

**Plant productivity and characterization of zeoponic substrates after three successive crops of radish (Raphanus sativus L.)**

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## 1   **Abstract**

2   The National Aeronautics and Space Administration (NASA) has developed a zeolite-  
3   based synthetic substrate, termed zeaponics. The zeoponic substrate (consisting of  
4   NH<sub>4</sub>- and K-exchanged clinoptilolite, synthetic apatite, and dolomite) provides all of  
5   the plant-essential nutrients through mineral dissolution and ion exchange, with only  
6   the addition of water. Previous studies have shown high productivity of wheat in  
7   zeoponic substrates; however, no experiments have been conducted on other crops.  
8   The objective of this study was to determine the productivity and nutrient uptake of  
9   radish (Raphanus sativus L.) grown in zeoponic substrates with three successive crops  
10   in the same substrate. Radish was chosen because of its sensitivities to NH<sub>4</sub><sup>+</sup>.  
11   Average fresh weights of edible roots were similar for radish grown in zeoponic  
12   substrates watered with deionized H<sub>2</sub>O (10.97 g/plant) and in potting mix control  
13   substrate irrigated with nutrient solution (10.92 g/plant). Average fresh weight  
14   production of edible roots for radish grown in same zeoponic substrate increased in  
15   yield over time with the lowest yield in the first crop (7.10 g/plant) and highest in the  
16   third crop (13.90 g/plant). The Ca plant tissue levels in radishes (1.8-2.9 wt. %)   
17   grown in zeoponic substrates are lower than the suggested sufficient range of 3.0-4.5  
18   wt. % Ca; however, the Ca level is highest (2.9 wt. %) in radishes grown in the third  
19   crop in the same zeoponic substrates. The higher radish yield in the third crop was  
20   attributed to a reduction in an NH<sub>4</sub>-induced Ca deficiency that has been previously  
21   described for wheat grown in zeoponic substrates. The P levels in plant tissues of  
22   radish grown in the zeoponic substrates ranged from 0.94-1.15 wt. %; which is  
23   slightly higher than the sufficient levels of 0.3-0.7 wt. %. With the exception of Ca  
24   and P, other macronutrient and micronutrient levels in radish grown in zeoponic  
25   substrates were well within the recommended sufficient ranges. After three successive

26 crops of radish growth, the zeoponic substrates had 52% of the original  $\text{NH}_4\text{-N}$  and  
27 78% of the original K remaining on zeolite exchange sites. Zeoponic substrates are  
28 capable of long-term productivity of radishes for space.

29

## 30 **Keywords**

31 Zeolite, Zeoponics, Clinoptilolite, Apatite, Radish

32

## 33 **1. Introduction**

34 On January 14, 2004, President Bush provided a strategic vision of space exploration  
35 beyond earth orbit to the National Aeronautics and Space Administration (NASA) and  
36 the American people. These long-duration missions to the Moon, Mars, and beyond  
37 will require life support systems that recycle and regenerate air, water, and food.  
38 Plants may be used in regenerative life-support systems to sequester carbon dioxide  
39 and generate oxygen through photosynthesis, produce potable water through  
40 transpiration, and produce food through plant growth and reproduction [1]. NASA's  
41 Johnson Space Center (JSC) has developed a zeolite-based synthetic substrate  
42 consisting of clinoptilolite, synthetic apatite, and dolomite to provide plant-essential  
43 nutrients, pH-buffer, aeration, moisture retention, and mechanical support for plant  
44 growth in regenerative life-support systems [2].

