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The fate of organic carbon in marine sediments - new insights from recent data and analysis

<u>Authors</u>: LaRowe, D.E.^{1*}, Arndt, S.², Bradley, J.A.^{3,4}, Estes, E. R.⁵, Hoarfrost, A.⁶, Lang, S.Q.⁷,
 Lloyd, K.G.⁸, Mahmoudi, N.⁹, Orsi, W.D.^{10,11}, Shah Walter, S. R.¹², Steen, A.D.^{8,13}, Zhao, R.¹⁴

- 7 *Corresponding author
- 8

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- 9 1. Department of Earth Sciences, University of Southern California, Los Angeles, CA, 90089
- 10 USA, phone:615-438-8646, fax: 213-740-8801, email: larowe@usc.edu
- 11 2. BGeosys, Department of Geosciences, Environment and Society, Université libre de
- 12 Bruxelles, Ave F.D. Roosevelt 50 CP160/02, 1050 Brussels, Belgium, phone: +32 (2) 650 22 13 ameil: Sandra Arndt @ulb as ha
- 13 17, email: <u>Sandra.Arndt@ulb.ac.be</u>
- 14 3. School of Geography, Queen Mary University of London, Mile End Road, London E1 4NS,
- 15 United Kingdom, phone: +44 (0)20 7882 8417, fax: +44 (0)20 7882 7032, email:
- 16 james.bradley@qmul.ac.uk
- 17 4. Interface Geochemistry, GFZ German Research Centre for Geosciences, 14473 Potsdam,
- 18 Germany.email: james.bradley@gfz-potsdam.de phone: +49 331 288-28971
- 19 5. International Ocean Discovery Program, Texas A&M University, 1000 Discovery Drive,
- College Station, TX, 77845, USA; estes@iodp.tamu.edu, Office phone: 979-458-2730, Fax: 979 845-4857
- 22 6. Department of Biochemistry and Microbiology, Rutgers University, School of Environmental
- and Biological Sciences, 76 Lipman Drive, Suite 218, New Brunswick, NJ 08901,
- 24 ahoarfrost@bromberglab.org
- 25 7. School of the Earth, Ocean, and Environment, University of South Carolina, 701 Sumter St.,
- 26 Columbia, SC, 29208, USA, Phone: (803)777-8832, Fax: (803)777-6610, <u>slang@geol.sc.edu</u>
- 27 8. Microbiology Department, University of Tennessee, 1311 Cumberland Avenue
- 28 307 Ken and Blaire Mossman Bldg. Knoxville TN 37996-1937 Phone: 865-974-4224, Email:
- 29 <u>klloyd@utk.edu</u>, Fax: 865-974-4007
- 30 9. Department of Earth & Planetary Sciences, McGill University, 3450 University Street,
- 31 Montreal, Quebec, Canada H3A 0E8, Tel: 514-398-2722, Fax: 514-398-4680, email:
- 32 <u>nagissa.mahmoudi@mcgill.ca</u>
- 10. Department of Earth and Environmental Sciences, Paleontology & Geobiology, Ludwig Maximilians-Universität München, 80333 Munich, Germany.
- 35 11. GeoBio-Center^{LMU}, Ludwig-Maximilians-Universität München, 80333 Munich, Germany
- 36 12. School of Marine Science and Policy, University of Delaware, 700 Pilottown Road, Lewes,
- 37 DE 19958, USA, phone: 302-645-4262, email: suni@udel.edu
- 38 13. Department of Earth and Planetary Sciences, University of Tennessee, 1412 Circle Drive
- 39 Knoxville, TN 37996-1410, Phone: (865) 974-4014, Email: <u>asteen1@utk.edu</u>
- 40 14. School of Marine Science and Policy, University of Delaware, Lewes, DE 19958, USA
- 41 Tel: +1 302-727-2918, Fax: 645-4007, Email: <u>ruizhao@udel.edu</u>

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47 Abstract

48 Organic carbon in marine sediments is a critical component of the global carbon cycle, and 49 its degradation influences a wide range of phenomena, including the magnitude of carbon 50 sequestration over geologic timescales, the recycling of inorganic carbon and nutrients, the 51 dissolution and precipitation of carbonates, the production of methane and the nature of the 52 seafloor biosphere. Although much has been learned about the factors that promote and hinder 53 rates of organic carbon degradation in natural systems, the controls on the distribution of organic 54 carbon in modern and ancient sediments are still not fully understood. In this review, we 55 summarize how recent findings are changing entrenched perspectives on organic matter 56 degradation in marine sediments: a shift from a structurally-based chemical reactivity viewpoint 57 towards an emerging acceptance of the role of the ecosystem in organic matter degradation rates. 58 That is, organic carbon has a range of reactivities determined by not only the nature of the organic 59 compounds, but by the biological, geochemical, and physical attributes of its environment. This 60 shift in mindset has gradually come about due to a greater diversity of sample sites, the molecular 61 revolution in biology, discoveries concerning the extent and limits of life, advances in quantitative 62 modeling, investigations of ocean carbon cycling under a variety of extreme paleo-conditions (e.g. greenhouse environments, euxinic/anoxic oceans), the application of novel analytical techniques 63 and interdisciplinary efforts. Adopting this view across scientific disciplines will enable additional 64 progress in understanding how marine sediments influence the global carbon cycle. 65 66

Keywords: organic carbon; marine sediments; reactivity; microorganisms; deep biosphere
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69 **1. Introduction**

70 To contextualize the recent biogeochemical breakthroughs that have been made regarding 71 the fate of organic carbon in marine sediments, the first section of this review consists of a 72 summary of its role in the global carbon cycle and how this has varied over space and time. 73 Secondly, we describe the sources of new data that are shaping the transformation in how we think 74 about organic carbon in the marine system. This is followed by a section on organic carbon in 75 adjacent environments such as hydrothermal systems and the ocean basement, and then a brief 76 overview of modelling advances. The topics covered in this communication are summarized in the 77 schematic shown in Figure 1.

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79 **1.1 Sedimentary organics and the global carbon cycle**

80 Organic carbon (OC) degradation in marine sediments is a critical component of the global 81 carbon cycle and is intricately linked to Earth's climate (Berner and Canfield, 1989; Berner, 1990; 82 Siegenthaler and Sarmiento, 1993; Archer and Maier-Reimer, 1994; Mackenzie et al., 2004; 83 Ridgwell and Zeebe, 2005; Ridgwell and Hargreaves, 2007; Hülse et al., 2017). The specific 84 consequences of this process on biogeochemical cycles vary considerably depending on location as well as the temporal and spatial scales of concern. In the upper few meters of relatively recently 85 deposited sediment, the oxidation of organic carbon controls the fluxes of oxygen and nutrients 86 87 across the sediment-water interface (SWI), ultimately impacting primary productivity in the water 88 column (Van Cappellen and Ingall, 1994; Lenton and Watson, 2000). As organic carbon is 89 aerobically oxidized, pore water pH is decreased, potentially leading to dissolution of calcium 90 carbonate and amplifying the direct effect of OC oxidation on the carbon cycle (Emerson and Bender, 1981; Hales and Emerson, 1996). Deeper in sediments, organic carbon that escapes 91 92 oxidation can still be microbially reduced to CH₄, and under some conditions, fuel the slow build-

93 up of methane hydrates (e.g. Wallmann et al., 2012). Although hydrates represent a relatively small 94 carbon reservoir, perturbations in temperature and/or pressure can render hydrates unstable, 95 potentially leading to sudden transfers of carbon back into the ocean-atmosphere system that would 96 have important consequences for global carbon cycling and climate (Ruppel and Kessler, 2017). 97 The small fraction of photosynthetically produced organic carbon that entirely escapes degradation 98 and thus is buried over longer timescales in sediments helps to modulate the long-term evolution 99 of atmospheric CO₂, has enabled oxygen to accumulate in the atmosphere (e.g. Berner, 2003) and 100 has led to the formation of large reservoirs of hydrocarbons that provide most of the energy that 101 humans use. Whether marine sediment organic carbon contributes to water column anoxia, 102 becomes trapped in a clathrate, is transformed into petroleum or ends up as atmospheric CO₂ is 103 ultimately based on the flux of particulate organic carbon (POC) to sediments and the many factors 104 that dictate its rate of degradation.

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106 **1.2 POC fluxes and degradation rates**

107 The concentrations of POC shown in Figure 2 illustrate that the flux of organic carbon to 108 and through sediments vary tremendously (Seiter et al., 2004; Wallmann et al., 2012; LaRowe et 109 al., 2020). In this figure, it can be seen that POC concentrations are highest in recently deposited 110 sediment underlying northern high-latitude and south-east Asian coastal regions and the Humboldt 111 and South Equatorial currents to the west of Peru. They are generally lowest in abyssal regions far 112 from land, and in older and deeper sediments. This span of concentrations is due to variable 113 depositional fluxes and degradation rates.

114 Field observations have revealed that rates of POC degradation are not constant (e.g. 115 (Canfield et al., 2005), leading to burial rates that vary significantly in space (e.g. Canfield, 1994; Blair and Aller, 2012) and time (e.g. Arthur et al., 1985). In fact, first-order rate constants for POC 116 117 degradation rates derived from field and laboratory data span over eight orders of magnitude, with older sediments having the smallest values (Middelburg, 1989). Many different factors have been 118 119 invoked to explain this variability. They include, but are not limited to, organic matter composition (e.g. Westrich and Berner, 1984; Hedges et al., 1988; Tegelaar et al., 1989; Cowie et al., 1992; 120 121 Canfield, 1994; Hedges and Keil, 1995), electron acceptor (EA) availability (e.g. Demaison and 122 Moore, 1980; Emerson, 1985; Canfield, 1994; Hedges and Keil, 1995; Dauwe et al., 2001), benthic 123 microbial community composition and functional capacities (e.g. Canfield, 1994; Arnosti, 2011; 124 Steen et al., 2019), microbial inhibition by specific metabolites (e.g. Aller and Aller, 1998), priming (e.g. Stevenson, 1986; Graf, 1992; Aller et al., 1996; Sun et al., 2002a; van Nugteren et 125 126 al., 2009), physical and physicochemical protection (e.g. Keil and Kirchman, 1994; Mayer, 1994; 127 Kennedy et al., 2002; Estes et al., 2019; Hemingway et al., 2019), sediment deposition rate (e.g. 128 Müller and Suess, 1979; Calvert and Pedersen, 1992b; Tromp et al., 1995) and macrobenthic 129 activity (e.g. Aller, 1982; Aller and Cochran, 2019; Middelburg, 2019).

130 The rates of POC degradation in marine sediments are first and foremost controlled by the 131 source and transport of organic carbon. A higher lateral and/or vertical (i.e. sedimentation) 132 transport rate not only increases the deposition flux but also influences the quality of the organic carbon deposited, since the degree of pelagic degradation during transport through the water 133 134 column - and thus the nutritive value of POC - is reduced with extended sinking times. A 135 compilation of deep sediment trap data reveals large regional variations in vertical export 136 efficiency (Lutz et al., 2002; Henson et al., 2012a; Wilson et al., 2012). For instance, the fraction 137 of the organic carbon export flux that reaches water depths greater than 1.5 km varies between 138 0.28 and 30% (5.7% average) of the POC that escapes the photic zone (Lutz et al., 2002). Although

139 differences in export efficiencies are most likely not attributable to a single process, factors 140 invoked to explain variations in export efficiency include sinking rates and ballast (e.g. Armstrong et al., 2002; Francois et al., 2002; Klaas and Archer, 2002; Lutz et al., 2002), the seasonality of 141 142 export production (Antia et al., 2001) and ecosystem structure (Buesseler et al., 2008; Lam et al., 143 2011; Henson et al., 2012a; Mayor et al., 2012). And while export of large particles from surface 144 waters has been extensively investigated, it has also been shown that very small particles (< $1 \mu m$) 145 contribute to the chemical composition of exported POC (Close et al., 2013). In addition to the 146 vertical transport of POC from the euphotic zone, the efficient lateral transport of POC in nepheloid 147 layers, benthic storms, mud waves, strong (bottom) water currents or mass wasting events can support an important fraction of the POC deposition flux to continental slope and abyssal 148 149 sediments in the vicinity of dynamic continental margins (e.g. Ohkouchi et al., 2002; Levin and 150 Gooday, 2003; Mollenhauer et al., 2003; Inthorn et al., 2006b; Mollenhauer et al., 2007; Kusch et 151 al., 2010; Henson et al., 2012b; Bao et al., 2019). The efficiency of terrestrial organic matter export 152 from land is even more difficult to predict, given variable controls on residence times in soils, 153 followed by widely varying degrees of alteration during transport through the 154 riverine/estuarine/deltaic system and across the continental shelf (Hedges et al., 1997; Schlünz 155 and Schneider, 2000; Bianchi, 2011; Blair and Aller, 2011; Bauer et al., 2013a; Regnier et al., 2013; Canuel and Hardison, 2016; Kandasamy and Nagender Nath, 2016). Organic carbon is also 156 157 produced in sediments, by phototrophs in shallow waters (Middelburg, 2018, 2019) and chemoautotrophically (Veuger et al., 2012; Sweetman et al., 2017). Although the exact 158 159 mechanisms controlling the high variability in POC degradation are not well understood, the 160 variety of environments that have been sampled and examined in recent years has provided new 161 insights to constrain which variables are most important in particular settings.

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163 **1.3 Observations: Increased diversity of sample sites and data types**

164 Over the last several decades, scientific drilling of the seafloor by the International Ocean Discovery Program (IODP) and its precursors, the Integrated Ocean Drilling Program (IODP), the 165 Ocean Drilling Program (ODP) and the Deep Sea Drilling Program (DSDP), have enabled critical 166 167 expansion of sampled environments and new data that have dramatically increased our knowledge of the distribution of organic carbon in marine sediments. The sample sites targeted by most of 168 169 these expeditions were motivated by scientific questions about the structure of ocean crust, past 170 climate, and tectonics, leading to drilling locations and sampling methods that were optimized for study of these topics. Although total organic carbon was regularly measured on these samples, 171 172 tools to study the agents of their transformation, i.e. microorganisms, were relatively rudimentary 173 until culture-independent methods (e.g. DNA and RNA sequencing) became widely available in 174 the early 2000's. Much of the new understanding of controls on organic carbon degradation in marine sediments has to do with recent insight into the role of microorganisms. As pointed out in 175 176 the ODP Leg 201 summary (D'Hondt et al., 2003), this is despite the fact that deep-sea drilling had revealed indirect evidence of microbial activity as early as the 1980s (Oremland et al., 1982; 177 178 Whelan et al., 1986; Tarafa et al., 1987) and, by the 1990s, microbial abundances (see Parkes et 179 al., 2014)) for a review). Remarkably, ODP Leg 201, the first ODP cruise dedicated to studying 180 microorganisms, did not sail until 2002. Since this time, a number of IODP expeditions have been focused on studying the limits to life: Expeditions 329 (South Pacific Gyre Subseafloor Life), 331 181 182 (Deep Hot Biosphere), 336 (Mid-Atlantic Ridge Microbiology), 337 (Deep Coalbed Biosphere off 183 Shimokita), 357 (Atlantis Massif Seafloor Processes: Serpentinization and Life), 370 184 (Temperature Limit of the Deep Biosphere off Muroto), and 385 (Guaymas Basin Tectonics and

Biosphere). This explicit exploration of microbial communities further informs the perspective
that the reactivity of organic carbon in marine sediments is an ecosystem property (Middelburg,
2018).

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189 **2. Ecosystem nature of the problem**

190 Organic carbon in natural systems is often referred to as being either labile or recalcitrant 191 (e.g. Hedges et al., 2000). Some have expended considerable effort expanding this classification 192 to intermediate states such as semi-labile and semi-recalcitrant (Hansell, 2013). This reductionist 193 classification system implies that the degradation rate of organic compounds is determined from 194 specific attributes of the chemical compounds themselves. While the reactivity of organic 195 compounds is certainly related to their chemical composition and structure, numerous factors have 196 been invoked to explain the reactivity of organic carbon that are not intrinsic to the compounds, 197 but rather the properties of the surrounding environment (e.g. see Hedges et al., 2000; Burdige, 198 2007b; Arndt et al., 2013; Middelburg, 2018, 2019 and references therein). Examples include 199 temperature, microbial community structure and benthic ecosystem, mineral types and surface 200 area, redox state, light, nutrient availability, pH, salinity, porosity, permeability, water content and 201 oxygen exposure time. Because these variables are biophysiochemical properties of the environment, the reactivity of organic matter is determined by the interaction between the 202 203 compound and its ecosystem, not just the chemical nature of the organic compounds. Indeed, a 204 particular organic compound can be extremely resistant to degradation under certain conditions, 205 and be rapidly degraded in a different physiochemical context. For example, aerobic heterotrophic 206 bacteria in incubation experiments have been shown to oxidize and incorporate ¹⁴C-free organic 207 carbon preserved in marine black shales for millions of years (Petsch et al., 2001). This idea of 208 ecosystem properties governing organic compound reactivity is not a new one (Middelburg et al., 209 1993; Canfield, 1994; Harvey et al., 1995; Mayer, 1995), but it is only recently becoming the new 210 paradigm in the soil science community (Marschner et al., 2008a; Kleber and Johnson, 2010; 211 Schmidt et al., 2011; Lehmann and Kleber, 2015), and there are indication that it should prevail 212 among sediment scientists as well (Eglington and Repeta, 2014; Middelburg, 2018, 2019). In fact, 213 we suggest that the terms *labile* and *recalcitrant* be retired and replaced with the single term 214 reactivity. Similarly, since the fate of organic carbon in natural systems can include oxidation to 215 CO₂, hydrolysis and fermentation into smaller organics, reduction to methane, adsorption to 216 surfaces, incorporation into biomass and conversion to petroleum products and complex organic compounds, we recommend that the more inclusive term *transformation* be used to refer to the 217 218 fate of organic carbon rather than the relatively restrictive term *mineralization*. In the following 219 sections, we highlight some of the recent research that seeks to determine how a variety of 220 ecosystem properties influence the rates of organic carbon degradation.

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222 **2.1 Source and transit path**

223 A large fraction of the organic carbon that is buried in marine sediments originates from 224 photosynthetic activity in the terrestrial (net primary productivity (NPP) = 56 Pg C yr⁻¹) or marine 225 biosphere (NPP = 49 Pg C yr⁻¹) (Field et al., 1998). In addition, organic carbon can be transported to sediments by other means including the weathering of ancient organic carbon in rocks (Blair et 226 al., 2003), the remobilization of organic matter from thawing permafrost (Mann et al., 2015) and 227 228 resuspension of already-deposited carbon (see below). Autotrophic and Chemoautotrophic 229 production as well as secondary production by microorganisms and animals can also contribute to the organic carbon deposition flux (Eglinton and Repeta, 2010; Middelburg, 2011; Lengger et al., 230

231 2019). Approximately two-thirds of terrestrially produced OC is rapidly degraded within soils or 232 glacial environments. The remainder that escapes immediate degradation is either partially 233 degraded or temporarily stored before being transported downstream with old, weathered OC 234 within lakes, streams, rivers, estuaries, deltas, fjords, and ultimately, the oceans (Regnier et al., 2013). An estimated 1.9 Pg C yr⁻¹ ±1.0 Pg C yr⁻¹ of total soil carbon (mostly POC and dissolved 235 organic carbon, DOC, but also dissolved inorganic carbon, DIC) is exported to inland waters. Only 236 237 0.45 Pg C yr⁻¹ of the terrestrial-derived OC reaches the coastal ocean and 0.1-0.35 Pg C yr⁻¹ makes 238 it to the open ocean (Bauer et al., 2013b; Regnier et al., 2013). Thus, the land-ocean transition zone 239 acts as an important modulator of OC fluxes that not only further degrades and transforms 240 terrestrial OC inputs, but also stores OC on short and long timescales (Canuel and Hardison, 2016). 241 However, the exact amounts of OC that are degraded and temporarily or permanently preserved 242 during transit from land to ocean remain unknown (Regnier et al., 2013).

243 Similar to terrestrially-derived organic carbon, a large fraction (80-90%) of the OC 244 produced in the surface ocean is rapidly oxidized (Dunne et al., 2007; Hansell and Carlson, 2015). 245 Just over one-quarter (27%) of OC produced in the marine photic zone is DOC that can be 246 transported to the deep ocean by convection and mixing. While most of the exported DOC is degraded at mid-depths, an estimated 0.1 Pg C yr⁻¹ is contributed to the large, apparently 247 248 unreactive, deep ocean DOC reservoir (~ 680 Pg) that persists through multiple ocean mixing 249 cycles (Hansell and Carlson, 2015). As POC sinks to the ocean floor, it can be laterally transported 250 by ocean currents (Eglinton and Repeta, 2004) as it is further degraded. Degradation during burial 251 further reduces this flux such that < 0.3% of the original exported flux is ultimately sequestered in 252 deep marine sediments (Hedges and Keil, 1995; Eglinton and Repeta, 2004; Burdige, 2007b; 253 Dunne et al., 2007; Middelburg and Meysman, 2007; Honjo et al., 2008).

254 In addition to the vertical transport from the photic zone to sediments, POC can also be 255 transported after deposition on the seafloor via bioturbation and lateral motion following the 256 resuspension of sediments into the water column. Bioturbation, the biological reworking of 257 sediments near the SWI by macrofauna such as polychaetes and bivalves (Rhoads, 1974a; Aller, 258 1982), can exert an important influence of organic carbon transformation (Meysman et al., 2006; 259 Middelburg, 2018) in the upper 20 cm of sediments (Boudreau, 1994, 1998). These animals can 260 have a complex impact on sediment POC – in some cases accelerating POC degradation through grazing, the redistribution of particles and reintroduction of electron acceptors and DOC, 261 262 (Kristensen, 1985; Rice, 1986; Kemp, 1988; Aller, 1994; Aller and Aller, 1998; Kristensen and 263 Holmer, 2001a; Kristensen et al., 2011) and, in other cases, depositing organics in the form of tube 264 casings that can inhibit organic transformation (Kristensen et al., 1992; Kristensen, 2001). 265 Generally, it seems that bioturbation accelerates POC degradation (Rhoads, 1974b; Findlay and 266 Tenore, 1982; Aller and Aller, 1986; Herman et al., 1999; Aller et al., 2001; Kristensen and 267 Holmer, 2001b; Kristensen and Kostka, 2005; Aller and Cochran, 2019). Furthermore, benthic 268 macrofauna, in addition to being a source of OC, can affect sediment resuspension (Aller and 269 Cochran, 2019) by altering the bulk properties of sediments (Eckman et al., 1981; Rhoads et al., 270 1984), which in turn influences their lateral transport.

Sediments, and the POC within, on continental shelves and slopes can also be mobilized after initial deposition and redeposited under open-ocean waters. Not only does this process move organic carbon laterally to parts of the seafloor that typically have low POC deposition rates (Jahnke et al., 1990; Walsh, 1991; Bauer and Druffel, 1998; Ransom et al., 1998a; Thomsen and van Weering, 1998; Jahnke and Jahnke, 2000; Romankevich et al., 2009; Hwang et al., 2010), but it has the effect of accelerating the oxidation of POC that was buried in margin sediments (see Section 2.5 for a discussion) (de Lange et al., 1987; Prahl et al., 1989; Cowie et al., 1995; Prahl et al., 1997; Cowie et al., 1998; Hoefs et al., 1998; Prahl et al., 2003). However, the scale of this phenomenon, as well as its impact on the global distribution of organic matter reactivity, remains unclear (Eglinton and Repeta, 2014) due to large uncertainties in lateral particle fluxes (Jahnke et al., 1990; Reimers et al., 1992; Inthorn et al., 2006a; Thullner et al., 2009).

