

URC97152

**Hypergravity Alters the Susceptibility of Cells to
Anoxia-Reoxygenation Injury**

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INTRODUCTION: Gravity is a physical force, much like shear stress or mechanical stretch, and should affect organ and cellular **function**. Researchers have shown that gravity **plays** a role in ventilation and blood flow distribution, gas exchange, alveolar size and mechanical stresses within the lung (1-3). Short exposure to **microgravity** produced marked alterations in lung blood flow and ventilation distribution while **hypergravity** exaggerated the regional differences in lung structure and **function** resulting in reduced ventilation at the base and no ventilation of the upper half of the lung (4). **Microgravity** also decreased metabolic activity in cardiac cells, **WI-38** embryonic lung cells, and human lymphocytes (5). Rats, in the tail-suspended head-down tilt model, experienced transient loss of lung water (6), contrary to an expected increase due to pooling of blood in the pulmonary **vasculature**. **Hypergravity** has also been found to increase the proliferation of several different cell lines (e.g., chick embryo **fibroblasts**) while decreasing cell motility (7) and slowing liver regeneration following partial **hepatectomy** (8). These studies show that changes in the gravity environment will affect several aspects of organ and cellular **function** and produce major change in blood flow and tissue/organ **perfusion**. However, these past studies have not addressed whether **ischemia-reperfusion** injury will be exacerbated or ameliorated by changes in the gravity environment, e.g., space flight. Currently, nothing is known about how gravity will affect the susceptibility of different lung and vascular **cells** to this type of injury. We conducted studies that addressed the following question: Does the susceptibility of lung **fibroblasts**, vascular smooth muscle and **endothelial cells** to **anoxia/reoxygenation** injury change following exposure to **hypergravity** conditions?

EXPERIMENTAL METHODS: Bovine aorta **endothelial (BAEC)** and primate smooth muscle(**SMC**) were obtained from the **NIGMS/Coriell Cell Repository**. Rat lung **fibroblasts (RFL)** were isolated from adult Sprague **Dawley** rats as previously reported (9). All cell lines were maintained in **DMEM** containing 10% fetal bovine serum. Confluent cultures of each **cell** line were subjected to centrifugation at **6G** for 24-48 hrs. Control cultures were not centrifuged or rotated. Cells were then placed under **anoxia** (5% CO_2 /balance N_2) for 2 hr with or without a 1 hr period of **reoxygenation**. The change in viable cell numbers was assessed by: measuring viable cells by **hemacytometer** counting of **trypan blue** stained **cells**, or using **MTT** assay for viable cells (**microtiter** plate assay). The effect of **hypergravity** on the expression of heat shock protein (**HSP60**) by **RFL** was evaluated by **immunocytochemical** staining using a **FITC-labeled** monoclonal antibodies (**Stress Gene**). **Subconfluent** cultures were fixed after 12 and 24 hr under **hypergravity** using 10%**formalin**, 0.1 % **triton X-100** in **PBS**. Cultures were incubated with the primary antibody for 1 hr, washed **3X** with **PBS**, incubated with **biotin** labeled anti **IgG** secondary antibody for 1 hr and subsequently stained with **fluorescein** conjugated **streptavidin**. Cells were viewed by fluorescence microscopy and photographed. Controls, stained with non-immune mouse **IgG**, showed no fluorescence under these conditions.

RESULTS AND DISCUSSION: Figure 1 shows the change in viable cell count, expressed as percent of controls, for **SMC** and **BAEC** as a **function** of time under **6G**. The solid line at 100% represents the level for cultures that were not treated with **anoxia** or **anoxia-reoxygenation**. **SMC** were found to have a 60% decrease in viable cells after 48 hr of **hypergravity** and 2-hr of **anoxia**. In the first 24 hr of **hypergravity**, the acute response of **SMC** seems to be an exacerbated injury under **anoxia** and **reoxygenation**. This acute phase is followed in the next 24 hr by a reversal of the exacerbated injury as cells adapt to **hypergravity**.

The results found for **BAEC** show a different pattern. Control (non-centrifuged) cells had decreased viability under **anoxia** with only a slight further decrease during the **reoxygenation** period. With **BAEC**, the acute response (first 24 hr) to **hypergravity** is an increase in viable cells under **anoxia-reoxygenation**. As seen with **SMC**, there is a reversal of this increase in the following 24 hr period