45

46 Zeoponic plant-growth systems have been defined as the cultivation of plants in  
47 artificial soils, which have zeolite as a major component [3]. In the JSC zeoponic  
48 substrate, the native clinoptilolite cations are exchanged for  $\text{NH}_4^+$  and  $\text{K}^+$  cations [4],  
49 which then become the primary N and K sources for plant nutrition. The dynamic  
50 equilibria that takes place between zeolite, apatite, and dolomite in a zeoponic system

51 are discussed in detail by Lai and Eberl [5], Ming and Allen [6,7], and Beiersdorfer et  
52 al. [8]. In brief, dissolution of the apatite and dolomite supplies  $\text{Ca}^{2+}$  to the soil  
53 solution. Through ion-exchange reactions, solution  $\text{Ca}^{2+}$  will remove  $\text{NH}_4^+$  and  $\text{K}^+$   
54 from zeolitic exchange sites, thus making the  $\text{NH}_4^+$  and  $\text{K}^+$  cations available for plant  
55 uptake. The goal of the zeoponic research efforts at JSC is to develop a solid  
56 substrate that will slowly deliver most of the plant-essential macronutrients (N, P, K,  
57 Ca, Mg, and S) and all of the plant-essential micronutrients (Zn, Fe, Cu, Mn, B, Mo,  
58 and Cl) for many growth seasons, with the remainder of the required macronutrients  
59 (H, O, and C) being supplied by the water and air used throughout a space habitat  
60 (Figure 1).

61

62 Initial plant-growth experiments with wheat grown in zeoponic substrates resulted in  
63 excellent vegetative growth [2]. However, when wheat plants were grown to maturity  
64 in another experiment with zeoponic substrates, seed yield was poor [9]. Plant-tissue  
65 analyses suggested that high P and low Ca concentrations were possible reasons for  
66 the low yield [9]. An additional experiment with wheat investigating the effects of  
67 adding nitrifying bacteria, dolomite and ferrihydrite to the zeoponic substrate resulted  
68 in better yields [10]; however only a cumulative effect was reported and it is not clear  
69 which additive had the most effect. Beiersdorfer et al. [8] and Gruener et al. [11]  
70 described a common-ion effect that occurs in zeoponic substrates amended with Ca-  
71 bearing minerals, where the solubility of the synthetic apatite is diminished by the  
72 presence of  $\text{Ca}^{2+}$  in solution from the dissolution of other Ca-bearing minerals. This  
73 resulted in higher  $\text{Ca}^{2+}$  and lower P in solution. McGilloway et al. [12] demonstrated  
74 the establishment of nitrifying bacteria populations in zeoponic substrates, using  
75 Nitrosomonas sp. and Nitrobacter sp. bacteria, and the successful oxidization of  $\text{NH}_4^+$

76 to  $\text{NO}_2^-$  and then to  $\text{NO}_3^-$  during a plant growth experiment with radish. Ammonium  
77 oxidation rates for the three week experiment ranged from 1.2 to 1.78  $\mu\text{g N g}^{-1}$   
78 substrate  $\text{h}^{-1}$  and  $\text{NO}_2^-$  oxidation rates were 0.38 to 0.74  $\mu\text{g N g}^{-1}$  substrate  $\text{h}^{-1}$ .

79

80 The purpose of the present paper is to expand upon previous zeoponic plant growth  
81 studies and report on a plant-growth experiment investigating the response of radish  
82 plants grown in clinoptilolite-apatite-dolomite substrates. Goyal et al. [13] have  
83 shown that growth of radish is inhibited when  $\text{NH}_4^+$  is the sole source of N. Addition  
84 of  $\text{NO}_3^-$  equivalent to 10% or more of the  $\text{NH}_4^+$  concentration alleviated the inhibitor  
85 effects, and  $\text{NO}_3^-$  apparently facilitated the assimilation of  $\text{NH}_4^+$  by radish plants.  
86 Fenn and Taylor [14] demonstrated the use of  $\text{Ca}^{2+}$  as a means to increase the rate of  
87  $\text{NH}_4^+$  absorption by radish. Vegetative growth increased in two greenhouse  
88 experiments as  $\text{Ca}^{2+}$  concentrations increased, all at a constant  $\text{NH}_4^+$  concentration.

