

Mineral-catalyzed formation of marine NO and N₂O on the anoxic early Earth

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Abstract

Microbial denitrification converts fixed nitrogen species into gases in extant oceans. However, it is unclear how such transformations occurred within the early nitrogen cycle of the Archean. Here we present experimental anoxic surface-catalyzed reduction of nitrite and nitrate via green rust and magnetite combined with diffusion and photochemical modeling. We find that in a Fe^{2+} -rich marine environment, Fe minerals could have catalyzed abiotic denitrification reactions leading to the formation of nitric oxide (NO) and nitrous oxide (N_2O). Nitrate did not exhibit reactivity in the presence of either mineral or aqueous Fe^{2+} , however, both minerals induced rapid nitrite reduction to NO and N_2O . While N_2O escaped into the atmosphere (63% of nitrite-nitrogen, with green rust as catalyst), NO remained associated with precipitates (7%) serving as a potential shuttle to the benthic ocean. The modeling suggests that marine N_2O emissions would have sustained 0.8-6 ppb of atmospheric N_2O without a protective ozone layer. Our findings add detail to the as yet incompletely documented Archean nitrogen cycle, implying a globally distributed process driven by chemical kinetics similar to those of modern enzymatically mediated conversions.

Prior to the rise of O₂ during the Great Oxidation Event (GOE), the Archean ocean
20 received oxides of nitrogen (NO_x) by deposition of abiotic nitrogen fixation products¹. As a
consequence, nitrite (NO₂⁻) and nitrate (NO₃⁻) reached seawater concentrations in the lower μM
range¹⁻⁴. While microbial denitrification plays a dominant role in modern oceans to recycle NO_x⁻
into gaseous phases, such pathways were not available in the Archean without active NO_x⁻
reductases. A non-enzymatic process reducing fixed NO_x⁻ into gases is chemodenitrification,
25 which is of particular interest because nitrous oxide (N₂O) can be a primary product⁵⁻⁷. The
Archean atmosphere prior to the GOE was likely dominated by N₂ and CO₂, with ppm-levels of
CO, CH₄, and H₂ (ref. ⁸). The introduction of even trace amounts of more oxidized gases, such as
N₂O, could have created a significant source of thermodynamic disequilibrium to drive primitive
ecosystems. Further, N₂O is a strong greenhouse gas and has been considered to induce a warmer
30 climate under a faint young Sun⁹. Stanton *et al.* first showed through experiments and modeling
that aqueous Fe²⁺ could have acted as a driver for chemodenitrification to form N₂O abiotically in
Proterozoic oceans¹⁰. Yet, N₂O production rates established for the early marine environment, thus
far, have been ~4 orders of magnitude lower than rates reported from isotope tracer measurements
in modern oxygen minimum zones (Table 1), underlining the catalytic importance of
35 denitrification enzymes for rapid nitrogen cycling. As a result, it has been hypothesized that N₂O
was virtually absent on the anoxic early Earth, especially without a protective ozone (O₃) layer to
compensate for elevated photolytic N₂O destruction¹¹⁻¹³. While under ferruginous conditions
aqueous Fe²⁺ has not stimulated N₂O production close to microbially mediated rates (Table 1),
mineral phases bear the potential to reach such rates and could have therefore provided efficient
40 pathways to channel fixed nitrogen back into the Archean atmosphere.

Here, we evaluated the role of the mineral-catalyzed formation of NO and N₂O as a marine source pathway under anoxic, ferruginous conditions. NO has been measured in mixtures of NO_x⁻ and highly concentrated Fe²⁺ solutions⁵ and in a MoS₂ suspension under a geoelectric current¹⁴, but it has only been speculated to be produced in Fe mineral suspensions^{6,7}. The ferrous-
45 ferric hydroxy salt carbonate green rust (GR, [Fe²⁺_(1-x) Fe³⁺_x (OH)₂]^{x+} · [(x/2)CO₃²⁻ · (m/2)H₂O]^{x-}) has been shown to precipitate from Archean seawater-analog solutions, consistent with thermodynamics predicting a predominance of GR in the Fe sink fraction along the water column prior to 2.5-2.7 Ga¹⁵. Settling GR particles could have provided a transport mechanism for trace compounds to the seafloor¹⁵, where reducing conditions partially converted GR to magnetite
50 (Fe₃O₄), an important constituent of banded iron formations. Because NO₂⁻ and NO₃⁻ can spontaneously react in presence of reduced Fe¹⁶⁻¹⁸, we tested the reactivity of the mixed-valence Fe minerals GR and magnetite with NO₂⁻ and NO₃⁻ at *low* (1-5 μM), *high* (20-35 μM), and *excessive* (200-350 μM) abundances, and characterized their potential to produce NO and N₂O at relevant rates for the Archean ocean-atmosphere system. To do so, we used mineral particle concentrations
55 recorded for modern analogues as a proxy for mineral abundance (*Supplementary Figs. 1-4*). We experimentally simulated ferruginous conditions with a simple end-member artificial seawater recipe using a 20% CO₂-HCO₃⁻ buffer devoid of sulfur. Subsequent flux and photochemical calculations suggest marine mineral-catalyzed NO and N₂O formation constituted a substantial source of gaseous nitrogen oxides on the anoxic early Earth.

60

<Table 1>

65 *Abiotic NO_x⁻ reduction under simulated Archean conditions*

We first compared the reactivity of NO₃⁻ and NO₂⁻ with GR, magnetite and aqueous Fe²⁺ separately. For all reductants, consumption of NO₃⁻ and production of N₂O was negligible (<0.03 nmol h⁻¹, Fig. 1a) even over a prolonged period of 100 days. In contrast, NO₂⁻ showed reactivity in the presence of both mineral catalysts, and was stable in controls with 500 μM aqueous Fe²⁺. The
70 contrasting reactivity of the nitrogen oxides is reflected in the solid phase Fe²⁺/Fe³⁺ ratio determined at the end of the experiment (Fig. 2), revealing a trend of higher mineral oxidation with increasing NO₂⁻ concentration. Magnetite and GR showed disparate potential to produce N₂O. Concomitant with a more rapid NO₂⁻ consumption, N₂O production rates were more than 10 times higher when GR was the catalyst (Fig. 1b). At GR surface areas observed for the Archean ocean
75 analogue Lake Matano, Indonesia²⁹ (~10⁻³ m² mL⁻¹, Fig. 1c, d), abiotic N₂O production rates were highly similar to (*low* NO₂⁻) or exceeding (*high* NO₂⁻) biotic production rates measured recently in the Eastern Tropical North Pacific^{19,20} (Table 1). As a reference, modern NO₂⁻ concentrations are near the bottom of the experimental range employed here, which extends higher to account for a range of possible Archean NO_x⁻ abundances (see discussion below).