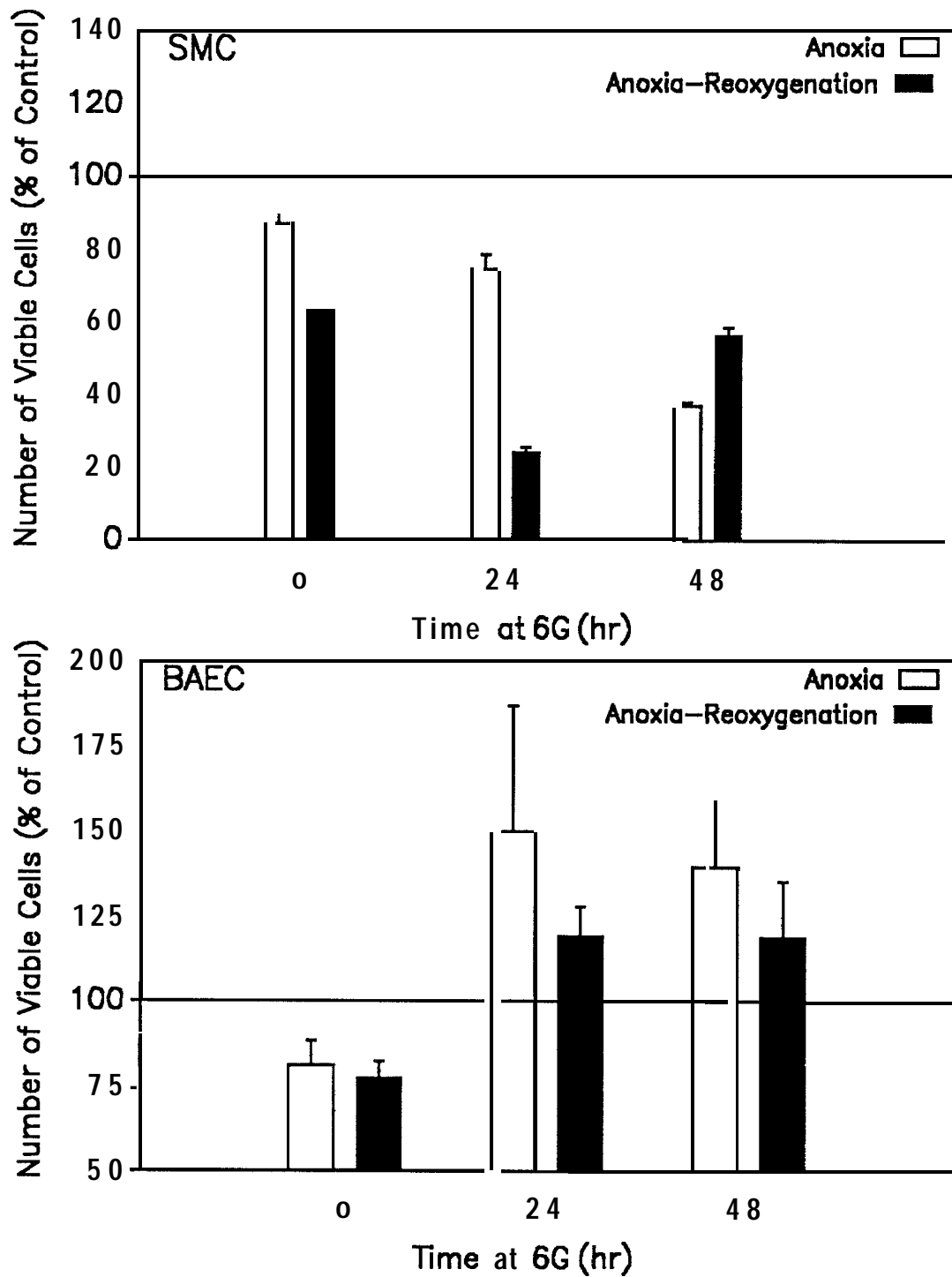


FIGURE 1. The response of SMC and BAEC to anoxia or anoxia-reperfusion injury following hypergravity treatment for 24-48 hr. Data is plotted as mean of the % of matched untreated (controls) normogravity or hypergravity cultures.

as the cells adapt to hypergravity. Although it is expected that more cell damage and loss of viability would result from the combined effect of anoxia and reoxygenation, we found that 2 hr of anoxia results in similar damage. Hypergravity worsened the damage for SMC but decreased the damage for BAEC, suggesting that protective mechanisms maybe differentially activated in vascular cells.

Figure 2 shows that results similar to those seen with BAEC were also found for RFL whether these cells were maintained in DMEM or PBS. When maintained in complete media (DMEM), these cells show an increase in viable cells with time in hypergravity when treated with anoxia alone. However, RFL maintained in PBS show a steady decrease with time in hypergravity. RFL behave similar to BAEC when placed under anoxia-reoxygenation. The acute response of RFL to hypergravity is a transient decrease in susceptibility to damage, i.e., increased viable cell counts, under anoxia-reoxygenation. Again this is followed by a reversal in the following 24 hr period as the cells adapt to hypergravity. This pattern of response was also found for RFL maintained in PBS.