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283 **2.2 Organic carbon structure and composition**

284 Although the majority of organic carbon inputs to marine sediments originate as POC, 285 these sources can be transformed and contribute to the DOC pool through microbial activities as 286 well as sorption/desorption from mineral surfaces. Chemically, both POC and DOC range in size 287 and complexity from simple monomers to mixtures of large, complex polymers as well as humic 288 substances (de Leeuw and Largeau, 1993; Hedges et al., 2000), the last of which is an ill-defined 289 molecular group of varying sizes that are solely defined by the extraction procedure used to isolate 290 them. Most POC and DOC in marine sediments cannot be chemically identified at the structural 291 level, leading to terms such as "molecularly uncharacterizable carbon, MUC" (Cowie et al., 1995; Prahl et al., 1997; Wakeham et al., 1997; Burdige and Gardner, 1998; Hedges et al., 2000; Burdige, 292 293 2007b; Burdige and Komada, 2015). Depending on the sample, anywhere from 50-90 wt% of OC 294 in marine sediments is referred to as MUC. Approximately 60-90% of pore water DOC has a 295 molecular weight of less than 3 kDa, and the compounds that can be identified tend to be short-296 chain organic acids such as acetate, amino acids, and carbohydrates (Burdige and Komada, 2015). 297 High-resolution techniques have shown that thousands of organic compounds are found in pore 298 water (e.g. Repeta et al., 2002; Koch et al., 2005; Schmidt et al., 2009; Fox et al., 2018), but many 299 fall into a rather unsatisfactory category known as CRAM, carboxyl-rich alicyclic molecules - a 300 poorly defined pool of organics thought to be less reactive than other DOC compounds (Hertkorn 301 et al., 2006).

302 As with DOC, the bulk of organic compounds in the particulate fraction of marine sediments (i.e. POC) is rarely identified and often said to be 'uncharacterizable', at least with 303 304 respect to partitioning it into compound classes such as amino acids, carbohydrates, lignin or lipids 305 (Cowie et al., 1995; Prahl et al., 1997; Wakeham et al., 1997; Hedges et al., 2000; Benner, 2002; 306 Koch et al., 2005; Burdige, 2007a; Mao et al., 2011; Ball and Aluwihare, 2014; Estes et al., 2019). 307 Although it is not clear why this is the case, one of the explanatory hypotheses, that organics attach 308 to mineral surfaces and condense into large macromolecules that are less accessible by 309 exoenzymes, e.g. (Hedges et al., 1988; Keil and Kirchman, 1994; Hedges et al., 2000), no longer 310 holds in the soil science community. Briefly, it was long thought that complex macromolecular 311 carbon compounds form in soils through abiotic processes and that these large compounds were especially resistant to biological degradation. More recently, however, there is a growing 312 313 consensus that these larger compounds are an artifact of extraction procedures and there is no 314 evidence for their presence *in situ* (Kelleher and Simpson, 2006; Lehmann and Kleber, 2015).

315 The structure and composition of organic matter serves as the first guidepost to determining 316 its reactivity. For instance, relatively weak bonds between monomers render a biopolymer more reactive over a wide range of environmental conditions, than aliphatic moieties cross-linked by 317 318 ether bonds (Tegelaar et al., 1989; de Leeuw and Largeau, 1993). At the most fundamental level, 319 this structural heterogeneity is responsible for the higher reactivities of most algal organic carbon 320 over most terrestrial organic matter (Canuel and Martens, 1996; Hedges and Oades, 1997; Dauwe 321 and Middelburg, 1998; Camacho-Ibar et al., 2003; Burdige, 2005; Dai and Sun, 2007), the 322 selective preservation of certain compounds in the geological record and the widely observed 323 decrease of organic carbon with depth and or transport/burial time (Goth et al., 1988; Tegelaar et 324 al., 1989). However, the relative importance of organic structure and composition in controlling 325 overall preservation/degradation remains unclear (see de Leeuw et al., 2006; Gupta et al., 2007; 326 Gupta, 2015). In addition, because of the extremely limited availability of compound-specific 327 kinetic data (e.g. Ming-Yi et al., 1993; Harvey and Macko, 1997; Sun et al., 2002b), a comprehensive knowledge of organic structure and composition does not help in quantifying 328 329 organic matter reactivity. Ultimately, linking the identity of individual organic compounds to their 330 reactivity would require knowledge of how many other biophysiochemical variables impact it as 331 well.

332333 2.3 Sediment matrix

334 Mineral surfaces are thought to slow or prevent the degradation of organic carbon, largely 335 based on observations that there is a positive correlation between mineral surface area and the 336 amount of organic matter in sediments (e.g. Mayer et al., 1985; Keil et al., 1994; Mayer, 1994). 337 Most hypotheses explaining this phenomenon rest upon the notion that extracellular enzymes 338 cannot access mineral-associated organic compounds. Compounds can associate with minerals via 339 a number of mechanisms, both physical and chemical, including 1) physisorption (physical 340 adsorption) in surface irregularities, 2) strong mineral surface bonding, 3) insertion into clay 341 interlayers, 4) formation of mineral-OC aggregates, 5) co-precipitation with Fe-oxides and other 342 authigenic minerals, 6) seclusion by exopolymeric substances (EPS) and 7) protection within 343 biogenic minerals such as diatom frustules (Keil and Hedges, 1993; Keil et al., 1994; Mayer, 1994; 344 Hedges and Keil, 1995; Ransom et al., 1997; Ransom et al., 1998b; Mayer, 1999; Arnarson and 345 Keil, 2001; Mayer and Xing, 2001; Kennedy et al., 2002; Ingalls et al., 2003; Pacton et al., 2007a; 346 Pacton et al., 2007b; Kennedy and Wagner, 2011; Lalonde et al., 2012; Hemingway et al., 2019). 347 Chemical preservation mechanisms, in particular, are likely important, since OC concentration 348 often correlates more strongly with the abundance of specific mineral classes such as smectite 349 clays (Ransom et al., 1998a), metal oxides (Lalonde et al., 2012; Roy et al., 2013; Barber et al., 350 2017) or tephra (Longman et al., 2019) rather than total mineral surface area, suggesting that the 351 specific chemical bonds formed by these minerals with organic compounds are essential to 352 decreased reactivity.

353 Given the large variety of organic compounds and minerals found in marine sediments, it 354 is probable that all of the mechanistic hypotheses noted above contribute to the long-term preservation of OC. The array of explanations is likely due to the fact that the samples used to 355 356 generate them have come from different places and therefore are characterized by a variety of 357 different biogeochemical conditions and timescales of preservation. Although there is a growing 358 appreciation of the complexity of mineral-organic reactions, disentangling which mechanism 359 operates under what set of environmental conditions is complicated by a lack of data that more 360 fully contextualizes mineralogical observations such as oxygen exposure times (OETs). For example a study examining how OETs affect mineral-organic associations in northeast Pacific 361 362 sediments (Arnarson and Keil, 2007) found that for OETs shorter than a year, OC was mostly in a 363 mineral-free state. For sediments with OETs ranging from years to decades, OC was largely in 364 mineral-organic aggregates. For longer OETs (centuries to millennia), the aggregates broke down and most OC was found sorbed to mineral surfaces or protected inside biogenic diatom frustules 365 (see Section 2.5 for a discussion on electron acceptor availability). 366

The exposure to a variety of geochemical conditions over varying timescales also impacts how solid phases can control organic carbon reactivity. For instance, microorganisms use Fe- and

369 Mn-oxide minerals to oxidize organic carbon in marine sediments (Ehrlich, 1971; Aller, 1980; 370 Berner, 1981; Jones, 1983; Burdige and Nealson, 1986; Lovley, 1991; Thamdrup, 2000; Hyun et al., 2017), especially where manganese- and iron-oxides are abundant or rapidly recycled due to 371 372 fluctuating redox conditions (Sørensen and Jeørgensen, 1987; Aller et al., 1990; Canfield et al., 373 1993). In fact, it has been estimated that 3% of the POC degraded in the top 50 cm of global marine 374 sediments is coupled to Fe-oxide reduction (Thullner et al., 2009), though this is spatially quite 375 variable (Burdige, 2012; Dale et al., 2015). Yet, these same mineral phases provide protection and 376 stabilization of organic matter (Lalonde et al., 2012; Johnson et al., 2015; Barber et al., 2017). 377 Substantially more OC is associated with Fe oxides via inner-sphere complexation in coastal environments with short OETs than in low OC, deep-sea sediment (Barber et al., 2017). 378 379 Experimental evidence additionally demonstrates that the composition of OC matters in 380 determining the balance between degradation and preservation by minerals: the presence of 381 disaccharides inhibits the abiotic degradation of protein by the manganese oxide mineral birnessite 382 (Reardon et al., 2018). Furthermore, carbon compounds can serve as templates for mineral 383 nucleation, and co-precipitate with minerals during mineral growth and aggregation (Mann et al., 384 1993; Moreau et al., 2007; Kleber et al., 2015 and references therein). Beyond redox chemistry, 385 (Taylor, 1995) showed that under some conditions sorbed proteins are hydrolyzed much faster 386 than dissolved proteins, indicating that surface sorption can in fact enhance organic carbon 387 degradation.

388 Generalizations about organic-mineral interactions are further complicated by the fact that 389 sediment types may vary on the centimeter scale and might be dominated by an array of mineral 390 types that have distinct physiochemical properties. Common sediment lithologies include chert, 391 siliceous ooze, carbonate, clay, silt and sand in the form of turbidites, volcaniclastics and limestone 392 (Plank and Langmuir, 1998). Sediments made of these various constituents respond differently to 393 increasing pressures and temperatures during diagenesis, leading to dissolution, precipitation, and 394 solid-phase reordering reactions (Prothero and Schwab, 2004) that will certainly impact organic-395 mineral associations. How specific mineral-organic associations change across environmental 396 gradients is not well known, but potentially crucial to understanding the fate of OC in marine 397 sediments. The common clay mineral smectite provides an illustrative example since it has been 398 shown to preferentially sequester organics, relative to chlorite-rich clays (Ransom et al., 1998a). 399 With as little as 1.3 MPa pressure (Hüpers and Kopf, 2012), the interlayer in smectite begins to 400 dewater and collapse, a process that also depends on temperature, the identities of interlayer cations 401 and the concentrations of cations present in solution (Ransom and Helgeson, 1995). If organics are 402 sequestered in this interlayer, they could be expelled during this transition and transported into 403 other parts of the sediment. Likewise, the reductive dissolution of metal oxides may liberate 404 chemically or physically adsorbed organics (Coppola et al., 2007). Other recent work suggests that 405 the salinity gradient experienced by smectite during transport from the terrestrial to marine 406 environments induces cation exchange reactions and the removal of associated pedogenic organic 407 carbon, followed by repopulation of the mineral surface with marine OC (Blattmann et al., 2019). 408 On the other end of the size spectrum, sandy sediments - half of continental shelf seafloor - allow 409 for water column POC to be pumped biologically or tidally into the subsurface. This reactive DOC stimulates organisms (Huettel et al., 2014) to consume it and more of the particulate organic 410 411 fraction, contributing to low POC in sandy sediments (Boudreau et al., 2001). 412

413 **2.4 Hydrolysis and Fermentation**

414 When free oxygen has been exhausted in sediments, OC is thought to be transformed 415 through a series of steps that include the extracellular enzymatic hydrolysis of large, complex 416 organic compounds into smaller ones that are then fermented into volatile fatty acids, H₂ and other 417 simple chemical species, some of which are then oxidized by microorganisms using electron 418 acceptors such as nitrate, metal-oxides and sulfate, or reduced to methane (Schulz, 2006). Thought 419 to be the rate limiting step in organic matter degradation, extracellular enzymes produced by 420 microorganisms are known to break large and/or complex organic compounds to supply 421 microorganisms with energy and nutrients such as nitrogen and phosphorous (Arnosti, 2011). 422 Extracellular enzymes are diverse and present in low concentrations in marine sediments (Steen et 423 al., 2019). Although temperature and pH are dominant controls on rates of enzyme-catalyzed 424 reactions, patterns of activity across ecosystem types do not necessarily correlate with such physio-425 chemical variables (Mahmoudi et al., 2020). It seems that the functional diversity of microbial 426 communities, nutrient availability and organic matter reactivity are likely to better explain patterns 427 of extracellular enzyme activities, with a notably strong correlation with particle composition, size 428 and abundance (Arnosti et al., 2014).

429 It is difficult to locate fermenting microorganisms in sediment columns since they do not 430 leave a distinct chemical trace of their activity (Nealson, 1997). A near-limitless number of organic 431 compounds can serve as reactants and products, and a considerable number of inorganic species 432 can be produced and consumed by both fermentative and non-fermentative processes (LaRowe 433 and Amend, 2019). In addition, many organisms can switch between fermentation and other 434 catabolic strategies, and macrofauna can also leave signatures reminiscent of fermentative 435 pathways as they partially digest organic matter during gut passage (McInerney et al., 2008; 436 Jochum et al., 2017). Consequently, the specific forms of fermentation and diversity of 437 fermentative mechanisms in marine sediments remain largely unknown.

438 Recent analyses of biomolecular data (see Section 2.6) from sediments confirm that 439 fermentation generally appears to be a widespread survival strategy for many cosmopolitan groups 440 of microorganisms in anoxic sediments. For example, Bathyarchaeota, Hadesarchaea, and the 441 Atribacteria that are common in marine sediments all utilize fermentative strategies - using 442 peptides, aldehydes, sugars and lignin as substrates (Lloyd et al., 2013; Baker et al., 2016; Nobu 443 et al., 2016; Orsi, 2018; Yu et al., 2018a). As has been often presumed, fermenting bacteria have 444 now been found throughout the anoxic sediment column across multiple geochemical zones (Orsi 445 et al., 2017; Beulig et al., 2018). Many bacteria with the capability to ferment (in particular alpha-446 and gamma-proteobacteria) have also been found in oxic marine sediment such as deep-sea red 447 clay, though they respire oxygen in these settings (Vuillemin et al., 2019). Eukaryotic 448 microorganisms are also involved in fermentation processes in sediments. For example, the 449 majority of H₂ produced in anoxic permeable sediments results from fermentation by eukaryotic 450 algae (Bourke et al., 2016). In deeper sediments down to at least 2,000 meters below the seafloor (mbsf), fermenting fungal cells can still persist (Ciobanu et al., 2014), suggesting that they have 451 the ability to contribute to H₂ production since this can be a product of their fermentative pathway 452 453 (Orsi, 2018). Finally, metagenomic and proteomic data taken from terrestrial settings suggest that 454 fermenting organisms constitute a large fraction of the Candidate Phyla Radiation, an uncultured 455 but geographically widespread and genetically diverse group of bacteria (Wrighton et al., 2012; 456 Wrighton et al., 2014; Anantharaman et al., 2016; Danczak et al., 2017).

457

458 **2.5 Electron acceptors**

Most sedimentary POC – and its hydrolysis and fermentation products – are consumed by
 microorganisms using an array of electron acceptors. The identities and concentrations of EAs are
 in turn determined by the composition of the overlying seawater and the types of mineral phases

462 that are deposited along with organic compounds. The principal EAs encountered in marine sediments, O₂, NO₃⁻, Mn(IV), Fe(III), SO₄²⁻ and CO₂, are traditionally thought to be consumed in 463 the order listed based on the idea that this sequence follows the order of decreasing Gibbs energy 464 465 yield of the corresponding organic matter oxidation reactions (Claypool and Kaplan, 1974; Froelich et al., 1979; Stumm and Morgan, 1996), though it should be noted that the range of Gibbs 466 467 energies for reactions involving these EAs can overlap depending on the environmental conditions 468 (LaRowe and Van Cappellen, 2011; LaRowe and Amend, 2014, 2015a). This hierarchy leads to 469 redox zonation in marine sediments - oxic sediments nearest the SWI, followed by so-called sub-470 oxic zones where NO_3^- and Mn(IV) reduction occurs, a ferrigenous layer if Fe(III)-bearing minerals are present, then a sulfidic layer where the bulk of SO_4^{2-} reduction takes place and finally 471 472 a methanogenic zone at the bottom of the sediment column. Not all of these redox zones will 473 necessarily be present in any given sediment column. The thickness of the zones can vary 474 dramatically (Glud, 2008; D'Hondt et al., 2015; Egger et al., 2018) and the order of them can 475 exhibit complex patterns (see Jørgensen et al., 2019). For instance, in many coastal sediments 476 where POC fluxes are high, the oxic zone might be vanishingly thin (Glud, 2008), while the oxic 477 layer in sediments underlying ocean gyres can penetrate tens of meters to the basement since POC 478 fluxes are so low (Røy et al., 2012; D'Hondt et al., 2015). In fact, D'Hondt et al. (2015) estimate 479 that 9-37% of the global sediment-basement interface is oxic (Figure 3a).

480 In an undetermined volume of the ocean crust, deep sediment layers can exhibit higher 481 concentrations of dissolved oxygen than in upper or middle layers due to the penetration and 482 circulation of deep, oxygenated seawater into unsedimented adjacent basaltic outcrops (Orcutt et 483 al., 2013b; Mewes et al., 2016; Kuhn et al., 2017). This subsediment, rapid movement of low-484 temperature seawater allows oxygen to diffuse upward from basement basalt into sediments, thus 485 creating a C-shaped O₂ curve in these sediments. An example of this is shown in Figure 4 for a 486 sediment pond near the mid-Atlantic Ridge, though it should be noted that these kinds of oxygen 487 profiles have also been observed in the North Pacific near a fracture zone (Mewes et al., 2016; 488 Kuhn et al., 2017). The upward transport of such microbial energy sources could be common 489 globally due to the vast number of seamounts that jut above the sediment-water interface (Wheat 490 et al., 2019 and references therein).

491 Other chemical compounds that can provide energy for microorganisms have been found 492 to be transported upward in sediments. In one such case near the Peru Margin, sulfate diffused 493 upward from a brine in the oceanic basement (D'Hondt et al., 2004; Parkes et al., 2005; Engelen 494 et al., 2008). In another, Cretaceous-aged organic-rich horizons support methane production that 495 seem to, in turn, provide energy for microbial communities in and above it, driving unexpected 496 distributions of EAs that differ from classical expectations (Arndt et al., 2006). In many anoxic 497 sedimentary settings, the reduced products of POC oxidation, compounds such as Fe²⁺, H₂S and 498 NH4⁺, diffuse upwards to be oxidized by chemolithotrophic microorganisms for energy. If redox 499 conditions oscillate, this process can cycle, creating the impression that POC is being transformed 500 more rapidly than it is (Thullner et al., 2009), though OC oxidation rates can be accelerated due to 501 these oscillations (Sun et al., 1993; Aller, 1994; Sun et al., 2002a; Caradec et al., 2004).

The deepest extent of the sulfate-reducing zone and thus the beginning of the methanogenic zone (sulfate-methane transition, SMT) varies widely on a global scale, and has been shown to strongly depend on sedimentation rates and associated organic matter burial fluxes (Berner, 1978; Borowski et al., 1999; Egger et al., 2018). In fact, Egger et al. (2018) recently used a compilation of 1,704 observations to correlate sedimentation rates with SMT depth to map the extent of the SMT globally (Figure 3b). It can be seen in this figure that SMT depth is spatially highly variable: in many shallow continental shelf and slope settings, SMT depth is < 1 mbsf, and up to 10 mbsf.
In deeper sediments, particularly water depths >2,000 m, SMT depth is at least 10 mbsf and in
many instances, hundreds of meters deep. The majority of sediments in the abyssal plain (>66%)
exhibit no SMT.

512 The preservation of POC in sediments is sometimes attributed to anoxia, and therefore 513 ascribed to the identities of the electron acceptors present (e.g. Demaison and Moore, 1980; 514 Emerson, 1985; Calvert and Pedersen, 1992a; Lee, 1992b; Aller, 1994; Canfield, 1994; Wignall, 515 1994), and to some extent the types of organic compounds deposited (Harvey et al., 1995; Harvey 516 and Macko, 1997; Sun et al., 1997; Bianchi et al., 2000; Grossi et al., 2001; Sun et al., 2002a). 517 Though there are many datasets and reports arguing for and against the idea that the presence of 518 free oxygen plays a decisive role in POC reactivity (see Hulthe et al., 1998), there seems to be a 519 growing consensus that the presence O₂ typically enhances organic carbon reactivity in many 520 marine sediments (Hartnett et al., 1998; Hedges et al., 1999; Keil and Cowie, 1999; Keil et al., 521 2004; Moodley et al., 2005; Cowie et al., 2009; Middelburg and Levin, 2009; Aller, 2014; 522 Eglington and Repeta, 2014; Keil et al., 2016). Notable exceptions to these observations include 523 studies that show that rates of POC degradation in anoxic sediments have been observed to be 524 nearly equal to (e.g. Henrichs and Reeburgh, 1987; Lee, 1992a; Kristensen and Holmer, 2001b) or 525 far exceeding those in oxic settings (Røy et al., 2012; D'Hondt et al., 2015). On a molecular level, 526 fatty acids can be degraded at similar rates independently of their degree of saturation in the 527 presence of oxygen, whereas unsaturated acids are preferentially degraded under anoxic conditions 528 (Harvey and Macko, 1997; Sun et al., 1997; Grossi et al., 2001). In sediments dominated by sulfate 529 reduction, measured cell specific sulfate-reduction rates vary by 10 orders of magnitude (Jørgensen et al., 2019). Furthermore, recent incubation experiments have built on other studies (e.g. Lee 530 531 1992) showing that the identity of the electron acceptors used during POC transformation 532 processes has little to no influence on the overall rate of organic carbon degradation (Beulig et al., 533 2018). Taken together, these studies suggest that the identity of the oxidant/reductant does not 534 necessarily determine the reactivity of organic carbon, but that O₂ has a special role among EAs 535 in evaluating the reactivity of POC.

536

537 **2.6 Microorganisms**

Although many factors influence the reactivity of organic carbon in sediments, one of the most unifying is microorganisms. The extent to which organic carbon reactivity is influenced by the taxonomic and functional distribution of microorganisms is only beginning to be explored, after an era in which little attention was paid to microbial life in sediments (see Section 1.3). In the last decade or so, there has been a revolution in understanding the number, identity, functional capabilities and extent of life in marine sediments, and their role in OC degradation.

545 **2.6.1 Abundance**

546 Microbial cell numbers correlate with mean sedimentation rate and distance from 547 continental landmasses (Kallmeyer et al., 2012), as well as with the amount of organic matter in 548 sediments (Jørgensen and Marshall, 2016). For surface sediments, those on continental margins contain $10^8 - 10^{10}$ cells cm⁻³, whereas those underlying oligotrophic gyres contain $10^5 - 10^7$ cells 549 cm⁻³ (Kallmeyer et al., 2012) (Figure 5). In most sediments, cell concentrations decrease with 550 551 increasing depth below the seafloor, according to a power law. In OC-poor, oxic sediments, cell concentrations drop below $\sim 10^2$ cells cm⁻³ between 10 and 15 mbsf (Kallmeyer et al., 2012; 552 Vuillemin et al., 2019). Cell abundance in OC-rich, anoxic sediments on continental shelves 553

remain relatively high at great depths in the sediment, and only fall to $\sim 10^2$ cells cm⁻³ between 1,000 – 2,500 mbsf (Kallmeyer et al., 2012).

556557 **2.6.2 Identity**

558 Rapid progress in high-throughput DNA sequencing and analytical tools over the last 559 decade has completely revised our understanding of the tree of life (Hug et al., 2016). Many of the 560 phylogenetic groups found in sediments are candidate phyla that are not yet cultivated, so their 561 role in organic matter transformation is not entirely clear (Lloyd et al., 2018). Though common 562 types of fungi, bacteria, and archaea have been found in sediments (Richards et al., 2012; Orsi et al., 2013), many new groups of microorganisms reside in the subsurface that appear to be endemic 563 564 to this environment. These endemic groups are repeatedly found to be relatively abundant in 565 subsurface settings (Anantharaman et al., 2016; Orsi, 2018), with widely differing diversity in oxic 566 and anoxic sediments (Orsi, 2018). In addition, sediments of varying redox state reveal evidence 567 of viruses (Engelhardt et al., 2015; Tully and Heidelberg, 2016; Bäckström et al., 2019; Cai et al., 2019), which may contribute to organic processing via lysis of microbial biomass (Danovaro et 568 569 al., 2008; Orsi, 2018), with archaea potentially being disproportionally lysed in surface sediments 570 under deep waters (Danovaro et al., 2016).

