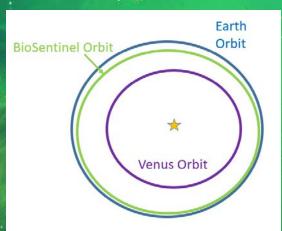
BioSentinel: optimizing growth conditions for improved yeast cell viability after long-term desiccation

Kaixin Cui^{1,2}, Sergio R. Santa Maria³, Sofia Massaro Tieze⁴, Lauren Liddell⁵, and Sharmila Bhattacharya⁶

¹Space Life Sciences Training Program (SLSTP), NASA Ames Research Center, ²University of Colorado Boulder, ³University of New Mexico, ⁴Blue Marble Space Institute of Science, ⁵Logyx LLC, ⁶NASA Ames Research Center

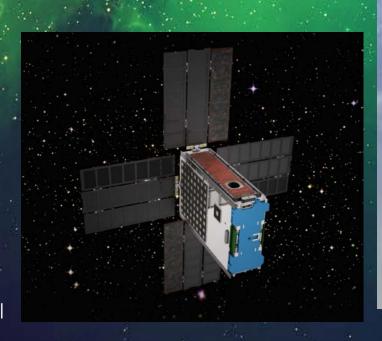
BioSentinel Overview

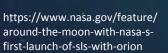
- Study the effects of deep space radiation on yeast, Saccharomyces cerevisiae
 - One type of damage generated from ionizing radiation in deep space double strand breaks (DSBs)
 - Yeast is suitable as a biological model
 - Eukaryotic, Robust analog for human cells
 - Tolerant to desiccation
 - Long duration mission: 12 months
 - Storage (pre-launch): ~6 to 9 months
 - DNA damage repair pathway is similar to that of humans
- Heliocentric orbit
- 1 of 13 secondary payloads on SLS EM1, December 2019
 - 1st biological experiment in deep space in 46 years
- ISS mission in LEO for lower radiation levels, microgravity control for deep space experiment





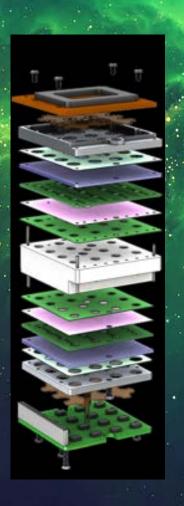
https://www.nasa.gov/mission_pages/station/main/index.html

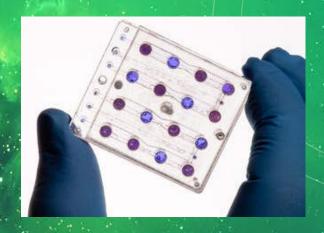




YEAST CELLS PREPARATION FOR SPACE

- Grown in liquid medium for 7 days
- Loaded and air dried into the wells of microfluidic card on Earth
- Mix of growth medium and metabolic indicator dye pumped into wells at time points to rehydrate the cells while in orbit
- 3-color LED detection system: measure changes in growth and metabolism resulting from ionizing radiation exposure
- Wild type control strain and a rad51 mutant strain that is defective for DNA damage repair (more sensitive)

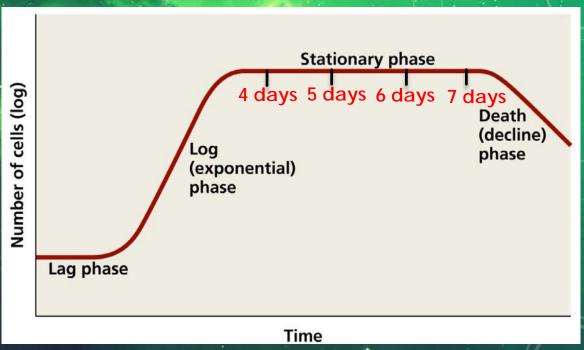






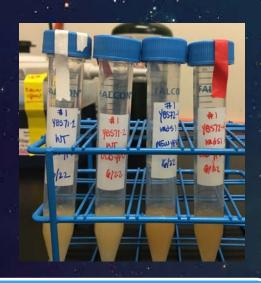
GOAL OF THE PROJECT

- Determine the growth period that improves desiccation tolerance but allows for retention of uniform radiation sensitivity
- An extended time in stationary phase: yeast are more tolerant to desiccation
- Excessive exposure in stationary phase
 - nitrogen starvation
 - loss of viability
 - heterogeneous cell population due to sporulation
 - Tetrads with 4 spores are formed
 - Indirect measure of stress
 - Spores are more tolerant to radiation



https://www.researchgate.net/figure/Typical-yeast-growth-curve-Saccharomyces-cerevisiae-grown-in-YPD-media-at-30C-for 12_fig1_235352009

