

CALMODULIN-DEPENDENT PROTEIN KINASE MEDIATES HYPERGRAVITY-INDUCED CHANGES IN F-ACTIN EXPRESSION BY ENDOTHELIAL CELLS,

**Felisha D. Love[‡], Caroline Melhado[‡], Francis Bosah[‡],
Sandra A. Harris-Hooker[‡] and Gary J. Sanford[‡]**

Departments of Biochemistry[‡] and Medicine[‡]

Morehouse School of Medicine, Atlanta, GA 30310

INTRODUCTION: A number of basic cellular functions, e.g., **electrolyte** concentration cell growth rate, glucose **utilization**, bone **formation**, response to **growth** stimulation and **exocytosis** are modified by **microgravity** or during **spaceflight** (1-4). Studies with intact animal during **spaceflights** have found lipid accumulations within the lumen of the **vasculature** and degeneration of the vascular **wall** (5). Capillary alterations with extensive **endothelial invaginations** were also seen (6). **Hemodynamic** studies have shown that there is a redistribution of **blood** from the **lower extremities** to the upper part of the **body**; this will alter vascular permeability, resulting in leakage into surrounding tissues (7). These studies indicate that changes in gravity will affect a number of physiological systems, including the **vasculature**. However, few studies have addressed the effect of **microgravity** on **vascular cell** function and metabolism. A major problem with ground based studies is that achieving a true **microgravity** environment for prolonged period is not possible. On the other **hand**, increasing gravity (i.e., **hypergravity**) is easily achieved. Several researchers have shown that **hypergravity** will increase the proliferation of several different cell lines (e.g., chick embryo **fibroblasts**) while decreasing cell motility (8) and slowing liver regeneration following partial **hepatectomy** (9). These studies suggest that **hypergravity** will alter the behavior of most cells. Several investigators have shown that **hypergravity** affects the expression of the early response genes (**c-fos** and **c-myc**) and the activation of several protein **kinases** (PK's) in cells (10,11). In this study we investigated whether **hypergravity** alters the expression of **f-actin** by aortic endothelial cells, and the possible role of protein **kinases** (**calmodulin**(II)-dependent and **PKA**) as mediators of these effects.

EXPERIMENTAL METHODS: BAECs were obtained from NIGMS/Coriell Cell Repository. Cells were maintained in **Dulbecco's** Modified Eagle's Medium supplemented with 10% fetal bovine serum and 1X antibiotics. We assessed **F-actin** expression in BAECs subjected to **centrifugation** in the presence or absence of protein **kinase** inhibitors (PKIs).

F-actin Expression When Subjected to Hypergravity: Subconfluent cultures were subjected to **hypergravity** (centrifuged at 6 and 12 G) for 24, 48 and 72 hr. The **cultures** were rinsed with phosphate buffered saline (PBS) and then fixed with 1% **glutaraldehyde**/0.1% **Triton X-100**/2.5% **Formalin** in PBS for 24 hours. **Following** fixation, the cells were stained with **FITC-phalloidin** and examined using phase and fluorescence microscopy. Controls were **treated** similarly except they were not subjected to **centrifugation**.

Effect of PKIs on F-actin Expression Under Hypergravity: The PKIs, **KT5926** and **KT5720**, were **added**, singly, to the medium of **subconfluent** cultures (2 μ L PKI/mL) and **centrifuged** at 6 and 12 G for 24, 48, and 72 hr. Cultures were stained and examined for **f-actin** expression as previously outlined. Controls were treated similarly except they were not subjected to **centrifugation**.

RESULTS AND DISCUSSION: Figure 1 shows **subconfluent** BAECs that were subjected to **hypergravity** (12G). The cells generally showed increased **F-actin** expression throughout the **cell** as a function of time under **hypergravity**. Optimal fluorescence was observed **after** 48 hr with a slight **decline** seen after 72 hr. Cells subjected to **6G** of **hypergravity** gave similar results. The controls, however, generally showed a decreased **F-actin** expression with increasing time of culture, with no fluorescence observed **after** 72 hr. The fluorescence observed for control cells

was localized to the nuclear region instead of throughout the cell as observed for hypergravity treated cells. There were no other morphological differences seen for either hypergravity-treated or control cells when examined by phase contrast microscopy. In this study, we only examine changes in f-actin. These findings suggest that hypergravity has a direct effect on f-actin levels in endothelial cells but could also affect other forms of actin. The latter possibility will be investigated in future studies.