80

<Fig. 1>

The conversion of NO₂⁻ to N₂O was not equimolarly balanced and we therefore considered accumulation of NO as intermediary species³⁰. After initially observing modest NO
85 accumulation, we injected concentrated hydrochloric acid in order to dissolve mineral particles and release any nitrogen bound to mineral surfaces. Indeed, the particle dissolution was followed by a spike in headspace NO (Fig. 1e). At least 92% of the total NO was bound to GR (97% was bound to magnetite). To rule out reaction of the acid with residual NO₂⁻, we conducted controls with

sulfuric acid and NO_2^- (*Supplementary Table 3*). Other controls evaluating the stability of NO in
90 the mineral suspension included acid injection at later time points in time-extended experiments.
NO yields at 220 hours ($< 0.5 \mu\text{M NO}_2^-$) were similar to those at 50 hours after NO_2^- addition
(*Supplementary Fig. 5, Supplementary Table 3*). We calculated the NO yield in respect to the total
 NO_2^- and, accordingly, 7% reacted to NO (over 63% that reacted to N_2O) with GR as catalyst and
87% reacted to NO (over 8% that reacted to N_2O) with magnetite as catalyst. The remaining
95 product pool could be N_2 (ref. ³⁰) and only constituted a minor fraction. Our data show that
mineral-associated NO, presumably in the form of nitrosyl species³¹, is a significant byproduct and
geochemically stable in presence of Fe mineral phases.

<Fig. 2>

100

N_2O accumulation triggered by marine emissions

To elucidate the impact of the experimentally derived production rates on N_2O dispersion
in seawater and emission to the Archean atmosphere, we combined diffusion models into a simple
flux balance model (*Supplementary Information*). We considered a 100 m deep ocean slab beneath
105 the photic zone as the part of the water column with a maximum overlap of NO_x^- ions from the
surface and GR formation fueled by upwelling Fe^{2+} -rich water from the depth. An estimate of the
GR particle concentration in the Archean ocean was informed by observed Fe particle numbers
from seven modern marine and freshwater anoxic basins (Fig. 1d), including Kabuno Bay (KB),
East Africa³². The peak Fe^{2+} concentration in the particulate fraction of KB waters ($11 \mu\text{M}$) closely
110 reflects concentrations used for the thermodynamic modeling of GR precipitation in Archean
seawater¹⁵, which is why we used the KB level as reference and enveloped that value with a
putative lower and upper boundary of 50% and 500%. Archean seawater Fe^{2+} could have reached

concentrations of 55 μM , as constrained by Fe^{2+} toxicities on cyanobacteria³³ and the appearance of herringbone and micritic carbonates^{34,35}. At that upper boundary and under *low* (conservative) NO_2^- conditions, net N_2O emissions conceivably reached 235 [98.1–413.9] $\text{nmol m}^{-2} \text{h}^{-1}$ (95% CI, *Supplementary Figs. 6 and 7, Supplementary Table 4*), implicating 4 times the modern marine average flux³⁶. Overall, N_2O emissions from the ferruginous ocean gradually increased with the GR particle concentration.

120 To simulate the effect of N_2O fluxes triggered by mineral catalysis into an anoxic Archean atmosphere, we applied a photochemical model³⁷. We probed four different N_2O fluxes corresponding to *low* and *high* NO_2^- levels and Fe^{2+} concentrations in GR phases around the KB reference (*Supplementary Table 4*). Although the upper boundary of our estimates implicates near-modern N_2O abundances in the troposphere (0-13 km altitude, Fig. 3), N_2O concentrations derived from *low* NO_2^- seawater levels, and predicted to be 0.8-6 ppb, are more realistic. These concentrations match mixing ratios in previous Proterozoic atmosphere models assuming 1% of present O_2 levels^{10,13}. A striking difference is, however, the inclusion of an O_3 protective layer in previous models, which was assumed to be present after the GOE, i.e. in the Proterozoic. Our photochemical model produced an ozone column of $\sim 9.6 \times 10^{14}$ molecules cm^{-2} , which is less than 130 the present-day level by 4 orders of magnitude and previously modeled Proterozoic levels by ~ 3 orders of magnitude. Thus, without the aid of O_3 shielding, marine mineral-catalyzed N_2O production generated tropospheric N_2O levels akin those in oxygenated atmospheres later in Earth's history.

135 <Fig. 3>

The main sink of N₂O is photolysis by radiation <230 nm⁴⁰ in the stratosphere, which governs the shape of the concentration profiles (Fig. 3a). In the Archean atmosphere, N₂O abundances could have been higher due to i) additional sources and ii) protective agents against UV light. For instance, high-energy particles from solar flares led to N₂O formation most active at higher altitudes and could complement surface sources⁴¹. These pathways were perhaps more pronounced in the Hadean or early Archean when the Sun was more active and the magnetic field openings were greater at the poles. Furthermore, in the presence of methane hazes, N₂O could have been shielded through strong UV absorption by fractal haze particles³⁹, which prolonged its lifetime. A sensitivity study using the photochemical model shows that, with a fractal haze, N₂O concentrations derived from *low* NO₂⁻ seawater levels can build up to 1-7 times the present-day atmospheric level (Fig. 3b). One may ask if this buildup of abiotic N₂O may constitute a “false positive” scenario for using N₂O as a biosignature gas for terrestrial exoplanets akin to the Archean Earth. We suggest that this would not be a true false positive because the high abundance of methane, which is the prerequisite of the fractal haze layer, is likely biogenic in the first place.

Despite the extent of tropospheric N₂O predicted by our model, effects on the planetary climate were probably modest because N₂O would not be sufficiently concentrated to significantly increase the infrared atmospheric opacity in CO₂ window regions. Especially in high CO₂ atmospheres, the N₂O radiative forcing is diminished since it may not occupy otherwise transparent infrared windows. The infrared window between 6 and 8 μm could potentially be filled by N₂O absorption, but this is dependent on atmospheric methane which absorbs at similar

wavelengths¹³. Since methane concentrations were likely >100 ppmv⁴², greenhouse warming by N₂O under our simulated conditions therefore contributed only weakly to a warmer climate under the faint young Sun of the Archean⁹.

Fe mineral reactions as significant sink for marine NO_x⁻

Because of prevailing ferruginous conditions, mineral reactions were likely limited by abiotic nitrogen fixation rates rather than mineral surface area. Previous seawater NO_x⁻ estimations (8.7-24,000 μM², 1-10,000 μM⁴³, <1 μM⁴ and 0.2-2 μM³⁸) contain a large uncertainty. For comparison, modern NO₂⁻ concentrations reach ~1.5 μM around oxygen minimum zones, but contrary to the Archean ocean, oxidoreductases effectively suppress NO₂⁻ accumulation⁴⁴. While previous estimates refer to the average bulk seawater composition, top-down gradients of NO_x⁻ or conical zones of higher NO_x⁻ concentrations beneath rainout areas contributed perhaps to a more heterogeneous distribution. In such zones, NO_x⁻ may have reached steady-state concentrations well above the *low* NO_x⁻ range (1-5 μM) designated here.