YEAST DESICCATION PROCEDURE







Grow cells in liquid medium for a varying number of days (4-7 days) at room temperature



Plate some cells and grow on a YPD agar plate for 3 days at 30°C

 Provides undesiccated control which is a baseline for cell viability



Load a 96-well plate with cells with Trehalose drying buffer and left to air dry at 23°C

YPD = Yeast Extract Peptone Dextrose

Trehalose Drying Buffer = sugar to help protect cells when drying

Did this procedure for both rad51 and wild type

YEAST REHYDRATING PROCEDURE

Place SC in the wells with dried cells, Wait 30 minutes

Place SC in the wells with dried cells, Wait 30 minutes



Mix and Plate on YPD agar plates



Mix and Place the mixture into another 96 well plate



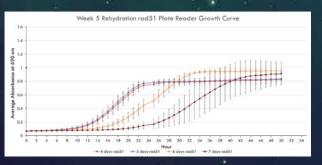
Wait 3 days at 30°C



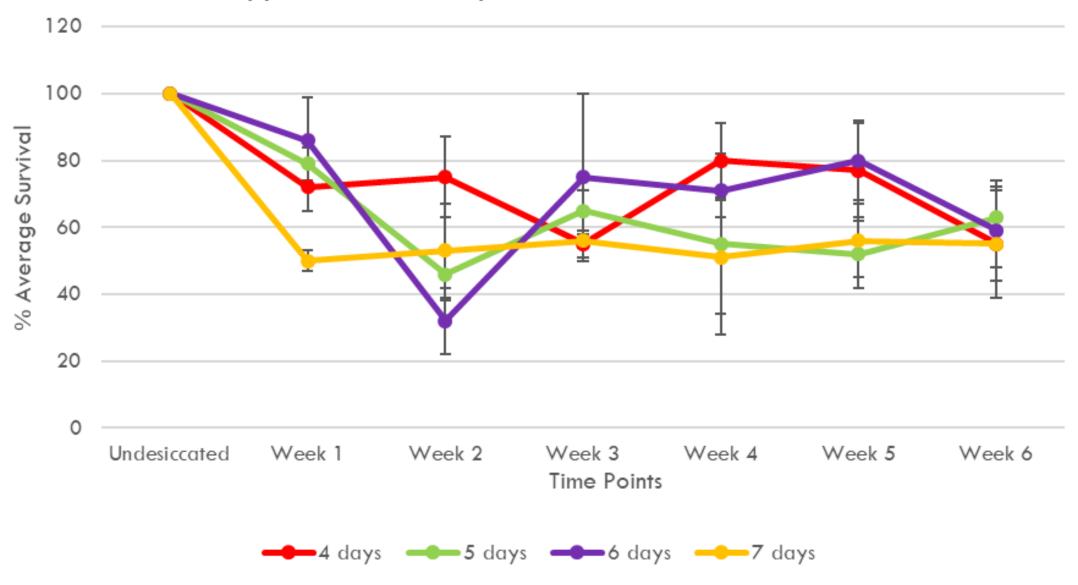
Start 24 hr plate reading, Observe growth curve at 690 nm absorbance which observes turbidity



