

N85-32715

V. Cohen: SULFUR TRANSFORMATIONS AT THE  
HYDROGEN SULFIDE/OXYGEN INTERFACE IN STRATIFIED  
WATERS AND IN CYANOBACTERIAL MATS

Stratified water bodies allow the development of several microbial plates along the water column. The microbial plates develop in relation to nutrient availability, light penetration, and the distribution of oxygen and sulfide. Sulfide is initially produced in the sediment by sulfate-reducing bacteria. It diffuses along the water column creating a zone of hydrogen sulfide/oxygen interface. In the chemocline of Solar Lake (Sinai) oxygen and sulfide coexist in a 0-10 cm layer that moves up and down during a diurnal cycle. The microbial plate at the chemocline is exposed to oxygen and hydrogen sulfide, alternating on a diurnal basis. The cyanobacteria occupying the interface switch from anoxygenic photosynthesis in the morning to oxygenic photosynthesis during the rest of the day. This activity results in a temporal build up of elemental sulfur during the day which disappears at night due to both oxidation to thiosulfate and sulfate by thiobacilli, and reduction to hydrogen sulfide by *Desulfurobacter* sp. and anaerobically respiring cyanobacteria.

High dark CO<sub>2</sub> fixation, which can be stimulated by sulfide, elemental sulfur, and thiosulfate in the presence of oxygen or nitrate, is found in the chemocline. Over 90 percent of the primary production in the stratified Solar Lake occurs under sulfide conditions because of the activities of several cyanobacteria, plates of *Chromatium* sp., and *Prostrebacillus* sp.. These sulfur bacteria exist above the major cyanobacterial plate of *Oscillatoria Izquierdoi* which is found at the deepest part of the hypolimnion. The relative contribution of anoxygenic photosynthesis to overall primary productivity is a function of light penetration to the hydrogen sulfide/oxygen interface layer. When only 1 percent of surface light reaches this layer, anoxygenic photosynthesis accounts for about 5 percent of the overall primary productivity whereas if 20 percent of the surface light reaches the chemocline (the case in Solar Lake), anoxygenic photosynthesis accounts for more than 90 percent of the photosynthetic carbon dioxide assimilation.

The study of the hydrogen sulfide/oxygen interface in sediments requires the use of microelectrodes for pO<sub>2</sub>, pH, pS<sup>2-</sup>, pH<sub>2</sub>, and pCO<sub>2</sub>. These electrodes, now used in several laboratories, were introduced to microbial ecology by N.P. Revsbech of Aarhus University in Denmark. Sharp gradients of all these redox parameters are observed in microscale (the top 1-10 mm of the sediment column). These result from intense microbial activities in this thin photic zone.

Diurnal fluctuations at the hydrogen sulfide/oxygen interface in sediment are much more pronounced than those of stratified water bodies, since the established gradients in sediment are very steep and diffusion in these dimensions is very fast. Because of this close

proximity, the diurnal fluctuations expose cyanobacteria to sulfide at night and sulfate-reducing bacteria to high concentrations of oxygen during the day.

The cyanobacteria cope with exposure to sulfide either by carrying out facultative anoxygenic photosynthesis or by performing oxygenic photosynthesis in the presence of sulfide. When pH<sub>2</sub> electrodes were introduced to the Fmax zone, a transient peak of H<sub>2</sub> was observed upon turning on the light, possibly indicating photolysis of water by cyanobacteria under these conditions. *Oscillatoria limnetica* were shown to produce H<sub>2</sub> in a CO<sub>2</sub> limited environment under both aerobic and anaerobic conditions.

Sulfate reduction was found to be enhanced in the light at the surface of the cyanobacterial mats. Microsulfate reduction measurements showed enhanced activity of sulfate reduction even under high oxygen concentrations of 300-800 μM. Apparent aerobic SO<sub>4</sub> reduction activity can be explained by the co-occurrence of H<sub>2</sub>. The physiology of this apparent sulfate reduction activity is currently being studied.