89

90 The first objective of this study, was to use the growth of radish plants, which are  
91 extremely sensitive to  $\text{NH}_4^+$  nutrition, as an indicator of the effectiveness of the  
92 addition of dolomite to create a common ion effect and increase the amount of  $\text{Ca}^{2+}$  in  
93 soil solution, and the addition of nitrifying bacteria to zeoponic substrates to oxidize  
94  $\text{NH}_4^+$  to  $\text{NO}_3^-$ . The second objective of this study was to determine the nutrient  
95 delivery availability of the zeoponic substrate after three successive radish crops.

96

## 97 **2. Experimental**

### 98 **2.1 Materials**

99 Clinoptilolite-rich tuff (Cp) from the Fort LeClède deposit, Sweetwater County,  
100 Wyoming and a synthetic hydroxyapatite produced in the laboratory were used as the

101 primary components of the zeoponic substrate. The Fort LeClède clinoptilolite is  
102 nearly monomineralic as determined by X-ray diffraction analysis [15], with only  
103 trace amounts of quartz present. The Cp was transformed into either an ammonium-  
104 exchanged (NH<sub>4</sub>-Cp) or a potassium-exchanged (K-Cp) form, using the method of  
105 Allen et al. [4]. The NH<sub>4</sub><sup>+</sup> and K<sup>+</sup> cation-exchange capacity (CEC) of the  
106 clinoptilolite-rich tuff were 207 cmol<sub>(c)</sub>/kg and 202 cmol<sub>(c)</sub>/kg, respectively,  
107 determined by a CsCl method described by Ming and Dixon [16].

108

109 The synthetic apatite (SA) is an agronutrient-substituted hydroxyapatite, produced in  
110 the laboratory according to the method of Golden and Ming [17]. In addition to Ca  
111 and PO<sub>4</sub>, the SA has Mg, SO<sub>4</sub>, and micronutrients (i.e., Zn, Fe, Cu, Mn, B, Mo, and  
112 Cl) incorporated in its structure [18,19,20,21]. The chemical composition of the SA is  
113 shown in Table 1.

114

115 Dolomite (Baker Grandol Regular #4, Baker Refractories, York, Pennsylvania, U. S.  
116 A.) was added as a pH buffer and additional Ca and Mg source for plant growth  
117 (Table 1). All materials (i.e., NH<sub>4</sub>-Cp, K-Cp, SA, dolomite) were crushed and sieved  
118 to a particle size of 0.5-1.0 mm.

119

## 120 2.2. Methods

121 The clinoptilolite (Cp), synthetic apatite (SA), and dolomite were combined to form  
122 the zeoponic substrate, consisting of 36 wt. % NH<sub>4</sub>-Cp; 36 wt. % K-Cp; 18 wt. % SA  
123 and 10 wt. % dolomite. Two control substrates were also developed: a potting mix  
124 control composed of peat, vermiculite, and perlite in a 1:1:1 volume ratio, and a K-  
125 exchanged Cp (K-Cp) control. The three substrates (i.e., Cp-SA-dolomite or zeoponic

126 substrate, potting mix control, K-Cp control) were placed in pots with dimensions of  
127 9.5 cm in diameter and 8.5 cm deep. The substrate volume in each pot was 440 ml  
128 and the average bulk density of the zeoponic substrates was 1 g/cm<sup>3</sup>. Thus, each pot  
129 contained 440 grams of zeoponic materials, or 440 ml of potting mix. The zeoponic  
130 substrates (i.e., Cp-SA-dolomite substrates) were inoculated with Nitrosomonas sp.  
131 and Nitrobacter sp. bacteria to establish a nitrification process, in which the NH<sub>4</sub><sup>+</sup>  
132 released from the NH<sub>4</sub>-Cp is oxidized to form NO<sub>3</sub><sup>-</sup> (see Henderson et al. [10],  
133 McGilloway et al. [12]).