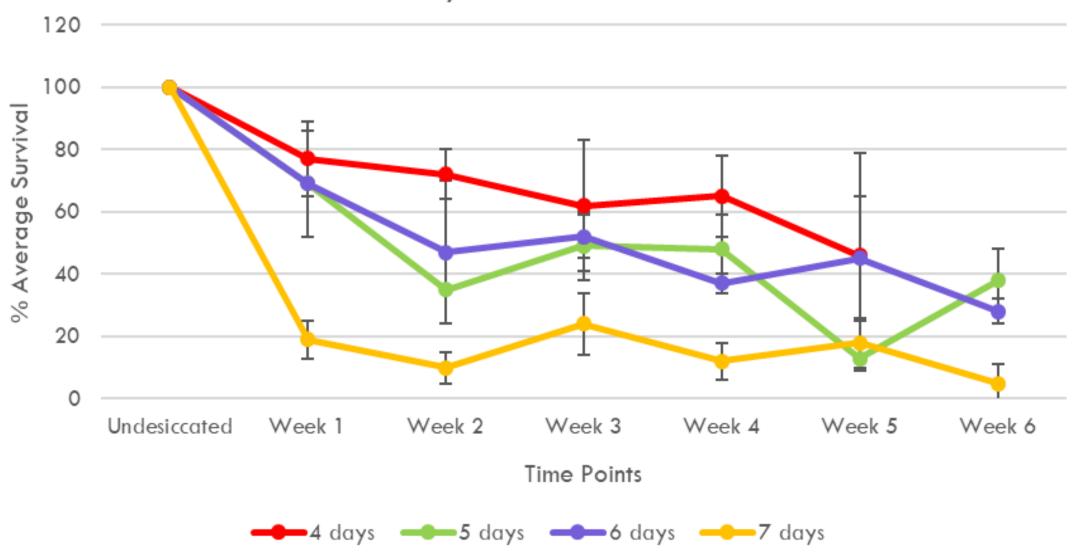
SC = Synthetic Complete

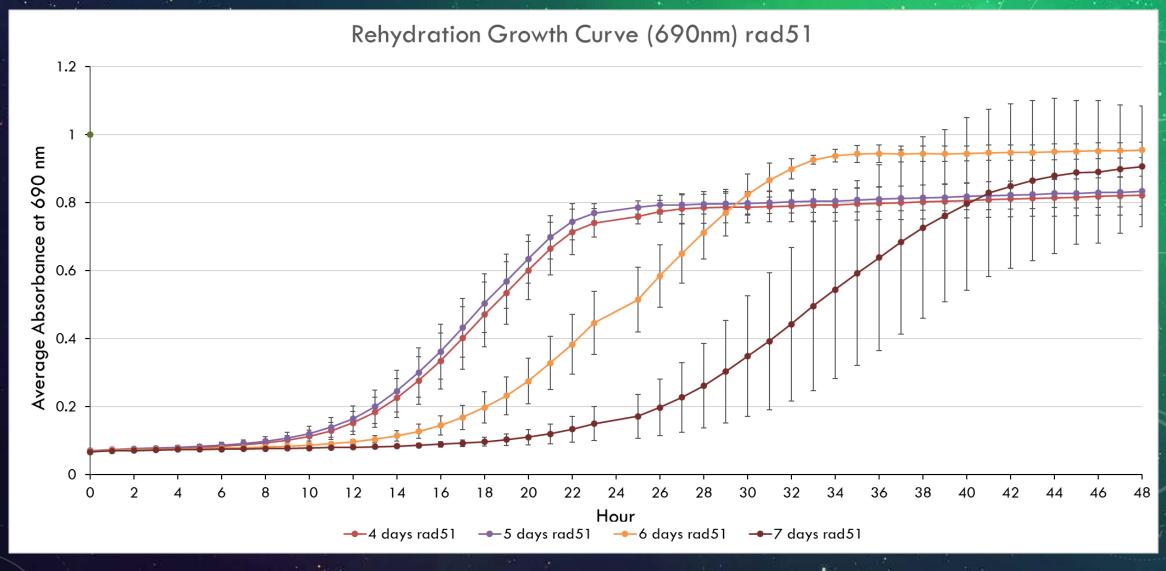


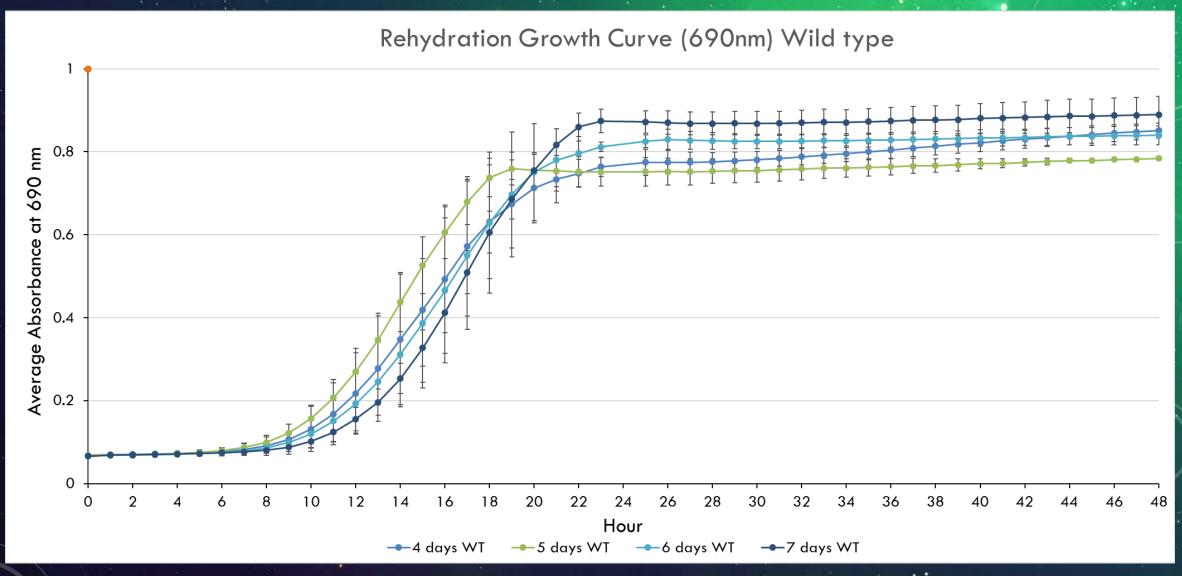