571

572 **2.6.3 Functional capabilities**

573 Before the molecular biology revolution, the functional capabilities of microbial 574 communities in sediments were inferred from pore water profiles, with little to no direct biological 575 information. For example, decreases in sulfate and increases in sulfide suggested microbial sulfate 576 reduction and amendment experiments revealed that the addition of fermentation end products (H₂, 577 acetate) stimulated rates of sulfate reduction (Goldhaber et al., 1977; Iverson and Jørgensen, 1985). 578 The ability to sequence and identify genes with known functions, and determine their expression 579 levels, allowed for the discovery of new types of microorganisms as well as new functional 580 capabilities (Biddle et al., 2006; Biddle et al., 2008), which could then be correlated to geochemical 581 profiles.

582 Knowledge of microbial functionality from environmental 'omics data (the term 'omics is 583 commonly used to refer to the analysis of DNA, RNA, metabolite and protein sequences extracted 584 from samples) can be inferred from a close similarity of protein encoding genes from environmental samples to those found in genomes of cultured microbes that have been assigned a 585 function based on biochemical experiments (de Bruijn, 2010). Marine sediment communities are 586 587 dominated by microorganisms that are not closely related to any current pure culture (Parkes et al., 2005; Biddle et al., 2006; Inagaki et al., 2006; Lloyd et al., 2018). Therefore, it is speculative 588 589 to assume that distant genetic similarities from the *in situ* microorganisms to cultured strains imply 590 that those genes are enabling microorganisms to perform the same function. A large number of 591 genes encoding hypothetical proteins in marine metagenomes, which are digital libraries of all the 592 DNA present from all the microorganisms in a natural sample, remain difficult to annotate with 593 information about their identities or functions. For instance, in the large TARA Oceans 594 metagenomic data set, only about 16% percent of DNA sequences that encoded a hypothetical 595 protein had a statistically significant similarity to proteins with an experimentally determined 596 function. Furthermore, 44% of the hypothetical proteins had no significant similarity to gene 597 families that share general biochemical functions (Sunagawa et al., 2015a). This problem is 598 difficult to solve using traditional bioinformatic approaches, which rely on comparing 599 environmental sequences to genes whose function has been identified.

600 A further complicating factor is that databases of known genes are dominated by well-601 characterized microorganisms, particularly from a single phylogenetic group (Proteobacteria), 602 whereas marine environments are dominated by uncultured organisms from diverse lineages 603 (Lloyd et al., 2018). Physiologies from some of these uncultured microbes have been inferred from 604 whole genome reconstructions in marine sediments (Lloyd et al., 2013), metatranscriptomics 605 showing which genes were being transcribed at the time of sampling (Orsi et al., 2013), 606 compositions of natural isotopes of biomass (Biddle et al., 2006; Shah et al., 2008; Meador et al., 607 2015), stable isotope probing (Morono et al., 2011; Trembath-Reichert et al., 2017), direct 608 measurement of metabolites (Bird et al., 2019) and tracking increases in a microbial group's cell 609 abundance during laboratory enrichment of natural marine sediments (Kevorkian et al., 2018; Yu 610 et al., 2018b). The limitations of these approaches are that DNA and RNA sequences from 611 environmental samples can only be given functional annotations based on their similarity to known 612 cultures. Therefore, truly novel functions cannot be determined from sequencing methods alone. 613 Heterologous expression and characterization can be used to identify novel functions (Cottrell et 614 al., 2005; Michalska et al., 2015; Wrighton et al., 2016), but such methods have only been applied 615 to enzymes with enough homology to a known protein to develop a hypothesis. Determining the functions of genes encoding truly novel "hypothetical" proteins will be very important for inferring 616 functions of uncultured microorganisms in marine sediments. 617

618 It should be noted that genome representation in databases represents one of the largest 619 issues for making correct assignments of protein encoding genes in environmental genomics 620 datasets. For example, the number of genomes from archaea in databases that derive from 621 subsurface environments has grown substantially in recent years (Lloyd et al., 2013; Spang et al., 622 2015; Anantharaman et al., 2016; Baker et al., 2016; Jungbluth et al., 2017; Dombrowski et al., 623 2018; Tully et al., 2018; Seitz et al., 2019). Before these genomes were available, protein encoding 624 genes expressed by archaea in marine sediments were estimated to be about 1% to 2% of total 625 genes expressed (Orsi et al., 2013). However, a re-analysis of that same data including these new archaeal genomes in the database showed that archaea actually express 25% of the total genes 626 627 (Orsi, 2018), clearly demonstrating a database bias. Thus, we imagine that as more genomes from 628 archaea are sequenced, their representation in environmental 'omics datasets will continue to 629 increase. The archaea are thus likely to be critically important for subsurface carbon cycling, as 630 predicted by earlier studies from the marine subsurface (Biddle et al., 2006; Lloyd et al., 2013).

631

632 **2.6.4 OC Degradation potential based on biomolecular data**

633 The advent of inexpensive nucleic acid sequencing technology combined with the 634 availability of user-friendly bioinformatics processing platforms has made it possible for nonspecialists in bioinformatics to substantiate biogeochemical work with 'omics data. 'Omics 635 approaches that have proven valuable in sediment biogeochemistry include single-cell genomics, 636 metagenomics and the related practice of "binning" metagenomic sequences into "metagenome-637 assembled genomes" (Albertsen et al., 2013), metaproteomics, and metabolomics (for a review 638 639 see Gutleben et al., 2018). These approaches, especially when used in concert, have enabled insight 640 into the mechanisms of organic matter degradation that would be impossible from purely 641 geochemical techniques. For instance, in deep sediments of the Baltic Sea, it appears that 642 Atribacteria have the potential to act as a keystone species, accessing a wide range of organic 643 carbon using a broad spectrum of extracellular enzymes, and then exporting the resulting free 644 amino acids, possibly supporting the rest of the microbial community (Bird et al., 2019).

645 Ideally, the integration of multiple molecular approaches will yield information on the 646 rates and potentials of organic matter oxidation by microbial communities. Progress along this 647 path has been made in studies of the nitrogen cycle, for which the genomic pathways of important 648 transformations (N₂ fixation, nitrification, etc.) are well-characterized and the relevant genes are 649 highly conserved (Pachiadaki et al., 2017). This information has been used to construct a "gene-650 centric" model of the nitrogen cycle in the Arabian Sea oxygen minimum zone, revealing 651 interactions between the cryptic sulfur cycle and the nitrogen cycle which would have been 652 difficult to observe using purely biogeochemical techniques (Reed et al., 2014).

653 The use of biomolecular data to better understand organic carbon cycling is a nascent area 654 of inquiry that presents researchers with many avenues of research to make sense of the 655 tremendous diversity and complexity of organic structures, and therefore enzymes that catalyze organic matter transformations. One aspect of this complexity is illustrated in Figure 6, which 656 657 shows that the phylogenetic diversity of putative secreted organic-degrading enzymes in anoxic 658 marine sediments spans the tree of life (Orsi, 2018). Peptidases provide a more specific example: 659 even though all peptidases catalyze essentially the same reaction, breaking a peptide bond, this can 660 be performed by 268 different structural families of proteins via eight separate catalytic mechanisms (Rawlings et al., 2016). Other categories of enzymes related to organic carbon 661 degradation are similarly diverse, obscuring relationships between enzyme structure and function 662 663 (Michalska et al., 2015). Because many enzymes relevant to OC degradation can catalyze reactions beyond those including their "preferred" substrates, caution must be used when inferring 664 degradation processes from genomic data or enzyme assays (Steen et al., 2015). It is also possible 665 that novel uncultured clades harbor catalytic enzymes not identifiable by current annotation 666 667 methods. Therefore, even if particular genes are present in an ecosystem, biomolecular data sets 668 alone might not be sufficient for making specific predictions about the rate, quantity and type of 669 organic carbon reacted in a given system. Lastly, recent experimental work using bioreactor incubations with marine sediments and different marine heterotrophs has directly demonstrated 670 671 species-to-species differences in the rate, quantity, and type of organic matter oxidized, illustrating that the reactivity of organic carbon is also a function of the microorganisms that are present and 672 673 active in a given environment (Mahmoudi et al., 2019).

674

675 **2.6.5 Energetic and power constraints**

676 Absent photosynthesis, all microbial energy is derived from catalyzing redox reactions, nearly all of which are ultimately driven by the degradation of organic carbon. The amount of 677 678 Gibbs energy available in a number of sedimentary settings has been determined (e.g. Schrum et 679 al., 2009; Wang et al., 2010; LaRowe and Amend, 2014; Teske et al., 2014), as have the metabolic 680 rates in a number of subsurface habitats (Orcutt et al., 2013a). However, the rate at which this 681 energy is used, microbial power utilization, is less well constrained, despite being critical for 682 understanding activity levels and growth state (Hoehler and Jørgensen, 2013; LaRowe and Amend, 2015a). In situ measurements of microbial power utilization are not yet feasible, but calorimetric 683 684 measurements carried out in the laboratory have begun to assess low rates of heat production from small numbers of cells taken from the subsurface (Robador et al., 2016). However, a growing 685 number of studies have computed power usage by combining geochemical data with modeling 686 687 tools (e.g. LaRowe and Amend, 2015a, b; Bradley et al., 2018b). In these studies, data gathered by 688 scientific drilling is used to inform POC degradation models (see Section 4) while the energetics 689 of organic carbon degradation can be constrained by either using the total molecular composition of dead microbial cells (Bradley et al., 2018b) or by relating the nominal oxidation state of organic
 carbon to its energy content (LaRowe and Van Cappellen, 2011).

692 The results of these power studies have shown that in low-energy sediments, such as those 693 underlying the South Pacific Gyre (SPG), microbial activity is limited, more or less, to maintaining 694 cellular integrity through biomolecular repair and replacement (Bradley et al., 2018a), a state akin 695 to dormancy (see Section 4.3). Thus, maintenance activities (the sum of activities that do not 696 produce growth) constitute a much greater fraction of total power utilization by microbial 697 communities in habitats where growth is minimal. Indeed, bioenergetic modelling of the SPG 698 sediments suggested that <0.1% of the power from organic carbon oxidation can be attributed to 699 growth, with maintenance accounting for the rest (Bradley et al., 2019).

700 Power calculations have also been used to constrain the cell-specific power requirement of 701 microorganisms in sediments. Assuming that all counted cells are involved in organic matter 702 degradation, minimal cell-specific power requirement of microbial cells in SPG sediments have been estimated to be around 1.9×10⁻¹⁹ W cell⁻¹ (LaRowe and Amend, 2015b). Assuming this value 703 704 for the non-growing cells in SPG sediments, the degradation of cells that died in the sediments, 705 necromass, was estimated to provide 2 to 13% of the power used by microbial communities in 706 shallow and relatively young sediments (<10,000 years) from SPG (Bradley et al., 2018b). In a 707 similar study, mean cell-specific metabolic rates of functional groups involved in nitrogen cycling 708 in oligotrophic North Pond sediments (on the western flank of the Mid-Atlantic Ridge) showed 709 that an increased power supply in transition zones between oxic and anoxic regimes may be 710 responsible for the revival of organisms from a maintenance state, and even for growth (Zhao et 711 al., 2019).

712 The lack of quantitative data accurately describing the distribution of various functional 713 groups of microorganisms in marine sediments has impeded the calculation of cell-specific power 714 requirements for particular catabolic groups. This has led to sometimes-untested assumptions 715 about the fraction of a community catalyzing a particular reaction (e.g. that ~10% of the total cells 716 in organic-rich sediments are sulfate reducers (Hoehler and Jørgensen, 2013)). Quantification 717 methods like marker-gene-based qPCR and FISH are useful means to measure the abundances of 718 various functional groups in marine sediments (Schippers and Neretin, 2006; Lever, 2013; 719 Buongiorno et al., 2017), though their application is limited by primer and probe biases.

One of the goals motivating calculation of cell-specific power requirements for different types of functional groups (e.g. Zhao et al., 2019) under a wide range of natural conditions is to determine the limits that the environment imposes on basal power requirements (Hoehler and Jørgensen, 2013). Knowing what the ultimate limits are would greatly facilitate the prediction of the standing stock of biomass of different functional groups in marine sediments on a global scale using geochemical and physical data (e.g. Bowles et al., 2014; Egger et al., 2018) and without the requirement for the collection, analysis and assembly of biological samples on such a vast scale.

727

728 **2.6.6 Microbial turnover rates**

Most early studies focusing on microorganisms in marine sediments were largely conducted in the upper tens of centimeters of sediment, which are both younger and often more carbon-rich than deeper sediments (see Figure 2). While some studies measured microbial activities in subsurface sediments as deep as 1.5 mbsf at the Peru Margin (Parkes et al., 1990), these studies generally depended on laboratory incubations to measure metabolic rates, which tend to overestimate the *in situ* rate. 735 In recent years, new approaches that are incubation-independent and sensitive to low 736 activities have revealed that a large, active, yet slow-growing microbial community inhabits the 737 deep subsurface. These microbes subsist on low power levels and have been ascribed some of the 738 slowest turnover times ever documented. Amino acid racemization modelling has been used to 739 estimate that some marine sediment microorganisms require hundreds to thousands of years to 740 grow or replace all of their biomass (Lomstein et al., 2012), though these estimates have been 741 adjusted to span years to hundreds of years (Braun et al., 2017). It should be noted that this work 742 was carried out in relatively young, organic-rich sediments, (especially when compared to ancient 743 sediments, e.g. from SPG). A different and perhaps more sensitive method, deuterium 744 incorporation into lipids, has been used to estimate microbial community turnover times from tens 745 to hundreds of years in relatively shallow (< 1 mbsf) subsurface sediments from the Baltic Sea 746 (Wegener et al., 2012), as well as in very deep (up to 2,000 mbsf) sediments from offshore Japan 747 (Trembath-Reichert et al., 2017). In all cases, these turnover times are several orders of magnitude 748 slower than the doubling times of laboratory cultures, which are typically on the order of hours to 749 days.

750 With such slow replication rates, it is unlikely that subsurface sedimentary microorganisms have experienced a sufficient number of generations for specific adaptations to have evolved and 751 spread through the community (Starnawski et al., 2017), calling into question whether these 752 753 organism scan be considered endemic (see Section 2.6.2). Under this scenario, it is more likely 754 that a community that is able to tolerate this increasing energy limitation is selected for as it is 755 buried deeper in the sediment column (Marshall et al., 2019). This idea is substantiated by evidence 756 that the microbial taxa that dominate deep subsurface sediments are common across a wide range 757 of locations (Petro et al., 2017) and are often found in surficial sediments (Walsh et al., 2015; 758 Starnawski et al., 2017), while the converse is not the case – surficial sediments contain microbial 759 taxa that are not found deeper in the sediment column. In fact, it has recently been shown that at 760 sites as diverse as the Indian Ocean and the Bering Sea, the operational taxonomic units that are 761 most abundant at depth are a subset of the local seafloor community (Kirkpatrick et al., 2019). 762 That is, net replication was not required to produce the microbial population observed in deep 763 sediment, which could instead have been produced by differential mortality rates. These lines of 764 evidence suggest that microbial community members with a slow-metabolizing survival strategy 765 are selected for in the energy-limited deep subsurface environment, though it is unclear that these 766 traits are passed on as in Darwinian selection. As noted above, the importance of energetic efficiency as a selective advantage for long-term microbial survival has been supported by 767 768 modelling (Bradley et al., 2019) and experimental (Vuillemin et al., 2019) work. Metabolomic and 769 transcriptional evidence has identified some specific mechanisms that likely underlie the slow-770 metabolizing survival strategy – for example, the use of chemical protectants to stabilize nucleic acids and proteins, and metabolic interdependencies among members of the microbial community 771 772 (Bird et al., 2019). However, many of these mechanisms - and the impact of these slow 773 metabolizers on the long-term carbon cycle - remain underexplored.

774

775 2.7 Sulfurization

Sulfurization is one of the mechanisms thought to be responsible for the preservation of organic carbon in marine sediments (for reviews see Sinninghe Damsté and De Leeuw, 1990; Werne et al., 2004; Amrani, 2014). Sulfurization is the process whereby sulfur atoms ultimately replace carbon atoms in organic compounds, producing organic sulfur compounds (OSC). OSCs are thought to be less accessible to microbial degradation since S replaces reactive functional 781 groups that tend to be the easiest parts for microorganisms and/or their exoenzymes to access 782 (Kohnen and Sinninghe Damsté, 1989). Sulfur has been found in a wide variety of organic 783 compound types in sediments (Werne et al., 2004), occupying the full range of sulfur oxidation 784 states (-2 to +6) (Amrani, 2014). The preservation potential of OSCs is based on a) observations 785 that the S:C in POC increases with depth in sedimentary systems (Francois, 1987; Eglinton et al., 786 1994; Hetzel et al., 2009; Amrani, 2014), b) the isotopic composition of organically bound S 787 (Francois, 1987; Mossmann et al., 1991), c) intense sulfurization of POC-rich Cretaceous and Jurassic deposits (van Kaam-Peters et al., 1997; Sinninghe Damsté et al., 1998; Van Kaam-Peters 788 789 et al., 1998; Kolonic et al., 2002; Kolonic et al., 2005; Böttcher et al., 2006; van Dongen et al., 790 2006; Hetzel et al., 2009; Raven et al., 2018; Raven et al., 2019) d) observations that many fossil 791 fuels and their precursors have high levels of OSCs (Sinninghe Damsté et al., 1989a; Sinninghe 792 Damsté et al., 1989b) and e) laboratory studies (Krein and Aizenshtat, 1994; Schouten et al., 1994).

793 Sulfurization is thought to proceed in anoxic environments in which sufficiently high 794 concentrations of POC and H₂S coexist in the absence of metals such as reactive Fe (Gransch and 795 Posthuma, 1974; Werne et al., 2004; Amrani, 2014), although there is some evidence that 796 sulfurization can happen in the presence of reactive iron species since it can lead to the formation 797 of polysulfides which in turn seem to enhance the incorporation of S atoms into organic structures 798 (Kohnen and Sinninghe Damsté, 1989; Werne et al., 2004; Heitmann and Blodau, 2006). 799 Somewhat paradoxically, sulfurization is thought to be an abiotic process, yet the requisite sulfide 800 is attributed to microbial sulfate reduction, leading to the observation that high organic content is 801 sometimes required for its preservation (Quijada et al., 2016).

802 Sulfurization takes place in a variety of environments on a range of timescales. Although 803 it was traditionally thought to be a relatively slow process taking place in sediments and petroleum 804 reservoirs (Sinninghe Damsté and De Leeuw, 1990; Werne et al., 2004), more recent research has 805 shown that it can take place in hydrothermal systems (Gomez-Saez et al., 2016), in sinking marine particles on a timescale of hours (Raven et al., 2016; Raven et al., 2019), and surface sediments 806 807 subjected to oscillating redox conditions (Jessen et al., 2017). It has been shown that such rapid 808 sulfurization of organic carbon likely exerted an important feedback on ocean redox geochemistry 809 and climate during the end of a major Cretaceous extinction event (OAE2), ultimately terminating 810 the extreme environmental conditions that caused it (Raven et al., 2019, Huelse et al., 2019). 811

812 2.8 Terrestrial studies

813 Investigations on the cycling of organic carbon in terrestrial settings offer valuable insights 814 to analogous processes within the marine subsurface. Moreover, terrestrial settings are easier to 815 access than deep marine environments, and thus they are more amenable to long term observations 816 and manipulation for field and laboratory-based experiments. Through such studies, the stability 817 of organic carbon has been found to be largely dependent on the complex interplay of the 818 physicochemical (e.g. OC-mineral interaction and aggregation, temperature, moisture, salinity, etc.) and biological (e.g. microbial community composition, nutrient availability, extracellular 819 820 enzyme production, etc.) properties of the environment rather than on the molecular properties of 821 the OC itself (e.g. elemental composition, presence of functional groups, molecular conformation, 822 etc.). Given that a substantial fraction of POC buried in marine sediments is of terrestrial origin 823 (Burdige, 2007a), understanding the factors governing OC stability in terrestrial ecosystems 824 provides valuable insight into the marine carbon cycle.

825 Within soils, certain organic compounds such as lignin have classically been viewed as 826 resistant to biodegradation due to their large and complex molecular structures. However, lignin 827 and lignocellulose compounds are no more likely to persist in soils than other organics given the proper conditions (Marschner et al., 2008b; Klotzbücher et al., 2011; Schmidt et al., 2011). This
is largely due to the activity of fungi, who play a critical role in the degradation of poorly reactive
organic substrates on land (Treseder and Lennon, 2015) and thus may also play a similar role in
marine sediments containing soil-derived organics. In fact, new experiments show that fungi living
in coastal marine sediments play a role in degrading lignocellulose at the land-sea interface
(Ortega-Arbulú et al., 2019).

834 The ability of microorganisms to degrade soil OC may, at times, also be restricted by 835 nutrient or energy limitations. This is especially pronounced in deep soil layers where turnover 836 times are on the order of thousands to tens of thousands of years (Schmidt et al., 2011). Recent 837 studies suggest that this long turnover time cannot be solely attributed to the chemical structure of 838 OC as there is not always a significant change in composition with depth (e.g. Fontaine et al., 839 2007). However, amendments with fresh OC to subsoil via root exudates or decomposition (often 840 called priming) has been shown to stimulate increased degradation of the old OC, indicating that 841 the fresh OC may provide a needed energy source to promote the activity of existing microbial 842 degraders (Fontaine et al., 2007; Marschner et al., 2008b; Kuzyakov, 2010). This mechanism has 843 also been shown to occur to marine OC as well (Steen et al., 2016). The mechanism behind priming 844 effects, however, is unclear. Common root exudates such as oxalic acid may enhance organic 845 carbon degradation by removing organic carbon from mineral surfaces, thereby increasing its 846 reactivity (Keiluweit et al., 2015). In the case of permafrost, which is also primarily composed of 847 old organic carbon, temperature has been found to be a dominant factor governing its degradation. 848 While frozen, decreased water availability and microbial and enzymatic activity lead to an increase 849 in OC stability. With rising temperatures, these limitations are largely erased and OC degradation 850 rates increase (Schuur et al., 2009; Schmidt et al., 2011) and references therein).

851 Molecular biological tools have revealed that anoxic freshwater sediments contain similar 852 microbial groups that exist in their marine counterparts (Vuillemin et al., 2018). The presence of 853 the common phylum, Bathyarchaeota, in deep terrestrial sediments that contain high amounts of 854 unreactive plant derived organic matter (e.g. lignocellulose) is thought to be due to their ability to use lignin as an energy source (Yu et al., 2018b). Bathyarchaeota might also be involved in the 855 856 turnover of terrestrially derived organic matter, such as lignin, in marine sediments in coastal 857 settings where land-derived OC is commonly deposited. Other bacterial "dark matter" groups that 858 are present in marine sediments such as the candidatus groups Latescibacteria, Omnitrophica, and 859 Parcubacteria have been shown to have similar rates of organic carbon turnover in freshwater sediments relative to more well-studied groups such as the Proteobacteria (Coskun et al., 2018). 860 861 Given their transcriptional activities in marine sediments (Orsi, 2018), these groups could also be 862 important for benthic carbon cycling in the ocean.