Our experimental data suggest rapid consumption of seawater NO_x⁻ (NO₂⁻ in particular) revealing a sink distributed over the pelagic ocean that has thus far not been accounted for. Previously proposed geochemical reactions that consumed dissolved NO_x⁻ species on the early Earth included reduction to ammonia⁴⁵ and reduction of NO₃⁻, NO₂⁻ and N₂ during water-rock interactions between hydrothermal fluids and the oceanic crust⁴⁶⁻⁴⁸. However, abiotic NO_x⁻ conversion to ammonia is not significant at pH ≤ 7.3 (ref. ⁴⁵), a regime that dominated the early oceans⁴⁹, and therefore restricted this NO_x⁻ sink to more alkaline environments, such as ultramafic-hosted hydrothermal vents. Because NO₃⁻ was photo-reduced to NO₂⁻ and mineral-catalyzed reduction of NO₂⁻ is several orders of magnitude faster than photo-reduction of NO₂⁻ (ref. ⁴),

reaction on Fe mineral surfaces constituted a sink for fixed nitrogen from both the NO_3^- and NO_2^- pool. We henceforth tested if NO_2^- consumption on mineral particles and in sediments exhausted seawater NO_2^- , possibly impeding mineral-catalyzed NO and N_2O production (*Supplementary*
185 *Information*). Assuming a combined NO_x flux of 2.5×10^{11} mol N yr^{-1} from lightning events⁵⁰, volcanic eruptions⁵¹, and meteoritic impacts⁵², GR particle surfaces equivalent to 200% KB draw NO_2^- concentration down to 8.4 μM . At higher GR abundances, the reactants became exhausted and any NO_2^- from NO_3^- photo-reduction or deposited to the ocean directly converted to NO and N_2O . Importantly, abiotic fluxes were co-regulated by the influx of NO_x into the ocean and the
190 amount of GR suspended in seawater. If N_2O formation dropped due to reactant limitation, atmospheric N_2O may have reached sub-ppb concentrations, but no lower than HNO-derived N_2O concentrations (>0.5 ppb, Fig. 3).

Effects of NO and N₂O on early anaerobic microbial life

195 With the assumption that individual denitrification steps evolved consecutively, our results provide a geochemical explanation for biochemical patterns known for over three decades. A primitive form of NO and N_2O reduction likely preceded aerobic respiration based on the structural resemblance of NO^{53} and $\text{N}_2\text{O}^{54-56}$ reductases (Nor and Nos) with components of cytochrome c oxidase, including hydrogen-bond- and metal-configuration, Ca ligation⁵⁷, and
200 phylogenetic analyses^{53,58}. This concept is supported by a simple make-up of the respiratory chains involving quinone-dependent Nor and membrane-bound Nos, as present in some gram(+) bacteria^{55,56}. Because of their dependence on a complex cytochrome c maturation system, NO_3^- and NO_2^- reductases probably evolved later⁵⁹.

In order to constrain the effects of N₂O on early microorganisms, we estimated dissolved
205 abundances based on atmosphere-to-seawater diffusion. Accordingly, at least 0.04-4.7 nM N₂O
were dissolved in the ferruginous seawater based on mere equilibration with the Archean air
without haze (*Supplementary Table 4*). True concentrations were likely higher in proximity around
actively N₂O-generating GR particles. Contemporary microbial N₂O consumption is based on
high-affinity enzymes adapted to low N₂O steady-state concentrations⁶⁰. For example, 30 nM N₂O
210 is readily metabolized in anoxic seawater⁶¹, but the critical threshold for marine N₂O respiration is
likely much lower (pers. commun., Sukhwan Yoon, KAIST).

<Fig. 4>

215 Under generally accepted Archean ocean conditions, N₂O reduction to N₂ is
thermodynamically favorable but kinetically hindered without enzymatic catalysis, making it an
ideal potential energy source. For instance, coupled to H₂ oxidation (H₂ as a simple and available
reductant), an affinity of up to 300 kcal per electron accepted by N₂O is significantly higher than
that of any other redox reaction tested. Moreover, GR-shuttling may have been effective for NO
220 molecules to reach benthic microbial life, acting as a biological electron sink⁵³. The availability of
NO was essential for early denitrifying microorganisms because marine sediments in the Archean
lacked nitrification as source of NO_x. The reduction of the available NO_x (Fig. 4a-c) achieves a
higher energy yield than magnetite reduction (Fig. 4d) and methanogenesis (Fig. 4e), underscoring
their benefit to microbial life in a sulfate-poor marine environment.

225 Conversely, NO and N₂O could have been damaging to microorganisms. The inhibitory
effects of the NO molecule are caused by radical production and nitrosative stress⁶², which forced

microbial cells to rapidly reduce NO using Nor-type enzymes. Given the negative effects of NO on phototrophs^{63,64} (15 nM NO halted growth), mineral-catalyzed NO formation could have corrupted the capability of phototrophic communities to harvest light energy. In contrast to its energetic
230 benefits, N₂O can bind and inactivate vitamin B₁₂^{65,66}. Among diverse roles as cofactor in essential cellular processes, vitamin B₁₂ is a key metabolite for methanogens because its cobalt(I) center serves as a methyl acceptor during methanol to methane transformation⁶⁷. For example, methanogens in tropical soils have shown metabolic sensitivity to N₂O as low as 4 nM⁶⁸ which could have feasibly accumulated in ferruginous waters (*Supplementary Table 4*). Whether NO and
235 N₂O had positive or negative net effects on early microorganisms remains elusive but, in either case, a mineral source of these compounds inevitably exerted selective pressure and shaped the composition of Archean microbial communities.

Our study suggests that a significant portion of NO_x⁻ in the ferruginous Archean ocean was diverted to NO and N₂O leading to more oxidized products compared to other sink pathways
240 (ending in N₂ or ammonia). Given GR phases at 100% KB equivalence, *low* seawater NO₂⁻ levels, and a 70% ocean cover of Earth's surface, N₂O emissions exported 2.1 Tg N yr⁻¹ into the Archean atmosphere. We regard any biological N₂O production (e.g., through ammonia oxidation) that would have added to this flux as insignificant in the reducing milieu of a ferruginous ocean. Considerable amounts of mineral surface-bound NO could have been transported down to the
245 seafloor by settling of GR precipitates. Assuming a particle sinking velocity of 50 m day⁻¹ (ref. ⁶⁹), 1.2 Tg N yr⁻¹ in the form of NO could have reached ocean sediments. Thus, the degree of thermodynamic disequilibria in the redox balance of the Archean nitrogen cycle may have been underestimated⁷⁰. We also caution to use N₂O as unique biosignature in exoplanet exploration. It is possible that conditions for the mineral-catalyzed N₂O production on early Earth-like exoplanets

250 are even more favorable, resulting in N₂O atmospheric concentrations exceeding modern ones
(higher abiotic nitrogen fixation rates, higher Fe mineral load). Signals from low-methane
atmospheres could be interpreted as false positives without any biological basis.