Wildtype Cell Viability- normalized with undesiccated



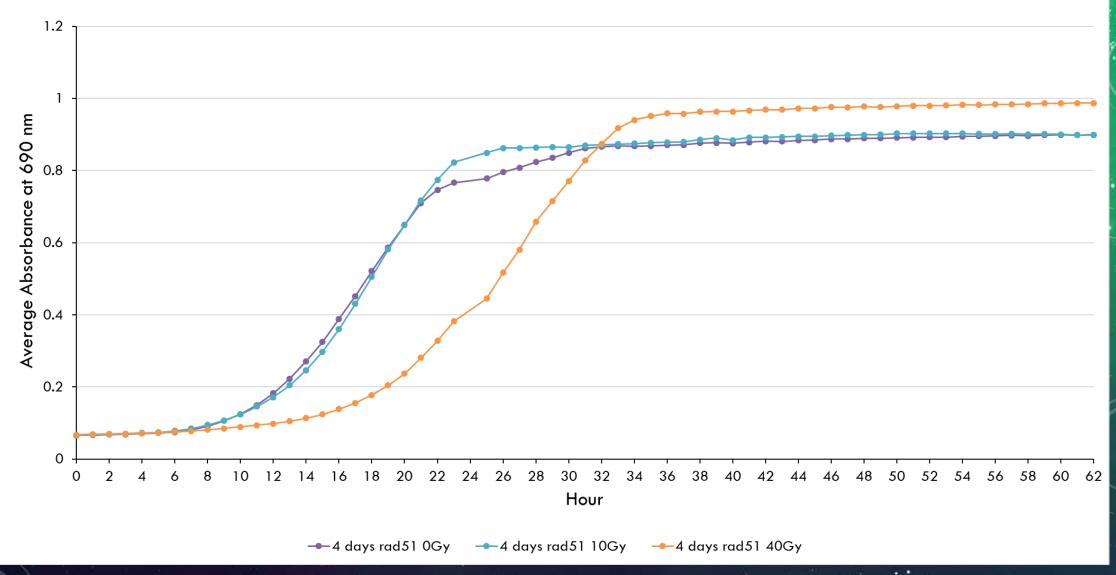
rad51 Cell Viability- normalized with undesiccated