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870

864 **3. Old, hot and deep organic carbon**

Most studies concerning the rates of POC degradation in marine sediments focus on the shallowest, most recently deposited material. However, it is becoming clear that deeply-buried organic carbon is actively being consumed and that fluid circulation in the ocean basement can impact OC reactivity through the modification of organic compounds as well as the composition of sediment pore water.

871 **3.1 Ancient sediments**

Throughout Earth's history there have been numerous periods of time when the deposition flux of organic carbon into sediments has been much higher than the present day (Pedersen and 874 Calvert, 1990). A fraction of this ancient OC persists in modern marine sediments within layers of 875 elevated OC concentrations (Condie et al., 2001; Jenkyns, 2010; Ohkouchi et al., 2015). Although 876 the details vary depending on the time period, there is evidence that large-scale tectonic processes 877 and volcanism are ultimately responsible for the relatively high concentrations of OC deposited in 878 the past (Berry and Wilde, 1978; Condie et al., 2001; Jenkyns, 2010; Trabucho-Alexandre et al., 879 2012; Ohkouchi et al., 2015). One such example is a section of Cretaceous-aged sediments located 880 in an area of the equatorial Atlantic known as the Demerara Rise, where drill cores from ODP Leg 881 207 have revealed shales that typically contain between 2 and 15 wt% POC in layers ranging from 56 to 94 meters thick (ShipboardScientificParty, 2004). Biogeochemical reaction transport 882 883 modelling has revealed that deep Demerara organic-rich strata likely host organisms that convert 884 the POC in these shales into CH₄ which is subsequently consumed by anaerobic methane oxidizing microbial consortia using SO₄²⁻ as the oxidant (Arndt et al., 2006). Although data on microbial 885 886 abundance are not available, it is likely that biomass concentration is higher in POC-rich sediment 887 layers as well as the adjacent sulfate-reducing zone than in the hundreds of meters of sediment 888 separating these layers from the SWI, in which the POC content is far lower. Supporting this 889 notion, cell abundances in core sections 1,500 to 2,500 mbsf near the Shimokita Peninsula, Japan, 890 are orders of magnitude higher in organic-rich lignite layers than surrounding, organic-poor 891 sediments (Inagaki et al., 2015). Although much of this deeply-buried organic matter looks to have 892 escaped microbial degradation, it seems as if it is actually hosting a very slowly metabolizing 893 community. Observations and diagenetic modeling results show that the organic matter in ancient, 894 deeply buried organic carbon-rich strata still provides a suitable substrate for ongoing microbial 895 respiration (Krumholz et al., 1997; Coolen et al., 2002; Krumholz et al., 2002; Moodley et al., 896 2005; Arndt et al., 2006; Arndt et al., 2009). Microbial biomass and cell activity have also been 897 shown to peak at redox transition zones, including at oxic-anoxic transition zones in North Pond 898 sediments (Zhao et al., 2019), and in deep (~90 m below the SWI) Peru Margin sediments 899 associated with ODP Leg 201 (site 1229) where there is a convergence of methane and sulfate, the 900 latter of which is diffusing upwards from a brine (Jørgensen et al., 2003).

901 The rate at which old, deeply buried organic carbon is metabolized by microorganisms in 902 marine sediments becomes extremely slow with depth (Middelburg, 1989), but as these organics 903 are exposed to sufficient pressure and temperature, they can be converted abiotically into 904 petroleum through a process known as catagenesis. Although it is thought that the principle zone 905 of oil formation in organic-bearing sediments occurs from ~ 50 - 160° C, with pressure playing a 906 lesser role, hydrocarbons and methane can be generated abiotically from complex organic matter 907 at lower temperatures (Tissot and Welte, 1984; Hunt, 1996). As noted by LaRowe et al. (2017a), 908 nearly 35% of the volume of marine sediments are above 60°C, and even if sediments do not 909 contain enough OC to be commercial sources of petroleum (TOC > 0.5% by weight), the remaining 910 organic carbon in them can still be converted to microbially accessible hydrocarbons through 911 abiotic processes. Some fraction of small-molecular weight hydrocarbons and CH₄ produced 912 abiotically in sedimentary basins could be expelled and migrate to regions that are more conducive 913 for life. Both petroleum and natural gas can migrate hundreds of kilometers from source rocks 914 (Selley, 1998), and there are certainly thermophilic anaerobes capable of oxidizing common 915 products of catagenesis such as alkanes and benzyl-compounds (Teske et al., 2014). In addition, water, CO₂ and H₂S can also be produced in large amounts during the catagenesis of Type II 916 917 kerogens (Tissot and Welte, 1984), fueling microbial activities with carbon and electron acceptors 918 and donors. Furthermore, it has been estimated that more CH₄ has been produced by methanogens 919 degrading petroleum reservoirs than there is primary CH₄ produced from catagenesis (Milkov,

2011). In total, $1.97 - 3.94 \times 10^{13}$ kg of free CH₄ generated from the microbial degradation of petroleum is estimated to exist in sedimentary basins (Milkov, 2011). Taken together, catagenesis could be fueling microbial communities deep in marine sediments of unknown size.

923

924 **3.2 Hydrothermal sediments**

925 Sediments that are deposited in regions near spreading centers or volcanic hot spots are 926 subjected to advecting hot fluids that transform organic molecules. The effects are particularly 927 pronounced near continental settings, such as the Guaymas Basin in the Gulf of California 928 (Simoneit and Lonsdale, 1982) and at Middle Valley on the Juan de Fuca Ridge, off the coast of 929 Washington State (Cruse and Seewald, 2006). The transformations that occur have been compared 930 to the formation of petroleum on geologic time scales (Simoneit and Lonsdale, 1982), but with 931 important differences due to the presence of water (Seewald, 2001). In laboratory experiments, 932 sediments heated in the presence of water typically release a large pulse of organic matter into the 933 fluids, followed by a slow decline in overall organic concentrations, presumably due to the 934 formation of degradation products such as CO_2 and CH_4 (Seewald et al., 1990; Lin et al., 2017). 935 Organic acids, acetate in particular, are some of the most abundant degradation products (Eglinton 936 et al., 1987; Fisher, 1987; Lundegard and Kharaka, 1994; Kawamura et al., 1996; Shebl and 937 Surdam, 1996; Seewald, 2001), although reactive amino acids, polysaccharides, and small peptides 938 have also been found to be released from Guaymas Basin sediments by heating (Martens, 1990; 939 Lin et al., 2017). These ancient petroleum-derived substrates are further catalyzed by subsurface 940 microbes (Pearson et al., 2005). Acetate and other low molecular weight organic compounds 941 including formate, lactate, methanol and ethanol have been identified in Guaymas Basin sediments 942 and were found to have a microbial rather than thermal source (Zhuang et al., 2019). Hydrothermal 943 alteration of DOC begins at temperatures as low as 68 °C (Hawkes et al., 2016), suggesting that 944 the hydrothermal influence may be widespread (LaRowe et al., 2017a).

945 946

947 **3.3 Ocean basement fluids**

948 Scientific drilling into the seafloor has revealed that the chemical constituents of fluids 949 circulating in the ocean crust basement, such as oxygen, diffuse into overlying sediments (Orcutt 950 et al., 2013b; Wheat et al., 2013) (see Figure 4). The implications of this for POC degradation in 951 sediments is only just being explored. The oceanic basement is generally considered a net sink for 952 marine OC (Lang et al., 2006; Shah Walter et al., 2018) with both microbiological and abiotic 953 removal mechanisms. In addition to the sediment column, deep-ocean DOC, POC and sedimentary 954 particles enter the crust with oceanic bottom water through exposed outcrops. Most of this fluid 955 flux occurs away from active ridge axes in older, cooler crust and is equivalent to about one fifth 956 of the global riverine flux into the ocean (Johnson and Pruis, 2003).

957 Recent studies have described the concentration and isotopic composition of DIC and DOC 958 in ocean basement fluids from naturally outflowing fluids from the Dorado Outcrop (McManus et 959 al., 2019) and fluids recovered from IODP CORK observatories on the flank of the Juan de Fuca 960 Ridge (Lin et al., 2019) and North Pond (Shah Walter et al., 2018) - all relatively cool settings 961 away from mid-ocean ridge spreading centers. Although fluid chemistry is variable on the flanks 962 of ridges, ranging from warm, anoxic fluids recovered from near the Juan de Fuca Ridge (Lang et 963 al., 2006; Lin et al., 2019) to oxygenated fluids that resemble bottom seawater at North Pond 964 (Meyer et al., 2016), fluid temperatures are low enough to allow for microbial activity to influence 965 the OC reservoir (McCarthy et al., 2011; Shah Walter et al., 2018; McManus et al., 2019).

966 Heterotrophic activity has been identified in these environments (e.g. Furnes et al., 2001; Lin et 967 al., 2015; Robador et al., 2015; Russell et al., 2016) and DOC concentrations in circulating 968 basement fluids are lower than in overlying bottom water. Compared to 35-45 µM in deep 969 seawater, cool hydrothermal DOC concentrations can be <15 µM (Lang et al., 2006; Lin et al., 970 2012; Shah Walter et al., 2018; Lin et al., 2019). This DOC removal has been attributed to selective oxidation on the basis of a concurrent loss of oxygen, the ¹⁴C content of the remaining organic 971 matter and characterization by NMR and FT-ICR-MS (LaRowe et al., 2017b; Shah Walter et al., 972 2018; Lin et al., 2019). The DOC removed in the crustal subsurface has a ¹⁴C age of up to 4,300 973 years at North Pond, indicating a long residence time in the open ocean before oxidation by 974 975 basement microorganisms, underscoring the importance of treating OC degradation as an ecosystem property. Chemoautotrophic DOC, isotopically and molecularly distinct from deep-976 977 ocean DOC, has also been identified in cool hydrothermal fluids, although their concentrations are 978 lower than DOC in bottom water, implying a slow production rate (McCarthy et al., 2011; Shah 979 Walter et al., 2018).

980 Fluids that pass through high temperature black smoker hydrothermal systems have DOC 981 concentrations that are approximately one-third that of deep seawater (Lang et al., 2006). Abiotic 982 removal pathways are dominant in regions of active hydrothermal venting where fluids are 983 intensely heated and can reach temperatures of 400°C. Thermal decomposition of OC to volatile 984 gases, CO₂, H₂ and CH₄, has been demonstrated experimentally (Siskin and Katritzky, 1991; 985 Seewald, 2001; McCollom and Seewald, 2003b, a) and is known to be an important loss 986 mechanism that "scrubs" fluids of deep-ocean DOC in high temperature reaction zones (Lang et 987 al., 2006; Hawkes et al., 2015), although DOC sorption to crustal surfaces is also possible 988 (Schwarzenbach et al., 2005). In subseafloor regions adjacent to high temperature venting, 989 oxygenated seawater can mix with reduced hydrothermal fluids, creating chemical disequilibria 990 that autotrophic microorganism can use to fuel primary production (McCollom and Shock, 1997). 991 These regions can be hot spots of organic matter production both within the fluids and within the 992 surrounding sediments (Karl et al., 1980; Lang et al., 2006; Wankel et al., 2011). DOC is seemingly 993 produced abiotically at vents hosted on ultramafic rocks, with elevated concentrations compared 994 to overlying bottom water (Lang et al., 2010). Most of this increase is thought to be due to the 995 abiotic formation of small organic acids (Lang et al., 2010; McDermott et al., 2015). A 996 combination of microbial, abiotic and sedimentary sources likely contributes to subseafloor DOC 997 pools, which if circulated in the basement from the ridges, could diffuse into overlying sediments, 998 potentially fueling communities.

1000 **4. Computational models**

999

1001 A theoretical understanding of the carbon cycle in marine sediments underpins much of the present knowledge and constraints on the burial and transformation of organic carbon in this 1002 1003 setting, both in the present day and throughout Earth's history. Numerical models have been used 1004 for decades within this framework to quantify how POC drives sediment diagenesis at particular 1005 locations (Berner, 1964; Lerman, 1971; Berner, 1980; Boudreau, 1997). These models can include 1006 the role of particular electron acceptors, secondary redox reactions, sorption and desorption, and 1007 microbial dynamics while simultaneously accounting for transport processes such a sedimentation, 1008 bioturbation, bioirrigation and the diffusion of solutes. A comprehensive review of reaction 1009 transport models and their use in marine sediments is provided in (Arndt et al., 2013), covering 1010 the formulation of various reaction and transport networks, their application to natural and 1011 engineered systems across a wide range of temporal and spatial scales, and the challenges and

limitations of implementing these models. The following sections highlight recent modeling
 advances with respect to marine sediments and new research directions that could further improve
 them.

1015

1016 **4.1 Representing pools of organic molecules**

It is both infeasible and ineffectual to explicitly resolve the true complexity of organic 1017 1018 carbon in marine sediments in numerical models. Rather, owing to its complexity and 1019 heterogeneity, OC is usually characterized by its apparent bulk reactivity. Therefore, numerical models for OC transformations in marine sediments must account for changes in both (1) 1020 1021 concentration and (2) reactivity of the bulk substrate. If more specific information describing how the proportions of compound types are changing were available, ideally, this would be quantified 1022 1023 as well. The concentration of organic carbon is usually defined in absolute terms as the sum of all 1024 of the various reactive fractions, and corresponds directly to conventional laboratory 1025 measurements of POC. Models of organic carbon reactivity can be broadly divided into two 1026 classes: discrete and continuum. Within discrete models, OC is attributed to either a single pool of 1027 concentration G (one-G) with a constant first-order degradation rate (Berner, 1980), or divided 1028 across a discrete number of pools representing various classes of reactivity (multi-G), where the 1029 apparent reactivity of the bulk organic matter is related to the reactivity of each class (Jørgensen, 1030 1978). Continuum models, alternatively, assume a continuous distribution of organic matter 1031 compounds across an infinite spectrum of reactivities (Aris, 1968; Ho and Aris, 1987; Boudreau 1032 and Ruddick, 1991). The choice of model formulation is generally governed by the overarching 1033 research question, the relevant spatial- and timescales, data availability, and mathematical expedience. Since these numerical formulations are abstracted from measurements, it can be 1034 1035 challenging to constrain reaction rate constants based on experimental data. Consequently, organic matter reactivity is traditionally constrained by inverse modeling of comprehensive sets of 1036 sediment depth profiles. 1037

1038

1039 4.2 Organic molecular data

1040 There are very few models describing the degradation of OC in sediments that use 1041 information about the molecular character of organic compounds. This is primarily because there are relatively few reports describing the chemical formulas and structures of marine sedimentary 1042 organic compounds in a way that could be parameterized in a model (see Section 2.2). The 1043 1044 modeling studies that have used molecular information have only tangentially addressed how this 1045 information impacts rates of marine OC degradation. Reaction transport modeling (Niggemann et 1046 al., 2007; Freitas et al., 2017) and kinetic modeling studies (Schouten et al., 2010) have explored 1047 how different degradation rates of specific biomarker compounds could influence the interpretation of past sea surface temperatures. The abundance and proportion of certain types of 1048 1049 amino acids in marine sediments has been used as an index for the degradation state of POC 1050 (Dauwe et al., 1999). In another study that evaluated the abundance of particular organic compounds, the Gibbs energy associated with the degradation of organic compounds was used to 1051 1052 explain patterns of biomarker degradation in sediments from the Southeast Atlantic ocean (Hernández-Sánchez et al., 2014). The Gibbs energies in this study are estimated based on the 1053 1054 oxidation state of the carbon in organic compounds, which is in turn calculated from the 1055 stoichiometry of the compounds (see LaRowe and Van Cappellen, 2011). LaRowe and Van 1056 Cappellen used this approach to argue that the rate of organic carbon degradation in anoxic marine sediments is retarded by the molecular character of the organic compounds. The rationale for this 1057

1059

is based on the idea that the rate of microbial catabolism is proportional to the Gibbs energy of that 1058 metabolism: less exergonic reactions are catalyzed more slowly than more exergonic reactions (see 1060 Jin and Bethke, 2003; LaRowe et al., 2012).

1061

1062 **4.3.** Microorganisms in models

Although the actions of microorganisms are implicitly accounted for within models of OC 1063 1064 degradation in marine sediments, they are rarely explicitly resolved within the mathematical 1065 formulae, i.e. as a separate state variable. This is mostly due to (a) model applications that are focused on geochemistry or biogeochemistry rather than microbiology, and (b) the uncertainties 1066 1067 concerning microbial growth, maintenance, death, and dormancy in marine sediments which must be resolved in order to accurately simulate microbial dynamics (Hoehler and Jørgensen, 2013; 1068 1069 Jørgensen and Marshall, 2016; Kempes et al., 2017; Bradley et al., 2018a). Nevertheless, 1070 (Boudreau, 1999) was the first to couple microbial processes to organic carbon diagenesis, and 1071 derive a mathematical basis between previously observed microbial biomass and organic carbon 1072 concentrations in sediments (Bird and Duarte, 1989). Later work incorporated explicit 1073 mathematical representation of microbial processes into reactive transport frameworks linking 1074 sediment redox gradients and reaction rates to microbial processes (Wirtz, 2003; Thullner et al., 1075 2005), the competition of different microbial groups for a common substrate (Thullner et al., 2007) 1076 and to assess the impact of transport processes on transient biomass distributions, anaerobic 1077 oxidation of methane rates and methane release fluxes from the sea floor (Dale et al., 2006; Regnier et al., 2011; Puglini et al., 2019). Formulations of microbial processes in sediment models have 1078 1079 been developed further to account for the relative importance of growth versus maintenance, and variable physiological states (i.e. dormancy) (Bradley et al., 2018a, 2019). 1080 1081

1082 **4.4** Application scale

1083 Despite marine sediments comprising a significant volume of the Earth's surface (LaRowe 1084 et al., 2017a), playing a dominant role in the global carbon budget on long time scales (Arndt et 1085 al., 2013), and hosting a significant fraction of Earth's living biomass (Kallmeyer et al., 2012), their treatment in global-scale models of the Earth's climate and biogeochemistry may often be 1086 1087 little more than a simple closure term for mass conservation (Soetaert et al., 2000; Hülse et al., 1088 2017; Lessin et al., 2018). In case they are explicitly resolved, the interactions between marine 1089 sediments and the overlying water column, i.e. benthic-pelagic coupling, are often neglected or 1090 crudely implemented in such global-scale models (Soetaert et al., 2000; Gehlen et al., 2006; Munhoven, 2007; Hülse et al., 2017). Even relatively simple reaction-transport models are more 1091 1092 typically applied to regional scales (Ruardij and Van Raaphorst, 1995; Luff and Moll, 2004; Arndt 1093 and Regnier, 2007) over idealized global ocean hypsometric transects or provinces (e.g. Soetaert et al., 1996; Thullner et al., 2009; Krumins et al., 2013) and over time-spans of thousands to 1094 1095 millions of years (e.g. Arndt et al., 2009; Krumins et al., 2013; Orcutt et al., 2013b). Only a very 1096 small number of global scale biogeochemical or Earth System models employ an explicit, 1097 vertically resolved, multi-component description of diagenetic dynamics (e.g. Heinze et al., 1999; 1098 Munhoven, 2007; Shaffer et al., 2008; Palastanga et al., 2011; Ilyina et al., 2013; Tjiputra et al., 1099 2013; Hülse et al., 2018a). Yet, due to the need to find computationally efficient analytical solutions 1100 to the diagenetic equations these descriptions generally rely on simplifying assumptions and/or are 1101 restricted to the upper few centimeters of the sediment. However, coupled models can provide 1102 important insights into ocean biogeochemical cycling and climate feedbacks. For instance, the recent coupling of a vertically resolved benthic model to the three-dimensional Earth System 1103

1104 Model CGENIE (Hülse et al., 2018b) has revealed that organic matter sulfurization reduces the 1105 extent and intensity of toxic euxinic conditions, and accelerates climate cooling on a scale that is globally significant during ocean anoxic event recovery (Hülse et al., 2019). Alternatively, large 1106 1107 ensemble runs of complex one-dimensional diagenetic models have also been used to derive 1108 transfer functions for specific target outputs such as benthic fluxes or benthic methane gas hydrates 1109 that have then been applied on a regional and global scale (Gypens et al., 2008; Dale et al., 2015; 1110 Capet et al., 2016). In another large-scale effort, (LaRowe et al., 2020) have developed a global-1111 scale model based on the analytical solution of the one-dimensional conservation equation for benthic organic carbon dynamics that reveals the 3-D distribution of marine sediment POC for 1112 1113 Quaternary-aged sediments (< 2.6 Ma) as well as rates of its degradation. Bradley et al. (in 1114 revision) have built on this model to quantitatively estimate the cell-specific power utilization of 1115 microorganisms transforming POC in global aerobic, sulfogenic and methanogenic sediment 1116 horizons.

1117

1118 **5. Outlook**

1119 Although it is well known that human activity is responsible for a rapid rise of atmospheric 1120 CO₂, it is unclear how this disturbance will impact the natural fluxes of carbon among major global reservoirs. In particular, it is still an open question how human-induced climate change will alter 1121 1122 the strength of the marine sedimentary carbon sink, and therefore control of atmospheric CO₂. 1123 Recent observations and model projections suggest that the impact of climate change on marine 1124 POC is likely going to be regionally heterogeneous (Passow and Carlson, 2012), and will include 1125 warming waters, disappearing sea ice, increased DIC content, lowered pH and altered fluxes of organic carbon into and through the water column (Levin and Le Bris, 2015; Sweetman et al., 1126 2017), particularly from terrestrial sources (Bauer et al., 2013b; Regnier et al., 2013). Each of these 1127 factors have potential implications for how organic carbon is delivered to and processed within 1128 1129 sediments. Yet, because benthic carbon dynamics are first and foremost controlled by the quantity 1130 and quality of OC that settles onto the seafloor, perhaps the most important factor in controlling the response of deep ocean (< 200 m) benthic carbon dynamics to projected environmental change 1131 is the biological carbon pump, the process by which organic carbon produced in the euphotic zone 1132 1133 is exported into the deep ocean. In addition, coastal benthic carbon cycling, in particular in 1134 nearshore depositional environments in the vicinity of large rivers such as, among others, the Arctic shelf, the Amazon shelf or the South China Sea, will also be affected by changes in 1135 1136 terrestrial inputs.

1137 The geologic record includes numerous examples of such climate change induced 1138 perturbations in the functioning of the biological carbon pump (Arthur et al., 1985; Kohfeld et al., 1139 2005; Ridgwell, 2011; John et al., 2014; Hülse et al., 2019). For instance, abundant black shales 1140 in the sedimentary record speak to periods when much or all of the world's bottom ocean waters 1141 were devoid of free O₂ (Jenkyns, 2010), likely due to a warmer climate, the paleogeography, 1142 enhanced nutrient supply and elevated marine primary productivity that in turn might have been maintained by benthic nutrient cycles perturbed by this bottom water anoxia (Ingall and Jahnke, 1143 1144 1994; Van Cappellen and Ingall, 1994). Ultimately, the widespread anoxic and euxinic depositional conditions enhanced organic carbon preservation such that atmospheric CO₂ and, 1145 1146 therefore, temperatures decreased and O₂ eventually returned to bottom waters, a process lasting 1147 tens to hundreds of thousands of years (Arthur et al., 1988; Kolonic et al., 2005; Jarvis et al., 2011; Hülse et al., 2019; Raven et al., 2019). Though there is a consensus that it is not currently possible 1148 to gauge how the biological pump will be altered in the next several decades (Pörtner et al., 2014), 1149

a number of studies are hinting at how climate change will impact the flux of carbon to marinesediments.