When BAEC and RFL have apparently adapted to hypergravity, both cell lines are not readily injured by anoxia-reoxygenation. In fact, RFL maintained in PBS, which has no energy source (e.g., glucose) was not found to have the maximum injury under hypergravity as expected. Under these conditions RFL responded similarly to cells in complete media. These results suggest that hypergravity may increase protective mechanisms in these cells that ameliorate possible damage from anoxia-reoxygenation. One possible protective mechanism that maybe induced by hypergravity is increased expression of stress proteins like heat shock proteins. Heat shock proteins have been shown to protect cells from damage and death under a number of different stresses, e.g., increased temperature or shear stress (10). We tested this possibility with RFL, as shown in figure 3. Within 12 hr under hypergravity, RFL's had clearly increased immunofluorescence for heat shock protein, HSP60 (figure 3A) compared to control cells (figure 3 C). By 24 hr, hypergravity resulted in an even more intense immunofluorescence, suggesting that HSP60 expression is stimulated for at least the initial 24 hr period. Preliminary data from similar studies with BAEC (data not shown) indicate that these cells also may increase their expression of heat shock proteins within the first 24 hr period of hypergravity. Further studies are planned to examine the possible mediation of hypergravity effects on cells by the heat shock protein family.

ACKNOWLEDGEMENTS: These studies were supported by grants from NASA NAG9-644 and NCCW-0085.

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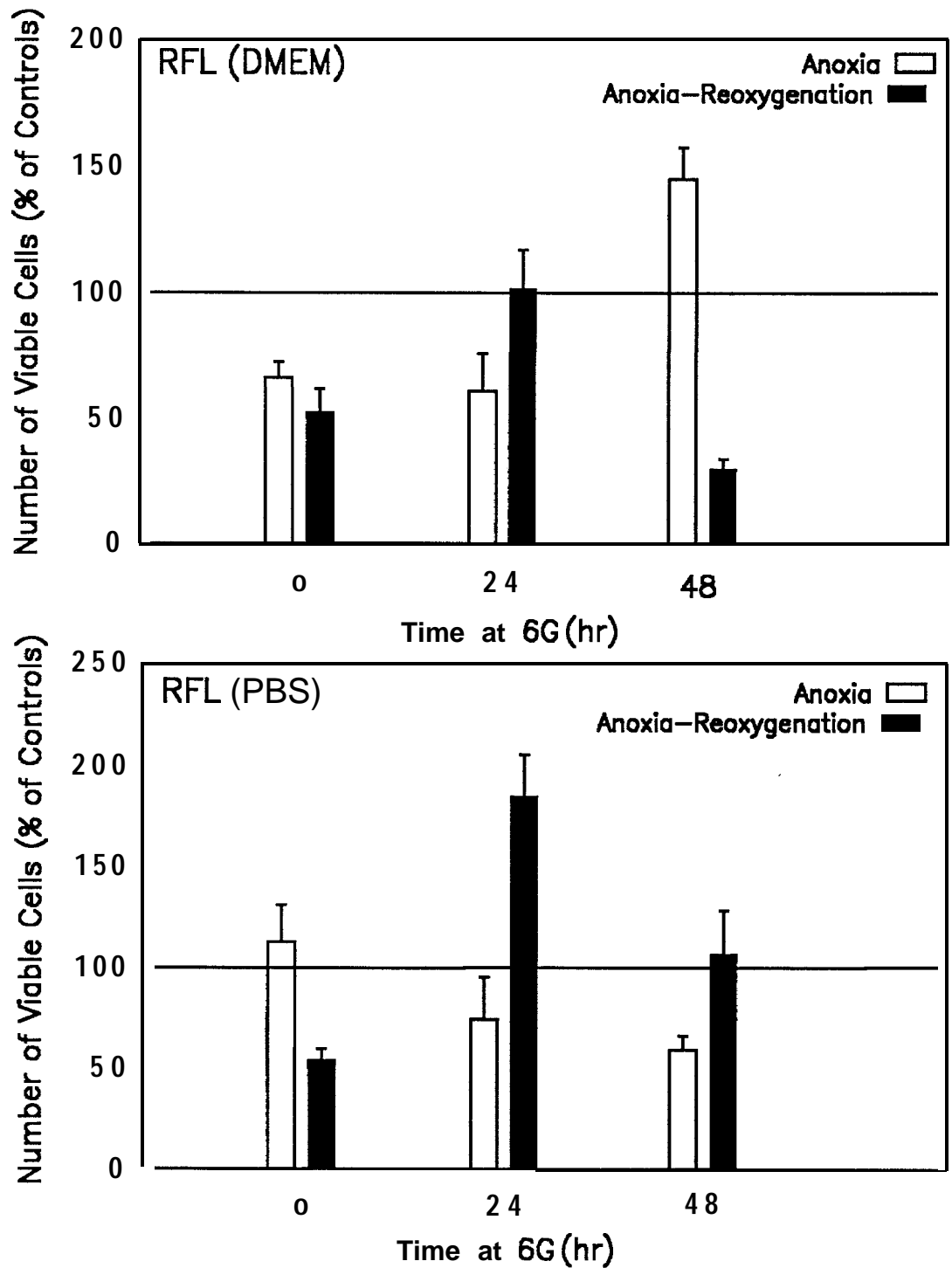
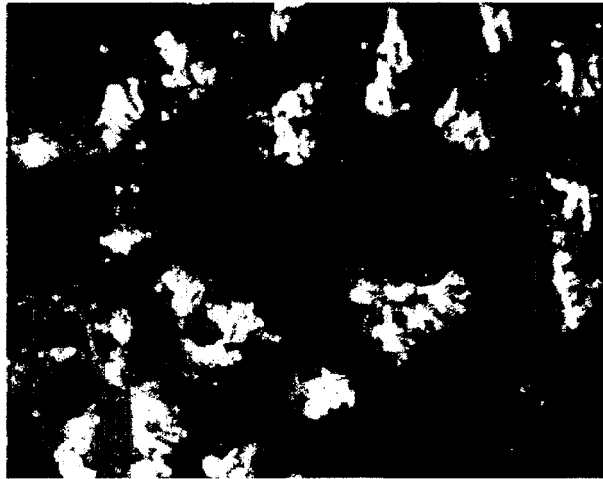
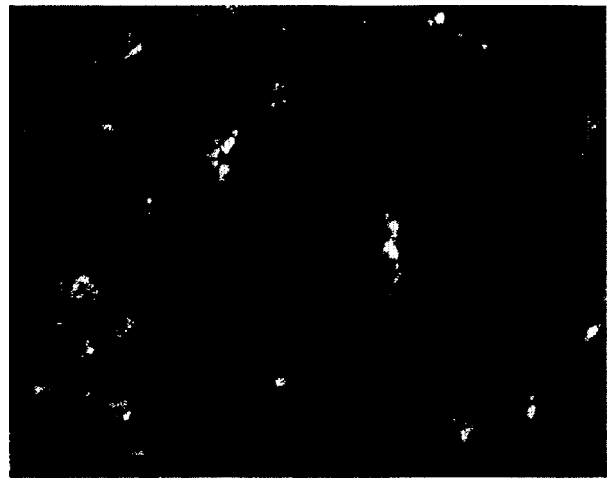