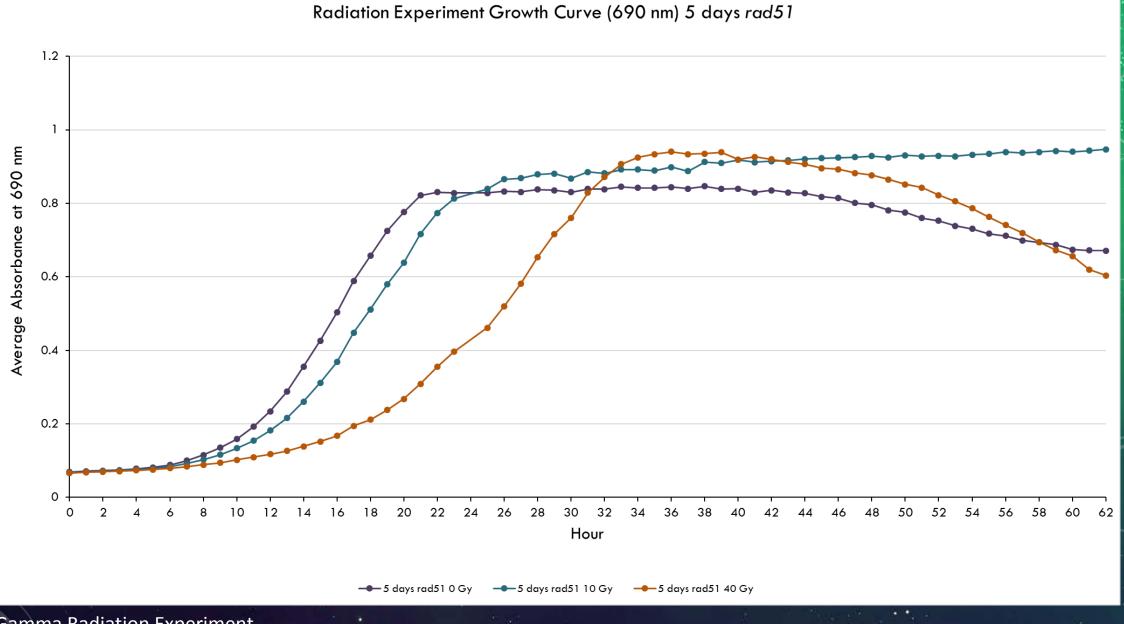


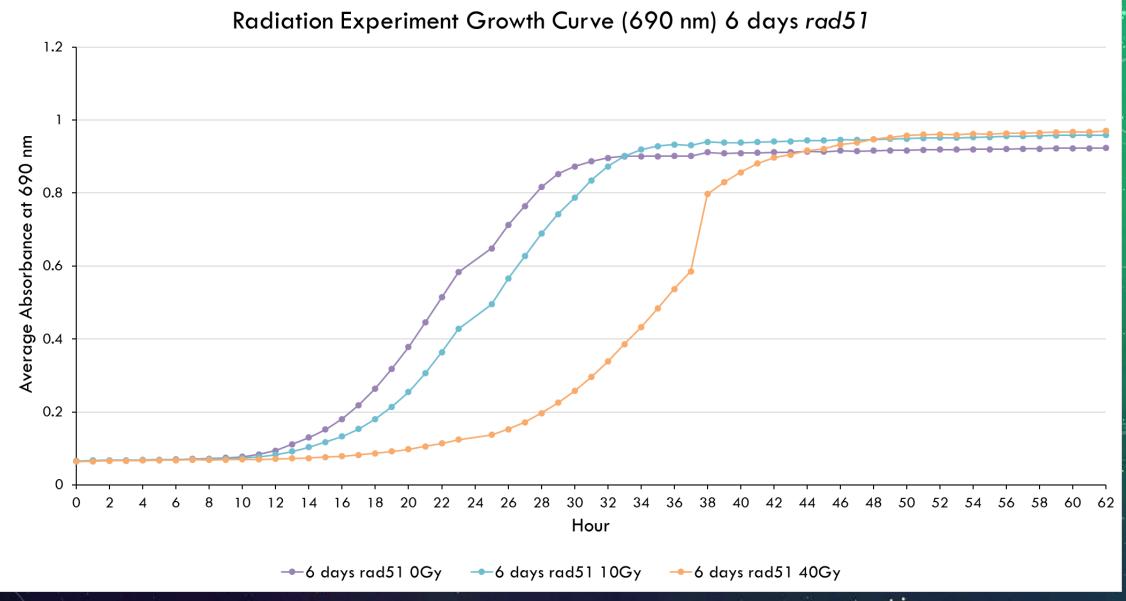




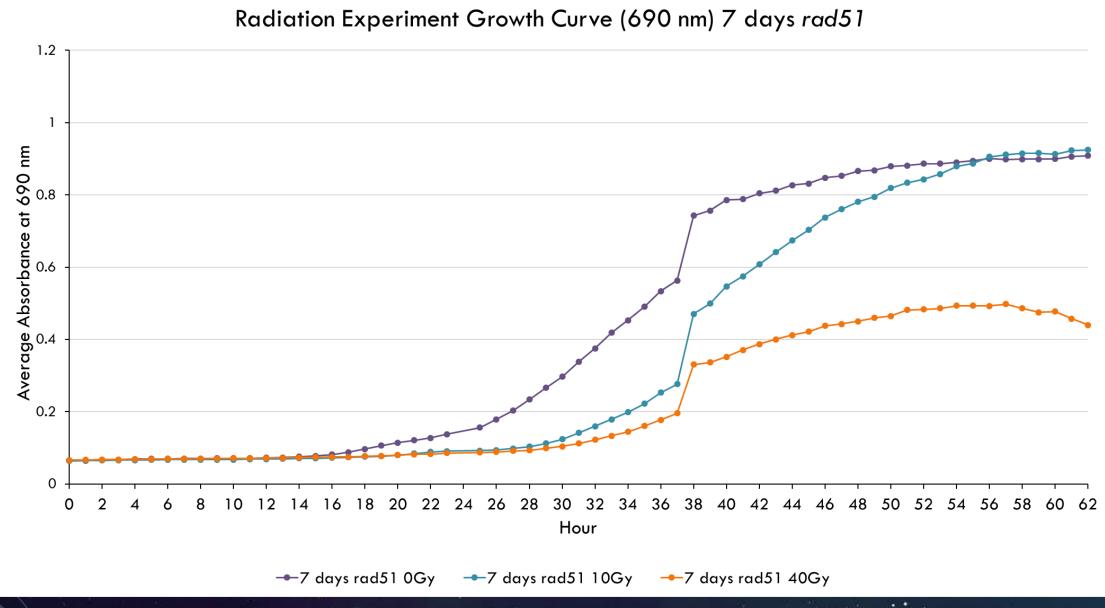


Gamma Radiation Experiment 4 days of growth *rad51*