1152 As a master variable for biogeochemical reactions, temperature will have likely have a 1153 significant impact on the reactivity of organic carbon in marine sediments. Ocean warming has 1154 already led to the expansion of oxygen minimum zones in the water column over the last 50 years 1155 (Schmidtko et al., 2017; Bertagnolli and Stewart, 2018; Breitburg et al., 2018), disrupting the role 1156 of bioturbation on POC reactivity. Warmer water seems to select for smaller plankton altering the 1157 export flux of POC to the seafloor (Morán et al., 2010) since smaller particles tend to have longer transit times to the seafloor. In high latitudes, the disappearance of sea ice, an increase in the length 1158 1159 of the growing season, fundamental changes to regional circulation (e.g. Atlantification) resulting in changing salinity, temperature and nutrient conditions will exert important, yet poorly known 1160 1161 impacts on ecosystem structure. In general, temperature changes are known to influence the 1162 structure and function of marine microbial communities (Sunagawa et al., 2015b), and, in addition 1163 to other environmental forces, virus-host relationships (Danovaro et al., 2008; Danovaro et al., 2011), which in turn can alter patterns of carbon sequestration (Guidi et al., 2016) in sediments. 1164 1165 The combination of warming, acidification, eutrophication and human activities such as bottom trawling (Hiddink et al., 2017) and seafloor mining (Orcutt et al., 2020) might lead to ecosystem 1166 destruction and/or many areas of the seafloor to become covered in microbial mats (de Bakker et 1167 1168 al., 2017; Ford et al., 2018), rather than bioturbated sediments. In addition, lower than normal pH 1169 cause some marine bacterioplankton to express genes for maintenance rather than growth (Bunse 1170 et al., 2016), thus slowing the flux of C to sediments. Although it is difficult to predict how it will 1171 impact the reactivity of organic carbon in sediments, e.g. (Isla and DeMaster, 2018), it is clear that climate change is altering the physiochemical variables that govern microbial behavior. Therefore, 1172 1173 attempts to better understand how carbon fluxes will respond to projected climate change and also 1174 how carbon fluxes have responded to past extreme climate and carbon cycle perturbations will require an ecosystem approach that includes the role of microorganisms (Cavicchioli et al., 2019). 1175

1176 The information summarized in this contribution supports the emerging view that organic matter reactivity in marine sediments is a complex function of biological, geochemical and 1177 1178 physical forces that vary from one part of the seafloor to another. Given the large variety of 1179 organic compounds, minerals, organisms, and environmental conditions found in marine sediments, it is undoubtedly true that all of the mechanistic hypotheses described in this review 1180 contribute in some way to the long-term preservation of organic carbon, with the relative 1181 importance of each changing with both time and space. Going forward, it will be critical that 1182 studies examining sedimentary organic carbon account for the whole array of biophysiochemical 1183 1184 factors that impact reactivity, thus providing the much needed interdisciplinary data sets required to advance our quantitative understanding and predictive capabilities. Disentangling which 1185 mechanism operates under what set of environmental conditions is a complicated task requiring 1186 1187 integration of measurements, laboratory experiments, quantitative modelling and an open mind. A 1188 community effort will be required to understand not only what determines organic carbon 1189 reactivity in marine sediments now, but how this will change in the future. Moreover, integrated 1190 approaches considering marine sediments in relation to the terrestrial and water column settings is 1191 needed to gain a truly global and comprehensive understanding of the carbon cycle.

1192

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1216 **Figure captions:**

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Figure 1. Schematic of topics discussed in this review: ecosystem factors that influence the reactivity of particulate organic carbon, POC, in marine sediments. The bulleted processes/variables and four categories are not necessarily independent of one another.

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Figure 2. Concentration of particular organic carbon (POC) at a) the sea floor, b) 1 meter below the sea floor and c) 10 meters below the seafloor, based on calculations by LaRowe et al. (2020) and data summarized in Wallmann et al (2012), which, for Holocene sediments, is taken from a compilation by (Seiter et al., 2004). Grey areas in c) indicate regions where Quaternary sediments (i.e. sediments deposited throughout the last 2.59 million years) are less than 10 m thick. The Quaternary cutoff is the temporal limit for the model used by LaRowe et al. (2020).

1228

Figure 3. Estimated a) regions of the seafloor where dissolved O_2 is modeled to be present throughout the sediment to the underlying oceanic basement and b) depth of the sulfate-methane transition (SMT) zone. The dark shading in a) refers to the minimum extent of modeled O_2 penetrating regions while the light shading, together with the dark shading, indicate the maximum extent, based on D'Hondt et al (2015). The white regions in b) denote regions where there is no SMT. All data for b) from Egger et al (2018).

1235

Figure 4. Oxygen concentration profiles as a function of depth in marine sediments and ocean basement crust from three IODP drill cores located on ~ 8 Ma flank of the mid-Atlantic Ridge (also known as North Pond – see Orcutt et al., 2013). The bottom panel contains a cross section schematic of what is thought to be the mechanism of O_2 delivery to basal sediments – upward diffusion from oxygenated fluid flowing rapidly in the basement. This fluid is chemically very

1241 similar to local bottom seawater and is likely introduced to the subsurface from locally outcropping

- basalt (see Meyer et al., 2016). Oxygen loss in flowing fluids is thought to result from bothdiffusion into sediments and consumption during microbial oxidation of DOC.
- 1244
- Figure 5. Calculated cell concentrations in marine sediment at a) the seafloor, b) 1 meter below the sea floor and c) 10 meter below the seafloor using the data compilation and approach described by Kallmeyer et al. (2012).
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Figure 6. Phylogenetic tree showing microbial groups containing genes encoding putatively secreted enzymes capable of degrading proteins and carbohydrates in anoxic marine sediments (modified from Orsi et al, 2018). The term CAZymes refers to carbohydrate-active enzymes (Lombard et al., 2013).

1254 **References**

- Albertsen, M., Hugenholtz, P., Skarshewski, A., Nielsen, K.L., Tyson, G.W. and Nielsen, P.H.
 (2013) Genome sequences of rare, uncultured bacteria obtained by differential coverage
 binning of multiple metagenomes. *Nature Biotechnology* 31, 533.
- Aller, J.Y. and Aller, R.C. (1986) Evidence for localized enhancement of biological associated
 with tube and burrow structures in deep-sea sediments at the HEEBLE site, western North
 Atlantic. Deep Sea Research Part A. Oceanographic Research Papers 33, 755-790.
- Aller, R., Aller, J. and Kemp, P. (2001) Effects of Particle and Solute Transport on Rates and
 Extent of Remineralization in Bioturbated Sediments, pp. 315-333.
- Aller, R.C. (1980) Diagenetic Processes Near the Sediment-Water Interface of Long Island Sound.
 II. Fe and Mn, in: Saltzman, B. (Ed.), *Advances in Geophysics*. Elsevier, pp. 351-415.
- Aller, R.C. (1982) Carbonate dissolution in nearshore terrigeneous muds—the role of physical and
 biological reworking. *Journal of Geology* 90, 79-95.
- Aller, R.C. (1994) Bioturbation and remineralization of sedimentary organic matter: effects of
 redox oscillation. *Chemical Geology* 114, 331-345.
- Aller, R.C. (2014) 8.11 Sedimentary Diagenesis, Depositional Environments, and Benthic
 Fluxes, in: Holland, H.D., Turekian, K.K. (Eds.), *Treatise on Geochemistry (Second Edition)*. Elsevier, Oxford, pp. 293-334.
- Aller, R.C. and Aller, J.Y. (1998) The effect of biogenic irrigation intensity and solute exchange
 on diagenetic reaction rates in marine sediments. *Journal of Marine Research* 56, 905-936.
- Aller, R.C., Blair, N.E., Xia, Q. and Rude, P.D. (1996) Remineralization rates, recycling, and storage of carbon in Amazon shelf sediments. *Continental Shelf Research* 16, 753-786.
- Aller, R.C., Charnock, H., Edmond, J.M., McCave, I.N., Rice, A.L. and Wilson, T.R.S. (1990)
 Bioturbation and manganese cycling in hemipelagic sediments. *Philosophical Transactions of the Royal Society of London. Series A, Mathematical and Physical* Sciences 331, 51-68.
- Aller, R.C. and Cochran, J.K. (2019) The Critical Role of Bioturbation for Particle Dynamics,
 Priming Potential, and Organic C Remineralization in Marine Sediments: Local and Basin
 Scales. Frontiers in Earth Science 7.
- Amrani, A. (2014) Organosulfur compounds: Molecular and isotopic evolution from biota to oil
 and gas. *Annual Review of Earth and Planetary Sciences* 42, 733-768.
- Anantharaman, K., Brown, C.T., Hug, L.A., Sharon, I., Castelle, C.J., Probst, A.J., Thomas, B.C.,
 Singh, A., Wilkins, M.J., Karaoz, U., Brodie, E.L., Williams, K.H., Hubbard, S.S. and

- Banfield, J.F. (2016) Thousands of micorbial genomes shed light on interconneecacted
 biogeochemical processes in an aquifer system. *Nature Communications* 7, Art. 13219.
- Antia, A., Koeve, W., Fischer, G., Blanz, T., Schulz-Bull, D., Scholten, J., Neuer, S., Kremling,
 K.K., J., Peinert, R., Hebbeln, D., Bathmann, U., Conte, M., Fehner, U. and Zeitzschel, B.
 (2001) Basin-wide particulate carbon flux in the Atlantic Ocean: regional export patterns
 and potential for atmospheric carbon sequestration. *Global Biogeochemical Cycles* 15, 845-862.
- Archer, D. and Maier-Reimer, E. (1994) Effect of deep-sea sedimentary calcite preservation on
 atmospheric CO2 concentration. *Nature* 367, 260-263.
- Aris, R. (1968) Prolegomena to the rational analysis of systems of chemical reactions, II. Some adenda. *Arch. Rational Mech. Analysis* 27, 356-364.
- Armstrong, R.A., Lee, C., Hedges, J.I., Honjo, S. and Wakeham, S.G. (2002) A new, mechanistic
 model for organic carbon fluxes in the ocean based on the quantitative association of POC
 with ballast minerals. *Deep-Sea Research Part II* 49, 219-236.
- Arnarson, T.S. and Keil, R.G. (2001) Organic-mineral interactions in marine sediments studied
 using density fractionation and X-ray photoelectron spectroscopy. *Organic Geochemistry* 32, 1401-1415.
- Arnarson, T.S. and Keil, R.G. (2007) Changes in organic matter-mineral interactions for marine
 sediments with varying oxygen exposure times. *Geochimica et Cosmochimica Acta* 71,
 3545-3556.
- Arndt, S., Brumsack, H.-J. and Wirtz, K.W. (2006) Cretaceous black shales as active bioreactors:
 A biogeochemical model for the deep biosphere encountered during ODP Leg 207
 (Demerara Rise). *Geochim. Cosmochim. Acta* 70, 408-425.
- Arndt, S., Hetzel, A. and Brumsack, H.-J. (2009) Evolution of organic matter degradation in
 Cretaceous black shales inferred from authigenic barite: A reaction-transport model. *Geochim. Cosmochim. Acta* 73, 2000-2022.
- Arndt, S., Jørgensen, B.B., LaRowe, D.E., Middelburg, J.B.M., Pancost, R.D. and Regnier, P.
 (2013) Quantifying the degradation of organic matter in marine sediments: A review and synthesis. *Earth Sci. Rev.* 123, 53-86.
- Arndt, S. and Regnier, P. (2007) A model for the benthic-pelagic coupling of silica in estuarine
 ecosystems: sensitivity analysis and system scale simulation. *Biogeosciences* 4, 331-352.
- Arnosti, C. (2011) Microbial extracellular enzymes and the marine carbon cycle. *Annual Review* of Marine Science 3, 401-425.
- Arnosti, C., Bell, C., Moorhead, D.L., Sinsabaugh, R.L., Steen, A.D., Stromberger, M.,
 Wallenstein, M. and Weintraub, M.N. (2014) Extracellular enzymes in terrestrial,
 freshwater, and marine environments: perspectives on system variability and common
 research needs. *Biogechemistry* 117, 5-21.
- Arthur, M.A., Dean, W.E., Pollastro, R., Scholle, P.A. and Claypool, G.E. (1985) A comparative geochemical study of two transgressive pelagic limestone units, Cretaceous western interior basin, U.S., in: Pratt, L.M. (Ed.), *Fine-Grained Deposits and Biofacies of the Cretaceous Western Interior Seaway: Evidence of Cyclic Sedimentary Processes*. Soc. For Sediment. Geol., Tulsa, OK, USA, pp. 16-27.
- Arthur, M.A., Dean, W.E. and Pratt, L.M. (1988) Geochemical and climatic effects of increased
 marine organic carbon burial at the Cenomanian/Turonian boundary. *Nature* 335, 714-717.
- Bäckström, D., Yutin, N., Jørgensen, S.L., Dharamshi, J., Homa, F., Zaremba-Niedwiedzka, K.,
 Spang, A., Wolf, Y.I., Koonin, E.V. and Ettema, T.J.G. (2019) Virus Genomes from Deep

- Sea Sediments Expand the Ocean Megavirome and Support Independent Origins of ViralGigantism. *mBio* 10, e02497-02418.
- Baker, B.J., Saw, J.H., Lind, A.E., Lazar, C.S., Hinrichs, K.-U., Teske, A.P. and Ettema, T.J.G.
 (2016) Genomic inference of the metabolism of cosmopolitan subsurface Archaea,
 Hadesarchaea. *Nature Microbiology* 1, 16002.
- Ball, G.I. and Aluwihare, L.I. (2014) CuO-oxidized dissolved organic matter (DOM) investigated
 with comprehensive two dimensional gas chromatography-time of flight-mass
 spectrometry (GC × GC-TOF-MS). Org. Geochem. 75, 87-98.
- Bao, R., Zhao, M., McNichol, A., Wu, Y., Guo, X., Haghipour, N. and Eglinton, T.I. (2019) On
 the Origin of Aged Sedimentary Organic Matter Along a River-Shelf-Deep Ocean
 Transect. *Journal of Geophysical Research: Biogeosciences* 124, 2582-2594.
- Barber, A., Brandes, J., Leri, A., Lalonde, K., Balind, K., Wirick, S., Wang, J. and Gélinas, Y.
 (2017) Preservation of organic matter in marine sediments by inner-sphere interactions
 with reactive iron. *Scientific Reports* 7, DOI:10.1038/s41598-41017-00494-41590.
- Bauer, J.E., Cai, W.-J., Raymond, P.A., Bianchi, T.S., Hopkinson, C.S. and Regnier, P.A.G.
 (2013a) The changing carbon cycle of the coastal ocean. *Nature* 504, 61-70.
- Bauer, J.E., Cai, W.-J., Raymond, P.A., Bianchi, T.S., Hopkinson, C.S. and Regnier, P.A.G.
 (2013b) The changing carbon cycle of the coastal ocean. *Nature* 504, 61.
- Bauer, J.E. and Druffel, E.R.M. (1998) Ocean margins as a significant source of organic matter to
 the deep open ocean. *Nature* 392, 482-485.
- Benner, R. (2002) Chemical Composition and Reactivity, in: Hansell, D.A., Carlson, C.A. (Eds.),
 Biogeochemistry of Marine Dissolved Organic Matter. Academic Press, San Diego, pp.
 59-90.
- Berner, R.A. (1964) An idealized model of dissolved sulfate distribution in recent sediments.
 Geochimica et Cosmochimica Acta 28, 1497-1503.
- Berner, R.A. (1978) Sulfate reduction and the rate of deposition of marine sediments. *Earth and Planetary Science Letters* 37, 492-498.
- Berner, R.A. (1980) *Early Diagenesis: A Theoretical Approach* Princeton Univ. Press, Princeton,
 N.J.
- Berner, R.A. (1981) Authigenic mineral formation resulting from organic matter decomposition
 in modern sediments. *Fortschritte der Mineralogie* 59, 117-135.
- Berner, R.A. (1990) Atmospheric Carbon Dioxide Levels Over Phanerozoic Time. *Science* 249, 1365
 1382-1386.
- Berner, R.A. (2003) The long-term carbon cycle, fossil fuels and atmospheric composition. *Nature* 426, 323-326.
- Berner, R.A. and Canfield, D.E. (1989) A new model for atmospheric oxygen over Phanerozoic
 time. *Amer. J. Sci.* 289, 333-361.
- Berry, W.B.N. and Wilde, P. (1978) Progressive ventilation of the oceans an explanation for the
 distribution of the lower paleozoic black shales. *American Journal of Science* 278, 257 275.
- Bertagnolli, A.D. and Stewart, F.J. (2018) Microbial niches in marine oxygen minimum zones.
 Nature Reviews Microbiology 16, 723-729.
- Beulig, F., Røy, H., Glombitza, C. and Jørgensen, B.B. (2018) Control on rate and pathway of
 anaerobic organic carbon degradation in the seabed. *PNAS* doi/10.1073/pnas.1715789115,
 6.

- Bianchi, T.S. (2011) The role of terrestrially derived organic carbon in the coastal ocean: A
 changing paradigm and the priming effect. *Proceedings of the National Academy of Sciences* 108, 19473.
- Bianchi, T.S., Johansson, B. and Elmgren, R. (2000) Breakdown of phytoplankton pigments in
 Baltic sediments: effects of anoxia and loss of deposit-feeding macrofauna. *Journal of Experimental Marine Biology and Ecology* 251, 161-183.
- Biddle, J.F., Fitz-Gibbon, S., Schuster, S.C., Brenchley, J.E. and House, C.H. (2008) Metagenomic
 signatures of the Peru Margin subseafloor biosphere show a genetically distinct
 environment. *Proceedings of the National Academy of Sciences* 105, 10583-10588.
- Biddle, J.F., Lipp, J.S., Lever, M.A., Lloyd, K.G., Sørenen, K.B., Anderson, R., Fredricks, H.F.,
 Elvert, M., Kelly, T.J., Schrag, D.P., Sogin, M.L., Brenchley, J.E., Teske, A., House, C.H.
 and Hinrichs, K.-U. (2006) Heterotrophic Archaea dominate sedimentary subsurface
 ecosystems off Peru. *PNAS* 103, 3846-3851.
- Bird, D.F. and Duarte, C.M. (1989) Bacteria–Organic Matter Relationship in Sediments: A Case
 of Spurious Correlation. *Canadian Journal of Fisheries and Aquatic Sciences* 46, 904-908.
- Bird, J.T., Tague, E.D., Zinke, L., Schmidt, J.M., Steen, A.D., Reese, B., Marshall, I.P.G.,
 Webster, G., Weightman, A., Castro, H.F., Campagna, S.R. and Lloyd, K.G. (2019)
 Uncultured Microbial Phyla Suggest Mechanisms for Multi-Thousand-Year Subsistence
 in Baltic Sea Sediments. *mBio* 10, e02376-02318.
- Blair, N.E. and Aller, R.C. (2011) The Fate of Terrestrial Organic Carbon in the Marine
 Environment. *Annual Review of Marine Science* 4, 401-423.
- Blair, N.E. and Aller, R.C. (2012) The Fate of Terrestrial Organic Carbon in the Marine
 Environment. *Annual Review of Marine Science* 4, 401-423.
- Blair, N.E., Leithold, E.L., Ford, S.T., Peeler, K.A., Holmes, J.C. and Perkey, D.W. (2003) The
 persistence of memory: the fate of ancient sedimentary organic carbon in a modern
 sedimentary system. *Geochimica et Cosmochimica Acta* 67, 63-73.
- Blattmann, T.M., Liu, Z., Zhang, Y., Zhao, Y., Haghipour, N., Montluçon, D.B., Plötze, M. and
 Eglinton, T.I. (2019) Mineralogical control on the fate of continentally derived organic
 matter in the ocean. *Science* 366, 742.
- Borowski, W.S., Paull, C.K. and Ussler, W. (1999) Global and local variations of interstitial sulfate
 gradients in deep-water, continental margin sediments: Sensitivity to underlying methane
 and gas hydrates. *Marine Geology* 159, 131-154.
- Böttcher, M.E., Hetzel, A., Brumsack, H.-J. and Schipper, A. (2006) Sulfur-iron carbon
 geochemistry in sediments of the Demerara Rise. *ODP Scientific Results* 207, 12.
- Boudreau, B.P. (1994) Is burial velocity a master parameter for bioturbation? *Geochimica et Cosmochimica Acta* 58, 1243-1249.
- Boudreau, B.P. (1997) Diagenetic models and their implementation : Modelling transport and
 reactions in aquatic sediments. Springer, Berlin.
- Boudreau, B.P. (1998) Mean mixed depth of sediments: The wherefore and the why. *Limnology and Oceanography* 43, 524-526.
- Boudreau, B.P. (1999) A theoretical investigation of the organic carbon-microbial biomass relation
 in muddy sediments. *Aquatic Microbial Ecology* 17, 181-189.
- Boudreau, B.P., Huettel, M., Forster, S., Jahnke, R.A., McLachlan, A., Middelburg, J.J., Nielsen,
 P., Sansone, F., Taghon, G., Van Raaphorst, W., Webster, I., Weslawski, J.M., Wiberg, P.
 and Sundby, B. (2001) Permeable marine sediments: Overturning an old paradigm. *Eos*,
- 1423 *Transactions American Geophysical Union* **82**, 133-136.

- Boudreau, B.P. and Ruddick, B.R. (1991) On a reactive continuum representation of organic
 matter diagenesis. *Amer. J. Sci.* 291, 507-538.
- Bourke, M.F., Marriott, P.J., Glud, R.N., Hasler-Sheetal, H., Kamalanathan, M., Beardall, J.,
 Greening, C. and Cook, P.L.M. (2016) Metabolism in anoxic permeable sediments is
 dominated by eukaryotic dark fermentation. *Nature Geoscience* 10, 30.
- Bowles, M.W., Mogollón, J.M., Kasten, S., Zabel, M. and Hinrichs, K.-U. (2014) Global rates of
 marine sulfate reduction and implication for sub-sea-floor metabolic activities. *Science*334, 889-891.
- Bradley, J.A., Amend, J.P. and LaRowe, D.E. (2018a) Bioenergetic controls on micorbial
 ecophysiology in marine sediments. *Frontiers in Microbiology* 9, Article 180.
- Bradley, J.A., Amend, J.P. and LaRowe, D.E. (2018b) Necromass as a limited source of energy
 for microorganisms in marine sediments *Journal of Geophysical Research: Biogeosciences* 1436
 123, 577-590.
- Bradley, J.A., Amend, J.P. and LaRowe, D.E. (2019) Survival of the fewest: Microbial dormancy
 and maintenance in marine sediments through deep time. *Geobiology* 17, 43-59.
- Bradley, J.A., Arndt, S., Amend, J.P., Burwicz, E., Dale, A.W., Egger, M. and LaRowe, D.E. (in
 revision) Widespread energy limitation to life in global subseafloor sediments. *Science* Advances.
- Braun, S., Mhatre, S.S., Jaussi, M., Røy, H., Kjeldsen, K.U., Pearce, C., Seidenkrantz, M.-S.,
 Jørgensen, B.B. and Lomstein, B.A. (2017) Microbial turnover times in the deep seabed
 studied by amino acid racemization modelling. *Scientific Reports* 7, 5680.
- Breitburg, D., Levin, L.A., Oschlies, A., Grégoire, M., Chavez, F.P., Conley, D.J., Garçon, V.,
 Gilbert, D., Gutiérrez, D., Isensee, K., Jacinto, G.S., Limburg, K.E., Montes, I., Naqvi,
 S.W.A., Pitcher, G.C., Rabalais, N.N., Roman, M.R., Rose, K.A., Seibel, B.A., Telszewski,
 M., Yasuhara, M. and Zhang, J. (2018) Declining oxygen in the global ocean and coastal
 waters. *Science* 359, eaam7240.
- Buesseler, K.O., Trull, T.W., Steinberg, D.K., Silver, M.W., Siegel, D.A., Saitoh, S.I., Lamborg,
 C.H., Lam, P.J., Karl, D.M., Jiao, N.Z., Honda, M.C., Elskens, M., Dehairs, F., Brown,
 S.L., Boyd, P.W., Bishop, J.K.B. and Bidigare, R.R. (2008) VERTIGO (VERtical
 Transport In The Global Ocean): a study of particle sources and flux attenuation in the
 North Pacific. *Deep-Sea Research Part II* 55, 1522–1539.
- Bunse, C., Lundin, D., Karlsson, C.M.G., Akram, N., Vila-Costa, M., Palovaara, J., Svensson, L.,
 Holmfeldt, K., González, J.M., Calvo, E., Pelejero, C., Marrasé, C., Dopson, M., Gasol,
 J.M. and Pinhassi, J. (2016) Response of marine bacterioplankton pH homeostasis gene
 expression to elevated CO2. *Nature Climate Change* 6, 483-487.
- Buongiorno, J., Turner, S., Webster, G., Asai, M., Shumaker, A.K., Roy, T., Weightman, A.,
 Schippers, A. and Lloyd, K.G. (2017) Interlaboratory quantification of Bacteria and
 Archaea in deeply buried sediments of the Baltic Sea (IODP Expedition 347). *FEMS Microbiology Ecology* 93.
- Burdige, D.J. (2005) Burial of terrestrial organic matter in marine sediments: A re-assessment.
 Global Biogeochemical Cycles 19.
- 1465 Burdige, D.J. (2007a) *Geochemistry of Marine Sediments*. Princeton University Press, Princeton.
- Burdige, D.J. (2007b) Preservation of organic matter in marine sediments: Controls, mechanisms
 and an imbalance in sediment organic carbon budgets? *Chem. Rev.* 107, 467-485.