134  
135 The investigation consisted of 6 different radish crop cycles, each lasting 21 days.  
136 Three radish crops were grown successively in the same zeoponic substrate, and then  
137 the experiment was repeated in time with three more crops in a new zeoponic  
138 substrate. The purpose of the three successive crops in the same substrate was to  
139 evaluate the capability of using zeoponic substrates for extended periods; although it  
140 was not the focus of this study to exhaust the nutrient capability of the zeoponic  
141 substrates. For each crop, there were eight pots per substrate treatment (i.e., zeoponic  
142 substrate, K-Cp control substrate, and potting mix control substrate), for a total of 24  
143 pots per crop. The pots were arranged in a randomized block design. Nine radish  
144 seeds (cv. Cherry Belle) were initially planted in each pot and thinned down to 3  
145 plants per pot 5 days after planting. The zeoponic substrates were watered with  
146 deionized water throughout the experiment, and the control substrates were watered  
147 with 1/2-strength Hoagland's potassium phosphate-based nutrient solution [22].

148  
149 The radish plants were grown in an environmental growth chamber (Environmental  
150 Growth Chambers, Chagrin Falls, Ohio, Model G-15) under the following average

151 daily conditions: 16 hr of daylight, 8 hr of darkness, temperature of 23°C, 65%  
152 relative humidity, and a photosynthetic photon flux that averaged 300  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ .  
153 All treatments were irrigated to excess 6 times daily with an automated watering  
154 system at an average rate of 20 ml/min, for durations of 28 s per irrigation at the  
155 beginning of the experiment up to 48 s per irrigation at peak plant water usage.  
156 Radishes were harvested at 21 days, where fresh weights of shoots, edible storage  
157 roots, and fibrous roots were measured. Oven-dried (70 °C for 72 hr) plant samples  
158 were weighed to determine the dry-matter production. Dried plant samples were  
159 ground to pass a 40 mesh sieve and sent to The University of Wisconsin at Madison  
160 soil and plant analysis laboratory for N, P, K, Ca, Mg, S, Zn, Fe, Cu, Mn, and B plant-  
161 tissue analysis. Macro- and micronutrients were measured by Inductively Coupled  
162 Plasma Optical Emission Spectroscopy (ICP-OES) and Mass Spectroscopy (ICP-MS)  
163 using acid digestion with nitric acid and hydrogen peroxide. Total nitrogen was  
164 measured using a Kjeldahl process with sulfuric acid and a metal catalyst.

165

166 After three successive crops of radish, the zeoponic substrate was dried, and the  
167 exchange cations (i.e.,  $\text{K}^+$  and  $\text{NH}_4^+$ ) on the clinoptilolite were determined by the  
168 method of Ming and Dixon [15]. Briefly, the cations on the exchange sites of  
169 clinoptilolite were determined by replacing them with  $\text{Cs}^+$  (0.5 M CsCl solution);  $\text{K}^+$   
170 and  $\text{NH}_4^+$  were subsequently determined by atomic absorption and ion-selective  
171 electrode techniques, respectively. The pH of the extraction solution was buffered by  
172 the Cp-SA-dolomite substrate ( $\text{pH} \approx 8.0$ ).

173

### 174 **3. Results**

175



### 176 3.1 Fresh weight production

177 Analysis of variance (ANOVA) was used to determine whether statistical differences  
178 exist between substrates on the basis of fresh weight/plant. ANOVA was significant  
179 for the fresh weight/plant (f ratio = 4.39; p = 0.013). Tukey-Kramer multiple  
180 comparison tests [23] were then performed to test the significance for all possible  
181 pairs (a significant F-test is a prerequisite for the Tukey-Kramer multiple  
182 comparison). For this experiment, statistical differences are defined at the 5%  
183 probability level.

