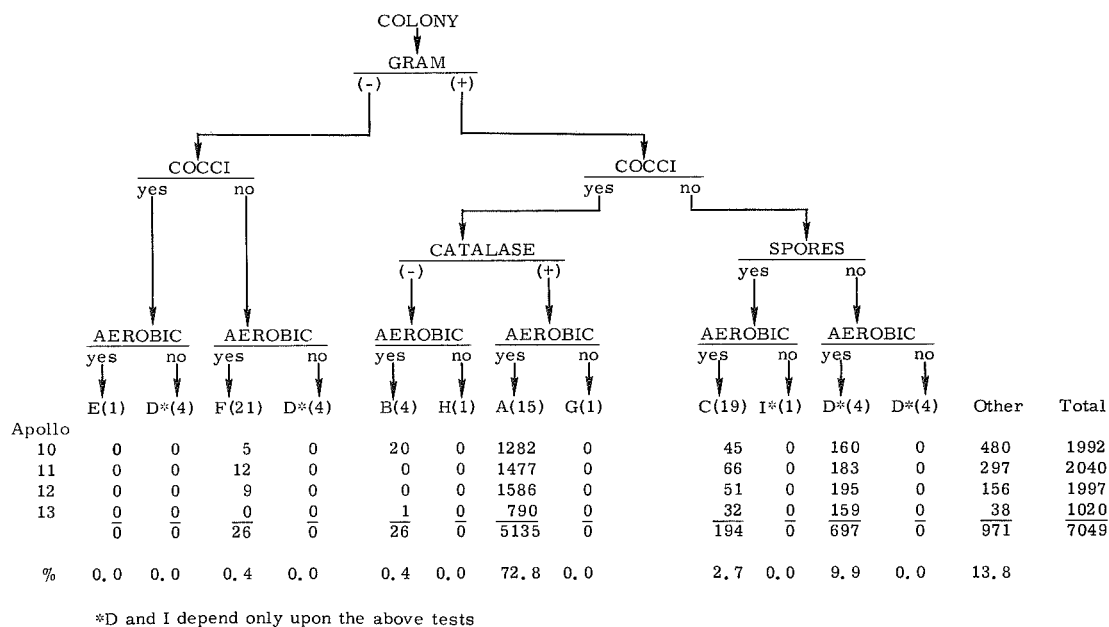


Figure 4. Flow Diagram of General Scheme in Yes/No Form

In Figure 4, schemes A through I are indicated by the appropriate letter with the number in parentheses designating the number of categories that the scheme leads to. In general, schemes themselves consist of matrices although there are exceptions. For example, it was sufficient for the Public Health Service Apollo purpose to record in Scheme D only the results of the four determining tests, e. g. anaerobic, non spore-forming, gram positive rods. Thus scheme D occurs in four places in the above tree, each containing only one identification category (itself) which depends solely upon the five previous tests. Thus, to proceed through the flow diagram (hereafter referred to as the tree), it is only necessary to know the results of five tests which have a yes/no nature. The fact that these outcomes are yes/no in nature is somewhat arbitrary and based upon the Public Health Service requirements for this particular identification process. For example, morphology is reported in a number of categories (see Appendix 1) but decisions are made based upon whether the sample was coccoidial or not. Similarly, spore forming ability is somewhat open ended in that it depends on the ability to detect a spore in a number of ways.

For a computer to be able to deal with the results of biological tests it has to be coded some way. In the Public Health system, the test results are recorded in a column on a card. Thus there is a natural numbering associated with the test, namely, the number of column of the card in which the result was recorded. The tests of primary interest for the tree are as follows:

Table 1. Primary Yes/No Test Results for General Scheme

	<u>Test</u>	<u>Col. No.</u>
1.	Gram Stain	18
2.	Cocci	17
3.	Catalase Reaction	41
4.	Spore Forming Ability	17
5.	Aerobic Growth	15

With the more or less natural conventions indicated above where +1, -1, and 99 were allowed as matrix entries, we have clear interpretations of the tests in Table 1. That is, catalase reaction positive and gram stain positive will be regarded as +1, as are the other three properties listed in Table 1, with negative reactions being -1. An inspection of Figure 4 reveals that not all of the five tests are always required. For example, a gram negative result indicates that the catalase reaction and the spore forming ability does not influence the outcome. Continuing with the conventions

indicated in the beginning of this section, we can use a 99 as a matrix entry where this situation prevails. With these interpretations the tree in Figure 4 can be written as a matrix as follows:

		Scheme (Supercategory)											
		A	B	C	D1	D2	D3	D4	E	F	G	H	I
Card Column (Test)	18	1	2	3	4	5	6	7	8	9	10	11	12
	17	1	1	1	-1	-1	1	1	-1	-1	1	1	1
	41	1	1	-1	1	-1	-1	-1	1	-1	1	1	-1
	17	1	-1	99	99	99	99	99	99	99	1	-1	99
	15	99	99	1	99	99	-1	-1	99	99	99	99	1
	15	1	1	1	-1	-1	1	-1	1	1	-1	-1	-1

Figure 5. Matrix Form of General Identification Scheme

In Figure 5 the rows represent the numbered test possibilities, while the columns represent the scheme which is to be used for the next stage in the identification. Thus, what originally appeared as the flow diagram and is now referred to as the tree can be represented as a matrix just as all of the other schemes involved in the Public Health Service identification system. This similarity allows each of the stages in the Public Health Service system to be handled by a computer in a uniform manner, as will be seen. Not only the tests but the schemes need to be indexed by some number readable by a computer. This can be arbitrary or could follow some preconceived pattern. In the case of the Public Health Service system there is a matrix which represents the tree resulting from the flow diagram itself. This is treated somewhat separately inasmuch as it is the first entry into any identification process. The remainder of the schemes are then numbered in their natural order, thus allowing for repetition; that is, A = 1, B = 2, C = 3, and D = 4 (note that D appears four times because it can appear four times from the matrix representing the flow diagram), E = 5, etc. There is the possibility that test results for an unknown are either incomplete or inconsistent with the entries in the matrix in Figure 4. Hence we have the possibility of no identification. No identification can occur in the case that there is no outcome consistent with the test results from the tree or in the case that there is an outcome where there is difficulty matching the matrix. This latter point will be elaborated in more detail later.

The process of converting the schemes themselves to a matrix for the computer identification is relatively simple compared to that of converting the flow diagram to a matrix. These schemes are more typical of those used by microbiologists in general; where a result is positive, +1 is entered, -1 is entered for a negative result, and 99 is used for a weak or variable result. Thus Scheme A (Baird-Parker) of Figure 3 appears as follows:

Identification Matrix A

	1	2	3	4	5	6	7	8	9	10	11	12	13	14
14	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	1
37	1	1	1	1	1	1	1	1	1	1	1	1	99	99
37	1	1	1	1	1	1	-1	-1	-1	-1	-1	-1	-1	-1
42	1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1
43	1	1	1	-1	-1	-1	-1	-1	-1	-1	-1	1	-1	-1
53	1	1	-1	1	1	1	1	1	1	1	-1	-1	-1	-1
36	-1	-1	-1	-1	-1	-1	-1	-1	-1	1	99	1	-1	-1
38	1	1	99	-1	1	99	-1	1	99	1	1	1	-1	-1
39	1	1	-1	99	1	99	99	1	1	1	1	1	-1	99
40	1	-1	-1	-1	-1	1	-1	-1	1	1	1	1	-1	-1

Figure 6. Computer Version of Scheme A Matrix

In Figure 6 the column headings indicate the organism category as used by the Public Health Service or Baird-Parker, while the left hand column indicates the entry on the card where the test result will be recorded by the Public Health Service. Similarly this is used for the other schemes. There are a few instances where this conversion procedure is not as straightforward as indicated above. Examples of these occurrences will be provided and discussed later.

With all of the identification procedure represented uniformly as matrices of the above type, it is, in principle, quite simple to make an identification. The appropriate data for an unknown are selected and compared with the columns of the matrix representing the tree to determine the next step in the process. There are, however, a few difficulties remaining. One such difficulty is that the data being compared with the columns in the above matrices must be of the same form as those entries, e.g. +, - or 99 representing indeterminate. The data required are in general either not recorded or given some value in the range 0, 1, ... 9. Some

means must be provided for the computer to determine, for example whether an entry of 3 in the Baird-Parker mannitol test should be counted as +, - or 99. To accomplish this, a series of transformations were provided which would convert such entries to the appropriate value for comparison in the above matrices. It should be noted that this conversion is of a different nature than the previous conversion of test results to entries in the matrices. Previously, an established outcome of a test was being interpreted as a +, - or an indeterminate test for classification (or matrix generation) purposes, whereas now observed data are being translated for comparison with the classification. In this latter case, an attempt is made to simulate the judgment that a microbiologist uses in taking, observing and interpreting the data.

Because of this fundamental difference the transformations were treated in as general and flexible a manner as possible. As part of the input to the program, each test that will be used in the identification process is listed on a card. On the same card each possible outcome that could be recorded by the Public Health Service microbiologists is listed, followed by its interpretation as +1, -1, or 99 as it is to be used in comparison with the matrices. Thus, it is an extremely simple matter to change an interpretation to improve the performance of the identification program. Since occasionally the same test is used in different portions of the identification process and interpreted in a different manner, the transformations have to be associated with the appropriate stage in the identification process.

An example of the above occurs in the tree where column 17 on the PHS card (Cellular Morphology) is used for a number of purposes. In the tree we need the results of 17 to determine cocci or not, that is the second row in the tree matrix; also it is possible that the PHS has checked spores, No. 1 in column 17. Thus, column 17 is used twice as an entry into the tree matrix. It is a bit more complicated than that because there are other tests, namely 65, 66 and 67 which are also used to determine the ability to sporulate. Thus, if, during the morphological examination, a spore is detected, 1 is checked in column 17 and the issue insofar as sporulation is concerned is settled. If something other than 1 is checked, tests 65, 66, and 67 are run. 67 involves a sporulation media. If there is growth on this media, No. 2 is checked and it is concluded that the sample is capable of sporulation. Similarly, in 66 (heatshock) the checking of a 2 indicates sporulation, and in 67 the checking of a 2 also indicates sporulation. Thus 17 really provides the entry for a

series of conditional inquiries to determine whether sporulation is possible. If so, a +1 is recorded as a result of the transformation. The figure below indicates this sequence as it is incorporated into the program.

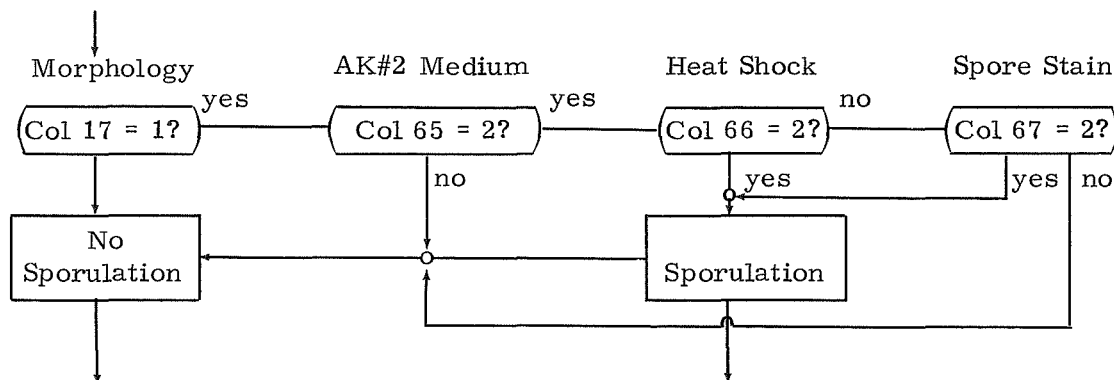


Figure 7. PHS Sequence for Detecting Sporulation

It should be noted that although the logic, illustrated by the above diagram, is incorporated into the program this could have been taken care of a number of other ways. For example, there could have been a separate column for sporulation. The same series of tests could have been run by the Public Health Service and recorded as + or -, in which case this particular bit of programming to fit the actual sequence of operation might have been avoided. However, there is no real problem and it can be regarded as part of the transformation of the data into a number which indicates whether sporulation was observed or not. It is, nevertheless, a place where the program departs from complete generality in simply comparing transformed data with the appropriate matrices, since the logic illustrated above is incorporated as part of the program.