- Burdige, D.J. (2012) Estuarine and coastal sediments coupled biogeochemical cycling, in: Laane,
 R., Middelburg, J.J. (Eds.), *Treatise on Estuarine and Coastal Science*. Academic Press,
 pp. 279-316.
- Burdige, D.J. and Gardner, K.G. (1998) Molecular weight distribution of dissolved organic carbon
 in marine sediment pore waters. *Marine Chemistry* 62, 45-64.
- Burdige, D.J. and Komada, T. (2015) Sediment Pore Waters, in: Hansell, D.A., Carlson, C.A.
 (Eds.), *Biogeochemistry of Marine Dissolved Organic Matter* 2ed. Academic Press, London, pp. 535-577.
- Burdige, D.J. and Nealson, K.H. (1986) Chemical and microbiological studies of sulfide-mediated
 manganese reduction. *Geomicrobiology Journal* 4, 361-387.
- Cai, L., Jørgensen, B.B., Suttle, C.A., He, M., Cragg, B.A., Jiao, N. and Zhang, R. (2019) Active
 and diverse viruses persist in the deep sub-seafloor sediments over thousands of years. *The ISME Journal* 13, 1857-1864.
- Calvert, S.E. and Pedersen, T.F. (1992a) Organic carbon accumulation and preservation in marine
 sediments: how important is anoxia, in: Whelan, J., Farrington, J.W. (Eds.), *Organic Matter: Productivity, Accumulation, and Preservation in Recent and Ancient Sediments*Columbia University Press, New York, pp. 231-293.
- Calvert, S.E. and Pedersen, T.F. (1992b) Organic carbon accumulation and preservation in marine
 sediments: How important is anoxia?, in: Whelan, J.K., Farrington, J.W. (Eds.),
 Productivity, accumulation and preservation of organic matter in recent and ancient sediments. Columbia University Press, New York, pp. 231-263.
- Camacho-Ibar, V.c.F., Aveytua-Alcázar, L. and Carriquiry, J.D. (2003) Fatty acid reactivities in
 sediment cores from the northern Gulf of California. *Organic Geochemistry* 34, 425-439.
- Canfield, D.E. (1994) Factors influencing organic carbon preservation in marine sediments. *Chem. Geol.* 114, 315-329.
- Canfield, D.E., Jørgensen, B.B., Fossing, H., Glud, R., Gundersen, J., Ramsing, N.B., Thamdrup,
 B., Hansen, J.W., Nielsen, L.P. and Hall, P.O.J. (1993) Pathways of organic carbon
 oxidation in three continental margin sediments. *Marine Geology* 113, 27-40.
- Canfield, D.E., Kristensen, E. and Thamdrup, B. (2005) Advances in Marine Biology: Aquatic
 Geomicrobiology. Elsevier Academic Press, San Diego.
- Canuel, E.A. and Hardison, A.K. (2016) Sources, Ages, and Alteration of Organic Matter in
 Estuaries. *Annual Review of Marine Science* 8, 409-434.
- Canuel, E.A. and Martens, C.S. (1996) Reactivity of recently deposited organic matter:
 Degradation of lipid compounds near the sediment-water interface. *Geochim. Cosmochim.* Acta 60, 1793-1806.
- Capet, A., Meysman, F.J.R., Akoumianaki, I., Soetaert, K. and Grégoire, M. (2016) Integrating
 sediment biogeochemistry into 3D oceanic models: A study of benthic-pelagic coupling in
 the Black Sea. *Ocean Modelling* 101, 83-100.
- Caradec, S., Grossi, V., Gilbert, F., Guigue, C. and Goutx, M. (2004) Influence of various redox
 conditions on the degradation of microalgal triacylglycerols and fatty acids in marine
 sediments. *Organic Geochemistry* 35, 277-287.
- Cavicchioli, R., Ripple, W.J., Timmis, K.N., Azam, F., Bakken, L.R., Baylis, M., Behrenfeld, M.J.,
 Boetius, A., Boyd, P.W., Classen, A.T., Crowther, T.W., Danovaro, R., Foreman, C.M.,
 Huisman, J., Hutchins, D.A., Jansson, J.K., Karl, D.M., Koskella, B., Mark Welch, D.B.,
- 1512 Martiny, J.B.H., Moran, M.A., Orphan, V.J., Reay, D.S., Remais, J.V., Rich, V.I., Singh,
- 1513 B.K., Stein, L.Y., Stewart, F.J., Sullivan, M.B., van Oppen, M.J.H., Weaver, S.C., Webb,

- 1514 E.A. and Webster, N.S. (2019) Scientists' warning to humanity: microorganisms and 1515 climate change. *Nature Reviews Microbiology* **17**, 569-586.
- Ciobanu, M.-C., Burgaud, G., Dufresne, A., Breuker, A., Rédou, V., Ben Maamar, S., Gaboyer,
 F., Vandenabeele-Trambouze, O., Lipp, J.S., Schippers, A., Vandenkoornhuyse, P.,
 Barbier, G., Jebbar, M., Godfroy, A. and Alain, K. (2014) Microorganisms persist at record
 depths in the subseafloor of the Canterbury Basin. *ISME J.*, doi:10.1038/ismej.2013.1250.
- Claypool, G.E. and Kaplan, I.R. (1974) The origin and distribution of methane in marine
 sediments, in: I.R.Kaplan (Ed.), *Natural Gases in Marine Sediments*. Plenum Press, New
 York, pp. 99-139.
- 1523 Close, H.G., Shah, S.R., Ingalls, A.E., Diefendorf, A.F., Brodie, E.L., Hansman, R.L., Freeman,
 1524 K.H., Aluwihare, L.I. and Pearson, A. (2013) Export of submicron particulate organic
 1525 matter to mesopelagic depth in an oligotrophic gyre. *PNAS* 110, 12565-12570.
- Condie, K.C., Des Marais, D.J. and Abbott, D. (2001) Precambrian superplumes and
 supercontinents: a record in black shales, carbon isotopes, and paleoclimates? *Precambrian Research* 106, 239-260.
- Coolen, M.J.L., Cypionka, H., Sass, A.M., Sass, H. and Overmann, J. (2002) Ongoing
 modification of Mediteerranean Pleistocene sapropels mediated by prokaryotes. *Science* 296, 2407-2410.
- Coppola, L., Gustafsson, Ö., Andersson, P., Eglinton, T.I., Uchida, M. and Dickens, A.F. (2007)
 The importance of ultrafine particles as a control on the distribution of organic carbon in
 Washington Margin and Cascadia Basin sediments. *Chemical Geology* 243, 142-156.
- Coskun, Ö.K., Pichler, M., Vargas, S., Gilder, S. and Orsi, W.D. (2018) Linking uncultivated
 microbial populations and benthic carbon turnover by using quantitative stable isotope
 probing. *Applied and Environmental Microbiology* 84, e01083-01018.
- Cottrell, M.T., Yu, L. and Kirchman, D.L. (2005) Sequence and expression analyses of Cytophaga like hydrolases in a Western arctic metagenomic library and the Sargasso Sea. *Applied and environmental microbiology* 71, 8506-8513.
- Cowie, G., Calvert, S., De Lange, G.J., Keil, R. and Hedges, J. (1998) Extents and implications of
 organic matter alteration at oxidation fronts in turbidites from the Madeira Abyssal Plain.
 Proceedings of the Ocean Drilling Program Science Results 157, 581-589.
- Cowie, G.L., Hedges, J.I. and Calvert, S.E. (1992) Sources and relative reactivities of amino acids,
 neutral sugars, and lignin in an intermittently anoxic marine environment. *Geochimica et Cosmochimica Acta* 56, 1963-1978.
- Cowie, G.L., Hedges, J.I., Prahl, F.G. and de Lange, G.J. (1995) Elemental and major biochemical
 changes across an oxidation front in a relict turbidite: An oxygen effect. *Geochimica et Cosmochimica Acta* 59, 33-46.
- Cowie, G.L., Mowbray, S., Lewis, M., Matheson, H. and McKenzie, R. (2009) Carbon and nitrogen elemental and stable isotopic compositions of surficial sediments from the Pakistan margin of the Arabian Sea. *Deep Sea Research Part II: Topical Studies in Oceanography* 56, 271-282.
- Cruse, A.M. and Seewald, J.S. (2006) Geochemistry of low-molecular weight hydrocarbons in
 hydrothermal fluids from Middle Valley, northern Juan de Fuca Ridge. *Geochimica et Cosmochimica Acta* 70, 2073-2092.
- D'Hondt, S., Inagaki, F., Zarikian, C.A., Abrams, L.J., Dubois, N., Engelhardt, T., Evans, H.,
 Ferdelman, T., Gribsholt, B., Harris, R., Hoppie, B.W., Hyun, J.-H., Kallmeyer, J., Kim,
 J., Lynch, J.E., McKinley, C.C., Mitsunobu, S., Morono, Y., Murray, R.W., Pockalny, R.,

- Sauvage, J., Shimono, T., Shiraishi, F., Smith, D.C., Smith-Duque, C.E., Spivack, A.J.,
 Steinsbu, B.O., Suzuki, Y., Szpak, M., Toffin, L., Uramoto, G., Yamaguchi, Y.T., Zhang,
 G., Zhang, X.-H. and Ziebis, W. (2015) Presence of oxygen and aerobic communities from
 sea floor to basement in deep-sea sediments. *Nat. Geosci.* 8, 299-304.
- D'Hondt, S., Jørgensen, B.B., Miller, D.J., Batzke, A., Blake, R., Cragg, B.A., Cypionka, H.,
 Dickens, G.R., Ferdelman, T., Hinrichs, K.U., Holm, N.G., Mitterer, R., Spivack, A.,
 Wang, G.Z., Bekins, B., Engelen, B., Ford, K., Gettemy, G., Rutherford, S.D., Sass, H.,
 Skilbeck, C.G., Aiello, I.W., Guerin, G., House, C.H., Inagaki, F., Meister, P., Naehr, T.,
 Niitsuma, S., Parkes, R.J., Schippers, A., Smith, D.C., Teske, A., Wiegel, J., Padilla, C.N.
 and Acosta, J.L.S. (2004) Distributions of microbial activities in deep subseafloor
 sediments. *Science* 306, 2216 2221.
- 1571 D'Hondt, S., Jorgensen, B.B., Miller, D.J. and Expedition 329 Scientists (2003) Leg 201 Summary.
 1572 Proc. ODP 201.
- 1573 Dai, J. and Sun, M.-Y. (2007) Organic matter sources and their use by bacteria in the sediments of
 1574 the Altamaha estuary during high and low discharge periods. *Organic Geochemistry* 38, 1 1575 15.
- 1576 Dale, A.W., Nickelsen, L., Scholz, F., Hensen, C., Oschlies, A. and Wallmann, K. (2015) A revised
 1577 global estimate of dissolved iron fluxes from marine sediments. *Global Biogeochemical* 1578 *Cycles* 29, 691-707.
- 1579 Dale, A.W., Regnier, P. and Van Cappellen, P. (2006) Bioenergetic controls on anaerobic
 1580 oxidation of methane (AOM) in coastal marine sediments: A theoretical analysis. *American* 1581 *Journal of Science* 306, 246-294.
- Danczak, R.E., Johnston, M.D., Kenah, C., Wrighton, K.C. and Wilkins, M.J. (2017) Members of
 the Candidate Phyla Radiation are functionally differentiated by carbon- and nitrogen cycling capabilities. *Microbiome* 5, 112.
- Danovaro, R., Corinaldesi, C., Dell'Anno, A., Fuhrman, J.A., Middelburg, J.J., Noble, R.T. and
 Suttle, C.A. (2011) Marine viruses and global climate change. *FEMS Microbiology Reviews* 35, 993-1034.
- Danovaro, R., Dell'Anno, A., Corinaldesi, C., Magagnini, M., Noble, R., Tamburini, C. and
 Weinbauer, M. (2008) Major viral impact on the functioning of benthic deep-sea ecosystems. *Nature* 454, 1084-1087.
- Danovaro, R., Dell'Anno, A., Corinaldesi, C., Rastelli, E., Cavicchioli, R., Krupovic, M., Noble,
 R.T., Nunoura, T. and Prangishvili, D. (2016) Virus-mediated archaeal hecatomb in the
 deep seafloor. *Science Advances* 2, e1600492.
- Dauwe, B. and Middelburg, J.J. (1998) Amino acids and hexosamines as indicators or organic
 matter degradation state in North Sea sediments. *Limnol. Oceanog.* 43, 782-798.
- Dauwe, B., Middelburg, J.J. and Herman, P.M.J. (2001) Effect of oxygen on the degradability of
 organic matter in subtidal and intertidal sediments of the North Sea area. *Marine Ecology Progress Series* 215, 13–22.
- Dauwe, B., Middelburg, J.J., Herman, P.M.J. and Heip, C.H.R. (1999) Linking diagenetic
 alteration of amino acids and bulk organic matter reactivity. *Limnology and Oceanography* 44, 1809-1814.
- de Bakker, D.M., van Duyl, F.C., Bak, R.P.M., Nugues, M.M., Nieuwland, G. and Meesters, E.H.
 (2017) 40 Years of benthic community change on the Caribbean reefs of Curaçao and Bonaire: the rise of slimy cyanobacterial mats. *Coral Reefs* 36, 355-367.

- 1605de Bruijn, F.J. (2010) Handbook of Molecular Microbial Ecology I: Metagenomics and1606Complementary Approaches. Wiley Blackwell, Hoboken, New Jersey.
- de Lange, G.J., Jarvis, I. and Kuijpers, A. (1987) Geochemical characteristics and provenance of
 late Quaternary sediments from the Madeira Abyssal Plain, N Atlantic. *Geological Society, London, Special Publications* 31, 147-165.
- de Leeuw, J.W. and Largeau, C. (1993) A review of macromolecular organic compounds that
 comprise living organisms and their role in kerogen, coal and petroleum formation, in:
 Engel, M.H., Macko, S.A. (Eds.), *Organic geochemistry: Principles and applications*.
 Plenum Press, New York.
- de Leeuw, J.W., Versteegh, G.J.M. and van Bergen, P.F. (2006) Biomacromolecules of algae and
 plants and their fossil analogues, in: Rozema, J., Aerts, R., Cornelissen, H. (Eds.), *Plants and Climate Change*. Springer Netherlands, Dordrecht, pp. 209-233.
- 1617 Demaison, G.J. and Moore, G.T. (1980) Anoxic environments and oil source bed genesis.
 1618 American Association of Petroleum Geologists Bulletin 64, 1179–1209.
- 1619 Dombrowski, N., Teske, A.P. and Baker, B.J. (2018) Expansive microbial metabolic versatility
 1620 and biodiversity in dynamic Guaymas Basin hydrothermal sediments. *Nature* 1621 *Communications* 9, 4999.
- Dunne, J.P., Sarmiento, J.L. and Gnanadesikan, A. (2007) A synthesis of global particle export
 from the surface ocean and cycling through the ocean interior and on the seafloor. *Global Biogeochem. Cycles* 21, GB4006, doi:4010.1029/2006GB002907.
- 1625 Eckman, J.E., Nowell, A.R.M. and Jumars, P.A. (1981) Sediment destabilization by animal tubes
 1626 (burrows). *Journal of Marine Research* 39, 361-374.
- Egger, M., Riedinger, N., Mogollon, J.M. and Jorgensen, B.B. (2018) Global diffusive fluxes of
 methane in marine sediments. *Nature Geoscience* 11, 421-425.
- Eglington, T.I. and Repeta, D.J. (2014) Organic matter in the contemporary ocean, in: Turekian,
 K.K., Holland, H.D. (Eds.), *Treatise on Geochemistry*, 2 ed. Elsevier, Amsterdam, pp. 151 189.
- Eglinton, T.I., Curtis, C.D. and Rowland, S.J. (1987) Generation of water-soluble organic-acids
 from kerogen during hydrous pyrolysis implications for porosity development.
 Mineralogical Magazine 51, 495-503.
- Eglinton, T.I., Irvine, J.E., Vairavamurthy, A., Zhou, W. and Manowitz, B. (1994) Formation and
 diagenesis of macromolecular organic sulfur in Peru margin sediments. *Organic Geochemistry* 22, 781-799.
- Eglinton, T.I. and Repeta, D.J. (2004) Organic Matter in the Contemporary Ocean, in: Holland,
 H.D., Turekian, K.K. (Eds.), *Treatise on Geochemistry*. Elsevier, Amsterdam.
- Eglinton, T.I. and Repeta, D.J. (2010) Organic Matter in the Contemporary Ocean, in: Holland,
 H.D., Turekian, K.K. (Eds.), *Geochemistry of Earth Surface Systems: A derivative of the Treatise on Geochemistry*. Elsevier Science & Technology, London, pp. 389-424.
- Eglinton, T.I. and Repeta, D.J. (2014) 8.6 Organic Matter in the Contemporary Ocean, in:
 Holland, H.D., Turekian, K.K. (Eds.), *Treatise on Geochemistry (Second Edition)*.
 Elsevier, Oxford, pp. 151-189.
- 1646 Ehrlich, H.L. (1971) Bacteriology of manganese nodules. V. Effect of hydrostatic pressure on
 1647 bacterial oxidation of MnII and reduction of MnO2. *Appl Microbiol* 21, 306-310.
- Emerson, S. (1985) Organic carbon preservation in marine sediments. , in: Sundquist, E.T.,
 Broecker, W.S. (Eds.), *The Carbon Cycle and Atmospheric CO2: Natural Variations* Archean to Present. AGU, Washington, D.C., pp. 78-87.

- Emerson, S. and Bender, M. (1981) Carbon fluxes at the sediment-water interface of the deep-sea
 calcium carbonate preservation. *J. Mar. Res.* 39, 139-162.
- Engelen, B., Ziegelmüller, K., Wolf, L., Köpke, B., Gittel, A., Cypionka, H., Treude, T.,
 Nakagawa, S., Inagaki, F., Lever, M.A. and Steinsbu, B.r.O. (2008) Fluids from the
 Oceanic Crust Support Microbial Activities within the Deep Biosphere. *Geomicrobiology Journal* 25, 56-66.
- 1657 Engelhardt, T., Orsi, W. and Jørgensen, B.B. (2015) Viral activities and life cycles in deep
 1658 subseafloor sediments. *Environmental Microbiology Reports* 7, 868-873.
- Estes, E.R., Pockalny, R., D'Hondt, S., Inagaki, F., Morono, Y., Murray, R.W., Nordlund, D.,
 Spivack, A.J., Wankel, S.D., Xiao, N. and Hansel, C.M. (2019) Persistent organic matter
 in oxic subseafloor sediment. *Nature Geoscience* 12, 126-131.
- Field, C.B., Behrenfeld, M.J., Randerson, J.T. and Falkowski, P. (1998) Primary Production of the
 Biosphere: Integrating Terrestrial and Oceanic Components. *Science* 281, 237-240.
- Findlay, S. and Tenore, K.E.N. (1982) Nitrogen Source for a Detritivore: Detritus Substrate Versus
 Associated Microbes. *Science* 218, 371.
- Fisher, J.B. (1987) Distribution and occurrence of aliphatic acid anions in deep subsurface waters.
 Geochimica et Cosmochimica Acta 51, 2459-2468.
- Fontaine, S., Barot, S., Barré, P., Bdioui, N., Mary, B. and Rumpel, C. (2007) Stability of organic
 carbon in deep soil layers controlled by fresh carbon supply. *Nature* 450, 277.
- Ford, A.K., Bejarano, S., Nugues, M.M., Visser, P.M., Albert, S. and Ferse, S.C.A. (2018) Reefs
 under Siege—the Rise, Putative Drivers, and Consequences of Benthic Cyanobacterial
 Mats. Frontiers in Marine Science 5.
- Fox, C.A., Abdulla, H.A., Burdige, D.J., Lewicki, J.P. and Komada, T. (2018) Composition of
 Dissolved Organic Matter in Pore Waters of Anoxic Marine Sediments Analyzed by 1H
 Nuclear Magnetic Resonance Spectroscopy. *Frontiers in Marine Science* 5.
- Francois, R. (1987) A study of sulphur enrichment in the humic fraction of marine sediments
 during early diagenesis. *Geochimica et Cosmochimica Acta* 51, 17-27.
- Francois, R., Honjo, S., Krishfield, R. and Manganini, S. (2002) Factors controlling the flux of
 organic carbon to the bathypelagic zone of the ocean. *Global Biogeochemical Cycles* 16,
 GB001722.
- Freitas, F.S., Pancost, R.D. and Arndt, S. (2017) The impact of alkeone degradation on U^{K'37}
 paleothermometry: A model-derived assessment. *Paleoceanography* 32, 648-672.
- Froelich, P.N., Klinkhammer, G.P., Bender, M.L., Luedtke, N.A., Heath, G.R., Cullen, D.,
 Dauphin, P., Hammond, D., Hartman, B. and Maynard, V. (1979) Early oxidation of
 organic matter in pelagic sediments of the eastern equatorial Atlantic: suboxic diagenesis. *Geochim. Cosmochim. Acta* 43, 1075-1090.
- Furnes, H., Muehlenbachs, K., Torsvik, T., Thorseth, I.H. and Tumyr, O. (2001) Microbial
 fractionation of carbon isotopes in altered basaltic glass from the Atlantic Ocean, Lau Basin
 and Costa Rica Rift. *Chemical Geology* 173, 313-330.
- Gehlen, M., Bopp, L., Emprin, N., Aumont, O., Heinze, C. and Ragueneau, O. (2006) Reconciling
 surface ocean productivity, export fluxes and sediment composition in a global
 biogeochemical ocean model. *Biogeosciences* 3, 521-537.
- 1693 Glud, R.N. (2008) Oxygen dynamics of marine sediments. *Marine Biology Research* 4, 243-289.
- 1694 Goldhaber, M.B., Aller, R.C., Cochran, J.K., Rosenfeld, K.K., Martens, C.S. and Berner, R.A.
- 1695 (1977) Sulfate reduction, diffusion and bioturbation in Long Island Sound sediments:
 1696 Report of the FOAM group. *American Journal of Science* 277, 193-237.