**Cohen, Y..** 1984. The Solar Lake cyanobacterial mats: Strategies of photosynthetic life under sulfide. In *Microbial Mats: Stromatolites*. (Y. Cohen, R.W. Castenholz, and H.O. Halvorson eds.), Alan R. Liss, New York, pp. 133-148.

**Cohen, Y.,** 1984. Microtechnique for in situ sulfate reduction measurement in proximity to oxygen. *Arch. Microbiol.*, (in press).

**Cohen, Y.,** 1984. Sulfate reduction under oxygen in cyanobacterial mats and its coupling to primary production. *Limnol. Oceanogr.*, (in press).

**Cohen, Y., Aizenstat, Z., Stoler, A., and Jørgensen, B.B..** 1986. Microbial geochemistry of Solar Lake Sinai. In *Biogeochemistry of Ancient and Modern Environments*. (U.N. Trudinger and M.R. Walter, eds.), Australian Academy of Science, Canberra, pp. 167-177.

**Cohen, Y. and Gack, E..** 1984. Fe<sup>2+</sup> dependent photosynthesis in cyanobacteria. *Nature*, (in press).

**Cohen, Y., Padan, E., and Shilo, M..** 1975a. Facultative anoxygenic photosynthesis in the cyanobacterium *Oscillatoria limnetica*. *J. Bacteriol.*, 127:855-861.

- Cohen, Y., Jørgensen, B.B., Padan, E., and Shilo, M.** 1975b. Sulphide-dependent anoxygenic photosynthesis in the cyanobacterium *Oscillatoria limnetica*. *Nature*, 257:489-491.
- Cohen, Y., Goldberg, M., Krumbein, W.E., and Shilo, M.** 1977a. Solar Lake (Sinai). I. Physical and chemical limnology, *Limnol. Oceanogr.*, 22:597-607.
- Cohen, Y., Krumbein, W.E., and Shilo, M.** 1977b. Solar Lake (Sinai). 2. Distribution of photosynthetic microorganisms and primary production, *Limnol. Oceanogr.*, 22:609-610.
- Jørgensen, B.B., Revsbech, N.P., and Cohen, Y.** 1983. Photosynthesis and structure of benthic microbial mats: Microelectrode and SEM studies of four cyanobacterial communities, *Limnol. Oceanogr.*, 28:1075-1093.
- Jørgensen, B.B., Revsbech, N.P., Blackburn, T.H., and Cohen, Y.** 1979. Diurnal cycle of oxygen and sulfide microgradients and microbial photosynthesis in a cyanobacterial mat sediment, *Appl. Environ. Microbiol.*, 38:46-58.
- Oren, A., Padan, E., and Malkin, S.** 1979. Sulfide inhibition of photosystem II in cyanobacteria (blue green algae) and tobacco chloroplasts, *Biochem. Biophys. Acta*, 546:270-279.
- Oren, A. and Shilo, M.** 1979. Anaerobic heterotrophic dark metabolism in the cyanobacterium *Oscillatoria limnetica*: sulfur respiration and lactate fermentation, *Arch. Microbiol.*, 122:77-84.
- Padan, E. and Cohen, Y.** 1982. Anoxygenic photosynthesis. In *The Biology of Cyanobacteria*, (M.C. Carr and R.A. Whitton, eds.), Blackwell Scientific, Oxford, pp. 215-235.
- Revsbech, N.P., Jørgensen, B.B., Blackburn, T.H., and Cohen, Y.** 1983. Microelectrode studies of the photosynthetic and O<sub>2</sub>, H<sub>2</sub>S and pH profiles of a microbial mat, *Limnol. Oceanogr.*, 28:1062-1074.
- Skyring, F.W.**, 1984. Sulfate reduction in marine sediments associated with cyanobacterial mats, Australia. In *Microbial Mats: Stromatolites*, (Y. Cohen, R.W. Castenholz, and H.O. Halvorson eds.), Alan R. Liss, New York, pp. 265-276.