<Fig. 5>

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We provide evidence that higher O₃ levels are not required to reach ppb N₂O
concentrations prior to the GOE and present a concept of a complete abiotic N₂O cycle closed by
mineral-catalyzed reactions (Fig. 5), hundreds of millions of years prior to the biological and more
efficient production and consumption of N₂O¹¹. Although the effects of abiotic N₂O production
260 were probably modest on the early climate, marine sources of NO and N₂O could have markedly
influenced the evolution and survival strategies of Archean life. Rather than being a mainly
biological invention, the reduced branch of the abiotic nitrogen cycle, driven by chemical kinetics
highly similar to those of modern enzymatically mediated conversions (Table 1), was seemingly
co-opted by early organisms.

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275 **Acknowledgments**

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Author Contributions

S.B., H.I. and H.C.-Q. developed the overall study objectives and the experimental design. S.B. performed the experiments. S.B., T.E. and S.R. conducted thermodynamic and diffusion modeling. R.H. created the photochemical model. S.B. and H.C.-Q. drafted the manuscript. All authors participated in final revisions of the paper.

Competing Interest

The authors have no competing interests to declare.

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Tables

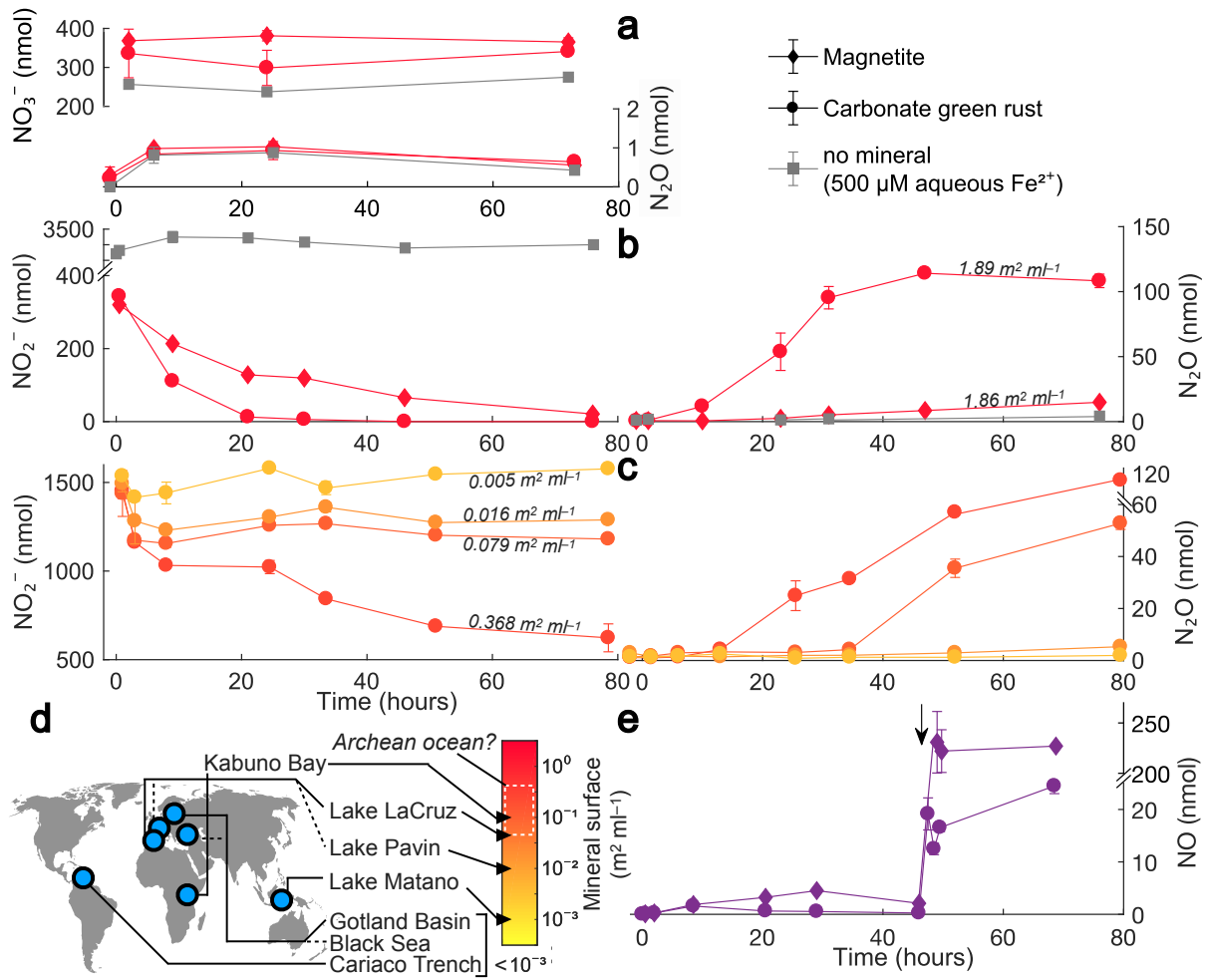
Table 1. N₂O and NO production rates of the modern and early marine environment and from cultured microbial species.

Earth period	System	N ₂ O production rates (nM days ⁻¹)	Reference
modern	Eastern Tropical North Pacific	0.5-5	19,20
modern	Arabian Sea	0.005-0.08	21
modern	Eastern Tropical South Pacific	6-43.2	22
modern	Eastern Tropical North Atlantic	1.5-3.5	23
modern	Peruvian coastal upwelling zone	5-49	24
modern	cultured <i>Nitrosomonas marina</i> C-113a	2-3	25
modern	cultured <i>Nitrosomonas marina</i> NM22	1.4-4.2	23
modern	cultured <i>Nitrosococcus oceani</i> NC10	1.2-3.7	23
modern	cultured <i>Nitrosopumilus maritimus</i> SCM1	0.4-24.3	23
early	simulated ferruginous ocean with aqueous Fe ²⁺	0.0001-0.01	10
early	simulated ferruginous ocean with GR particles	0.3-17.9*	this study

Earth period	System	NO production rates (nM days ⁻¹)	Reference
modern	Equatorial Pacific, near 170° West	34.6-103.7	26
modern	Yellow Sea	129.6	27
modern	cultured <i>Nitrosopumilus maritimus</i> SCM1	4,800	28
early	simulated ferruginous ocean with GR particles	≥737.5**	this study

310 *Derived from incubations with *low* NO₂⁻ abundances and 0.005-0.366 m² ml⁻¹ green rust surface. **Estimate of total (free and mineral-associated) NO based on particle dissolution after 50 hours in incubations with *high* NO₂⁻ abundances.