Subconfluent BAECs subjected to hypergravity in the presence of the PKI - KT5926, which is a specific inhibitor of calmodulin(II)-dependent protein kinase, exhibited increased f-actin expression with increasing time under 6G is shown in Figure 2. Similar results were observed for cells subjected to 12G hypergravity. The fluorescence, observed under either 6 or 12G, was especially noted at cell-to-cell junctions as well as for the nuclear regions. The controls for PKI-KT5926 showed a slight increase in f-actin expression with increasing time under 6G. The protein kinase inhibitor KT5720 did not affect f-actin expression by BAEC under any of the conditions studied. Again, no other morphological changes were observed when examined by phase contrast microscopy. These results indicate that, of the two protein kinases examined, only the calmodulin(II)-dependent protein kinase has a mediator roles for cells subjected to hypergravity. Since calmodulin(II)-dependent protein kinase is activated normally by either increased intracellular Ca^{+2} concentration or inositol-3-phosphate, these may also mediate hypergravity-induced cellular changes. However, with the complexity for signal transduction mechanisms, it is not possible with the present study, to venture farther in possible signaling events.

ACKNOWLEDGEMENTS: This study was supported by grants NASA NAG9-644 and NCCW-008.

REFERENCES

1. Rijken PJ, et al. (1991). *Aviat Space Environ Med* 62:32-36.
2. Kumei Y, et al. (1989). *J Cell Sci* 93:221-226.
3. Cogoli A, Tschogg A and Fuchs-Bislin P (1984). *Science* 225:228-230.
4. Gruener R and Hoeger G (1990). *Am. J Physiol* 258: C489-C494.
5. Doty S, E Holton, G Durnova and A Kaplansky. (1990) *FASEB J* 4:16.
6. Philpott D, I Popova, K Kate, et al. (1990) *FASEB J* 4:73.
7. Nixon J, R Murray, C Byrant, et al. (1979). *J Appl Physiol* 46:541.
8. Tschopp A and Cogoli A, *Experientia* 39, 1323-1329, 1983
9. Kropacova K, et al., *The Physiologists* 31, S75-S76, 1988
10. Nose K and Shibamura M, *Exp. Cell Res.* 211:168-70, 1994
11. DeGroot KP, et al., *Exp. Cell Res.* 197:87-90, 1991.