There is one other area in which the program departs from the generality that has been indicated thus far. This also involves the entry in column 17. Column 17 is used to record the occurrence of yeasts, molds, actinomycetes, and streptomycetes which are observed during the morphological examination. Thus, in the event that one of the items 3 through 6 appears in column 17, an immediate identification of the appropriate category is made rather than proceeding through the matching of matrices with data which would not have been taken inasmuch as the sample had already been identified.

Having set up the above procedure to simulate the identification process as carried out by the Public Health Service, it would seem a simple matter to identify a sample. One simply takes the data, transforms them appropriately, makes the comparison until a match is found in the matrix representing the flow diagram, and then proceeds to the appropriate scheme where he selects a new set of data, transforms that, and obtains a match with the right column in that scheme to make an identification. Unfortunately, biological systems are highly variable and it is optimistic to expect exact agreement with the average of a collection of preceding experience for a given sample. As a result, rules have to be developed which would allow for this variation which is usually expressed as a number of disagreements in the above comparisons. The rules adopted were those that have been developed by the Public Health Service Spacecraft Bioassay Laboratory at Cape Kennedy in performing the Apollo sampling and identification. Before the rules are examined, it should be noted that a disagreement between the transformed data and a scheme may happen in a number of ways. First, it may happen that the transformed data disagree with any column in the matrices in one place or test. This can also happen for two, three or more distinct columns, indicating that a number of microorganism categories are equally close (only one disagreeing test) to the "category" represented by the test data. Similarly it may turn out that the test data disagree in two test places with one or more microorganism categories. The rules finally adopted by the Public Health Service were that an identification was made when there was an exact match, or there were not more than three single disagreements (with no exact match) — that is, there was a choice between three microorganism categories which disagreed at one place with the test results, or there was a single microorganism category which disagreed in two places and none that disagreed in fewer places. These possibilities are summarized in Table 2.

No. of Test Disagreements	No. of Categories	Computer Statement of RESULT
0		EXACT IDENT
1	1	ONE DISAGREEMENT
1	2	2 SINGLE DISAGREEMENTS
1	3	3 SINGLE DISAGREEMENTS
2	1	TWO DISAGREEMENTS
Other		NO ID POSSIBLE

Table 2. Computer Statement for Possibilities of Classification

This, of course, allowed for the possibility of multiple identifications, that is, as many as three equally likely microorganism categories. Thus there is the final problem of choice between these categories.

In the event that more than one equally likely microorganism category is arrived at by the computer, there are a number of options for choosing between these multiple identifications. One such option would be to use the statistics accumulated during the identification of some 7,049 samples by the Public Health Service and to select among the indicated possibilities in the same manner that the Public Health Service has previously used. In certain instances it develops that there is a natural choice between the alternatives based upon an analysis of the tests and the associated scheme. There are, of course, other methods of making a decision between the selected categories. However, the two mentioned above are those used in this particular system. At present the program simply prints out all possibilities in a manner that will be illustrated in the next section. The following section will discuss in detail the above two proposals for making a decision.

In this section, then, the PHS scheme was analyzed for incorporation into the abstract approach previously discussed. In addition, the rules and conventions adopted by the PHS and to be used in computerized identifications were defined and discussed.

The Program Itself - BUGID

In the preceding two sections, an abstract approach to the problem of simulating the identification process has been outlined and means described for applying it to the problem of identifying organisms taken from Apollo missions. It was seen that the Public Health Service Identification system lends itself very well to the abstract approach. It is largely composed of keys which can be represented by matrices of a standard form. Furthermore, in the process of using their system, the Public Health Service has developed conventions which are easily implemented in a program. Most of the conventions that are to be used have been discussed. The system that has been programmed and checked out with the Apollo 13 data is called BUGID for Bug Identification.

Initially, it was hoped that a program could be written which was entirely independent of the particular identification scheme to which it was applied. In practice this turned out to be not quite possible although it was very nearly achieved. There are a few instances (as will be seen) where BUGID is dependent upon the Public Health Service system. These instances involve, first, the possibility of an immediate identification, e.g. yeasts, molds, actinomycetes, and streptomycetes, where the program looks for the right digit in the appropriate column before proceeding. The other major instance where scheme instructions are not read into the program occurs in a routine which simulates the Public Health Service search for sporulation.

The heart of BUGID consists of two sub-routines called SETUP and MATCH. SETUP is the routine that selects the appropriate data entries and transforms them for comparison with the identification matrix being used. MATCH (which normally immediately follows SETUP) is the sub-routine which takes the vector consisting of the data transformed by SETUP (IVEC) and compares it with the matrix in the hopes of obtaining a suitably close match. Here, suitably close match means one that satisfies the conditions of Table 2.

The flow diagram of BUGID is presented as Figure 8. Figure 9 has the flow diagram of the sub-routine SETUP, while Figure 10 contains that of MATCH.

In Figure 8 the two exceptional parts of the main program BUGID are indicated. Before the program is run through, a check for immediate identification is made as indicated. The numbers in this portion of the program flow chart are actually part of the program and would have to be modified if any change in the procedures were made. Although this modification is trivial, it is a place where parameters of the identification scheme are not supplied as generalized inputs. Similarly, the section of the program indicated by sporulation is peculiar to the program, and the program would have to be modified internally if there were any change in these procedures. Neglecting these exceptional portions, one can see from Figure 8 that the program consists of repeatedly calling SETUP and then MATCH. As can be imagined, SETUP simply selects the appropriate data and applies the transformation to the data that the operator supplied to the program as input. MATCH simply compares the transformed vector with the columns of the correct matrix in order to determine if there is a sufficient agreement in order to make an identification. Again, this matrix is part of the input to the program. The manner in which the transformations and matrices are