- Gomez-Saez, G.V., Niggemann, J., Dittmar, T., Pohlabeln, A.M., Lang, S.Q., Noowong, A.,
 Pichler, T., Wörmer, L. and Bühring, S.I. (2016) Molecular evidence for abiotic
 sulfurization of dissolved organic matter in marine shallow hydrothermal systems. *Geochimica et Cosmochimica Acta* 190, 35-52.
- Goth, K., de Leeuw, J.W., Püttmann, W. and Tegelaar, E.W. (1988) Origin of Messel Oil Shale
 kerogen. *Nature* 336, 759-761.
- Graf, G. (1992) Benthic–pelagic coupling: a benthic view. *Oceanography and Marine Biology* **30**, 149-190.
- Gransch, J.A. and Posthuma, J. (1974) On the origin of sulphur in crudes, in: Tissot, B., Bienner,
 F. (Eds.), *Advances in Organic Geochemistry 1973*, Paris, pp. 727-739.
- Grossi, V., Blokker, P. and Sinninghe Damsté, J.S. (2001) Anaerobic biodegradation of lipids of
 the marine microalga Nannochloropsis salina. *Organic Geochemistry* 32, 795-808.
- 1709 Guidi, L., Chaffron, S., Bittner, L., Eveillard, D., Larhlimi, A., Roux, S., Darzi, Y., Audic, S., 1710 Berline, L., Brum, J.R., Coelho, L.P., Espinoza, J.C.I., Malviya, S., Sunagawa, S., Dimier, 1711 C., Kandels-Lewis, S., Picheral, M., Poulain, J., Searson, S., Stemmann, L., Not, F., 1712 Hingamp, P., Speich, S., Follows, M., Karp-Boss, L., Boss, E., Ogata, H., Pesant, S., Weissenbach, J., Wincker, P., Acinas, S.G., Bork, P., de Vargas, C., Iudicone, D., Sullivan, 1713 M.B., Raes, J., Karsenti, E., Bowler, C., Gorsky, G. and Tara Oceans Consortium, C. 1714 1715 (2016) Plankton networks driving carbon export in the oligotrophic ocean. Nature 532, 1716 465-470.
- Gupta, N.S. (2015) Plant Biopolymer–Geopolymer: Organic Diagenesis and Kerogen Formation.
 Frontiers in Materials 2.
- Gupta, N.S., Briggs, D.E.G., Collinson, M.E., Evershed, R.P., Michels, R., Jack, K.S. and Pancost,
 R.D. (2007) Evidence for the in situ polymerisation of labile aliphatic organic compounds
 during the preservation of fossil leaves: Implications for organic matter preservation.
 Organic Geochemistry 38, 499-522.
- Gutleben, J., Chaib De Mares, M., van Elsas, J.D., Smidt, H., Overmann, J. and Sipkema, D. (2018)
 The multi-omics promise in context: from sequence to microbial isolate. *Critical Reviews in Microbiology* 44, 212-229.
- Gypens, N., Lancelot, C. and Soetaert, K. (2008) Simple parameterisations for describing N and P
 diagenetic processes: Application in the North Sea. *Progress in Oceanography* 76, 89-110.
- Hales, B. and Emerson, S. (1996) Calcite dissolution in sediments of the Ontong-Java Plateau: In
 situ measurements of pore water O2 and pH. *Global Biogeochemical Cycles* 10, 527-541.
- Hansell, D.A. (2013) Recalcitrant dissolved organic carbon fractions. *Annu. Rev. Mar. Sci.* 5, 421445.
- Hansell, D.A. and Carlson, C.A. (2015) *Biogeochemistry of marine dissolved organic matter*, 2
 ed. Academic Press, London.
- Hartnett, H.E., Keil, R.G., Hedges, J.I. and Devol, A.H. (1998) Influence of oxygen exposure time
 on organic carbon preservation in continental margin sediments. *Nature* 391, 572-574.
- Harvey, H.R. and Macko, S.A. (1997) Kinetics of phytoplankton decay during simulated
 sedimentation: changes in lipids under oxic and anoxic conditions. *Org. Geochem.* 27, 129140.
- Harvey, H.R., Tuttle, J.H. and Bell, J.T. (1995) Kinetics of phytoplankton decay during simulated
 sedimentation: Changes in biochemical composition and microbial activity under oxic and
 anoxic conditions. *Geochim. Cosmochim. Acta* 59, 3367-3377.

- Hawkes, J.A., Hansen, C.T., Goldhammer, T., Bach, W. and Dittmar, T. (2016) Molecular
 alteration of marine dissolved organic matter under experimental hydrothermal conditions. *Geochimica et Cosmochimica Acta* 175, 68-85.
- Hawkes, J.A., Rossel, P.E., Stubbins, A., Butterfield, D.A., Connelly, D.P., Achterberg, E.P.,
 Koschinsky, A., Chavagnac, V., Hansen, C.T., Bach, W. and Dittmar, T. (2015) Efficient
 removal of recalcitrant deep-ocean dissolved organic matter during hydrothermal
 circulation. *Nature Geoscience* 8, 856-860.
- Hedges, J.I., Clark, W.A. and Cowie, G.L. (1988) Fluxes and reactivities of organic matter in a
 coastal marine bay. *Limnology and Oceanography* 33, 1137-1152.
- Hedges, J.I., Eglington, G., Hatcher, P.G., Kirchman, D.L., Arnosti, C., Derenne, S., Evershed,
 R.P., Kögel-Knaber, I., De Leeuw, J.W., Littke, R., Michaelis, W. and Rullkötter, J. (2000)
 The molecularly-uncharacterized component of nonliving organic matter in natural
 environments. *Organic Geochemistry* 31, 945-958.
- Hedges, J.I., Hu, F.S., Devol, A.H., Hartnett, H.E., Tsamakis, E. and Keil, R.G. (1999)
 Sedimentary organic matter preservation: A test for selective degradation under oxic conditions. *American Journal of Science* 299, 529-555.
- Hedges, J.I. and Keil, R.G. (1995) Sedimentary organic matter preservation: an assessment and
 speculative hypothesis. *Mar. Chem.* 49, 81-115.
- Hedges, J.I., Keil, R.G. and Benner, R. (1997) What happens to terrestrial organic matter in the
 ocean? Org. Geochem. 27, 195-212.
- Hedges, J.I. and Oades, J.M. (1997) Comparative organic geochemistries of soils and marine
 sediments. *Org. Geochem.* 27, 319-361.
- Heinze, C., Maier-Reimer, E., Winguth, A.M.E. and Archer, D. (1999) A global oceanic sediment
 model for long-term climate studies. *Global Biogeochemical Cycles* 13, 221-250.
- Heitmann, T. and Blodau, C. (2006) Oxidation and incorporation of hydrogen sulfide by dissolved
 organic matter. *Chemical Geology* 235, 12-20.
- Hemingway, J.D., Rothman, D.H., Grant, K.E., Rosengard, S.Z., Eglington, T.I., Derry, L.A. and
 Galy, V.V. (2019) Mineral protection regulates long-term global preservation of natural
 organic carbon. *Nature* 570, 228-231.
- Henrichs, S.M. and Reeburgh, W.S. (1987) Anaerobic mineralization of marine sediment organic
 matter: rates and the role of anaerobic processes in the oceanic carbon economy.
 Geomicrobiology Journal 5, 191-237.
- Henson, S., Sanders, R. and Madsen, E. (2012a) Global patterns in efficiency of particulate organic
 carbon export and transfer to the deep ocean. *Global Biogeochemical Cycles* 26,
 GB004099.
- Henson, S.A., Sanders, R. and Madsen, E. (2012b) Global patterns in efficiency of particulate
 organic carbon export and transfer to the deep ocean. *Global Biogeochemical Cycles* 26.
- Herman, P.M.J., Middelburg, J.J. and Van de Koppel, J. (1999) Ecology of estuarine
 macrobenthos. Advances in Ecology Research 29, 195-240.
- Hernández-Sánchez, M.T., LaRowe, D.E., Deng, F., Homoky, W.B., Browning, T.J., Martin, P.,
 Mills, R.A. and Pancost, R.D. (2014) Further insights into how sediment redox status
 controls the preservation and composition of sedimentary biomarkers. *Org. Geochem.* 76,
 220-234.
- Hertkorn, N., Benner, R., Frommberger, M., Shmitt-Kopplin, P., Witt, M., Kaiser, K., Kettrup, A.
 and Hedges, J.I. (2006) Characterization of a major refractory component of marine
 dissolved organic matter. *Geochimica et Cosmochimica Acta* 70, 2990-3013.

- Hetzel, A., Böttcher, M.E., Wortmann, U.G. and Brumsack, H.-J. (2009) Paleo-redox conditions
 during OAE 2 reflected in Demerara Rise sediment geochemistry (ODP Leg 207). *Palaeogeography, Palaeoclimatology, Palaeoecology* 273, 302-328.
- Hiddink, J.G., Jennings, S., Sciberras, M., Szostek, C.L., Hughes, K.M., Ellis, N., Rijnsdorp, A.D.,
 McConnaughey, R.A., Mazor, T., Hilborn, R., Collie, J.S., Pitcher, C.R., Amoroso, R.O.,
 Parma, A.M., Suuronen, P. and Kaiser, M.J. (2017) Global analysis of depletion and
 recovery of seabed biota after bottom trawling disturbance. *Proc Natl Acad Sci U S A* 114,
 8301-8306.
- Ho, T.C. and Aris, R. (1987) On apparent second-order kinetics *Amer. Inst. Chem. Eng. J.* 33, 1050-1051.
- Hoefs, M.J.L., Sinninghe Damsté, J.S., De Lange, G.J. and de Leeuw, J.W. (1998) Changes in
 kerogen composition across an oxidation front in Madeira Abyssal Plain turbidites as
 revealed by pyrolysis GC-MS. *Proceeding of the Ocean Drilling Program, Scientific Results* 157, 591-607.
- Hoehler, T.M. and Jørgensen, B.B. (2013) Micorbial life under extreme energy limitation. *Nat. Rev. Microbiol.* 11, 83-94.
- Honjo, S., Manganini, S.J., Krishfield, R.A. and Francois, R. (2008) Particulate organic carbon
 fluxes to the ocean interior and factors controlling the biological pump: A synthesis of
 global sediment trap programs since 1983. *Progress in Oceanography* 76, 217-285.
- Huettel, M., Berg, P. and Kostka, J.E. (2014) Benthic Exchange and Biogeochemical Cycling in
 Permeable Sediments. *Annual Review of Marine Science* 6, 23-51.
- Hug, L.A., Baker, B.J., Anantharaman, K., Brown, C.T., Probst, A.J., Castelle, C.J., Butterfield,
 C.N., Hernsdorf, A.W., Amano, Y., Ise, K., Suzuki, Y., Dudek, N., Relman, D.A., Finstad,
 K.M., Amundson, R., Thomas, B.C. and Banfield, J.F. (2016) A new view of the tree of
 life. *Nature Microbiology* 1, 16048.
- Hülse, D., Arndt, S., Daines, S., Regnier, P. and Ridgwell, A. (2018a) OMEN-SED 1.0: a novel,
 numerically efficient organic matter sediment diagenesis module for coupling to Earth
 system models. *Geosci. Model Dev.* 11, 2649-2689.
- Hülse, D., Arndt, S., Daines, S., Regnier, P. and Ridgwell, A. (2018b) OMEN-SED 1.0: a novel,
 numerically efficient organic matter sediment diagenesis module for coupling to Earth
 system models. *Geoscience Model Development* 11, 2649-2689.
- Hülse, D., Arndt, S. and Ridgwell, A. (2019) Mitigation of Extreme Ocean Anoxic Event
 Conditions by Organic Matter Sulfurization. *Paleoceanography and Paleoclimatology* 34, 476-489.
- Hülse, D., Arndt, S., Wilson, J.D., Munhoven, G. and Ridgwell, A. (2017) Understanding the
 causes and consequences of past marine carbon cycling variability through models. *Earth- Science Reviews* 171, 349-382.
- Hulthe, G., Hulth, S. and Hall, P.O.J. (1998) Effect of oxygen on degradation rate of refractory
 and labile organic matter in continental margin sediments. *Geochimica et Cosmochimica* Acta 62, 1319-1328.
- 1828 Hunt, J.M. (1996) Petroleum Geochemistry and Geology, second ed. Freeman.
- Hüpers, A. and Kopf, A.J. (2012) Effect of smecetite dehydration on pore water geochemistry in
 the shallow subduction zone: An experimental approach. *Geochemistry, Geophysics, Geosystems* 13, Q0AD26.

- Hwang, J., Druffel, E.R.M. and Eglinton, T.I. (2010) Widespread influence of resuspended
 sediments on oceanic particulate organic carbon: Insights from radiocarbon and aluminum
 contents in sinking particles. *Global Biogeochemical Cycles* 24.
- Hyun, J.H., Kim, S.H., Mok, J.S., Cho, H., Lee, T., Vandieken, V. and Thamdrup, B. (2017)
 Manganese and iron reduction dominate organic carbon oxidation in surface sediments of
 the deep Ulleung Basin, East Sea. *Biogeosciences* 14, 941-958.
- Ilyina, T., Six, K.D., Segschneider, J., Maier-Reimer, E., Li, H. and Núñez-Riboni, I. (2013)
 Global ocean biogeochemistry model HAMOCC: Model architecture and performance as
 component of the MPI-Earth system model in different CMIP5 experimental realizations. *Journal of Advances in Modeling Earth Systems* 5, 287-315.
- Inagaki, F., Hinrichs, K.-U., Kubo, Y., Bowles, M.W., Heuer, V.B., Hong, W.-L., Hoshino, T., 1842 1843 Ijiri, A., Imachi, H., Ito, M., Kaneko, M., Lever, M.A., Lin, Y.-S., Methé, B.A., Morita, S., 1844 Morono, Y., Tanikawa, W., Bihan, M., Bowden, S.A., Elvert, M., Glombitza, C., Gross, D., Harrington, G.J., Hori, T., Li, K., Limmer, D., Liu, C.-H., Murayama, M., Ohkouchi, 1845 1846 N., Ono, S., Park, Y.-S., Phillips, S.C., Prieto-Mollar, X., Purkey, M., Riedinger, N., 1847 Sanada, Y., Sauvage, J., Snyder, G., Susilawati, R., Takano, Y., Tasumi, E., Terada, T., Tomaru, H., Trembath-Reichert, E., Wang, D.T. and Yamada, Y. (2015) Exploring deep 1848 1849 microbial life in coal-bearing sediment down to ~2.5 km below the ocean floor. Science 1850 349, 420-424.
- Inagaki, F., Nunoura, T., Nakagawa, S., Teske, A., Lever, M.A., Lauer, A., Suzuki, M., Takai, K.,
 Delwiche, M., Colwell, F.S., Nealson, K.H., Horikoshi, K., D'Hondt, S. and Jørgensen,
 B.B. (2006) Biogeographical distribution and diversity of microbes in methane hydratebearing deep marine sediments on the Pacific Ocean Margin. *PNAS* 103, 2815-2820.
- Ingall, E. and Jahnke, R. (1994) Evidence for enhanced phosphorus regeneration from marine
 sediments overlain by oxygen depleted waters. *Geochimica et Cosmochimica Acta* 58, 2571-2575.
- Ingalls, A.E., Lee, C., Wakeham, S.G. and Hedges, J.I. (2003) The role of biominerals in the
 sinking flux and preservation of amino acids in the Southern Ocean along 170°W. *Deep-* Sea Research II 50, 713-738.
- Inthorn, M., Mohrholz, V. and Zabel, M. (2006a) Nepheloid layer distribution in the Benguela
 upwelling area offshore Namibia. *Deep Sea Research Part I: Oceanographic Research Papers* 53, 1423-1438.
- Inthorn, M., Wagner, T., Scheeder, G. and Zabel, M. (2006b) Lateral transport controls
 distribution, quality, and burial of organic matter along continental slopes in high productivity areas. *Geology* 34, 205-208.
- Isla, E. and DeMaster, D.J. (2018) Labile organic carbon dynamics in continental shelf sediments
 after the recent collapse of the Larsen ice shelves off the eastern Antarctic Peninsula: A
 radiochemical approach. *Geochimica et Cosmochimica Acta* 242, 34-50.
- Iverson, N. and Jørgensen, B.B. (1985) Anaerobic methane oxidation rates at the sulfate-methane
 transition in marine sediments from Kattegat and Skagerrak (Denmark). *Limnology and Oceanography* 30, 944-955.
- Jahnke, R.A. and Jahnke, D.B. (2000) Rates of C, N, P and Si recycling and denitrification at the
 US Mid-Atlantic continental slope depocenter. *Deep Sea Research Part I: Oceanographic Research Papers* 47, 1405-1428.
- Jahnke, R.A., Reimers, C.E. and Craven, D.B. (1990) Intensification of recycling of organic matter
 at the sea floor near ocean margins. *Nature* 348, 50-54.

- Jarvis, I., Lignum, J.S., Gröcke, D.R., Jenkyns, H.C. and Pearce, M.A. (2011) Black shale
 deposition, atmospheric CO2 drawdown, and cooling during the Cenomanian-Turonian
 Oceanic Anoxic Event. *Paleoceanography* 26.
- Jenkyns, H.C. (2010) Geochemistry of oceanic anoxic events. *Geochem. Geophys. Geosys.* 11, 003004 doi:03010.01029/02009GC002788.
- Jessen, G.L., Lichtschlag, A., Ramette, A., Pantoja, S., Rossel, P.E., Schubert, C.J., Struck, U. and
 Boetius, A. (2017) Hypoxia causes preservation of labile organic matter and changes
 seafloor microbial community composition (Black Sea). *Science Advances* 3, e1601897.
- Jin, Q. and Bethke, C.M. (2003) A new rate law describing microbial respiration. *Appl. Environ. Microbiol.* 69, 2340-2348.
- Jochum, L.M., Chen, X., Lever, M.A., Loy, A., Jørgensen, B.B., Schramm, A. and Kjeldsen, K.U.
 (2017) Depth distribution and assembly of sulfate-reducing microbial communities in marine sediments of Aarhus Bay. *Applied and Environmental Microbiology* 83, e01547-01517.
- John, E.H., Wilson, J.D., Pearson, P.N. and Ridgwell, A. (2014) Temperature-dependent
 remineralization and carbon cycling in the warm Eocene oceans. *Palaeogeography, Palaeoclimatology, Palaeoecology* 413, 158-166.
- Johnson, H.P. and Pruis, M.J. (2003) Fluxes of fluid and heat from the oceanic crustal reservoir.
 Earth Planet. Sci. Lett. 216, 565-574.
- Johnson, K., Purvis, G., Lopez-Capel, E., Peacock, C., Gray, N., Wagner, T., März, C., Bowen,
 L., Ojeda, J., Finlay, N., Robertson, S., Worrall, F. and Greenwell, C. (2015) Towards a
 mechanistic understanding of carbon stabilization in manganese oxides. *Nature Communications* 6, 7628.
- Jones, J.G. (1983) A note on the isolation and enumeration of bacteria which deposit and reduce
 ferric iron. *Journal of Applied Bacteriology* 54, 305-310.
- Jørgensen, B.B. (1978) Comparison of Methods for the Quantification of Bacterial Sulfate
 Reduction in Coastal Marine-Sediments .2. Calculation from Mathematical-Models.
 Geomicrobiology Journal 1, 29-47.
- Jørgensen, B.B., D'Hondt, S.L. and Miller, D.J. (2003) Leg 201 Synthesis: Controls on micorbial
 communities in deeply buried sediments. *Proc. IODP, Scientific Results* 201, 45.
- Jørgensen, B.B., Findlay, A.J. and Pellerin, A. (2019) The biogeochemical sulfur cycle of marine
 sediments. *Frontiers in Microbiology* 10, Article 849.
- Jørgensen, B.B. and Marshall, I.P.G. (2016) Slow microbial life in the seabed. *Annual Reviews in Marine Science* 8, 311-332.
- Jungbluth, S.P., Amend, J.P. and Rappé, M.S. (2017) Metagenome sequencing and 98 microbial
 genomes from Juan de Fuca Ridge flank subsurface fluids. *Scientific Data* 4, 170037.
- Kallmeyer, J., Pockalny, R., Adhikari, R.R., Smith, D.C. and D'Hondt, S. (2012) Global
 distribution of microbial abundance and biomass in subseafloor sediment. *PNAS* 109, 16213–16216.
- Kandasamy, S. and Nagender Nath, B. (2016) Perspectives on the Terrestrial Organic Matter
 Transport and Burial along the Land-Deep Sea Continuum: Caveats in Our Understanding
 of Biogeochemical Processes and Future Needs. *Frontiers in Marine Science* 3.
- Karl, D.M., Wirsen, C.O. and Jannasch, H.W. (1980) Deep-sea primary production at the
 Galapagos hydrothermal vents. *Science* 207, 1345-1347.
- Kawamura, K., Tannenbaum, E., Huizinga, B.J. and Kaplan, I.R. (1996) Volatile organic-acids
 generated from kerogen during laboratory heating. *Geochemical Journal* 20, 51-59.

- Keil, R.G. and Cowie, G.L. (1999) Organic matter preservation through the oxygen-deficient zone
 of the NE Arabian Sea as discerned by organic carbon:mineral surface area ratios. *Marine Geology* 161, 13-22.
- Keil, R.G., Dickens, A.F., Arnarson, T., Nunn, B.L. and Devol, A.H. (2004) What is the oxygen
 exposure time of laterally transported organic matter along the Washington margin?
 Marine Chemistry 92, 157-165.
- Keil, R.G. and Hedges, J.I. (1993) Sorption of organic matter to mineral surfaces and the
 preservation of organic matter in coastal marine sediments. *Chemical Geology* 107, 385 388.
- Keil, R.G. and Kirchman, D.L. (1994) Abiotic transformation of labile protein to refractory protein
 in sea water. *Marine Chemistry* 45, 187-196.
- Keil, R.G., Montlucon, D.B., Prahl, F.G. and Hedges, J.I. (1994) Sorptive presevation of labile
 organic matter in marine sediments. *Nature* 370, 549-552.
- Keil, R.G., Neibauer, J.A., Biladeau, C., van der Elst, K. and Devol, A.H. (2016) A multiproxy
 approach to understanding the "enhanced" flux of organic matter through the oxygendeficient waters of the Arabian Sea. *Biogeosciences* 13, 2077-2092.
- Keiluweit, M., Bougoure, J.J., Nico, P.S., Pett-Ridge, J., Weber, P.K. and Kleber, M. (2015)
 Mineral protection of soil carbon counteracted by root exudates. *Nature Climate Change*5, 588.
- Kelleher, B.P. and Simpson, A.J. (2006) Humic Substances in Soils: Are They Really Chemically
 Distinct? *Environmental Science & Technology* 40, 4605-4611.
- Kemp, P.F. (1988) Bacterivory by benthic ciliates: significance as a carbon source and impact on
 sediment bacteria. *Marine Ecology Progress Series* 49, 163-169.
- Kempes, C.P., van Bodegom, P.M., Wolpert, D., Libby, E., Amend, J. and Hoehler, T. (2017)
 Drivers of Bacterial Maintenance and Minimal Energy Requirements. *Frontiers in Microbiology* 8.
- Kennedy, M.J., Pevear, D.R. and Hill, R.J. (2002) Mineral surface control of organic carbon in
 black shale. *Science* 295, 657-660.
- Kennedy, M.J. and Wagner, T. (2011) Clay mineral continental amplifier for marine carbon
 sequestion in a greenhouse ocean. *PNAS* 108, 9776-9781.
- Kevorkian, R., Bird, J.T., Shumaker, A. and Lloyd, K.G. (2018) Estimating population turnover
 rates by relative quantification methods reveals microbial dynamics in marine dediment.
 Applied and Environmental Microbiology 84, e01443-01417.
- Kirkpatrick, J.B., Walsh, E.A. and D'Hondt, S. (2019) Microbial Selection and Survival in
 Subseafloor Sediment. *Frontiers in Microbiology* 10.
- Klaas, C. and Archer, D. (2002) Association of sinking organic matter with various types of
 mineral ballast in the deep sea: implications for the rain ratio. *Global Biogeochemical Cycles* 16, GB001765.
- Kleber, M., Eusterhues, K., Keiluweit, M., Mikutta, C., Mikutta, R. and Nico, P.S. (2015) Mineral organic associations: Formation, properties, and relevance in soil environments. *Advances in Agronomy* 130, 1-140.
- Kleber, M. and Johnson, M.G. (2010) Advances in understanding the molecular structure of soil
 organic matter: implications for interactions in the environment. *Adv. Agron.* 106, 77-142.
- Klotzbücher, T., Kaiser, K., Guggenberger, G., Gatzek, C. and Kalbitz, K. (2011) A new conceptual model for the fate of lignin in decomposing plant litter. *Ecology* 92, 1052-1062.