184

185 Fresh weights of edible roots were similar for radish grown in zeoponic substrates  
186 watered with deionized H<sub>2</sub>O and in potting mix substrate irrigated with ½-strength  
187 Hoagland's solution (Tables 2 & 3). The average fresh weights/plant for the three  
188 successive crops grown in the zeoponic and potting mix control substrates were not  
189 statistically different based on Tukey-Kramer tests; however, radish fresh weight  
190 grown in the K-Cp control substrate was significantly lower than those grown in the  
191 zeoponic and potting mix substrates. Although the average radish fresh weights of  
192 edible roots for all three crops together were similar for zeoponic and potting mix  
193 substrates (Table 2); there were differences in fresh weights between successive crops  
194 for the zeoponic substrates (i.e., differences between crops 1, 2, and 3, Table 3). Fresh  
195 weight production was lower in the first crop and highest in the third crop for radish  
196 grown in zeoponic substrates, i.e., the third successive radish crop grown in the same  
197 zeoponic substrate. Fresh weight production in potting mix substrates was highest in  
198 the first crop (Table 3). The production of radishes in the K-Cp control substrate was  
199 slightly higher in the second and third crops.

200

### 201 3.2 Harvest index

202 Harvest indexes (i.e., weight of fresh edible radish compared to the fresh weight of  
203 the entire plant) were slightly lower in the zeoponic substrates; although there were no  
204 substantial differences for harvest indexes between the three substrates (Table 4). The  
205 harvest indexes for the second and third crops in the same zeoponic substrates were  
206 higher than the first crop.

207

### 208 3.3 Plant-nutrient analysis

209 Average plant-tissue concentrations of the macronutrients (N, P, K, Ca, Mg, S) and  
210 the micronutrients (Zn, Fe, Cu, Mn, B) are listed in Tables 5 and 6, respectively, for  
211 21-day old radish plants grown in zeoponic and control substrates. All radishes  
212 grown in the three substrates had similar macronutrient (i.e., primary and secondary  
213 nutrients) compositions, except for Ca where the composition of Ca was noticeably  
214 lower in the first and second crops of radish grown in the zeoponic substrates  
215 compared to radishes grown in the potting mix and K-Cp substrates (Table 5).  
216 Another noticeable difference was slightly higher P in radishes grown in the zeoponic  
217 substrates compared to the potting mix and K-Cp control substrates (Table 5).

218

219 Micronutrient compositions were similar for all three substrates, with the exception of  
220 lower Mn in radishes grown in zeoponic substrates (Table 6).

221

### 222 3.4 Characterization of clinoptilolite exchange sites

223 Prior to plant growth experiments, the NH<sub>4</sub>-exchanged clinoptilolite had an NH<sub>4</sub>-  
224 cation exchange capacity (NH<sub>4</sub>-CEC) of 207 cmol<sub>(c)</sub>/kg, and the K-exchanged  
225 clinoptilolite had a K-CEC of 202 cmol<sub>(c)</sub>/kg. The average NH<sub>4</sub> was 107 cmol<sub>(c)</sub>/kg,

226 and the average K was 158 cmol<sub>(c)</sub>/kg on zeolitic exchange sites after three successive  
227 crops of radish growth. Thus, after three successive crops of radish growth, the  
228 zeoponic substrates had 52% of the original NH<sub>4</sub>-N and 78% of the original K  
229 remaining on zeolite exchange sites.