Figure



315 Fig. 1

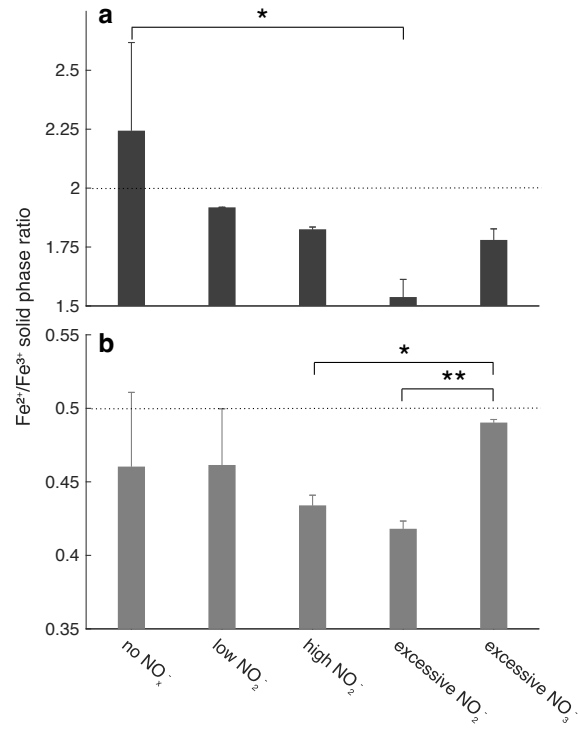
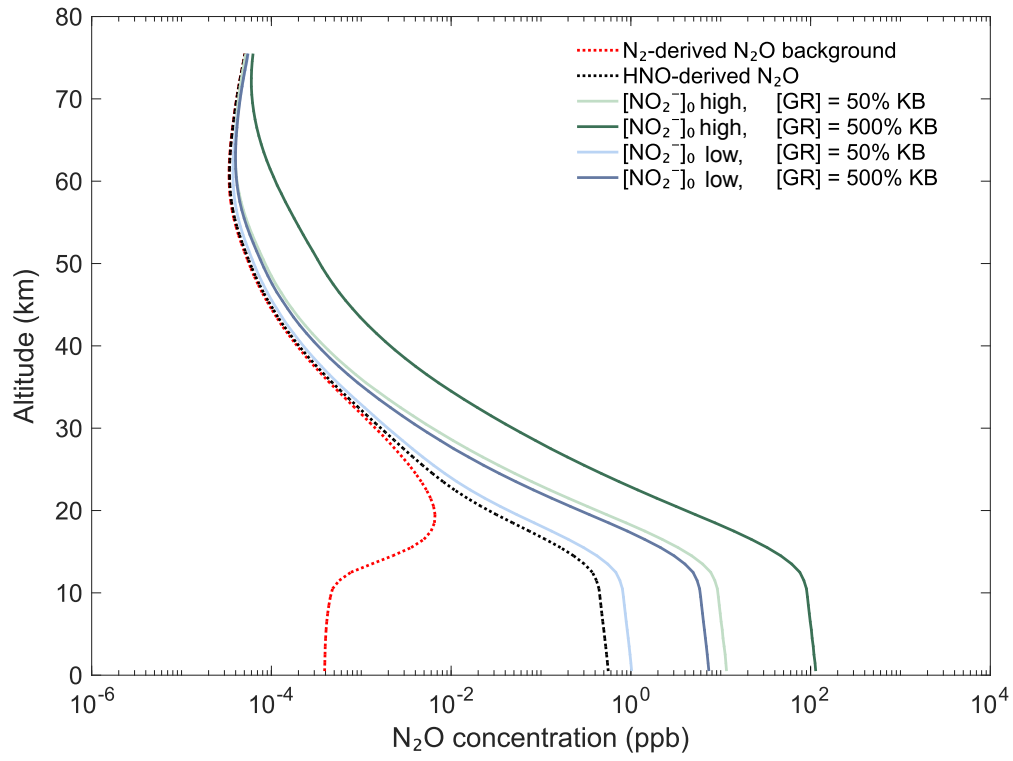