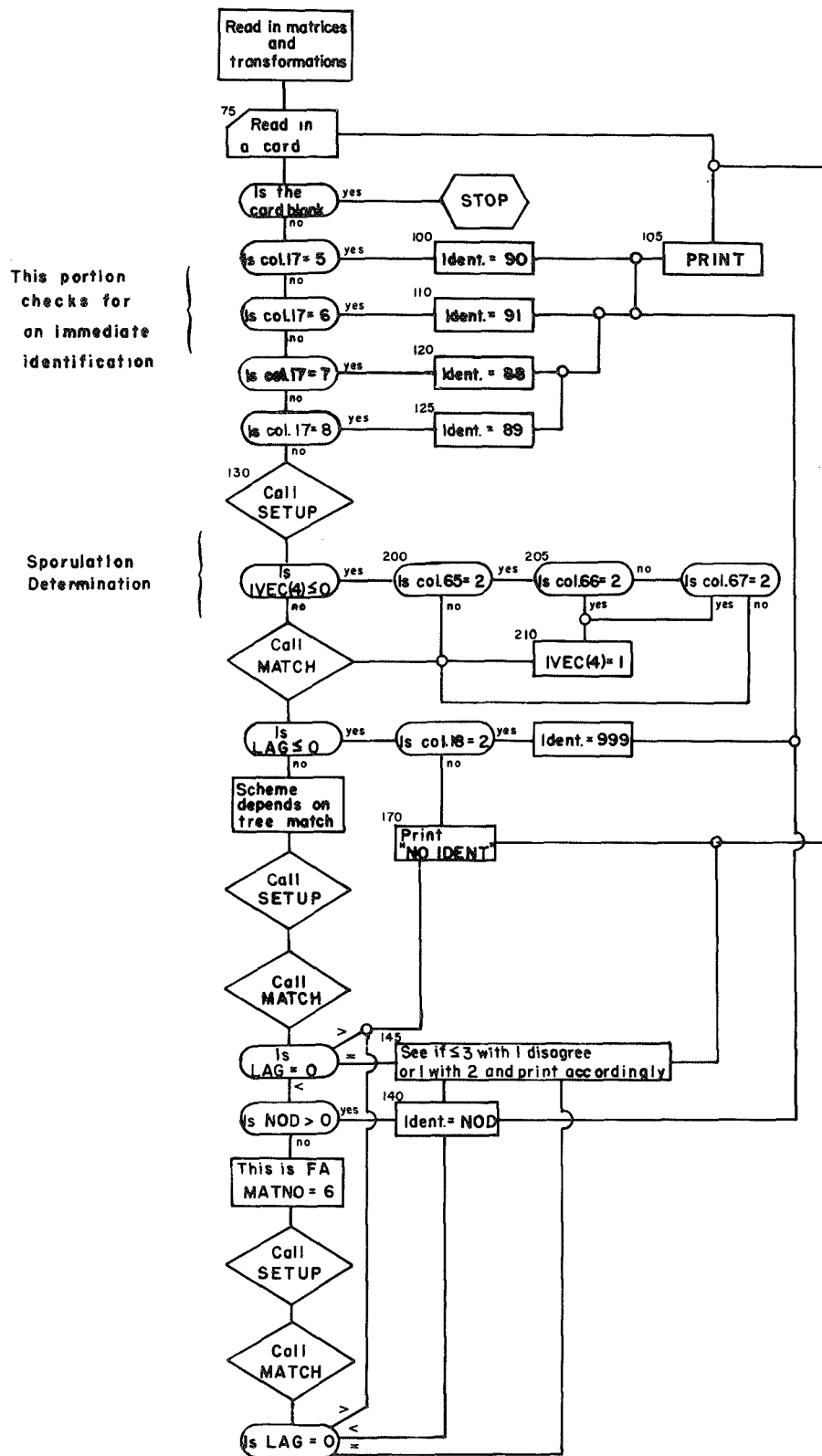


Figure 8. Flow Diagram - BUGID, Main Program

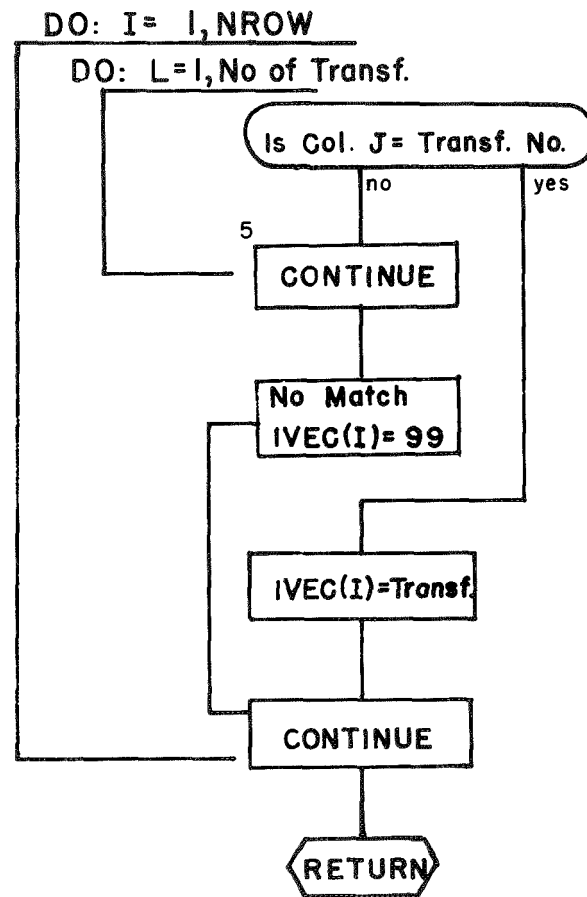


Figure 9. Flow Diagram - Sub-routine SETUP

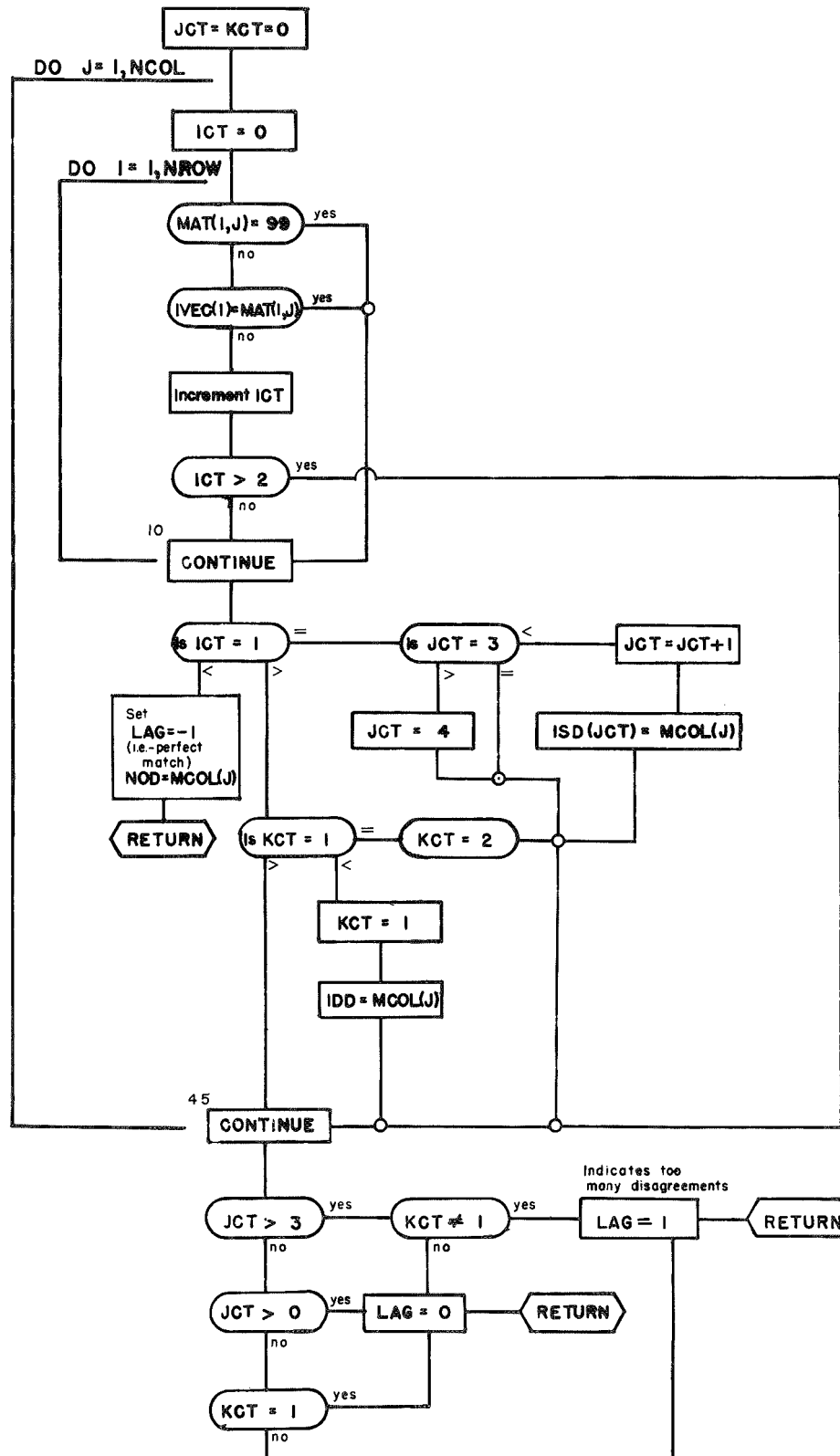


Figure 10. Flow Diagram - Sub-routine MATCH

For each matrix or associated scheme, the first line contains two digits providing the number of rows and the number of columns as labeled on the right hand side. The number of the column in the data card from which the data are required by the corresponding row of the matrix is listed in order on the next line. For example, the fourth row for the matrix of Scheme A requires the data from column 42 as punched by the PHS. The next row labels the columns of the matrix. These numbers are, in general, the identification number of the microorganism category used by the Public Health Service. For example, column 9 is coded 009 Micrococcus Subgroup 3. The transformations to which the raw data are subjected are listed immediately below the matrix. One transformation is given for each entry for the corresponding row of the matrix. Thus, row 10 of the matrix for Scheme A uses the data from column 40 on the card, which will be the result of the Baird-Parker mannitol test. Transformation 10 (found in the right hand column) indicates by the first number in the row that there are five possible outcomes that can be recorded. The subsequent integers taken pair-wise indicate that 3 is a positive result and should be regarded as +1. Similarly, 4 is positive, while the recorded result of 1, 2 or 5 would be negative and transformed to a -1. It should be noted that the number of transformations must equal the number of rows of the associated matrix.

SETUP is a sub-routine which performs the transformation on the data required for identification in a particular matrix. It reads the appropriate columns on the data card, applies the indicated transformations to this data, and obtains a resulting IVEC for the sample. This IVEC will consist of a series of numbers, one for each row of the matrix. The numbers will either be +1, -1, or 99. From an inspection of Figure 9 it can be seen that if an entry in a specific column is not an admissible one, that is, not one listed in the transformation for that column, a 99 will be inserted in IVEC. Thus a missing or incorrect test will be recorded as 99. As previously mentioned, a 99 in IVEC is treated differently than a 99 in the matrix where it represents an indeterminate or variable test rather than incomplete or erroneous information as it does in IVEC. This point will be illustrated by example later.

The other major sub-routine of the identification program, MATCH, is diagrammed in Figure 10. Essentially the function of MATCH is to compare IVEC as prepared by SETUP with the columns of the associated matrix. Hopefully a match

with one of the columns is obtained or at least sufficiently few disagreements (subject to the conventions of Table 2) are obtained so that an identification can be made.

As MATCH is called, three flags, ICT, JCT, and KCT, are set equal to zero. ICT is used during the comparison of IVEC with the column of the matrix to see whether an identification with the column is possible. JCT is used to keep track of the score of IVEC against each column with respect to single disagreements. KCT is similarly used to score the comparison for double disagreements. No index is required to keep track of exact agreement with a column since, in a well designed key, this can only occur for a unique column. Having set the flags, the first part of MATCH compares IVEC with the columns of the matrix MAT then being considered by running down the rows of MAT. As can be seen, if the entry in MAT under consideration is 99, this counts as an agreement. Then it simply continues to the next comparison. If, on the other hand, it is not 99, the corresponding entry in IVEC must equal the entry in MAT for agreement. If not, ICT is incremented. After being incremented, ICT is checked to see whether it is worthwhile proceeding with the remainder of the column. Recalling the columns which are candidates for possible identification must have two or fewer disagreements, a candidate is rejected whenever ICT is greater than two. If so, the sub-routine proceeds to the next column as a candidate. If not, it continues until all rows have been examined or the column is ruled out as a potential identification. Previously it was mentioned that 99s in a matrix were treated differently than 99s in the transformed data IVEC. From Figure 10 and the above it can be seen that, during the comparison process, if a 99 occurs in the matrix it does not count as a disagreement. On the other hand, if the matrix entry under consideration is not 99, exact agreement is required. That is, a 99 appearing in IVEC will count as a disagreement unless the corresponding entry in MAT is also 99 in which case it didn't make any difference. Thus the program has the ability to cope with weak or indeterminate tests appearing in a key while, at the same time, discriminating against missing or inadequate laboratory data.

The next portion of MATCH keeps score on the columns of MAT that have been or are being compared with IVEC. This is done by examining the value of the flag ICT as each column comparison is completed. From the above it can be seen that ICT will be equal to zero only if no disagreements occurred with IVEC in the column being compared. If this occurs, an exact identification is made and MATCH is exited.

Since $ICT = 1$ would indicate a single disagreement of which three possibles are allowed by convention, such a column is a candidate unless there are more than three. Thus, for $ICT = 1$, the value of JCT is determined relative to three. If it is not more than three, it is incremented and the sub-routine proceeds to the next column. Finally, there is the possibility of a single column having two disagreements. When a column has two disagreements, ICT is greater than one and less than three, namely two. In this case, KCT is examined to see if it is one. If it is already one, there are now two columns having two disagreements and an identification is not possible. On the other hand, if KCT is still zero, it is incremented to one and the sub-routine proceeds through the remainder of the columns.

In the event that an identification has not been made but the sub-routine has compared the transformed IVEC with each column of the matrix MAT, there is a final decision process of whether an identification can be made or not. The manner in which this is done is indicated in the bottom of Figure 10. Essentially, flags JCT and KCT are examined. In the event that JCT is greater than 3 (more than three single disagreements) and JCT is not equal to 1 (no single double disagreement), the flag is set equal to 1, which indicates too many disagreements with no identification possible. The other possibilities are similarly examined, and the sub-routine returns to the main program with the appropriate identification or identifications of the IVEC representing data from a sample.

The printed output of the program consists of the identification matrices that were read in and a tabulation of the identifications that have been made. A sample of this tabulated output is provided in Figure 11.

As can be seen from Figure 11, the basic identification line contains the sample identification, the date and the serial number from the data card, and the organism category (ORGANISM) that the program has assigned the sample to. Finally, the identification that was made by the Public Health Service and recorded on the card is listed for comparison purposes. Above this line are one or two lines which list the vector that was used in the matrix or matrices for the identification; that is, IVEC, as defined above, is printed. Thus, we see for Sample #4, which is listed as D0004, that the IVEC that resulted from the transformed data which was used for comparison in the tree matrix consisted of [1, 1, 1, -1, 1] as shown in entry D0004.

ID	DATE	SN	ORGANISM	PHS
VECTOR	1 -1	1 -1 1		
X0001	032770	2	37	-0
VECTOR	1 1	1 -1 1		
VECTOR	-1 1	1 -1 -1 -1 -1 -1 -1 1		
X0002	032770	2	6	-0
VECTOR	1 1	1 -1 -1		
VECTOR	-1 1	-1 -1 -1 -1 -1 1 1 -1		
X0003	032770	1	8 11	-0
VECTOR	1 1	1 -1 1		
VECTOR	-1 1	-1 -1 -1 -1 -1 -1 1 -1		
X0004	032770	1	7 13 14	-0
VECTOR	1 1	1 -1 1		
VECTOR	1 -1	-1 -1 1 1 -1 -1 1 -1		
X0005	032770	1	14	-0
VECTOR	1 1	-1 -1 1		
VECTOR	99 99	99 99 99 99 99 99 99		
X0006	032770	1	NO ID, B	-0
VECTOR	1 -1	1 -1 1		
D0002	032770	1	37	37
VECTOR	1 -1	1 -1 1		
D0003	032770	1	37	37
VECTOR	1 1	1 -1 1		
VECTOR	-1 1	1 -1 1 1 -1 1 1 -1		
D0004	032770	1	2	2
VECTOR	1 -1	-1 -1 1		
D0005	032770	1	37	37

Figure 11. BUGID Sample Output

This leads to Scheme A, and the IVEC used for comparison with the matrix for Scheme A reads [-1, 1, 1, -1, 1, 1, -1, 1, 1, -1]•. Finally, the identification of organism Category 2 was made, which agrees with that of the Public Health Service. As mentioned above, it was desirable to retain information about the level at which an identification was made (that is, the number of disagreements). This was done by printing in the first column under ORGANISM the identifications that were made by an exact identification. Three further columns were reserved for those identifications made with one disagreement; this identification is one of three which were possible. Finally, there is a column between these last three columns and the column in which the Public Health Service identification is printed that is reserved for an identification by means of one double disagreement. In the case that no identification was possible, a NO ID is printed in the ORGANISM column. It is convenient for checkout purposes to analyze why no identification was possible. As a result,

the computer also prints the letter designating the last matrix that it was using when it concluded that there was no possibility of an identification. These possibilities can be seen by an examination of Figure 11.

The above has explained the general operation of the program with a detailed description of the flow through the basic program. Similarly, the major sub-routines, SETUP and MATCH, have been described. A complete list of the program with the matrices and transformations presently used as input will be found in Appendix 2. To further illustrate the operation of the program, a sample data card will be used so that the steps taken by the program to assign an identification can be followed in detail.