230

#### 231 **4. Discussion**

232 Three crops of radish were grown successively in the same zeoponic substrate, and  
233 there were no significant differences for the fresh weight produced in zeoponic  
234 substrates irrigated with deionized H<sub>2</sub>O compared to the potting mix control  
235 substrates irrigated with nutrient solution. The major difference in fresh weight of the  
236 radish edible root occurred between the first crop and the second crop, where the  
237 average fresh weight increased by 68% in the second crop. The average fresh weight  
238 of the edible root in the third crop continued the trend of increasing mass, with a 17%  
239 increase over the second crop. The highest yield in the third crop may be due to more  
240 Ca<sup>2+</sup> available for plant uptake than during the first crop grown in the zeoponic  
241 substrate. The Ca plant tissue levels in radishes (1.8-2.9 wt. %) grown in zeoponic  
242 substrates are lower than the suggested sufficient range of 3.0-4.5 wt. % Ca for  
243 radishes [24]. However, the Ca level is highest (2.9 wt. %) in radishes grown in the  
244 third crop in the same zeoponic substrates. Fenn et al. [14] have shown the addition of  
245 Ca, when using urea, has also produced increased radish plant N-use efficiency and  
246 production in the field. Previous studies with wheat have also shown that NH<sub>4</sub><sup>+</sup>  
247 released from the exchange sites of the zeolite competed with the uptake of Ca<sup>2+</sup> in  
248 wheat plants and resulted in a an NH<sub>4</sub>-induced Ca deficiency [8,9,10,11,25]. The N  
249 levels in zeoponic radishes (6.2-7.1 wt. %) are slightly higher than the recommended  
250 range (3.0 to 6.0 wt. %) for the sufficient growth of radish. Nitrifying bacteria were

251 added to the zeoponic substrates prior to plant growth to enhance the conversion of  
252  $\text{NH}_4^+$  to  $\text{NO}_3^-$  based upon the studies of Henderson et al. [10] and McGilloway et al.  
253 [12]. Other studies have shown that  $\text{NH}_4\text{-N}$  inhibited the growth of radish; hence, it is  
254 important to convert  $\text{NH}_4^+$  to  $\text{NO}_3^-$  or use  $\text{NO}_3$  fertilizers to promote production of  
255 radish [13]. The combination of higher Ca and  $\text{NO}_3$  in “soil” solution for the third  
256 crop is likely responsible for the higher radish yield. The highest Ca contents in the  
257 third crop may be due to more Ca uptake by the plant and a reduction in  $\text{NH}_4$ -induced  
258 Ca deficiency (e.g., common ion effect of Ca from dolomite and apatite as described  
259 by [8]).

260

261 The higher yields in the second and third radish crops grown in the zeoponic  
262 substrates are also supported by higher harvest indexes; i.e., 0.62-0.63 for the second  
263 and third crops compared to a harvest index of 0.56 in the first crop (Table 4). The  
264 lower harvest index in the first crop may reflect a luxury uptake of N; which would  
265 result in the production of more leaves compared to edible root.

266

267 Nutrient uptake by radishes grown in the zeoponic substrate was sufficient for  
268 optimum growth with the exceptions of the Ca and N as discussed earlier and P. The  
269 P levels in the zeoponic substrates ranged from 0.94-1.15 wt. %; which is slightly  
270 higher than the sufficient levels of 0.3-0.7 wt. % [24]; however, there were no  
271 noticeable decreases in plant productivity for the luxury uptake of P. The other  
272 macronutrient and micronutrient levels in radish grown in zeoponic substrates were  
273 well within the recommended sufficient ranges [24].

274

275 A majority of the  $K^+$  and  $NH_4^+$  remained on the zeolitic exchange sites after the  
276 growth of 3 crops of radish in the same zeoponic substrate. Similar results have been  
277 reported for the extensive growth of wheat in zeoponic substrates where nearly 80 %  
278 of the  $NH_4^+$  and  $K^+$  remained on the zeolitic exchange sites after plant growth [26].