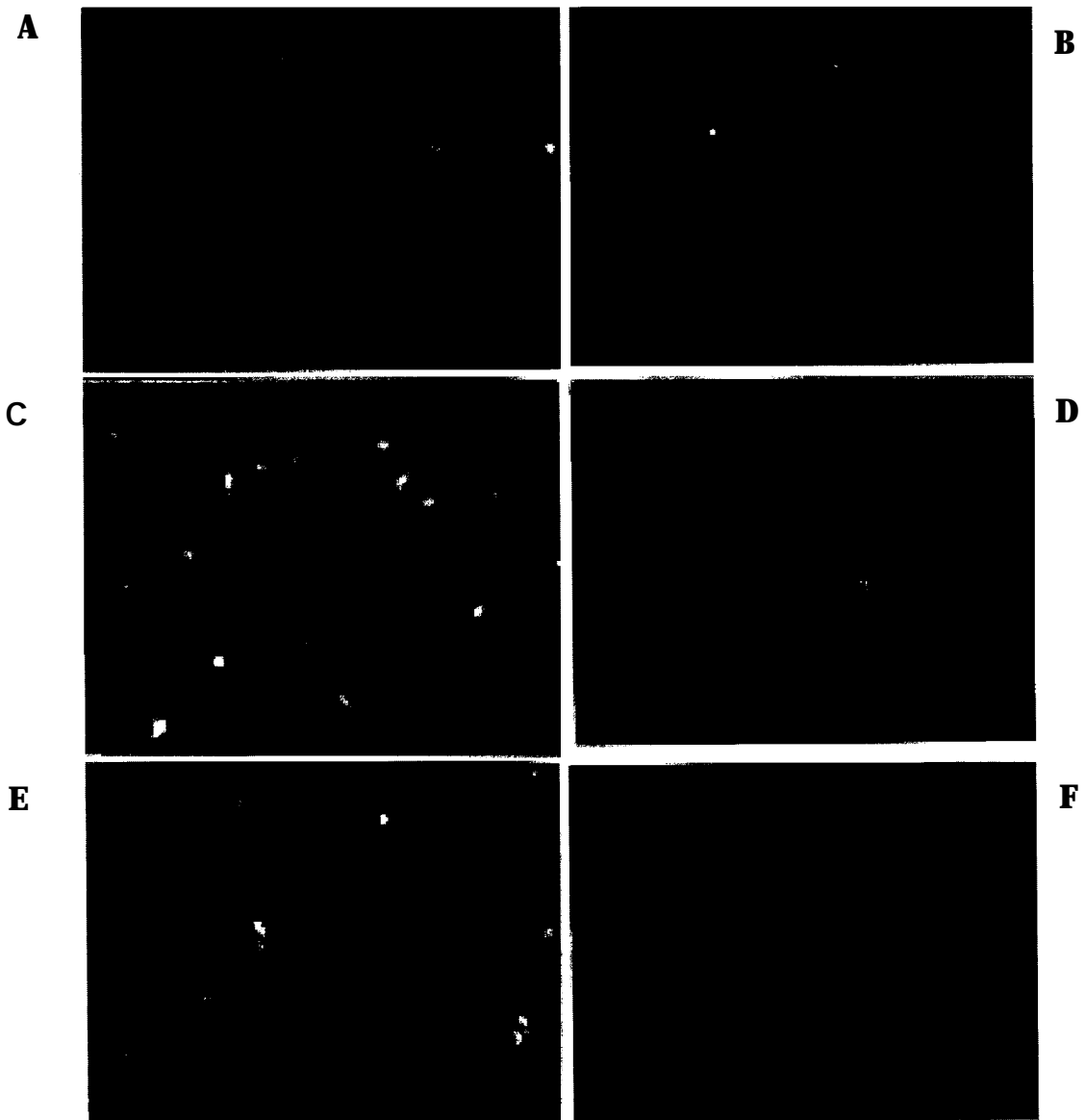


Figure 1. Fluorescence micrographs of BAEC after 24, 48 and 72 hr under control (A, C & E) and 12G hypergravity (B, D & F) conditions. Cultures are stained for f-actin using FITC-labeled phalloidin.

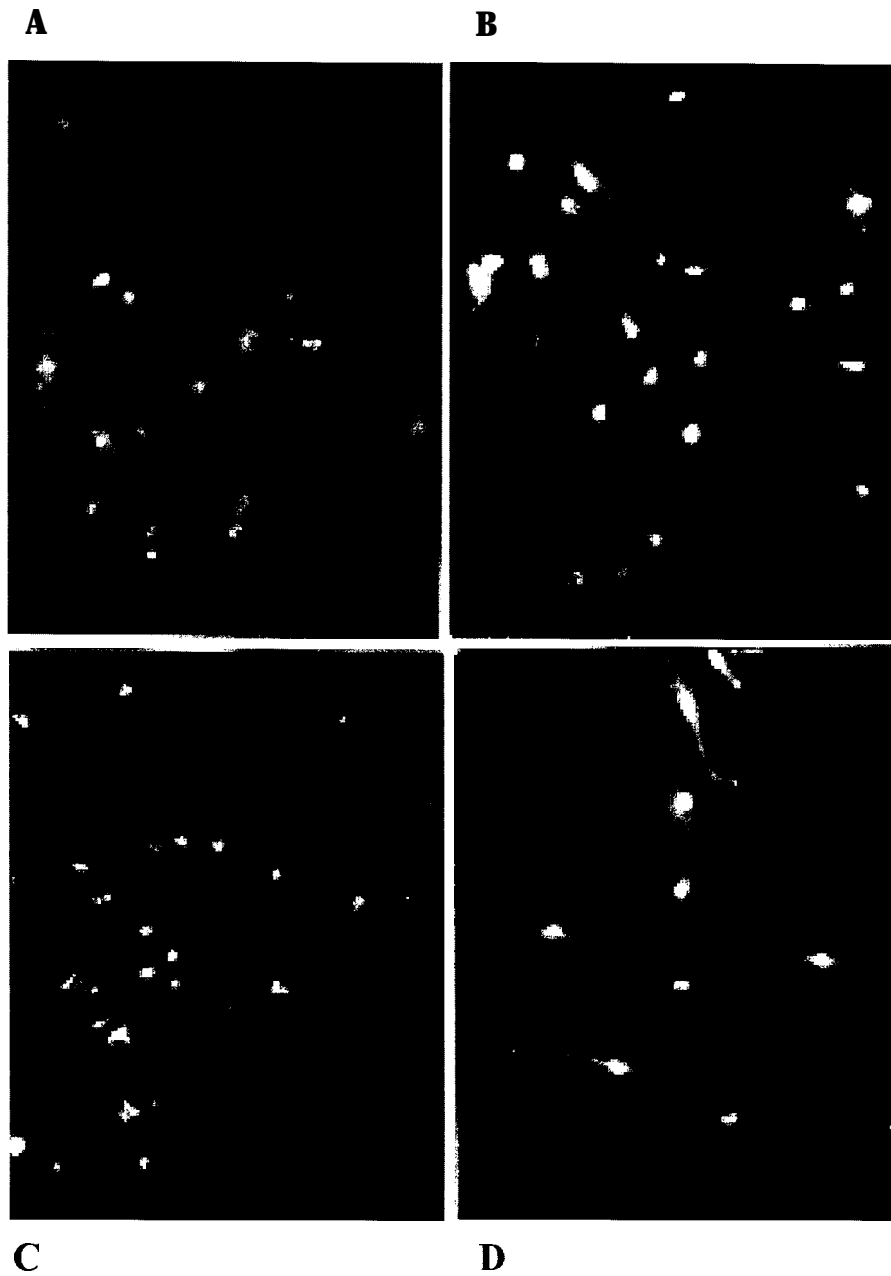


Figure 2. Fluorescence micrographs of BAEC treated with PKIKT5926 (for calmodulin(II)-dependent protein kinase) after 24 and 48 hr under control (A & C) and 6G hypergravity (B & D) conditions. Cultures are stained for f-actin using FITC-labeled phalloidin.