Gamma Radiation Experiment 6 days of growth *rad51*



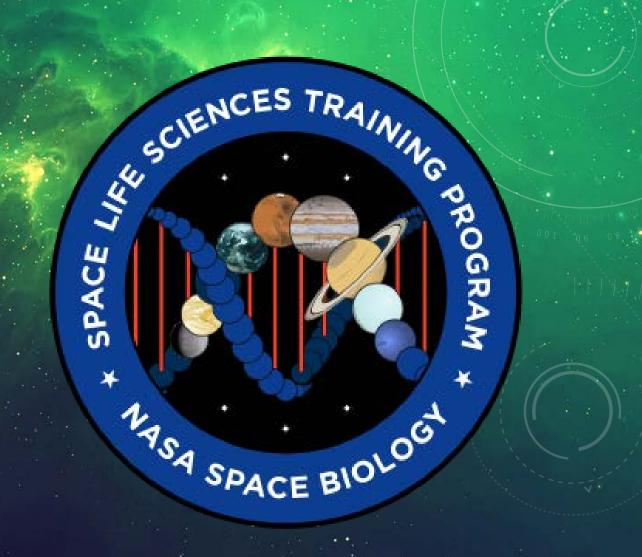
Gamma Radiation Experiment
7 days of growth *rad51*