279

## 280 **5. Conclusions**

281 Radish yields in zeoponic substrates were equivalent to yields in control substrates  
282 irrigated with nutrient solutions. Zeoponic substrates provided all of the plant  
283 essential nutrients required for the growth of radish. A high percentage of  $NH_4^+$  and  
284  $K^+$  cations remained on the zeolite exchange sites after three successive crops,  
285 although the amount of  $NH_4^+$  and  $K^+$  remaining on the exchange sites depends on a  
286 number of factors (e.g., volume of substrate, leaching, number of growth cycles), and  
287 could possibly be altered with the recycling of leachate. The increasing trend in  
288 radish crop yield after three successive crops (62 total days) and supporting data from  
289 other experiments (e.g., zeoponic wheat experiments, 270 total days), also  
290 demonstrate the potential for long term nutrient delivery of zeoponic substrates. This  
291 longevity is especially important for space missions to Mars, where missions lasting  
292 from 500-1000 days will need some amount of food production to either supplement  
293 or sustain food supplies for the astronaut crews. NASA's zeoponic substrate has been  
294 successfully flown on Space Shuttle missions [27] in plant growth chambers that are  
295 prototypes for those planned for Mars spacecraft. Likewise, zeoponic substrates  
296 could be used at a permanent outpost on the Moon, where resupply missions will  
297 likely only occur every six months.

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**Table 1.** Chemical composition of the synthetic apatite and dolomite used in zeoponic substrates.

Major oxides	Synthetic <sup>a</sup> apatite	Dolomite <sup>b</sup>
	-----wt. %-----	
CaO	46.6	35.03 (0.15) <sup>c</sup>
P <sub>2</sub> O <sub>5</sub>	34.6	-
MgO	2.77	17.66 (0.12)
SiO <sub>2</sub>	2.56	-
SO <sub>3</sub>	2.46	-
CO <sub>2</sub>		46.98 (0.02)
Fe <sub>2</sub> O <sub>3</sub>	1.76	-
FeO	-	0.31 (0.08)
ZnO	0.06	-
MnO	0.06	0.02 (0.08)
CuO	0.02	-
Total	-	100 (0.00)

<sup>a</sup>Hydroxyapatite produced in the laboratory by the method of Golden and Ming [17].

<sup>b</sup>Dolomite analysis by Beiersdorfer et al. [8].

<sup>c</sup>Numbers in parentheses represent one standard deviation (n=25; where n = # of analyses)

**Table 2.** Average total fresh weight/plant of the edible roots for radish grown in zeoponic substrates.

	Number	Mean	Std. Dev.	Std. Err. Mean
		----- wt. % -----		
Zeoponic substrate	144	10.97 <sup>a</sup>	4.91	0.41
Potting mix control substrate	144	10.92 <sup>a</sup>	5.55	0.46
K-clinoptilolite control substrate	144	9.50 <sup>b</sup>	3.78	0.32

<sup>a</sup>There is no statistical difference based on Tukey-Kramer tests [23] at a 5% probability level between plants grown in the zeoponic substrate and plants grown in the potting mix control substrate.

<sup>b</sup>There is a statistical difference based on Tukey-Kramer tests [23] at a 5% probability level between plants grown in K-clinoptilolite control substrate and plants grown in the other two substrates.

**Table 3.** Average fresh weight/plant of the edible roots for radish grown in zeoponic substrates.

Root fresh weight	Zeoponic substrate	Potting mix control substrate	K-clinoptilolite control substrate
	-----mass (g)-----		
Crop 1	7.10 (2.87) <sup>a</sup>	15.27 (3.56)	8.06 (3.52)
Crop 2	11.92 (2.07)	8.56 (3.50)	10.55 (1.99)
Crop 3	13.90 (2.56)	8.96 (1.99)	9.89 (1.37)

<sup>a</sup>Numbers in parentheses represent one standard deviation.

**Table 4:** Average harvest index for radish grown in zeoponic and control substrates for 21 days.

Root fresh weight	Zeoponic substrate	Potting mix control substrate	K-clinoptilolite control substrate
	----- <sup>o</sup> -----		
Crop 1	0.56 (0.08) <sup>a</sup>	0.65 (0.03)	0.60 (0.14)
Crop 2	0.63 (0.06)	0.59 (0.09)	0.68 (0.03)
Crop 3	0.62 (0.05)	0.64 (0.06)	0.66 (0.04)

<sup>a</sup>Numbers in parentheses represent one standard deviation.