Fig. 2



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Fig. 3

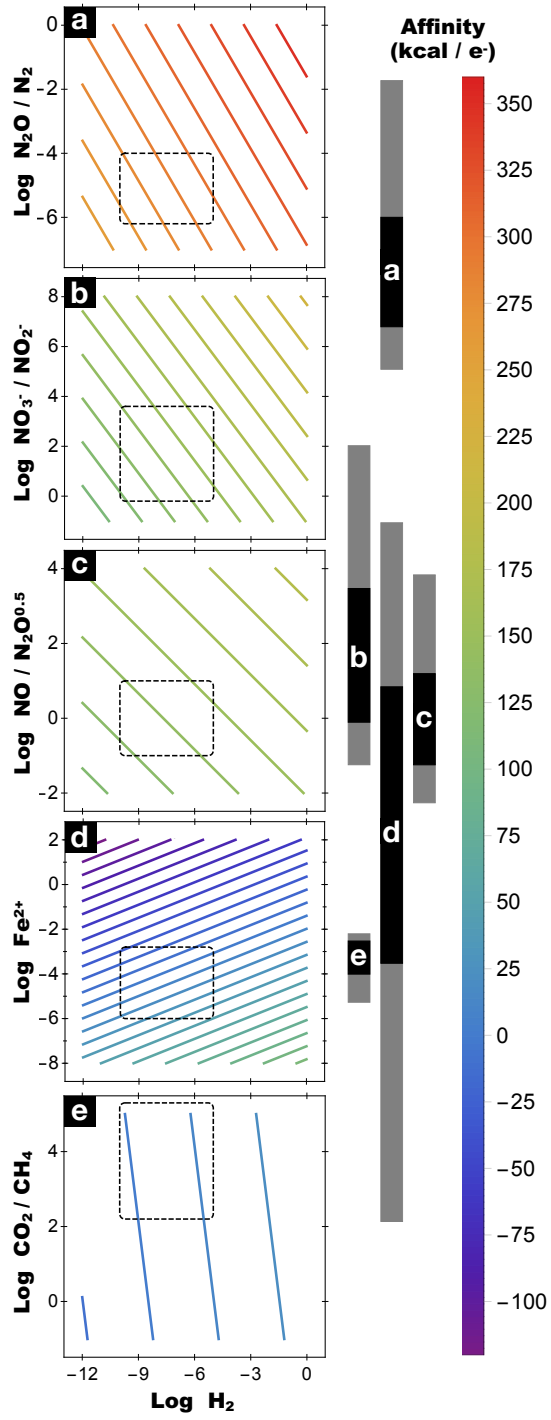


Fig. 4

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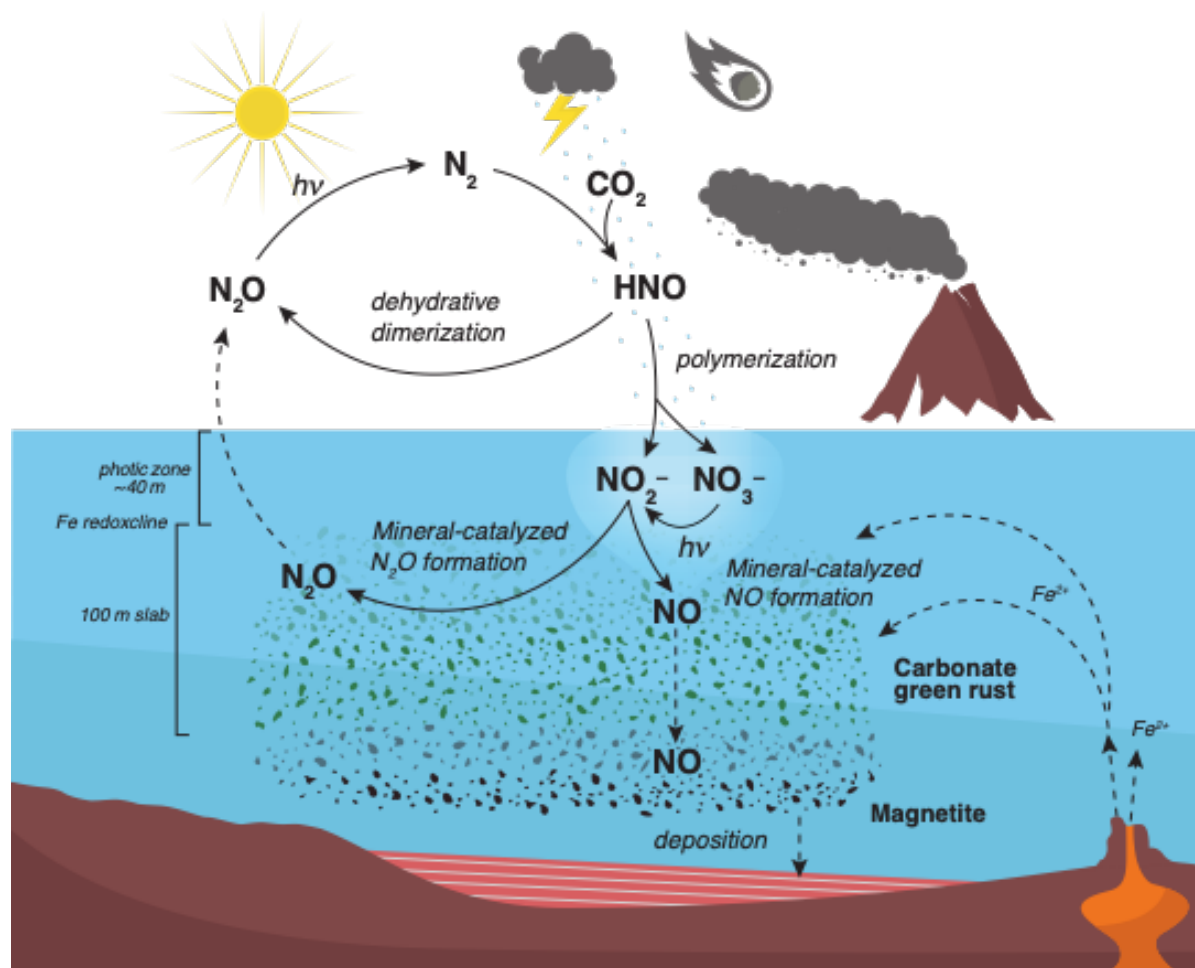


Fig. 5

Figure Legends/Captions

Fig. 1. Molecular NO_x^- consumption and associated NO and N_2O production with Fe minerals or aqueous Fe^{2+} . Dissolved NO_3^- and NO_2^- in anoxic artificial seawater solution were
 335 quantified concomitantly to NO and N_2O in the headspace. The results from incubations with initial NO_x^- concentrations in the *high* (20-35 μM) range are shown (all incubation results in *Supplementary Table 1*). Y-axes depict total quantities (gas + liquid phase) and are sometimes interrupted by breaks to better illustrate changes. **a**, NO_3^- amended microcosms with GR and magnetite. **b**, NO_2^- amended microcosms with GR and magnetite. **c**, NO_2^- reduction and N_2O
 340 production under varying GR mineral surface. After initial rapid NO_2^- consumption, the reaction may become mineral surface-limited hindering the reduction of more NO_2^- . **d**, Range of GR surface area was informed by naturally occurring Fe mineral particles in modern Archean ocean analogues (*Supplementary Information*). **e**, NO formation with 33 μM initial NO_2^- , mineral surface identical to **b**. Arrow indicates addition of concentrated hydrochloric acid resulting in subsequent
 345 mineral particle dissolution and outgassing of NO. Data are presented as mean values \pm *SD* (n = 3).

Fig. 2. Solid phase ratio of reduced and oxidized Fe in GR (a) and magnetite (b). Data were collected after acid digestion at the end of incubations. The dotted lines indicate stoichiometric GR and magnetite, respectively. For total Fe abundances, see *Supplementary Table 2*. **P* = 0.02, ***P* = 0.006, two-sided Student's *t* test. Columns indicate mean values and error bars denote one *SD*
 350 based on n = 2 (a) and n = 3 (b).

Fig. 3. Atmospheric N₂O under influence of mineral-catalyzed N₂O production in the Archean ocean. **a**, Abundances are based on fluxes assuming GR phases in KB equivalents with *low* and *high* seawater NO₂⁻ levels. Background N₂O formation occurred via the reaction N₂ + O(¹D) + M → N₂O + M and O(¹D) was produced by photodissociation of CO₂ with photons more energetic than 167 nm. N₂O can also be derived directly through dehydrative dimerization of HNO that did not polymerize into NO₃⁻ or NO₂⁻ (ref. ³⁸). N₂O profiles do not account for N₂O from the HNO pathway. **b**, Atmospheric N₂O abundances when a fractal haze layer is included in the photochemical model. We adopt the fractal haze predicted by ref. ³⁹ for the Archean atmosphere, which has an optical depth of ~10 at 200 nm and only ~0.6 at 500 nm. Bars at the right illustrate relative haze absorption based on data from ref. ³⁹. N₂O builds up to much higher abundances because of the strong shielding effect of haze.

Fig. 4. Affinity landscapes. **a**, Reduction of N₂O gas by molecular hydrogen (H₂). **b**, Reduction of NO₃⁻ by H₂. **c**, Reduction of NO by H₂. **d**, Magnetite reduction by H₂ at pH 7.3. **e**, CO₂ reduction by H₂ (methanogenesis). The colored curves in each plot represent lines of constant affinity, set 10 kcal apart. Black boxes represent regions of likely species activities; i.e., each of these regions is aligned on the scale bar to the right for easier comparison between reactions. More detail to the reaction stoichiometry in the *Supplementary Information*.

Fig. 5. Schematic depiction of mineral-catalyzed NO and N₂O formation at the junction of the early nitrogen and iron cycle. Heat shock reactions, as stimulated by galactic cosmic rays, meteoritic impact plumes, volcanic and thunderstorm lightening, produced the central precursor HNO, which dimerized directly to N₂O or polymerized into NO₂⁻ and NO₃⁻ (ref. ³⁸). These nitrogen oxides became interspersed into the surface ocean as plumes by rainout. Any NO₃⁻ constituted a source of NO₂⁻ through photo-reduction⁴. Upwelling Fe²⁺ precipitated into Fe oxyhydroxides and GR in the Fe redoxcline. Driven by Fe mineral phases, NO₂⁻ was reduced to N₂O. The abiotic nitrogen cycle was closed by photolytic destruction of emitted N₂O to N₂. As a byproduct, NO molecules remained bound at mineral particles as nitrosyl. This way, GR may have served as NO shuttle enabling transport into the deep ocean. Sinking GR transformed into Fe²⁺ carbonates, Fe²⁺ silicates, and magnetite, which then were deposited. Dashed lines mark diffusive or gravitational transport, whereas solid lines indicate chemical reactions.