For this illustration, Sample No. 4 of the Apollo 13 data will be used. A listing of this data card is given below as Figure 12 with a heading which makes it convenient to read the columns.

10	20	30	40	50	60	70	80
	A TTT		AAAAATAA	A		(TTT)	
123456789012345678901234567890123456789012345678901234567890							
D00040327701182122			14331212	2			0021

Figure 12. Listing of Sample No. 4, Serial D0004

Referring to Figure 8, it is seen that the first operations after reading in the program are to see if the card is blank, and then, if it is not, to see if column 17 has a 5, 6, 7 or 8. If column 17 has such an entry, an immediate identification is possible. As can be seen, the entry in column 17 for Sample No. 4 is 2, so the program then calls SETUP in order to construct IVEC for comparison with the tree matrix. Here, SETUP will use the transformation associated with the tree matrix. This matrix and the transformations are reproduced as Figure 13, and the columns on the data card used are designated "T" above.

5	12																NR, NC	TREE
18	17	41	17	16													ROWS	TREE
1	2	3	4	5	6	7	8	9	10	11	12						COLS	TREE
1	1	1	-1	-1	1	1	-1	-1	1	1	1						ROW 1	TREE
1	1	-1	-1	-1	-1	-1	-1	-1	1	1	-1						ROW 2	TREE
1	-1	99	99	99	99	99	99	99	99	1	-1	99					ROW 3	TREE
99	99	1	99	99	-1	-1	99	99	99	99	1						ROW 4	TREE
1	1	1	-1	-1	1	-1	1	1	-1	-1	-1						ROW 5	TREE
2	2	1	1	-1													TR1	TREE
4	2	1	1	-1	3	-1	4	-1									TR2	TREE
2	2	1	1	-1													TR3	TREE
4	3	1	1	-1	2	-1	4	-1									TR4	TREE
2	1	1	2	-1													TR5	TREE

Figure 13. Input Identification Data for General Scheme or Tree

As previously described, the second row of Figure 13 lists in order the data card columns (or tests) associated with the rows of the matrix and transformations. As can be seen, in this case, there are five rows of transformations and columns listed, although one column (17) is listed twice. SETUP proceeds to construct IVEC as follows: First, column 18 of the data in Figure 12 is selected. The entry is a 2; the transformation (TR 1 of Figure 13) indicates there are two possible entries in this column, namely 2 and 1, where 2 is to be changed to a 1 and a 1 is regarded as a negative result and, hence, is changed to a -1. Thus SETUP replaces the 2 in column 18 by 1 in IVEC. SETUP then proceeds, similarly, with column 17. The entry in column 17 is a 2 which, in this case, is replaced by a +1. By continuing through all five cases, the results indicated in Table 4 are obtained.

(i)	Col. No.	Data Entry	IVEC (i)
1	18	2	1
2	17	2	1
3	41	2	1
4	17	2	-1*
5	16	1	1

*Note: Although 2 in column 17 was a positive result for TR 2, it was a negative result in TR 4.

Table 4. Result of SETUP Applied with Tree Transformations to Data of Figure 9

After the program has constructed IVEC by means of SETUP for the tree, (see Figure 8) it queries "Is IVEC(4) ≤ 0 ?". In this instance, it is, and hence column 65 is examined to see if it is 2. From Figure 12, where columns 65, 66 and 67 are shown (TTT), one can see that it is not, and hence the program proceeds to call MATCH.

Since MATCH simply searches for a sufficiently close match between the columns of a matrix and IVEC, it is convenient to illustrate the situation by Figure 14.

	A	B	C	D1	D2	D3	D4	E	F	G	H	I	IVEC
	1	2	3	4	5	6	7	8	9	10	11	12	
18	1	1	1	-1	-1	1	1	-1	-1	1	1	1	1
17	1	1	-1	1	-1	-1	-1	1	-1	1	1	-1	1
41	1	-1	99	99	99	99	99	99	99	1	-1	99	1
17	99	99	1	99	99	-1	-1	99	99	99	99	1	-1
16	1	1	1	-1	-1	1	-1	1	1	-1	-1	-1	1

Figure 14. Tree Matrix and IVEC for Sample No. 4

MATCH first sets the flags, JCT, KCT and ICT, all equal to 0 and proceeds to make a comparison down the columns with the matrix MAT. Proceeding with the first column in Figure 14, it will ask in the first row if the entry MAT (1, 1) = 99. If not, as in this case, it asks "Is IVEC(1) = MAT (1, 1)?". In this case, it continues since 1 = 1. It will do so throughout the entire first column, the only difference being the 99 appearing in row 4 which does not count as a disagreement. Completing the first column, MATCH inquires whether ICT has been incremented. Since ICT has not been and is still $0 < 1$, it will set LAG = -1 and return to the main program having determined the next step. When LAG was set equal to -1, NOD was also set equal to MCOL(1), the column in which the perfect match was obtained. Thus, it is known which scheme matrix is used for the next iteration, in this case the first or A.

Since, in the main program, LAG is < 0 , the program proceeds to decide on the scheme based on the match with the tree vector. It now considers Scheme A whose matrix and transformations are reproduced as Figure 15. It now calls SETUP to prepare IVEC as before but now for Scheme A. Again, the columns on the data card of interest for Scheme A are listed in the second row of Figure 15. These are also

														NR, NC				MA				
10	14																					
14	37	37	42	43	53	36	38	39	40													
1	2	3	4	5	6	7	8	9	10	11	12	13	14					COLS	MA			
-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	1					ROW1	MA			
1	1	1	1	1	1	1	1	1	1	1	1	1	99	99					ROW2	MA		
1	1	1	1	1	1	-1	-1	-1	-1	-1	-1	-1	-1					ROW3	MA			
1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1					ROW4	MA		
1	1	1	-1	-1	-1	-1	-1	-1	-1	-1	1	-1	-1					ROW5	MA			
1	1	-1	1	1	1	1	1	1	1	-1	-1	-1	-1					ROW6	MA			
-1	-1	-1	-1	-1	-1	-1	-1	-1	1	99	1	-1	-1					ROW7	MA			
1	1	99	-1	1	99	-1	1	99	1	1	1	-1	-1					ROW8	MA			
1	1	-1	99	1	99	99	1	1	1	1	1	-1	99					ROW9	MA			
1	-1	-1	-1	-1	1	-1	-1	1	1	1	1	-1	-1					ROW10	MA			
		9	6	1	1	-1	2	-1	3	-1	4	-1	5	-1	7	-1	8	-1	9	-1	TR1	MA
		6	3	1	0	-1	1	-1	2	-1	4	1	5	-1							TR2	MA
		6	4	1	0	-1	1	-1	2	-1	3	-1	5	-1							TR3	MA
		2	2	1	1	-1													TR4	MA		
		2	2	1	1	-1													TR5	MA		
		2	2	1	1	-1													TR6	MA		
		5	3	1	4	1	1	-1	2	-1	5	-1							TR7	MA		
		5	3	1	4	1	1	-1	2	-1	5	-1							TR8	MA		
		5	3	1	4	1	1	-1	2	-1	5	-1							TR9	MA		
		5	3	1	4	1	1	-1	2	-1	5	-1							TR10	MA		

shown as "A" columns in Figure 2. These tests are labeled as rows for matrix A. Exactly as in the case of the tree, these columns are read, transformed by the above transformation and listed as the appropriate component of IVEC.

*Note: Test 37 is used two times in this scheme.

Having formed IVEC for Scheme A, MATCH returns to the main program which calls for MATCH for Scheme A.

Identification Matrix A															IVEC
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	
14	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	1	-1
37	1	1	1	1	1	1	1	1	1	1	1	1	99	99	1
37	1	1	1	1	1	1	-1	-1	-1	-1	-1	-1	-1	-1	1
42	1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1
43	1	1	1	-1	-1	-1	-1	-1	-1	-1	-1	1	-1	-1	1
53	1	1	-1	1	1	1	1	1	1	1	-1	-1	-1	-1	1
36	-1	-1	-1	-1	-1	-1	-1	-1	-1	1	99	1	-1	-1	-1
38	1	1	99	-1	1	99	-1	1	99	1	1	1	-1	-1	1
39	1	1	-1	99	1	99	99	1	1	1	1	1	-1	99	1
40	1	-1	-1	-1	-1	1	-1	-1	1	1	1	1	-1	-1	-1

Figure 16. Scheme A Matrix and IVEC for Sample No. 4

As before, it is convenient to illustrate the situation by means of a figure, Figure 16. MATCH then commences to compare IVEC with the entries in MAT column by column by running down the rows. As can be seen by an inspection of Figure 16, $IVEC(3) \neq MAT(1,3)$ and hence ICT will be incremented at this point. Again, ICT will be incremented when comparing $IVEC(9)$ with $MAT(1,9)$. This is a possible candidate for a single double disagreement. As MATCH completes the first column, $ICT = 2$. Referring again to Figure 16, it can be seen that there will be no disagreements during the comparison with the second column. Thus ICT for this column will be $0 < 1$, and again MATCH will assign $LAG = -1$ [for an exact agreement, $NOD = MCOL(2)$] and return to the main program.

In the main program, since $LAG < 0$ at this point, the program proceeds to inquire whether $NOD > 0$, and since $NOD = 2 > 0$, the program prints the identification of NOD which equals 2.

For comparison, the output of BUGID for Sample No. 4 is listed below as Figure 17.

	ORGANISM										PHS
VECTOR	1	1	1	-1	1						
VECTOR	-1	1	1	-1	1	1	-1	1	1	-1	
D0004	032770	1									2

Figure 17. BUGID Output for Sample No. 4

The vectors from the computer output can be compared with the corresponding IVECs determined above. As can be seen, both the computer and the Public Health Service identified this sample as belonging to microorganism category 2. The computer's identification was based on an exact match as would be expected from the above.

Lunar Information System

As mentioned in the Introduction, the interest in a computerized approach to bacterial identification arose through the Apollo sampling program. Such a possibility was envisioned (3) during the original design of the Lunar Information System. Feasibility had been established using the existing Lunar Information System and operating the identification system with the results of the sub-routine, QUALSUM, in the information system. After the feasibility had been demonstrated with this approach, it was hoped that a program could be written which would fit in the Lunar Information System, perhaps as originally conceived. The result of this consideration was the approach of the present report and the associated program which could be inserted into that portion of the Lunar Information System known as QUAL.

Between Apollo 12 and Apollo 13 there were many changes to the procedures used by the Public Health Service in identifying microorganisms. These changes were based upon previous experience in the program which indicated the types of organisms likely to be found. As a result of this experience, many new categories were added under Aerobic Gram Negative rods, Scheme F, and a new identification scheme written. In addition, since no Clostridium species had ever been found, Scheme I (the anaerobic, spore forming, gram positive rods) was condensed to "Clostridium species". These and the other changes necessitated revisions in the reporting format. The form included in Appendix 1 is the one presently being used. These changes greatly influenced the design of the present program in motivating the retention of sufficient flexibility to be able to incorporate any such future changes.

In addition to not operating from the Lunar Information System program QUALSUM, which requires an identification to be made before its utilization, the present program has numerous other advantages. One of these is the above mentioned capability of easy change; aside from providing a means for keeping up with

the evolving programs such as the Lunar Information System, this capability has many other uses. One of the minor modifications, the keys, can be incorporated with ease in order to ascertain their relative effectiveness. Also, since indeterminate tests and test results are being treated somewhat differently in the program, there is the option of modifying either of these at will in an effort to obtain a more efficient or accurate system. The program itself is sufficiently general that it also has application to entirely different systems. It could, for example, be modified with little effort to use a system developed specifically for other applications such as clinical microbiology.