**Table 5.** Average plant-tissue analyses for macronutrients (wt. %) for radish grown in zeoponic substrates and control substrates after 21 days.

<b>Macronutrient</b>			
Treatment	Crop 1	Crop 2	Crop 3
-----wt. %-----			
<b>Nitrogen</b>			
Zeoponics	6.97 (0.44) <sup>a</sup>	6.19 (0.39)	7.10 (1.70)
Potting Soil	6.20 (0.14)	5.56 (0.47)	6.32 (1.28)
K-Clino	6.10 (0.21)	5.49 (0.50)	6.24 (1.27)
<b>Phosphorus</b>			
Zeoponics	1.00 (0.01)	1.15 (0.01)	0.94 (0.05)
Potting Soil	0.86 (0.15)	0.86 (0.03)	0.70 (0.03)
K-Clino	0.59 (0.06)	0.55 (0.01)	0.55 (0.01)
<b>Potassium</b>			
Zeoponics	4.87 (1.12)	4.41 (0.74)	4.29 (1.20)
Potting Soil	4.72 (0.89)	4.39 (1.51)	3.94 (1.27)
K-Clino	5.60 (1.17)	4.77 (1.83)	4.57 (1.39)
<b>Calcium</b>			
Zeoponics	1.80 (0.05)	2.42 (0.23)	2.93 (0.24)
Potting Soil	2.20 (0.68)	2.98 (0.08)	3.67 (0.07)
K-Clino	3.27 (0.57)	3.25 (0.02)	3.05 (0.15)
<b>Magnesium</b>			
Zeoponics	0.96 (0.06)	0.93 (0.06)	0.81 (0.10)
Potting Soil	1.60 (0.13)	1.51 (0.60)	1.03 (0.27)
K-Clino	0.68 (0.04)	0.74 (0.16)	0.74 (0.18)
<b>Sulfur</b>			
Zeoponics	1.08 (0.01)	0.76 (0.03)	0.71 (0.02)
Potting Soil	1.31 (0.02)	1.44 (0.06)	1.35 (0.06)
K-Clino	1.12 (0.05)	1.27 (0.04)	1.13 (0.03)

<sup>a</sup>Numbers in parentheses represent one standard deviation.

**Table 6.** Average plant-tissue analyses for micronutrients (mg/kg) for radish grown in zeoponic substrates and control substrates after 21 days.

<b>Micronutrient</b>				
Treatment		Crop 1	Crop 2	Crop 3
		----- mg/kg -----		
<b>Zinc</b>				
Zeoponics		33 (1) <sup>a</sup>	33 (3)	30 (3)
Potting Soil		29 (6)	19 (4)	20 (4)
K-Clino		35 (6)	26 (3)	25 (4)
<b>Iron</b>				
Zeoponics		136 (23)	131 (6)	106 (13)
Potting Soil		160 (35)	142 (19)	93 (34)
K-Clino		120 (22)	90 (20)	93 (27)
<b>Manganese</b>				
Zeoponics		55 (6)	61 (3)	68 (3)
Potting Soil		139 (6)	130 (27)	97 (26)
K-Clino		106 (6)	83 (2)	60 (2)
<b>Copper</b>				
Zeoponics		8 (3)	7 (0)	7 (1)
Potting Soil		8 (4)	7 (1)	7 (2)
K-Clino		8 (2)	6 (1)	8 (2)
<b>Boron</b>				
Zeoponics		75 (5)	70 (2)	41 (9)
Potting Soil		56 (2)	57 (4)	54 (4)
K-Clino		69 (3)	63 (7)	60 (1)

<sup>a</sup>Numbers in parentheses represent one standard deviation.



## Figure Captions

**Figure 1.** Dynamic equilibria for NASA's zeoionic plant-growth system. Plant growth nutrients are slowly released from synthetic apatite (SA) and dolomite by dissolution and from clinoptilolite (Cp) by ion-exchange reactions.

Figure(s)