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560

Methods

565 *Mineral synthesis.* Carbonate green rust was synthesized according to the following. In brief, 18.2 M Ω -cm water was made anoxic by boiling and sparging with CO₂/N₂ (20:80). An anoxic 0.4 M Fe²⁺/Fe³⁺ solution (0.1 L) was prepared by mixing 7.42 g of FeSO₄·7 H₂O (>99%, Sigma Aldrich) and 2.66 g of anhydrous Fe₂(SO₄)₃ (97%, Sigma Aldrich). An alkaline, anoxic solution of 0.466 M Na₂CO₃ (Fisher Scientific) in 0.8 M NaOH (Fisher Scientific) (0.1 L) was prepared by dissolving
570 NaOH first under a constant stream of CO₂/N₂ gas, after which Na₂CO₃ was added. Both solutions were stirred continuously at 500 rpm. The alkaline solution was injected into the Fe²⁺/Fe³⁺ solution using CO₂/N₂ flushed syringes. Precipitation occurred immediately and the suspension was stirred in the dark for 24 hours. Magnetite was synthesized as nanoparticles following previously described protocols^{71,72}.

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Mineral harvest. Green rust and magnetite precipitates were washed in an anaerobic chamber (0.5% H₂ in N₂, Coy Laboratory Products) using a vacuum filtration unit (Nalgene, Mfr # 130-4020) and 0.45 μ m cellulose-acetate filter membranes (Sartorius). Anoxic 18.2 M Ω -cm water was poured onto the precipitates for a minimum of 8 cycles (pouring followed by extracting water).

580 When filtrate flow ceased to a minimum, the wet mineral paste was removed from the filter. Wet carbonate green rust (73.8% water content) and wet magnetite (47.2% water content) were added to microcosms. The wet minerals were also used for mineralogical characterization (XRD and microscopy). For BET analysis, a defined amount of precipitate was dried in small boxes filled with drierite and placed into the anaerobic chamber. Dry weight was constant after 3-5 days.

585

X-ray diffractometry (XRD). Powder X-ray diffractometry was conducted on a subset of samples after synthesis and washing of the mineral products. A glycerol smear was prepared with 10 mg sample in the anaerobic chamber and sealed in a glass vial to prevent oxidation prior to analysis. To collect the XRD data, the sample was removed from the vial and spread across a quartz zero
590 background plate that was placed onto a horizontal stage in a Bruker D-5000 diffractometer (Bruker, Germany) equipped with a Co K α X-ray tube (30 kV, $\lambda = 1.791\text{\AA}$). Signal peaks were compared to reference diffractograms in the RRUFF database. Instrument broadening was determined by a standard polycrystalline alumina sample. X-ray diffractometry data was analyzed with the CrystalDiffract software version 6.8.2 for Mac.

595

Scanning- and transmission-electron microscopy (SEM, TEM). Samples for SEM imaging were mounted on aluminum pin stubs with double-sided carbon tabs in the glove box and transported in a sealed jar with anoxic atmosphere to the microscope. Samples were not sputter-coated. Images were taken on an XL30 ESEM-FEG (Philips) operated at a 30 kV accelerating voltage and a 21 pA
600 beam current. TEM samples were dispersed on a Lacey Carbon film using 200 mesh copper grids. The instrument used was a CM200 Field Emission Microscope at 200KV (Philips) with $C_s = 1.2$ mm and a PTP resolution of 0.25 nm. Imaging was done on a Gatan Orius CCD system.

Brunauer-Emmett-Teller (BET) surface area measurements. For analysis of the BET surface area,
605 dried mineral precipitate was weighed inside the anaerobic chamber and added to a Florence glass flask that was closed with a rubber stopper for transport to the instrument. BET surface in replicate samples ($n = 3$ for GR, $n = 2$ for magnetite) was quantified using N₂ gas on a Tristar II 3020 analyzer (Micromeritics Inc.). The instrument has a limit of detection of 1 m². Our results

(Supplementary Table 4) are roughly consistent with previously determined BET values of 47 m² g⁻¹ for GR⁷³ and 95 m² g⁻¹ for magnetite⁷⁴.

We performed calculations of the mineral surface area for both minerals to supplement our measurements. To derive the mineral density, we calculated the average crystallite size by inserting the full width at half maximum, as determined from the diffractograms and the Bragg angle of the GR 0 0 3 reflection and of the magnetite 3 1 1 reflection, into the Scherrer equation. Mineral density was calculated using the formula $\rho = (M \times Z) / (V_c \times 0.60225)$ where M is molar mass, Z is the number of molecules per crystallite and V_c is crystallite size. The density and the grain volume as measured by TEM/SEM were used to calculate the grain mass. The final value in m² g⁻¹ was derived from the grain mass and the grain surface area (TEM/SEM). For magnetite, we calculated a surface area of 88.8 m² g⁻¹. For green rust, we calculated a minimum approximation (surface of green rust sheets only) of 27.6 m² g⁻¹ and a maximum approximation (including surface between sheets) of 538.5 m² g⁻¹. Thus, our measured value for magnetite is somewhat lower than the calculated and literature value. The measured value for green rust is in good agreement with the literature value and at the lower end of the calculated range.

Incubation conditions. Interactions of nitrogen oxides with Fe minerals were tested in anoxic microcosms designed to mimic Archean ocean conditions as closely as possible. Borosilicate glass bottles (30-120 mL) were closed with thick butyl rubber stoppers and a headspace of 20% CO₂ in N₂ was used throughout the experiment. All glassware was washed with 2 M HCl prior to use. The liquid phase constituted one third of the microcosm total volume. We used a published recipe for artificial Archean seawater⁷⁵ and omitted any sulfur species (0.56 M NaCl, 0.01 M NaHCO₃). The pH was initially set to 7.3 using a CO₂-HCO₃⁻ buffer. We boiled 18.2 MΩ·cm water and sparged it