Inasmuch as the feasibility program ran off QUALSUM, it was very dependent upon the Schemes A through I since QUALSUM had already sorted the data into these schemes based upon the PHS assignment of identification categories. In fact, it searched QUALSUM for a scheme and then performed the identification based upon the fact that the Public Health Service had identified the sample as belonging to a category in Scheme A. The present program, on the other hand, takes the raw data and makes the identification. In the case of the data for Apollo 13, the Public Health Service had made an identification, so it was possible to make a comparison between the computer and the Public Health Service results. It is of interest to compare this with the results obtained during the feasibility study. Table 6 presents such a statistical summary. Here, when multiple identifications were made by the computer, agreement was found if the identification of the PHS was included among them.

From the inspection of Table 6, it is apparent that there has been much improvement between Apollo 11 and Apollo 13. It should be noted that a higher percentage of the Apollo 13 identifications are made with no disagreements and there is much less choosing between a number of equally likely categories. In addition, there has been an improvement in the performance of the computerized identification. With Apollo 11 there was disagreement in 187 cases, or 12.7 percent. With Apollo 13, for the Scheme A data, there was agreement in 735 out of 793 samples identified by the Public Health Service, indicating disagreement in 58 cases, or 7.4 percent.

One point of interest is the large fraction of those samples which were identified as belonging to category No. 7 (Micrococcus sub-group 1) by means of three single disagreements, one each between 7, 13, and 14. It is further of interest that there are no instances of two single disagreements. From the distribution

illustrated in Table 6, the possible existence of a distinct category of Micrococcus might be postulated. It is of interest that test results that give rise to this occurrence of 7, 13, 14 in the Baird-Parker scheme happen in a unique manner, and this is discussed in some detail later.

Apollo 11 Data - Scheme A																	
Organism Category																	
No. of:	1	2	3	4	5	6	7	8	9	10	11	12	13	14	996	Total	
Disagree Categ.																	
0	12	261	4	59	78	77	41	11	14		13		11	0		581	
1 1	7	0	0	157	128	40	16	2	1		1		49	1		412	
1 2	3	82	0	18	2	30	13	67	18		9		5	0		247	
1 3	5	1	2	3	0	1	108	0	0		0		0	0		120	
2 1	0	0	0	2	0	0	2	2	1		2		2	0		11	
No ID	0	3	0	2	1	5	26	19	2		0		27	0		85	
Not A	1	8	0	1	6	1	1	2	0		1		0	0		21	
Total	28	355	6	242	215	154	207	103	36		26		104	1		1477	
Disagree	4	1	1	23	5	3	20	1	10		2		11	0		81	5.5%
No ID	0	3	0	2	1	5	26	19	2		0		27	0		85	5.7%
Not Scheme A	1	8	0	1	6	1	1	2	0		1		0	0		21	1.4%
Total	5	12	1	26	12	9	47	22	12		3		38	0		187	12.7%

Apollo 13 Data - Scheme A																	
Organism Category																	
No. of:	1	2	3	4	5	6	7	8	9	10	11	12	13	14	996	Total	
Disagree Categ.																	
0	0	142	0	8	17	21	31	20	45	0	3	0	116	0		403	
1 1	0	0	0	20	17	24	12	2	2	0	0	0	58	0		135	
1 2	0	7	1	17	0	2	0	50	12	0	0	0	21	0		110	
1 3	0	0	1	3	0	0	71*	0	8	0	0	0	0	0		83	
2 1	0	0	0	0	0	0	0	1	0	0	0	0	0	0		1	
															2	2	
No ID A																0	
Total	0	149	2	48	34	47	114	73	67	0	3	0	195	0	2	734	

*Note: Of these 71, 70 were 7, 13, 14 combinations.

Table 6. Summary of Computerized Identification of Apollo 11 and 13 Data

The incorporation of the present program in QUAL as part of the Lunar Information System would provide a number of advantages to the program. One of these

advantages would be the uniformity that is inherent in a computer operation. For example, every sample would be treated in exactly the same manner. Data would be interpreted in a repeatable manner each time. In the event of an inquiry, the process whereby an identification was made can be recovered and gone over step by step for review. There is a price paid for this, of course, in that the resolution of marginal cases by microbiological judgment is minimized. This occurs in the present largely in the case of equally likely categories determined by the computer which has to determine one particular one to record. Fortunately, the Apollo experience of these occurrences follows a pattern which can be simulated by the computer. That is, the computer would simulate previous experience in the Apollo Lunar inventory program. These decisions can be written down and many are straightforward. For example, in the event that a double disagreement with 8 and 11 occurs, 8 is selected. Similarly, between 2 and 4 the choice has always been category 4. Also, in the situation where a single disagreement occurs between category 7, 13, and 14, in every case 7 is selected by the Public Health Service.

Another advantage to having such a program as an integral part of the Lunar Information System would be the reduction of load placed on the microbiologists of the Public Health Service. They could devote themselves to taking the appropriate data, and the computer would then appropriately categorize them. More time could thus be spent developing more efficient schemes or systems for identification which could easily be adopted into the Lunar Information System.

There are, on the other hand, some restrictions inherent in using such a system. For such a program to work, data must be taken and recorded on a card before the program can even get started. Implicit in this is the fact that the appropriate data must be taken. That is, the microbiologist must have some rough idea of the identification in order to perform the appropriate tests. It would appear that rules of thumb could be developed so that the appropriate data would be taken with a minimal amount of effort. In fact, an inspection of the listing of the Apollo data tends to indicate that such rules have already been developed on the basis of experience and are currently being used.

To summarize, the incorporation of the identification program in QUAL would provide the Lunar Information System and Apollo sampling program with a number of advantages. Some of these would include the uniform manner in which identifications

are made along with the ability to reconstruct the data and decisions involved. It would also give great flexibility to the microbiologist in modifying, improving, or adding new identification procedures, and at the same time provide means by which their utility might be assessed. The system could possibly be applied to a variety of similar situations including possible non-Apollo applications. Another advantage might be the possibility of a natural extension of the Lunar Information System to planetary considerations.

Applications

The system that has been discussed was developed for Apollo applications and the program written for inclusion in the Lunar Informations System. Thus, at least one application was in mind during its development. However other applications became apparent as it evolved. As has been mentioned, the program is a useful tool for microbiologists in addition to its utility in the Apollo program. It could be used to determine key efficiency (that is, to assess how efficient tests are in an identification process) in addition to other possible applications. This section will consider some of these alternative uses of this approach to identification.

To illustrate some of these, the results obtained in the Apollo 13 data for the Baird-Parker scheme, or Scheme A, will be used. That part of Table 6 which contains the results is reproduced as Table 7 below.

		Apollo 13 Data														996	Total
No. of:	Disagree Categ.	1	2	3	4	5	6	7	8	9	10	11	12	13	14		
0		0	142	0	8	17	21	31	20	45	0	3	0	116	0		403
1	1	0	0	0	20	17	24	12	2	2	0	0	0	58	0		135
1	2	0	7	1	17	0	2	0	50	12	0	0	0	21	0		110
1	3	0	0	1	3	0	0	71*	0	8	0	0	0	0	0		83
2	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0		1
																2	2
	No ID A																0
	Total	0	149	2	48	34	47	114	73	67	0	3	0	195		2	734

*Note: Of these 71, 70 were 7, 13, 14 combinations

Table 7. Statistical Summary of BUGID Results with Apollo 13

Table 7 compares the results of the computerized identification with those of the Public Health Service for the Baird-Parker isolates on Apollo 13. The nomenclature in the left hand column has already been discussed. It is simply the listing of the ways in which the computer can obtain a result, as was presented in Table 2. In addition to obtaining information as to how well the computer agreed with the Public Health Service, other facts can be gathered from Table 7. For example, although the Public Health Service data give only the number of the identification made, the computer data give the distribution of identifications in terms of how close they were to the exact test sequence hoped for for the organism category. If the microorganisms were well behaved, the categories were well defined and very good tests were available, it might be hoped, in the best of all possible worlds, that, given a microorganism, appropriate tests would be run and they would agree exactly with those specified by the scheme for identifying this class of microorganisms. In reality this does not occur for a number of reasons. It is common for at least one test to differ from that expected for the microorganism at hand. This can be seen in Table 7 by inspecting the row listing the number of times that an organism was identified even though it disagreed in one test with the category in which it was identified. It is seen that there were 135 such occurrences. This particular situation does not unduly complicate the identification process, but when it is extended so that there is one disagreement with each of two or three categories, a choice must be made between the categories. The extent to which this reaching occurs in order to obtain an identification provides an indication of how convincing the identification is. Furthermore, having done this for a large number of samples, the results, as tabulated in Table 7, provide a measure of how well the identification process is performing. Looking at the totals listed in the right hand column of Table 7, it can be seen that most of the identifications were made when there was an exact agreement with the test results, a somewhat fewer number where there was one test neglected, a still fewer number where one test was neglected and a choice had to be made between two categories, and so forth. This would seem to indicate that the Baird-Parker scheme is quite good at identifying the Staphylococcus and Micrococcus into the categories assigned. It would be possible to use large collections of data to make judgments about the efficiencies of schemes and their associated tests by using reasoning similar to that above.

The totals examined above provide a measure of the scheme as a whole. If the body of Table 7 is examined, a few departures from the above pattern are noticed.

One in particular stands out as an anomaly. This occurs in category No. 7 (Micrococcus, sub-group 1) where the distribution of sample identification for the 114 samples in this category begins with 31 exact agreements with the expected test results, and then drops to 12 identifications made with one single disagreement (one test result was not as anticipated). There are no double disagreements. Then there are an unusually large number of 71 samples identified on the basis of three single disagreements. For these last 71, then, a choice had to be made between three alternative categories, each of which had one test result which differed from that sequence of tests provided by the sample. This unusual change in the trend discussed above seemed worthy of further analysis for, if nothing else, it may have been a peculiarity in the program. Upon inspection, it developed that the vast majority of these 71 (in fact 70) with three single disagreements were choices between categories 7, 13, 14. Figure 18 presents the test results expected for these three categories as given in the Baird-Parker scheme which was presented in Figure 1.

Organism category	7	13	14	Vector	Disagrees
Baird-Parker subgroup	1	7	8		
Data entry					
14	-	-	+	-	← w 14
37	+	±	±	+	
37	-	-	-	-	
42	-	-	-	-	
43	-	-	-	-	
53	+	-	-	-	← w 7
36	-	-	-	-	
38	-	-	-	-	
39	V	-	±	+	← w 13
40	-	-	-	-	

Figure 18. Pertinent Excerpts from the Baird-Parker Scheme with Commonly Occurring Vector Assigned as Category 7

It can be seen that a single disagreement with three test sequences as tabulated in Figure 18 can occur in only one way. The test results giving the vector (denoted "Vector") which disagrees in this manner are also tabulated along with an indication of which category disagreement occurs for a given test. It should be noted that the Baird-Parker glucose test is used twice. This test result is recorded in column 37 of the Public Health Service reporting form, and, hence, data are read from that

column twice although a different transformation is applied to the two readings in order to obtain the Baird-Parker results.

The 70 data cards representing these 7, 13, 14 combinations were extracted from the Apollo 13 data and listed. This listing is given as Table 8. In Table 8 the columns used for the tree are headed by a T and, similarly, those used by Baird-Parker Scheme A by an A. As can be seen from an inspection, the only variation in the relevant data is in column 14 which is used to record the pigment coloration. There, 8 (white) or 9 (yellow) is recorded with the single exception of one 5 (orange). Thus, not only is IVEC unique, but the raw data are remarkably uniform for those samples which give rise to the possibility 7, 13, 14. It was possible that the transformations converted the data in some peculiar manner so as to give rise to such a high incidence of samples falling in this particular category having a multiple choice. It appears from Table 8 that this is not the case and that, moreover, some 59.8 per cent of the samples identified as belonging to Microorganism category 7 (Baird-Parker Micrococcus sub-group 1) in fact do not quite fit the Baird-Parker classification.