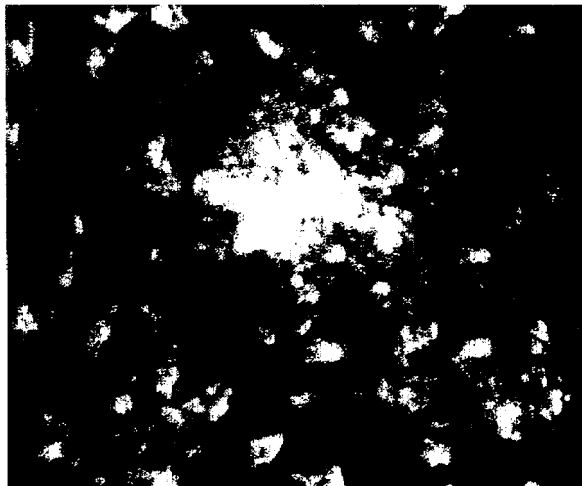
FIGURE 2. The response of RFL, maintained in complete media (DMEM) or in PBS, to anoxia or anoxia-reperfusion injury following hypergravity treatment for 24-48 hr. Data is plotted as mean of the 1% of matched untreated (controls) normogravity or hypergravity cultures.



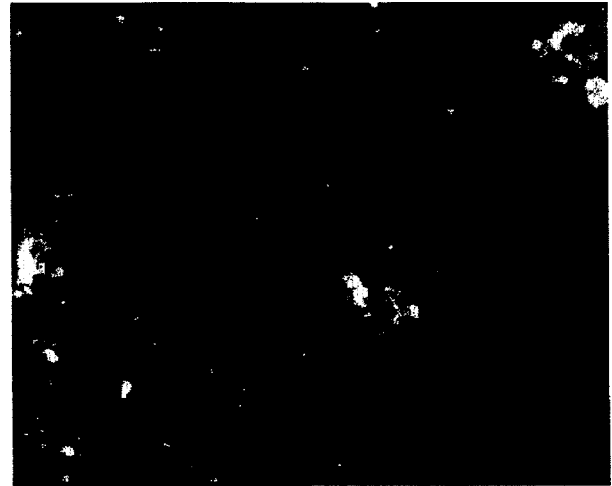
A



C



B



D

Figure 3. Expression of heat shock protein (HSP60) by RFL subjected to 12 and 24 hr of hypergravity (A & B) or normogravity (C & D).