with CO₂/N₂ (20:80) while it was cooling on ice. Salts were added during the sparging. The anoxic solution was then dispensed with a pipetor into microcosms in an anaerobic chamber (0.5% H₂ in N₂, Coy Laboratory Products). Nitrate and nitrite stock solutions were prepared with their
635 respective sodium salts NaNO₃ (≥99%, Fisher Scientific) and NaNO₂ (≥97%, Acros Organics) and dissolved in artificial seawater. The solutions were then sparged with N₂ and filter-sterilized (0.8/0.2 μm pore size, VWR) in the anaerobic chamber. Controls with aqueous Fe²⁺ were prepared in anoxic artificial seawater and FeCl₂ (≥99%, Fisher Scientific). Mineral surface area was controlled by the mineral mass added. Wet minerals were weighed in the anaerobic chamber
640 and distributed into microcosms using ethanol-washed plastic spatulas. Prior to start of the incubations with the injection of the NO_x⁻ solution, mineral agglomerates were dissipated in an ultrasonic bath (Ultrasonic cleaner 2510, Branson Ultrasonics). Microcosms were shaken at 250 rpm in the dark and at room temperature over the whole duration of the experiment. Controls were incubated in the anaerobic chamber to test for potential leaking through stoppers, which did not
645 occur. To dissolve mineral particles during incubations, 1.6 mL of concentrated 12N HCl or 37N H₂SO₄ was slowly injected through bottle septa to 10 mL mineral-seawater suspension. Dissolution of solid particles occurred instantly and the liquid first turned turbid orange-green and then clear green-yellow (HCl) or light yellow (H₂SO₄) within 15-20 hours. The last measurement of NO in the headspace was taken when the liquid was clear (*Supplementary Fig. 5*).

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Dissolved nitrite, nitrate, ammonium, ferrous and ferric iron measurements. All dissolved analytes were quantified spectrometrically with plate assays. Nitrite in solution was quantified with the Griess reagent (Promega, Kit G2930). Nitrate was first reduced to nitrite by vanadium(III) chloride and then quantified as nitrite⁷⁶. Ammonium production was verified with the salicylic acid assay⁷⁷.

655 To determine the $\text{Fe}^{2+}/\text{Fe}^{3+}$ solid phase ratio, mineral particles were settled, after which supernatant artificial seawater was removed from the microcosms. Anoxic acidic extraction of green rust and magnetite was conducted by completely dissolving mineral samples in 6 M HCl, followed by incubation at 70 °C for 24 hours under 100% N_2 headspace to prevent abiotic oxidation⁷¹. The samples were then further diluted 1:10 in 1 M HCl. Ferrous and ferric ions in the extracts were
660 measured by reaction with ferrozine after the method of Stookey⁷⁸.

N₂O gas measurements. To quantify N_2O production, 200 μL of headspace gas was sampled with a gas-tight syringe (VICI Precision Sampling) and injected into a gas chromatograph (GC, SRI Instruments) equipped with an electron-capture detector (ECD). Two continuous HayeSep-D
665 columns were kept at 90°C (oven temperature) and N_2 (UHP grade 99.999%, Praxair Inc.) was used as carrier gas. The ECD current was 250 mV and the ECD cell was kept at 350°C. The N_2O measurements were calibrated using customized standard mixtures (Scott Specialty Gases, accuracy $\pm 5\%$) over a range of 0.25–100 ppmv. Gas accumulation in the microcosms was monitored over time. Gas concentrations were corrected using Henry's law and the dimensionless
670 concentration constant $k_H^{cc}(\text{N}_2\text{O}) = 0.6112$ to account for gas partitioning into the aqueous phase at 25°C.

NO gas measurements. Nitric oxide (NO) was quantified in the microcosm headspace with a chemiluminescence-based analyzer (LMA-3D NO_2 analyzer, Unisearch Associates Inc.).
675 Headspace gas (50 μL) was withdrawn with a CO_2 - N_2 -flushed gas-tight syringe and injected into the analyzer. The injection port was customized to fit the injection volume and consisted of a T-junction with an air filter at one and a septum at the other end. An internal pump generated

consistent airflow. In short, sample NO was oxidized to NO₂ by a CrO₃ catalyst. The NO₂ flew across a fabric wick saturated with a Luminol solution. Luminol was obtained from Drummond
680 Technology Inc. (Canada). Readings were corrected for background NO₂ every 15 minutes (“zeroing”). Shell airflow rate was kept at 500 mL min⁻¹ and the span potentiometer was set to 8. Measurements were calibrated with a 0.1 ppm NO (in N₂) standard (<0.0005 ppm NO₂, Scott-Marine Inc.) over a range of 5–1,000 ppbv. Gas concentrations were corrected using Henry’s law and the dimensionless concentration constant $k_H^{cc}(\text{NO}) = 0.0465$ to account for gas partitioning
685 into the aqueous phase at 25°C.

Flux modeling. The partial fluxes of the overall flux balance $\Phi_{\text{sed}} + \Phi_{\text{par}} = \Phi_{\text{atm}}$ were normalized to a 100 m vertical slab with 1 m² basis. The upper 100 m ocean water are typically considered as well-mixed, hence, reactant and catalyst are homogeneously distributed in that space. Depending
690 on the water depth, a portion of the NO_x could reach the ocean floor, which is why we added a generic sediment flux (Φ_{sed}) to the balance equation. The dominant flux would emerge from floating GR mineral particles (Φ_{par}) that are distributed along the water column. For simplicity, we assume that all GR had aged into magnetite in the sediment and all floating particles were GR phases. We refer to the *Supplementary Information* for a more detailed description of the
695 derivation of the partial fluxes, the error propagation method, and thermodynamic calculations.

Photochemical modeling. We have used an atmospheric photochemistry model to simulate the effects of N₂O emission into an anoxic atmosphere akin to the Archean Earth’s condition. The photochemistry model used^{37,79} has been validated by computing the atmospheric compositions of
700 present-day Earth and Mars, as the outputs agreed with the observations of major trace gases in

Earth's and Mars' atmospheres⁸⁰. For example, Figure 2-2 of ref. ⁸⁰ shows that the model reproduces the vertical profile of the mixing ratio of N₂O in modern Earth's atmosphere using a globally averaged emission rate (1×10^9 molecules cm⁻² s⁻¹). For this work, we simulate a 1-bar atmosphere of 95% N₂ and 5% CO₂ to approximate the anoxic and CO₂-rich environment of the
705 Archean Earth⁸¹. We have used the UV and optical spectrum of the present-day Sun to drive the photochemical model because the solar flux in 200-230 nm, which causes the photodissociation of N₂O, depends weakly on the age⁸². We have updated the absorption cross sections of CO₂ and H₂O using the latest experimental results⁸³. We assume a surface temperature of 288 K and a stratospheric temperature of 200 K, and adopt the eddy diffusion coefficient derived from the
710 number density profiles of trace gases on Earth⁸⁴. The photochemistry model includes a comprehensive reaction network for O, H, C, N, and S species including sulfur and sulfuric acid aerosols, and includes volcanic outgassing of CO, H₂, SO₂, and H₂S. The outgassing rate is not high enough to produce a H₂SO₄ aerosol layer in the atmosphere and these gases do not impact the lifetime of N₂O significantly.

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Data availability

All data are provided in the manuscript and the supplementary materials, and are available as raw data on the figshare platform (<https://doi.org/10.6084/m9.figshare.20740204.v2>) or upon request
720 from the authors.

Code availability

The code for the photochemical model will be shared by the authors upon request.

Methods-only References

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