CONCLUSION

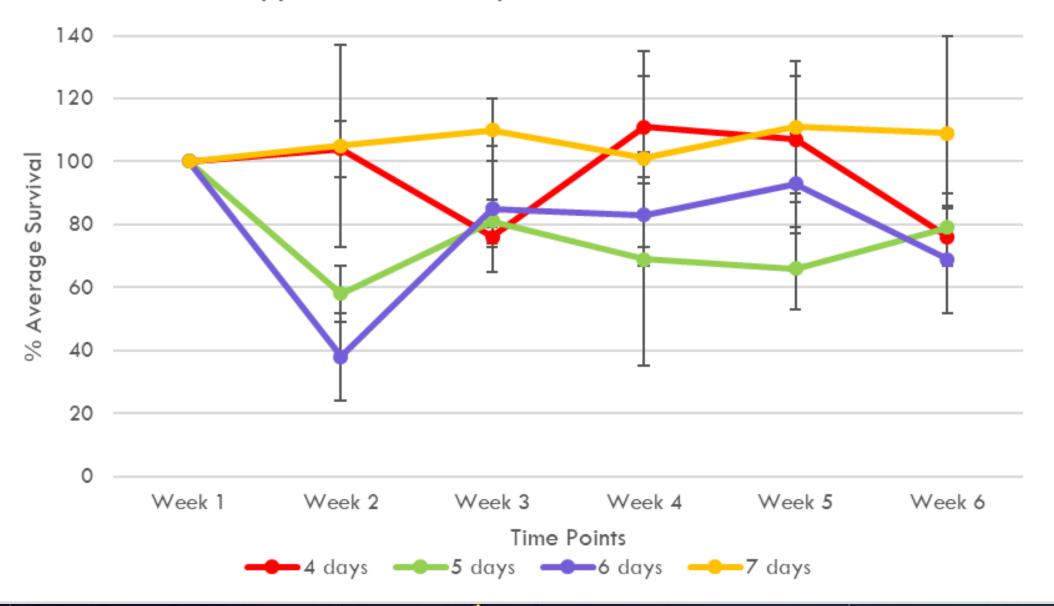
- Currently 4, 5 and 6 days of growth is a better option than 7 days of growth
- Over time this may change
- Continue to take data points in the next few months
- The experiment may be repeated to confirm results

ACKNOWLEDGMENTS

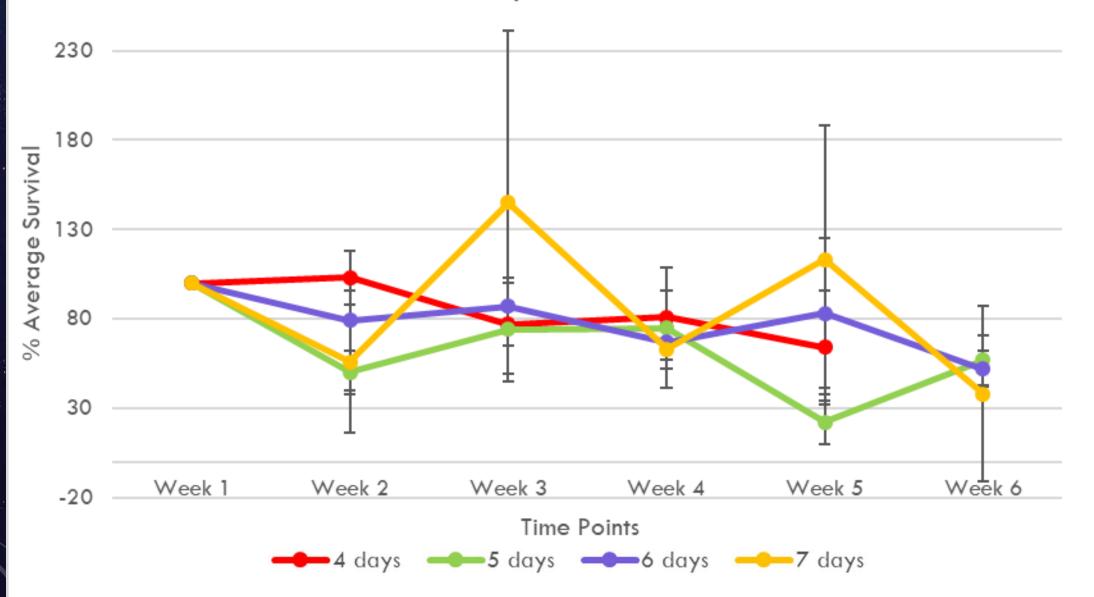
- > Sergio R. Santa Maria
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- ▶ Lauren Liddell
- Sharmilla Bhattachayra
- > Tristan Caro
- > Aimee Johnson
- > SLSTP Management