It is of interest that Baird-Parker (10) indicates that although a number of schemes have been proposed for dividing the general Staphylococcus and Micrococcus into species, "groups" or "subgroups", at present none of the classifications is acceptable to all taxonomists. Considering the Micrococcus genus, it is generally agreed there are two well-defined species M. luteus and M. roseus, which correspond to his subgroup 7 and 8 respectively or the Public Health Service categories 13 and 14. Baird-Parker suggests "Undoubtedly other species will be recognized eventually and these will be given specific names when they have been adequately defined." In view of the two kinds of samples occurring in the Apollo data and the high incidence of the one discussed above, it might be worth considering these as separate groups. A worthy investigation would be the development of a test to further distinguish between the two types of Micrococcus sub-group 1.

Thus, some applications, in addition to the immediate Apollo usage, have been indicated. This approach provides a tool for the microbiologists which, in addition to simulating the identification process, can be used to form judgments about the efficiency of a particular scheme or process. It, furthermore, can be used to ascertain how well a classification scheme fits a collection of data.

1234567890123456789012345678901234567890123456789012345678901234567890			
A TTT	AAAAATAA	A	(TTT)
D00110327701182122	13131211	1	0071
D00240327702182122	13131211	1	0071
D00280327702182122	13131211	1	0071
D00300327702182122	13131211	1	0071
D00320327702152122	13131211	1	0071
D00350327702182122	13131211	1	0071
D00720327703182122	13131211	1	0071
D00730327703182122	13131211	1	0071
D00780327703182122	13131211	1	0071
D00820327703182122	13131211	1	0071
D00840327703182122	13131211	1	0071
D01180327703182122	13131211	1	0073
D01480327703182122	13131211	1	0074
D01660327701192122	13131211	1	0075
D01670327701192122	13131211	1	0075
D02000327702192122	13131211	1	0076
D02440330702182122	13131211	1	0072
D02560330702192122	13131211	1	0072
D02750330702182122	13131211	1	0072
D02780330702192122	13131211	1	0072
D03150330703192122	13131211	1	0072
D03280330703182122	13131211	1	0072
D03350330703192122	13131211	1	0072
D03400330703182122	13131211	1	0072
D03710403701192122	13131211	1	0071
D03850403703192122	13131211	1	0071
D03860403703192122	13131211	1	0071
D03870403703192122	13131211	1	0071
D03890403703192122	13131211	1	0071
D03940403703192122	13131211	1	0071
D03980403703192122	13131211	1	0071
D04010403703192122	13131211	1	0071
D04080403703192122	13131211	1	0071
D04110403703192122	13131211	1	0071
D04140403703192122	13131211	1	0071
D04160403703192122	13131211	1	0071
D04170402703192122	13131211	1	0071
D04200403703192122	13131211	1	0071
D04230403703192122	13131211	1	0071
D04250403703192122	13131211	1	0071
D04260403703192122	13131211	1	0071
D04300403703192122	13131211	1	0071
D04310403703192122	13131211	1	0071
D04320403703192122	13131211	1	0071
D04430403703192122	13131211	1	0072
D04860403703152122	13131211	1	0072
D04870403703192122	13131211	1	0072
D05140403703192122	13131211	1	0072
D05300403703192122	13131211	1	0072
D05310403703192122	13131211	1	0072
D05500403703152122	13131211	1	0072
D05830403702182122	13131211	1	0073
D06070403702182122	13131211	1	0073
D06190403701191122	13131211	1	0074
D06270403701192122	13131211	1	0074
D07190407701182122	13131211	1	0072
D07450407701192122	13131211	1	0072
D07540407701192122	13131211	1	0072
D07670407701182122	13131211	1	0072
D07770407701182122	13131211	1	0072
D08380407703192122	13131211	1	0072
D08560407701182122	13131211	1	0073
D09820410701182122	13131211	1	0071
D09920410701192122	13131211	1	0071
D10020410701192122	13131211	1	0071
D10040410701192122	13131211	1	0071
D10100410701192122	13131211	1	0071
D10470410703182122	13131211	1	0071
D10480410703182122	13131211	1	0071
D10500410703182122	13131211	1	0071
1234567890123456789012345678901234567890123456789012345678901234567890			
A TTT	AAAAATAA	A	(TTT)

Table 8. Listing of Apollo 13 Data Cards for 7, 13, 14 Combinations Identified as 7

Conclusions

In the process of writing a program to perform identification for the Public Health Service Apollo Program, a novel approach to the simulation of the identification process was developed. In the development of this approach, a system was designed which could easily be used by practicing microbiologists and easily modified without major changes to the system itself. The design was such that the system can incorporate the spectrum of traditional identification techniques ranging from the binary selection process (such as trees) to keys which are typically presented as test-organism matrices, and it can encompass a variety of other methods.

The resulting program is intended to be included as part of the Lunar Information System. It obtains agreement with the Public Health Service on 92 percent of their data and will have a natural and easy extension to similar programs concerned with NASA planetary missions. In addition to relieving the Public Health Service of some of the burden of microbial identification, it provides a tool which can be used to improve this aspect of their tests.

The system can be used to assess existing or proposed microbiological identification procedures and to indicate directions in which they might be improved. It has the property of formalizing many of these procedures so that standard criteria can be established.

In addition to the above largely NASA-oriented applications, it has potential if applied to microbiological research. The above formal "standardized" rules or agreements about the level or criteria for identification could be used as a measure of grouping similar microorganisms. An example of this is Micrococcus Cohn, sub-group I, discussed above.

Not only does this approach have immediate application to NASA problems and demonstrated possibilities in microbiological research, but it may also find a wide variety of other applications in use. Interest has been expressed on the part of the Public Health Service in broader applications within their program.

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Appendix 1

Organism Categories and Data Recording Form

KEY NUMBER OF ORGANISMS

IBM CODES FOR COLUMNS 77 - 79 ON MICROBIAL IDENTIFICATION DATA SHEET

A Catalase positive, gram positive, aerobic cocci

- 001 Staphylococcus, Subgroup I
- 002 Staphylococcus, Subgroup II
- 003 Staphylococcus, Subgroup III
- 004 Staphylococcus, Subgroup IV
- 005 Staphylococcus, Subgroup V
- 006 Staphylococcus, Subgroup VI

- 007 Micrococcus, Subgroup 1
- 008 Micrococcus, Subgroup 2
- 009 Micrococcus, Subgroup 3
- 010 Micrococcus, Subgroup 4
- 011 Micrococcus, Subgroup 5
- 012 Micrococcus, Subgroup 6
- 013 Micrococcus, Subgroup 7
- 014 Micrococcus, Subgroup 8
- 996 Atypical Micrococcaceae

B Catalase negative, gram positive, aerobic cocci

- 015 Enterococcus
- 016 Lactic
- 017 Viridans
- 018 Pyogenic

C Gram positive, sporeforming, aerobic rods

- 019 B. alvei
- 020 B. badius
- 021 B. brevis
- 022 B. cereus
- 023 B. circulans
- 024 B. coagulans
- 025 B. firmus
- 026 B. laterosporus
- 027 B. lentus
- 028 B. licheniformis
- 029 B. macerans
- 030 B. megaterium
- 031 B. pantothenicus
- 032 B. polymyxa
- 033 B. pulvifaciens
- 034 B. pumilus
- 034 B. spaericus
- 036 B. subtilis
- 998 Atypical Bacillus spp.

KEY NUMBER OF ORGANISMS (Cont.)

- D Aerobic and anaerobic non sporeforming gram positive rods, anaerobic gram negative cocci, and anaerobic gram negative rods: test reactions only
 - 037 Aerobic, non sporeforming gram positive rods
 - 038 Anaerobic, non sporeforming gram positive rods
 - 039 Aerobic gram negative cocci
 - 040 Anaerobic gram negative rods

- E Aerobic gram negative cocci
 - 041 *Neisseria* spp.

- F Aerobic gram negative rods
 - 045 *Pseudomonas*, Group I
 - 049 *Pseudomonas*, Group II
 - 050 *Pseudomonas*, Group III
 - 051 *Pseudomonas*, Group IV
 - 052 *Aeromonas* spp.
 - 053 *Bibrio* spp.
 - 044 *Flavobacterium* spp.
 - 042 *Achromobacter* spp.
 - 043 *Alcaligenes* spp.
 - 054 *Escherichia*
 - 055 *Klebsiella-Enterobacter*
 - 056 *Salmonella*
 - 057 *Arizona*
 - 058 *Citrobacter*
 - 059 *Shigella*
 - 060 *P. vulgaris*
 - 061 *P. morganii*
 - 062 *P. mirabilis*
 - 063 *P. rettgeri*
 - 064 *Providencia*
 - 065 Unidentified gram negative rods

- G 046 Gram positive, catalase positive anaerobic cocci

- H 047 Gram positive, catalase negative anaerobic cocci

- I 048 *Clostridium* spp.

- J 088 *Actinomycetes*

- K 089 *Streptomycetes*

- L 090 Yeasts

- M 091 Molds

- N 999 Lost in process

MICROBIAL IDENTIFICATION

(Circle Code Number that Applies)		AREA:
(1-5) Identification Number 		(28) NITRATE (ANAEROBIC) ND 0 + 1 2
(6-11) Date 		(29) GLUCOSE (ANAEROBIC) ND 0 NG 1 G 2
(12) Sample Number 		(30) P.R. Sucrose ND 0 A 3 NC 1 AG 4 NG 2 ALK 5
(13) Sample Treatment Aerobic 1 Anaerobic 2 Heat Sh. Aerobic 3 Heat Sh. Anaerobic 4 Blood Agar Aerobic 5 Blood Agar Anaerobic 6 MacConkey 7 Mycophil 8		(31) P.R. Xylose ND 0 A 3 NC 1 AG 4 NG 2 ALK 5
(14) Pigment Black 1 Pink 6 Brown 2 Red 7 Green 3 White 8 Grey 4 Yellow 9 Orange 5		(32) O.F. Glucose ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(15) Translucent 1 Opaque 2 Soluble pigment 3		(33) Gas - 1 + 2
(16) Growth characteristics Aerobic 1 Anaerobic 2		(34) O.F. Maltose ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(17) CELLULAR MORPHOLOGY Rods 1 SPORES 3 YEASTS 5 ACTINOM. 7 COCCI 2 PLEO. 4 MOLDS 6 STREPTOM. 8		(35) B.P. Arabinose ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(18) Gram Stain - 1 + 2		(36) B.P. Glucose ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(19) P.R. Aesculin ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(37) B.P. Lactose ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(20) P.R. Arabinose ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(38) B.P. Maltose ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(21) P.R. Glucose ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(39) B.P. Mannitol ND 0 AO 3 NC 1 AF 4 NG 2 ALK 5
(22) P.R. Glycerol ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(40) Catalase ND 0 - 1 + 2
(23) MALONATE BROTH ND 0 + 3 NC 1 NG 2		(41) Coagulase ND 0 - 1 + 2
(24) P.R. Lactose ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(42) Phosphatase ND 0 - 1 + 2
(25) P.R. Maltose ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(43) Hemolysis ND 0 Beta 2 Alpha 1 Gamma 3
(26) P.R. Mannitol ND 0 A 3 NC 1 AG 4 NG 2 ALK 5		(44) Litmus Milk ND 0 Acid 2 NC 1 ALK 3
(27)		(45) Stormy - 1 + 2
		(46) Soft 1 Hard 2 No curd 3
		(47) Red. Litmus - 1 + 2
		(48) Peptonization - 1 + 2
		(49) Gelatin ND 0 - 1 + 2
		(50) Starch ND 0 - 1 + 2
		(51) Oxidase ND 0 - 1 + 2
		(52) V. P. ND 0 - 1 + 2
		(53) M. R. ND 0 - 1 + 2
		(54) Indol ND 0 - 1 + 2
		(55) Citrate ND 0 - 1 + 2
		(56) Nitrate ND 0 - 1 + 2
		(57) TSI ND 0 S+ 2 S- 1 ALK 3
		(58) B- 1 B+ 2 ALK 3
		(59) H2S 1 H2S+ 2
		(60) No gas 1 Gas 2
		(61) Optochin ND 0 - 1 + 2
		(62) Urease ND 0 - 1 + 2
		(63) Motility (SIM) ND 0 - 1 + 2
		(64) AK#2 ND 0 NG 1 G 2
		(65) Heat Shock S- 1 S+ 2
		(66) H- 1 H+ 2
		(67) Peptone Broth ND 0 Gas 2 NG 1 No Gas 3
		(68) 10°C ND 0 NG 1 G 2
		(69) 45°C ND 0 NG 1 G 2
		(70) 6.5% NaCl ND 0 NG 1 G 2
		(71) 500C ND 0 NG 1 G 2
		(72) 0.1% M.B. Milk ND 0 NG 1 G 2
		(73) MacConkey ND 0 P 2 NG 1 G 3
		(74) Flagella ND 0 - 1 + 2
		(75) Capsule ND 0 - 1 + 2
		(76-79) Key Number
		(80) Portion of Spacecraft Sampled CMI 1 IU 5 LAI 2 S48 6 LAE 3 SLA 7 LDE 4
		REMARKS

HSM 8.27 (NCDC)
4-69

Appendix 2

Listing of BUG ID and Identification Input Cards

```

BUGID,T100,CM100000.
ACCOUNT,E9935,D1741,A0003010,G1700,RT,KUNC.
FUN,S,,,,,37777.
MAP,ON.
REDUCE.
PRESET,NIND.
LGO.
'
PROGRAM BUGID(INPUT,OUTPUT)
DIMENSION IAGR(100),IDOWN(5)
DIMENSION IAGR(5,100)
COMMON/A/NROW(6),MROW(6,20),ICARD(80),NTRAN(6,20),MTRAN(6,20,10),
1 MTRA2(6,20,10),IVEC(20)
COMMON/B/MAT(6,20,20),NCOL(6),MCOL(6,20),LAG,ISD(3),IDD,NOD,JCT
1 ,KCT
DIMENSION IDMT(6)
DATA IDMT/4HTREE,4HA ,4HB ,4HC ,4HF ,4HFA /
NMT=6
NRT=20
NCT=20
DO 4 I=1,5
DO 4 J=1,100
4 IAGR(I,J)=0
C LEFT INDEX--1 IS EXACT AGREEMENT
C 2 IS ONE SINGLE DISAGREEMENT
C 3 IS TWO SINGLE DISAGREEMENTS
C 4 IS THREE SINGLE DISAGREEMENTS
C 5 IS ONE DOUBLE DISAGREEMENT
C 1 IS USED ALSO TO TALLY THE NO IDS.
C RIGHT INDEX--1-91 ARE REGULAR OUTCOMES
C 92 IS 996
C 93 IS 998
C 94 IS 999
C 95 IS NO ID IN TREE MATRIX
C 96 IS NO ID IN MATRIX A
C 97 IS NO ID IN MATRIX B
C 98 IS NO ID IN MATRIX C
C 99 IS NO ID IN MATRIX F
C 100 IS NO ID IN MATRIX FA
5 FORMAT(14I5)
C NM IS NUMBER OF MATRICES TO SET UP. THE FIRST MATRIX IS FIRST TREE.
READ 5,NM
IF(NM .LE. NMT) GO TO 15
10 FORMAT(*1NUMBER OF MATRICES IS*I3*--*I3* ALLOWED. CHANGE DIMENSIO
1NS OF MAT,NCOL,NROW,MCOL, MROW, NTRAN AND MTRAN. ALSO CHANGE NMT.
2*)
PRINT 10,NM,NMT
STOP
15 DO 65 IM=1,NM
READ 5,NR,NC
IF(NR .LE. NRT) GO TO 25
20 FORMAT(*1NUMBER OF ROWS FOR MATRIX*I2* IS*I3*--*I3* ALLOWED. CHAN
1GE DIMENSIONS OF MAT, IVEC, MROW, NTRAN AND MTRAN. ALSO CHANGE NR
2T.*)

```

```

        PRINT 20,IM,NR,NRT
        STOP
25      IF(NC .LE. NCT) GO TO 35
30      FORMAT(*1NUMBER OF COLUMNS FOR MATRIX*I2* IS*I3*--*I3* ALLOWED. C
        CHANGE DIMENSIONS OF MAT AND MCOL. ALSO CHANGE NCT.*)
        PRINT 30,IM,NC,NCT
        STOP
35      NROW(IM)=NR
        NCOL(IM)=NC
        READ 40,(MROW(IM,IR),IR=1,NR)
        READ 40,(MCOL(IM,IC),IC=1,NC)
40      FORMAT(36I3)
        DO 45 IR=1,NR
        READ 40,(MAT(IM,IR,IC),IC=1,NC)
45      CONTINUE
50      FORMAT(*1IDENTIFICATION MATRIX *A4//)
        PRINT 50,IDMT(IM)
        IF(IM-1) 52,52,54
51      FORMAT(10X,1HA,3X,1HB,3X,1HC,2X,2HD1,2X,2HD2,2X,2HD3,2X,2HD4,
1      3X,1HE,3X,1HF,3X,1HG,3X,1HH,3X,1HI)
52      PRINT 51
53      FORMAT(7X,20I4)
54      PRINT 53,(MCOL(IM,IC),IC=1,NC)
55      FORMAT(1X,I4,2X,20I4)
        DO 60 IR=1,NR
        PRINT 55,MROW(IM,IR),(MAT(IM,IR,IC),IC=1,NC)
60      CONTINUE
        DO 62 IR=1,NR
61      FORMAT(I5,20I3)
        READ 61,NTR,(MTRAN(IM,IR,I),MTRA2(IM,IR,I),I=1,NTR)
        NTRAN(IM,IR)=NTR
62      CONTINUE
65      CONTINUE
        IDB=1
        IDF=5
        IDTB=6
        IDTE=11
        ISN=12
        ICM=17
        IDIS=0
        MANY=0
69      FORMAT(5H1 ID,4X,4HDATE,2X,2HSN,40X,8HORGANISM//)
        PRINT 69
70      FORMAT(A1,75I1,I3,I1)
75      READ 70,(ICARD(I),I=1,78)
        MANY=MANY+1
C      A BLANK CARD WILL STOP PROGRAM
        IF(ICARD(2)) 76,76,85
76      IF(ICARD(3)) 77,77,85
77      IF(ICARD(4)) 78,78,85
78      IF(ICARD(5)) 79,79,85
79      IF(ICARD(6)) 80,80,85
80      PRINT 81
81      FORMAT(1H1,4X,5HEXACT,2X,8H1 SINGLE,2X,8H2 SINGLE,2X,8H3 SINGLE,

```

```

1 2X,8H1 DOUBLE,5X,5HTOTAL)
  DO 88 J=1,100
    ISUM=0
    DO 87 I=1,5
87   ISUM=ISUM+IAGR(I,J)
88   IACR(J)=ISUM
    DO 92 I=1,5
      ISUM=0
      DO 91 J=1,100
91     ISUM=ISUM+IAGR(I,J)
92     IDOWN(I)=ISUM
82     FORMAT(1X,I4,I5,5I10)
      DO 83 J=1,91
        PRINT 82,J,(IAGR(I,J),I=1,5),IACR(J)
83     CONTINUE
      J=996
      PRINT 82,J,(IAGR(I,92),I=1,5),IACR(92)
      J=998
      PRINT 82,J,(IAGR(I,93),I=1,5),IACR(93)
      J=999
      PRINT 82,J,(IAGR(I,94),I=1,5),IACR(94)
84     FORMAT(1X,A4,I5,5I10)
      DO 86 J=95,100
        JI=J-94
        PRINT 84, IDMT(JI),(IAGR(I,J),I=1,5),IACR(J)
86     CONTINUE
      ISUM=0
      DO 94 J=1,100
94     ISUM=ISUM+IACR(J)
      JSUM=0
      DO 96 I=1,5
96     JSUM=JSUM+IDOWN(I)
      IF(JSUM-ISUM) 98,99,98
97     FORMAT(32H TOTALS DO NOT AGREE--ACROSS IS ,I5,9H DOWN IS,I5)
98     PRINT 97,ISUM,JSUM
      GO TO 102
93     FORMAT(5X,I5,5I10)
99     PRINT 93,(IDOWN(I),I=1,5),ISUM
101    FORMAT(31H THE NUMBER OF DISAGREEMENTS IS,I5)
102    PRINT 101, IDIS
      MANY=MANY-1
103    FORMAT(*NUMBER OF INPUT CARDS IS*I6)
      PRINT 103,MANY
      CALL EXIT
85    IF(ICARD(ICM)-4) 130,130,90
90    IF(ICARD(ICM)-6) 100,110,115
95    FORMAT(1X,A1,4I1,2X,6I1,2X,I1,40X,I3,14X,I3)
C    PERFECT MATCH
100    ICT=90
105    IF(ICT-ICARD(77)) 117,106,117
106    IF(ICT-91) 114,114,107
107    IF(ICT-996) 109,108,109
108    LCT=92
      GO TO 116

```

```

109 IF(ICT-998) 112,111,112
111 LCT=93
    GO TO 116
112 IF(ICT-999) 117,113,117
113 LCT=94
    GO TO 116
114 LCT=ICT
116 IAGR(1,LCT)=IAGR(1,LCT)+1
    GO TO 118
117 IDIS=IDIS+1
118 PRINT 95,(ICARD(1),I=IDB,IDE),(ICARD(1),I=IDTB,IDTE),
1 ICARD(ISN),ICT,ICARD(77)
    GO TO 75
110 ICT=91
    GO TO 105
115 IF(ICARD(ICM)-7) 110,120,125
120 ICT=88
    GO TO 105
125 ICT=89
    GO TO 105
C    SET UP TRFE VECTOR
130 CALL SETUP(1)
    MATNO=1
    IF(IVEC(4)) 200,200,220
200 IF(ICARD(65)-2) 220,205,220
205 IF(ICARD(66)-2) 215,210,215
210 IVEC(4)=1
    GO TO 220
215 IF(ICARD(67)-2) 220,210,220
220 CALL MATCH(1)
    IF(LAG) 225,221,221
221 IF(ICARD(18)) 222,222,170
222 ICT=990
    GO TO 105
C    PERFECT MATCH
225 IF(NOD-1) 235,230,235
C    SCHEME A
230 MATNO=2
    GO TO 135
235 IF(NOD-2) 245,240,245
C    SCHEME B
240 MATNO=3
    GO TO 135
245 IF(NOD-3) 255,250,255
C    SCHEME C
250 MATNO=4
    GO TO 135
255 IF(NOD-9) 265,260,265
C    SCHEME F
260 MATNO=5
    CALL SETUP(MATNO)
    CALL MATCH(MATNO)
    IF(LAG) 261,145,170
261 IF(NOD) 262,262,140

```

```

C      SCHEMF FA
262  MATNO=6
      GO TO 135
265  IF(NOD=8) 275,270,275
C      E
270  ICT=41
      GO TO 105
275  IF(NOD=11) 285,280,285
C      H
280  ICT=47
      GO TO 105
285  IF(NOD=10) 295,290,295
C      G
290  ICT=46
295  IF(NOD=12) 305,300,305
C      I
300  ICT=62
      GO TO 105
305  IF(NOD=4) 315,310,315
C      D--39
310  ICT=39
      GO TO 105
315  IF(NOD=5) 325,320,325
C      D--40
320  ICT=40
      GO TO 105
325  IF(NOD=6) 335,330,335
C      D--37
330  ICT=37
      GO TO 105
335  IF(NOD=7) 170,340,170
C      D--38
340  ICT=38
      GO TO 105
135  CALL SFTUP(MATNO)
      CALL MATCH(MATNO)
      IF(LAG) 140,145,170
C      PERFECT MATCH
140  ICT=NOD
      GO TO 105
145  IF(JCT=3) 146,146,155
146  IF(JCT) 147,147,160
147  IF(KCT) 170,170,155
150  FORMAT(1X,A1,4I1,2X,6I1,2X,I1,52X,I3,2X,I3)
155  IF(IDD=ICARD(77))158,156,158
156  IF(IDD=91) 157,157,158
157  IAGR(5,IDD)=IAGR(5,IDD)+1
      GO TO 151
158  IDIS=IDIS+1
151  PRINT 150,(ICARD(I),I=IDB,IDE),(ICARD(I),I=IDTB,IDTE),
1    ICARD(ISN),IDD,ICARD(77)
      GO TO 75
159  FORMAT(1X,A1,4I1,2X,6I1,2X,I1,43X,3I3,5X,I3)
C      MCT=1 MEANS AGREEMENT, SO TALLY