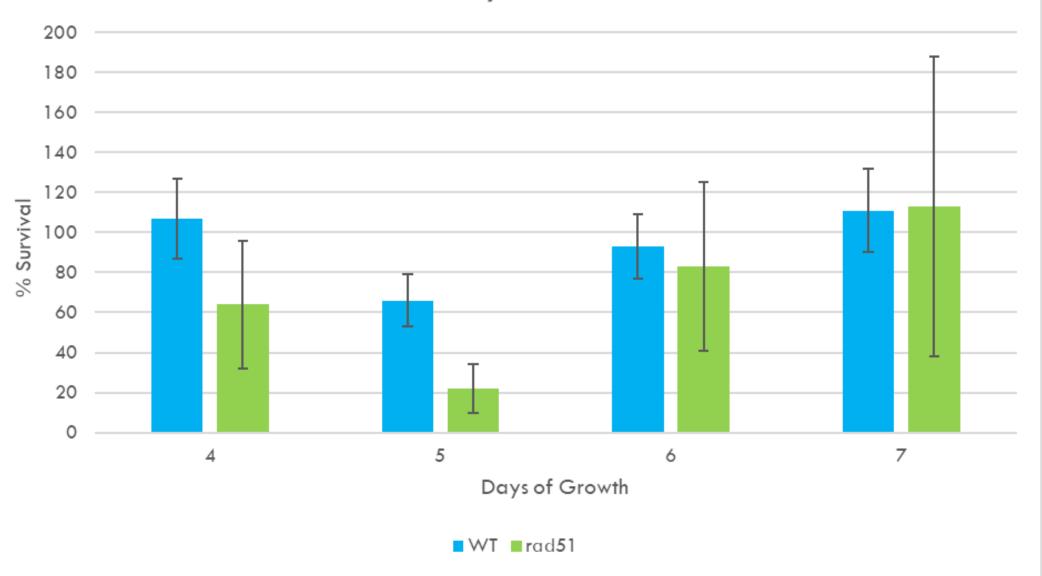
Wildtype Cell Viability- normalized with Time 0

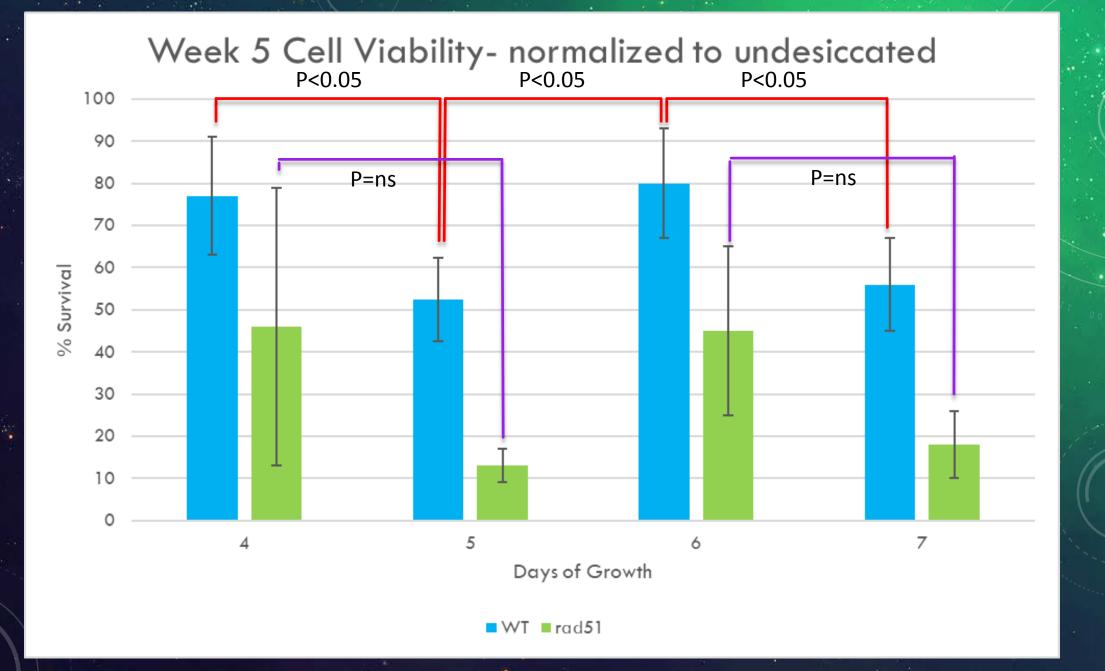


rad51 Cell Viability-normalized with Time 0



Week 5 Cell Viability- normalized to Time 0





Radiation Experiment Plate Reader Growth Curve