```

```

C      MCT=0 MEANS NO AGREEMENT
160    MCT=0
      DO 352 I=1,JCT
      IF(ISC(I)-ICARD(77)) 352,351,352
351    MCT=1
352    CONTINUE
      IF(MCT) 357,357,353
357    IDIS=IDIS+1
      GO TO 356
353    LCT=ICARD(77)
      IF(LCT-91) 354,354,356
354    IAGR(JCT+1,LCT)=IAGR(JCT+1,LCT)+1
356    GO TO(163,164,166),JCT
161    FORMAT(1X,A1,4I1,2X,6I1,2X,I1,43X,2I3,8X,I3)
162    FORMAT(1X,A1,4I1,2X,6I1,2X,I1,43X,I3,11X,I3)
163    PRINT 162,(ICARD(I),I=IDB,IDE),(ICARD(I),I=IDTB,IDTE),ICARD(ISN),
      1 (ISC(I),I=1,JCT),ICARD(77)
      GO TO 75
164    PRINT 161,(ICARD(I),I=IDB,IDE),(ICARD(I),I=IDTB,IDTE),ICARD(ISN),
      1 (ISC(I),I=1,JCT),ICARD(77)
      GO TO 75
166    PRINT 159,(ICARD(I),I=IDB,IDE),(ICARD(I),I=IDTB,IDTE),ICARD(ISN),
      1 (ISC(I),I=1,JCT),ICARD(77)
      GO TO 75
165    FORMAT(1X,A1,4I1,2X,6I1,2X,I1,40X,7HNO ID. ,A4,6X,I3)
170    GO TO (380,350,380,360,380,380),MATNO
C      SCHEME A WHICH DOES NOT MEET CRITERION
350    IF(ICARD(17)-2) 380,355,380
355    ICT=996
      GO TO 105
C      SCHEME C WHICH DOES NOT MEET CRITERION
360    IF(ICARD(17)-1) 365,375,365
365    IF(ICARD(17)-3) 370,375,370
370    IF(ICARD(17)-4) 380,375,380
375    ICT=998
      GO TO 105
380    PRINT 165,(ICARD(I),I=IDB,IDE),(ICARD(I),I=IDTB,IDTE),ICARD(ISN),
      1 IDMT(MATNO),ICARD(77)
      IAGR(1,94+MATNO)=IAGR(1,94+MATNO)+1
      GO TO 75
      END

```

```

SUBROUTINE MATCH(MATNO)
COMMON/A/NROW(6),MROW(6,20),ICARD(80),NTRAN(6,20),MTRAN(6,20,10),
1 MTRA2(6,20,10),IVFC(20)
COMMON/B/MAT(6,20,20),NCOL(6),MCOL(6,20),LAG,ISD(3),IDD,NOD,JCT
1 ,KCT
C   LAG=-1 MEANS PERFECT AGREEMENT--LAG=0 MEANS IDENT.--LAG=1 MEANS NO IDENT.
NC=NCOL(MATNO)
NR=NROW(MATNO)
JCT=KCT=0
DO 45 J=1,NC
  ICT=0
  DO 10 I=1,NR
    IF(MAT(MATNO,I,J)-99)4,10,4
4   IF(IVFC(I)-MAT(MATNO,I,J)) 5,10,5
5   ICT=ICT+1
    IF(ICT-2) 10,10,45
10  CONTINUE
    IF(ICT-1) 40,15,25
C   SINGLE DISAGREEMENT
15  IF(JCT-3) 21,20,45
20  JCT=4
    GO TO 45
21  JCT=JCT+1
    ISD(JCT)=MCOL(MATNO,J)
    GO TO 45
C   DOUBLE DISAGREEMENT
25  IF(KCT-1) 35,30,45
30  KCT=2
    GO TO 45
35  KCT=1
    IDD=MCOL(MATNO,J)
    GO TO 45
C   PERFECT MATCH
40  LAG=-1
    NOD=MCOL(MATNO,J)
    RETURN
45  CONTINUE
    IF(JCT-3) 60,60,50
50  IF(KCT-1) 55,65,55
C   TOO MANY DISAGREEMENTS
55  LAG=1
    RETURN
60  IF(JCT) 70,70,65
65  LAG=0
    RETURN
70  IF(KCT-1) 55,65,55
    END

```

```

      SUBROUTINE SFTUP(MATNO)
      COMMON/A/NROW(6),MROW(6,20),ICARD(80),NTRAN(6,20),MTRAN(6,20,10),
1  MTRA2(6,20,10),IVEC(20)
      N=NROW(MATNO)
      DO 15 I=1,N
C      J IS THE NUMBER OF COLUMN ON CARD.
      J=MROW(MATNO,I)
      K=NTRAN(MATNO,I)
C      MAKE TRANSFORMATION
      IS=ICARD(J)
      DO 5 L=1,K
      IF(IS-MTRAN(MATNO,I,L)) 5,10,5
5      CONTINUE
C      EXIT HERE MEANS NO MATCH--THEREFORE 99
      IVEC(I)=99
      GO TO 15
C      MATCH
10     IVEC(I)=MTRA2(MATNO,I,L)
15     CONTINUE
20     FORMAT(* VECTOR*20I3)
      PRINT 20,(IVEC(I),I=1,N)
      RETURN
      END

```

```

6
5 12
18 17 41 17 16
1 2 3 4 5 6 7 8 9 10 11 12
1 1 1 -1 -1 1 1 -1 -1 1 1 1
1 1 -1 1 -1 -1 -1 1 -1 1 1 -1
1 -1 99 99 99 99 99 99 99 1 -1 99
99 99 1 99 99 -1 -1 99 99 99 99 1
1 1 1 -1 -1 1 -1 1 1 -1 -1 -1
2 2 1 1 -1
4 2 1 1 -1 3 -1 4 -1
2 2 1 1 -1
4 3 1 1 -1 2 -1 4 -1
2 1 1 2 -1

```

```

10 14
14 37 37 42 43 53 36 38 39 40
1 2 3 4 5 6 7 8 9 10 11 12 13 14
-1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1 1
1 1 1 1 1 1 1 1 1 1 1 1 99 99
1 1 1 1 1 1 -1 -1 -1 -1 -1 -1 -1 -1
1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1 -1
1 1 1 -1 -1 -1 -1 -1 -1 -1 -1 1 -1 -1
1 1 -1 1 1 1 1 1 1 1 -1 -1 -1 -1
-1 -1 -1 -1 -1 -1 -1 -1 -1 1 99 1 -1 -1
1 1 99 -1 1 99 -1 1 99 1 1 1 -1 -1
1 1 -1 99 1 99 99 1 1 1 1 1 -1 99
1 -1 -1 -1 -1 1 -1 -1 1 1 1 1 -1 -1
9 6 1 1 -1 2 -1 3 -1 4 -1 5 -1 7 -1 8 -1 9 -1
6 3 1 0 -1 1 -1 2 -1 4 1 5 -1
6 4 1 0 -1 1 -1 2 -1 3 -1 5 -1
2 2 1 1 -1
2 2 1 1 -1
2 2 1 1 -1
5 3 1 4 1 1 -1 2 -1 5 -1
5 3 1 4 1 1 -1 2 -1 5 -1
5 3 1 4 1 1 -1 2 -1 5 -1
5 3 1 4 1 1 -1 2 -1 5 -1

```

```

8 4
69 70 71 73 47 45 44 44
15 16 17 18
1 1 99 -1
1 -1 99 -1
1 -1 -1 -1
1 1 -1 -1
1 1 1 1
-1 1 -1 -1
99 99 -1 1
99 1 99 99
2 2 1 1 -1
2 2 1 1 -1

```

```

NR,NC TREE
ROWS TREE
COLS TREE
ROW 1 TREE
ROW 2 TREE
ROW 3 TREE
ROW 4 TREE
ROW 5 TREE
TR1 TREE
TR2 TREE
TR3 TREE
TR4 TREE
TR5 TREE
NR,NC MA
COLS MA
ROW1 MA
ROW2 MA
ROW 3 MA
ROW4 MA
ROW5 MA
ROW6 MA
ROW7 MA
ROW8 MA
ROW9 MA
ROW10 MA
TR1 MA
TR2 MA
TR3 MA
TR4 MA
TR5 MA
TR6 MA
TR7 MA
TR8 MA
TR9 MA
TR10 MA
NR,NC MB
ROWS MB
COLS MB
ROW1 MB
ROW2 MB
ROW3 MB
ROW4 MB
ROW5 MB
ROW6 MB
ROW7 MB
ROW8 MB
TR1 MB
TR2 MB

```

TR3	MB
TR4	MB
TR5	MB
TR6	MB
TR7	MB
TR8	MB
NR,NC	MC
ROWS	MC
COLS	MC
ROW1	MC
ROW2	MC
ROW3	MC
ROW4	MC
ROW5	MC
ROW6	MC
ROW7	MC
ROW8	MC
ROW9	MC
ROW10	MC
TR1	MC
TR2	MC
TR3	MC
TR4	MC
TR5	MC
TR6	MC
TR7	MC
TR8	MC
TR9	MC
TR10	MC
NR,NC	MF
ROWS	MF
COLS	MF
ROW1	MF
ROW2	MF
ROW3	MF
ROW4	MF
ROW5	MF
ROW6	MF
ROW7	MF
ROW8	MF
ROW9	MF
ROW10	MF
ROW11	MF
ROW12	MF
TR1	MF
TR2	MF
TR3	MF
TR4	MF
TR5	MF
TR6	MF
TR7	MF
TR8	MF
TR9	MF
TR10	MF

4	5	1	2	-1	3	-1	4	-1				
5	1	1	2	-1	3	-1	4	-1	5	-1		
12	11											
23	38	53	54	55	56	58	59	61	60	63	64	
54	55	56	57	58	59	60	61	62	63	64		
-1	1	-1	1	-1	-1	-1	-1	-1	-1	-1	-1	
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1	-1	-1	-1	-1	99	1	1	-1	1	1		
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1	1	-1	99	99	-1	1	-1	1	-1	-1		
1	1	1	1	1	1	1	1	1	1	1		
1	1	1	1	1	-1	1	1	1	1	1		
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1	99	1	1	1	-1	1	1	1	1	1		
3	3	1	1	-1	2	-1						
5	3	1	4	1	5	1	1	-1	2	-1		
2	2	1	1	-1								
2	2	1	1	-1								
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2	2	1	1	-1								
2	2	1	1	-1								
2	2	1	1	-1								

TR11	MF
TR12	MF
NR,NC	FA
ROWS	FA
COLS	FA
ROW1	FA
ROW2	FA
ROW3	FA
ROW4	FA
ROW5	FA
ROW6	FA
ROW7	FA
ROW8	FA
ROW9	FA
ROW10	FA
ROW11	FA
ROW12	FA
TR1	FA
TR2	FA
TR3	FA
TR4	FA
TR5	FA
TR6	FA
TR7	FA
TR8	FA
TR9	FA
TR10	FA
TR11	FA
TR12	FA

Appendix 3

Sample BUG ID Results for Apollo 13

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D0003	032770	1	37	37
D0004	032770	1	2	2
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D0026	032770	2	996	13
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D0058	032770	2		8 11

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13		13
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	7 13 14	7
2		2
	6	6
	7 13 14	7
	6	6
13		13

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D0300	033070	2

	8 11	8
13		13
	3	4
2		2
996		4
	13	13
996		7
13		13
2		2
2		2
	13	13
4		4
13		13
	13	13
	2 4	4
2		2
	13	13
5		5
13		13
	8 11	8

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