- Koch, B.P., Witt, M., Engbrodt, R., Dittmar, T. and Kattner, G. (2005) Molecular formulae of marine and terrigenous dissolved organic matter detected by electrospray ionization Fourier transform ion cyclotron resonance mass spectrometry. *Geochim. Cosmochim. Acta* 69, 3299-3308.
- Kohfeld, K.E., Quéré, C.L., Harrison, S.P. and Anderson, R.F. (2005) Role of Marine Biology in
 Glacial-Interglacial CO<sub>2</sub> Cycles. *Science* 308, 74.
- Kohnen, M.E.L. and Sinninghe Damsté, J.S. (1989) Early incorporation of polysulfides in
 sedimentary organic matter. *Nature* 341, 640-641.
- Kolonic, S., Damsté, J.S.S., Böttcher, M.E., Kuypers, M.M.M., Kuhnt, W., Beckmann, B. and
 Wagner, G.S.T. (2002) Geochemical characterization of Cenomanian/Turonian black
 shales from the Tarfaya Basin (SW Morocco): Relationships between paleoenvironmental
 conditions and early sulfurization of sedimentary organic matter. *Journal of Petroleum Geology* 25.
- Kolonic, S., Wagner, T., Forster, A., Sinninghe Damsté, J.S., Walsworth-Bell, B., Erba, E.,
 Turgeon, S., Brumsack, H.-J., Chellai, E.H., Tsikos, H., Kuhnt, W. and Kuypers, M.M.M.
 (2005) Black shale deposition on the northwest African Shelf during the
 Cenomanian/Turonian oceanic anoxic event: Climate coupling and global organic carbon
 burial. *Paleoceanography* 20.
- Krein, E.B. and Aizenshtat, Z. (1994) The formation of isoprenoid sulfur compounds during
 diagenesis: simulated sulfur incorporation and thermal transformation. *Organic Geochemistry* 21, 1015-1025.
- Kristensen, E. (1985) Oxygen and Inorganic Nitrogen Exchange in a "Nereis virens" (Polychaeta)
 Bioturbated Sediment-Water System. *Journal of Coastal Research* 1, 109-116.
- Kristensen, E. (2001) Impact of polychaetes (Nereis spp. and Arenicola marina) on carbon
 biogeochemistry in coastal marine sediments[†]. *Geochemical Transactions* 2, 92.
- Kristensen, E., Andersen, F.Ø. and Blackburn, T.H. (1992) Effects of benthic macrofauna and
 temperature on degradation of macroalgal detritus: The fate of organic carbon. *Limnology and Oceanography* 37, 1404-1419.
- Kristensen, E. and Holmer, M. (2001a) Decomposition of plant materials in marine sediment
 exposed to different electron acceptors (O2, NO3-, and SO42-), with emphasis on
 substrate origin, degradation kinetics, and the role of bioturbation. *Geochimica et Cosmochimica Acta* 65, 419-433.
- 2001Kristensen, E. and Holmer, M. (2001b) Decomposition of plant materials in marine sediment2002exposed to different electron acceptors $(O_2, NO_3^-, and SO_4^{2-})$, with emphasis on substrate2003origin, degradation kinetics, and the role of bioturbation *Geochimica et Cosmochimica*2004Acta 65, 419-433.
- Kristensen, E. and Kostka, J.E. (2005) Macrofaunal burrows and irrigation in marine sediment:
 microbiological and biogeochemical interactions. *AGU Coast. Estuar. Studies* 60, 125-157.
- Kristensen, E., Mangion, P., Tang, M., Flindt, M.R., Holmer, M. and Ulomi, S. (2011) Microbial
 carbon oxidation rates and pathways in sediments of two Tanzanian mangrove forests.
 Biogeochemistry 103, 143-158.
- Krumholz, L., Harris, R. and Suflita, J. (2002) Anaerobic microbial growth from components of
 Cretaceous shales. *Geomicrobiol. J.* 19, 593-602.
- Krumholz, L., McKinley, J., Ulrich, G. and Suflita, J. (1997) Confined subsurface micorbial
 communities in Cretaceous rock. *Nature* 386, 64-66.

- Krumins, V., Gehlen, M., Arndt, S.V.C., P. and Regnier, P. (2013) Dissolved inorganic carbon
 and alkalinity fluxes from coastal marine sediments: model estimates for different shelf
 environments and sensitivity to global change. *Biogeosciences* 10.
- Kuhn, T., Versteegh, G.J.M., Villinger, H., Dohrmann, I., Heller, C., Koschinsky, A., Kaul, N.,
 Ritter, S., Wegorzewski, A.V. and Kasten, S. (2017) Widespread seawater circulation in
 18–22 Ma oceanic crust: Impact on heat flow and sediment geochemistry. *Geology* 45,
 799-802.
- Kusch, S., Rethemeyer, J., Schefuß, E. and Mollenhauer, G. (2010) Controls on the age of vascular
 plant biomarkers in Black Sea sediments. *Geochimica et Cosmochimica Acta* 74, 7031 7047.
- Kuzyakov, Y. (2010) Priming effects: Interactions between living and dead organic matter. *Soil Biology and Biochemistry* 42, 1363-1371.
- Lalonde, K., Mucci, A., Ouellet, A. and Gelinas, Y. (2012) Preservation of organic matter in sediments by iron. *Nature* **483**, 198-200.
- Lam, P.J., Doney, S.C. and Bishop, J.K.B. (2011) The dynamic ocean biological pump: Insights from a global compilation of particulate organic carbon, CaCO3, and opal concentration profiles from the mesopelagic. *Global Biogeochemical Cycles* **25**, GB3009.
- Lang, S.Q., Butterfield, D.A., Lilley, D.A., Johnson, H.P. and Hedges, J.I. (2006) Dissolved
 organic carbon in ridge-axis and ridge-flank hydrothermal systems. *Geochim. Cosmochim.* Acta 70, 3830-3842.
- Lang, S.Q., Butterfield, D.A., Schulte, M., Kelley, D.S. and Lilley, M.D. (2010) Elevated
 concentrations of formate, acetate and dissolved organic carbon found at the Lost City
 hydrothermal field. *Geochim. Cosmochim. Acta* 74, 941-952.
- LaRowe, D.E. and Amend, J.P. (2014) Energetic constraints on life in marine deep sediments, in:
 Kallmeyer, J., Wagner, K. (Eds.), *Life in Extreme Environments: Microbial Life in the Deep Biosphere*. de Gruyter, Berlin, pp. 279-302.
- 2040LaRowe, D.E. and Amend, J.P. (2015a)Catabolic rates, population sizes and2041doubling/replacement times of microorganisms in the natural settings. Am. J. Sci. 315, 167-2042203.
- LaRowe, D.E. and Amend, J.P. (2015b) Power limits for microbial life. *Front. Extr. Microbiol.* 6,
 Article 718 doi: 710.3389/fmicb.2015.00718
- LaRowe, D.E. and Amend, J.P. (2019) The energetics of fermentation in natural settings.
 Geomicrobiology Journal 36, 492-505.
- LaRowe, D.E., Arndt, S., Bradley, J.A., Burwicz, E.B., Dale, A.W. and Amend, J.P. (2020)
 Organic carbon and microbial activity in marine sediments on a global scale throughout
 the Quaternary. *EarthARXiv* https://doi.org/10.31223/osf.io/fj5tz.
- LaRowe, D.E., Burwicz, E.B., Arndt, S., Dale, A.W. and Amend, J.P. (2017a) The temperature and volume of global marine sediments. *Geology* **45**, 275-278.
- LaRowe, D.E., Dale, A.W., Amend, J.P. and Van Cappellen, P. (2012) Thermodynamic limitations on microbially catalyzed reaction rates. *Geochim. Cosmochim. Acta* **90**, 96-109.
- LaRowe, D.E., Koch, B.P., Robador, A., Witt, M., Ksionzek, K. and Amend, J.P. (2017b)
 Identification of organic compounds in ocean basement fluids. *Organic Geochemistry* 113, 124-127.
- LaRowe, D.E. and Van Cappellen, P. (2011) Degradation of natural organic matter: A thermodynamic analysis. *Geochim. Cosmochim. Acta* **75**, 2030-2042.

- Lee, C. (1992a) Controls on organic carbon preservation: the use of stratified water bodies to
 compare intrinsic rates of decomposition in oxic and anoxic systems. *Geochimica et Cosmochimica Acta* 56, 3323-3335.
- Lee, C. (1992b) Controls on organic carbon preservation: The use of stratified water bodies to
 compare intrinsic rates of decomposition in oxic and anoxic systems. *Geochimica et Cosmochimica Acta* 56, 3323-3335.
- Lehmann, J. and Kleber, M. (2015) The contentious nature of soil organic matter. *Nature* **528**, 60-68.
- Lengger, S.K., Rush, D., Mayser, J.P., Blewett, J., Schwartz-Narbonne, R., Talbot, H.M.,
 Middelburg, J.J., Jetten, M.S.M., Schouten, S., Sinninghe Damsté, J.S. and Pancost, R.D.
 (2019) Dark carbon fixation in the Arabian Sea oxygen minimum zone contributes to
 sedimentary organic carbon (SOM). *Global Biogeochemical Cycles* 33, 1715-1732.
- Lenton, T.M. and Watson, A.J. (2000) Redfield revisited 1. Regulation of nitrate, phosphate, and
 oxygen in the ocaen. *Global Biogeochem. Cycles* 14, 225-248.
- Lerman, A. (1971) Time to Chemical Steady-States in Lakes and Ocean, *Nonequilibrium Systems in Natural Water Chemistry*. AMERICAN CHEMICAL SOCIETY, pp. 30-76.
- Lessin, G., Artioli, Y., Almroth-Rosell, E., Blackford, J.C., Dale, A.W., Glud, R.N., Middelburg,
 J.J., Pastres, R., Queirós, A.M., Rabouille, C., Regnier, P., Soetaert, K., Solidoro, C.,
 Stephens, N. and Yakushev, E. (2018) Modelling Marine Sediment Biogeochemistry:
 Current Knowledge Gaps, Challenges, and Some Methodological Advice for
 Advancement. *Frontiers in Marine Science* 5.
- Lever, M.A. (2013) Functional gene surveys from ocean drilling expeditions a review and perspective. *FEMS Microbiology Ecology* **84**, 1-23.
- Levin, L.A. and Gooday, A.J. (2003) The deep Atlantic Ocean, in: Tyler, P.A. (Ed.), *Ecosystems* of the deep oceans. Elsevier, Amsterdam, pp. 111-178.
- Levin, L.A. and Le Bris, N. (2015) The deep ocean under climate change. *Science* **350**, 766-768.
- Lin, H.-T., Amend, J.P., LaRowe, D.E., Bingham, J.-P. and Cowen, J.P. (2015) Dissolved amino
 acids in oceanic basaltic basement fluids. *Geochim. Cosmochim. Acta* 164, 175-190.
- Lin, H.-T., Cowen, J.P., Olson, E.J., Amend, J.P. and Lilley, M.D. (2012) Inorganic chemistry,
 gas compositions and dissolved organic carbon in fluids from sedimented young basaltic
 crust on the Juan de Fuca Ridge flanks. *Geochim. Cosmochim. Acta* 85, 213-227.
- Lin, H.-T., Repeta, D.J., Xu, L. and Rappé, M.S. (2019) Dissolved organic carbon in basalt-hosted
 deep subseafloor fluids of the Juan de Fuca Ridge flank. *Earth and Planetary Science Letters* 513, 156-165.
- Lin, Y.-S., Koch, B.P., Feseker, T., Ziervogel, K., Goldhammer, T., Schmidt, F., Witt, M.,
 Kellermann, M.Y., Zabel, M., Teske, A. and Hinrichs, K.-U. (2017) Near-surface Heating
 of Young Rift Sediment Causes Mass Production and Discharge of Reactive Dissolved
 Organic Matter. *Scientific Reports* 7, 44864.
- Lloyd, K.G., Schreiber, L., Petersen, D.G., Kjeldsen, K.U., Lever, M.A., Steen, A.D.,
 Stepanauskas, R., Richter, M., Kleindienst, S., Lenk, S., Schramm, A. and Jørgensen, B.B.
 (2013) Predominant archaea in marine sediments degrade detrital proteins. *Nature* 496,
 215.
- Lloyd, K.G., Steen, A.D., Ladau, J., Yin, J. and Crosby, L. (2018) Phylogenetically Novel
 Uncultured Microbial Cells Dominate Earth Microbiomes. *mSystems* 3, e00055-00018.

- Lombard, V., Golaconda Ramulu, H., Drula, E., Coutinho, P.M. and Henrissat, B. (2013) The
 carbohydrate-active enzymes database (CAZy) in 2013. *Nucleic Acids Research* 42, D490 D495.
- Lomstein, B.A., Langerhuus, A.T., D'Hondt, S., Jørgensen, B.B. and Spivack, A.J. (2012)
 Endospore abundance, micorbial growth and necromass turnover in deep sub-seafloor
 sediment. *Nature* 484, 101-104.
- Longman, J., Palmer, M.R., Gernon, T.M. and Manners, H.R. (2019) The role of tephra in
 enhancing organic carbon preservation in marine sediments. *Earth-Science Reviews* 192, 480-490.
- Lovley, D.R. (1991) Dissimilatory Fe(III) and Mn(IV) reduction. *Microbiological Reviews* 55, 2113 259-287.
- Luff, R. and Moll, A. (2004) Seasonal dynamics of the North Sea sediments using a three dimensional coupled sediment–water model system. *Continental Shelf Research* 24, 1099 1127.
- Lundegard, P.D. and Kharaka, Y.K. (1994) Distribution and Occurrence of Organic Acids in
 Subsurface Waters. Springer, Berlin.
- Lutz, M., Dunbar, R. and Caldeira, K. (2002) Regional variability in the vertical flux of particulate
 organic carbon in the ocean interior. *Global Biogeochemical Cycles* 16, GB001383.
- Mackenzie, F.T., Lerman, A. and Andersson, A.J. (2004) Past and present of sediment and carbon
 biogeochemical cycling models *Biogeosciences* 1, 11-32.
- Mahmoudi, N., Enke, T.N., Beaupré, S.R., Teske, A.P., Cordero, O.X. and Pearson, A. (2019)
 Illuminating microbial species-specific effects on organic matter remineralization in marine sediments. *bioRxiv*, 705087.
- Mahmoudi, N., Hagen, S.M., Hazen, T.C. and Steen, A.D. (2020) Patterns in extracellular enzyme
 activity and microbial diversity in deep-sea Mediterranean sediments. *Deep Sea Research Part I: Oceanographic Research Papers*, 103231.
- Mann, P.J., Eglinton, T.I., McIntyre, C.P., Zimov, N., Davydova, A., Vonk, J.E., Holmes, R.M.
 and Spencer, R.G.M. (2015) Utilization of ancient permafrost carbon in headwaters of
 Arctic fluvial networks. *Nature Communications* 6, 7856.
- Mann, S., Archibald, D.D., Didymus, J.M., Douglas, T., Heywood, B.R., Meldrum, F.C. and
 Reeves, N.J. (1993) Crystallization at inorganic-organic interfaces: Biominerals and
 biomometic synthesis. *Science* 261, 1286-1292.
- Mao, J., Tremblay, L. and Gagné, J.-P. (2011) Structural changes of humic acids from sinking
 organic matter and surface sediments investigated by advanced solid-state NMR: Insights
 into sources, preservation and molecularly uncharacterized components. *Geochimica et Cosmochimica Acta* 75, 7864-7880.
- Marschner, B., Brodowski, S., Dreves, A., Gleixner, G., Gude, A., Grootes, P.M., Hamer, U.,
 Heim, A., Jandl, G., Ji, R., Kaiser, K., Kalbitz, K., Kramer, C., Leinweber, P., Rethemeyer,
 J., Schäffer, A., Schmidt, M.W.I., Schwark, L. and Wiesenberg, G.L.B. (2008a) How
 relevant is recalcitrance for the stabilization of organic matter in soils? *J. Plant Nutr. Soil Sci.* 171, 91-110.
- Marschner, B., Brodowski, S., Dreves, A., Gleixner, G., Gude, A., Grootes, P.M., Hamer, U.,
 Heim, A., Jandl, G., Ji, R., Kaiser, K., Kalbitz, K., Kramer, C., Leinweber, P., Rethmeyer,
 J., Schäffer, A., Schmidt, M.W.I., Schwark, L. and WIesenberg, G.L.B. (2008b) How
 relevant is recalcitrance for the stabilization of organic matter in soils? *J. Plant Nutr. Soil Sci.* 171, 91-110.

- Marshall, I.P.G., Ren, G., Jaussi, M., Lomstein, B.A., Jørgensen, B.B., Røy, H. and Kjeldsen, K.U.
 (2019) Environmental filtering determines family-level structure of sulfate-reducing
 microbial communities in subsurface marine sediments. *The ISME Journal* 13, 1920-1932.
- 2152 Martens, C.S. (1990) Generation of short chain organic acid anions in hydrothermally altered 2153 sediments of the Guaymas Basin, Gulf of California. *Applied Geochemistry* **5**, 71-76.
- Mayer, L.M. (1994) Surface area control of organic carbon accumulation in continental shelf
 sediments. *Geochim. Cosmochim. Acta* 58, 1271-1284.
- 2156 Mayer, L.M. (1995) Sedimentary organic matter preservation: an assessment and speculative 2157 synthesis—a comment. *Marine Chemistry* **49**, 123-126.
- Mayer, L.M. (1999) Extent of coverage of mineral surfaces by organic mattre in marine sediments.
 Geochimica et Cosmochimica Acta 63, 207-215.
- Mayer, L.M., Rahaim, P., Guerin, P., Macko, S.A., Watling, L. and Anderson, F.E. (1985)
 Biological and granulometric controls on organic matter of an intertidal mudflat. *Estuarine*,
 Coastal and Shelf Science 20, 491-503.
- Mayer, L.M. and Xing, B. (2001) Organic carbon-surface area-clay relationships in acid soils. *Soil Science Society of America Journal* 65, 250-258.
- Mayor, D.J., Thornton, B., Hay, S., Zuur, A.F., Nicol, G.W., McWilliam, J.M. and Witte, U.F.M.
 (2012) Resource quality affects carbon cycling in deep-sea sediments. *The ISME Journal*6, 1740-1748.
- McCarthy, M.D., Beaupré, S.R., Walker, B.D., Voparil, I., Guilderson, T.P. and Druffel, E.R.M.
 (2011) Chemosynthetic origin of ¹⁴C-depleted dissolved organic matter in a ridge-flank
 hydrothermal system. *Nat. Geosci.* 4, 32-36.
- McCollom, T.M. and Seewald, J.S. (2003a) Experimental constraints on the hydrothermal
 reactivity or organic acids and acid anions: I. Acetic acid, acetate, and valeric acid.
 Geochim. Cosmochim. Acta 67, 3645-3664.
- McCollom, T.M. and Seewald, J.S. (2003b) Experimental constraints on the hydrothermal
 reactivity or organic acids and acid anions: I. Formic acid and formate. *Geochim. Cosmochim. Acta* 67, 3625-3644.
- McCollom, T.M. and Shock, E.L. (1997) Geochemical constraints on chemolithoautotrophic
 metabolism by microorganisms in seafloor hydrothermal systems. *Geochim. Cosmochim.* Acta 61, 4375 4391.
- McDermott, J.M., Seewald, J.S., German, C.R. and Sylva, S.P. (2015) Pathways for abiotic organic
 synthesis at submarine hydrothermal fields. *Proceedings of the National Academy of Sciences* 112, 7668-7672.
- McInerney, M.J., Struchtemeyer, C.G., Sieber, J., Mouttaki, H., Stams, A.J.M., Schink, B., Rohlin,
 L. and Gunsalus, R.P. (2008) Physiology, ecology, phylogeny, and genomics of
 microorganisms capable of syntrophic metabolism. *Annals of New York Academy of Science* 1125, 58-72.
- McManus, J., Wheat, C.G. and Bach, W. (2019) Carbon cycling in low-temperature hydrothermal
 systems: The Dorado Outcrop. *Geochimica et Cosmochimica Acta* 264, 1-12.
- Meador, T.B., Bowles, M., Lazar, C.S., Zhu, C., Teske, A. and Hinrichs, K.-U. (2015) The archaeal
 lipidome in estuarine sediment dominated by members of the Miscellaneous
 Crenarchaeotal Group. *Environmental Microbiology* 17, 2441-2458.
- Mewes, K., Mogollón, J.M., Picard, A., Rühlemann, C., Eisenhauer, A., Kuhn, T., Ziebis, W. and
 Kasten, S. (2016) Diffusive transfer of oxygen from seamount basaltic crust into overlying

- sediments: An example from the Clarion–Clipperton Fracture Zone. *Earth and Planetary Science Letters* 433, 215-225.
- Meyer, J.L., Jaekel, U., Tully, B.J., Glazer, B.T., Wheat, C.G., Lin, H.-T., Hsieh, C.-C., Cowen,
 J.P., Hulme, S.M., Girguis, P.R. and Huber, J.A. (2016) A distinct and active bacterial
 community in cold oxygenated fludis circulating beneath the western flank of the MidAtlantic Ridge. *Scientific Reports* 6, doi: 10.1038/srep22541.
- Meysman, F.J.R., Middelburg, J.J. and Heip, C.H.R. (2006) Bioturbation: a fresh look at Darwin's
 last idea. *Trends in Ecology & Evolution* 21, 688-695.
- Michalska, K., Steen, A.D., Chhor, G., Endres, M., Webber, A.T., Bird, J., Lloyd, K.G. and
 Joachimiak, A. (2015) New aminopeptidase from "microbial dark matter" archaeon. *The FASEB Journal* 29, 4071-4079.
- 2205 Middelburg, J.J. (1989) A simple rate model for organic matter decomposition in marine 2206 sediments. *Geochem. Cosmochim. Acta* **53**, 1577-1581.
- 2207 Middelburg, J.J. (2011) Chemoautotrophy in the ocean. *Geophysical Research Letters* **38**.
- Middelburg, J.J. (2018) Reviews and syntheses: to the bottom of carbon processing at the seafloor.
 Biogeosciences 15, 413-427.
- Middelburg, J.J. (2019) Marine Carbon Biogeochemistry: A Primer for Earth System Scientists,
 in: Lohmann, G., Mysak, L.A., Notholt, J., Rabassa, J., Unnithan, V. (Eds.). SpringerOpen,
 Cham, Switzerland, p. 118.
- Middelburg, J.J. and Levin, L.A. (2009) Coastal hypoxia and sediment biogeochemistry.
 Biogeosciences 6, 1273-1293.
- 2215 Middelburg, J.J. and Meysman, F.J.R. (2007) Burial at sea. *Science* **316**, 1294-1295.
- Middelburg, J.J., Vlug, T., Jaco, F. and van der Nat, W.A. (1993) Organic matter mineralization
 in marine systems. *Global and Planetary Change* 8, 47-58.
- Milkov, A.V. (2011) Worldwide distribution and significane of secondary microbial methane
 formed during petroleum biodegradation in conventional reservoirs. *Org. Geochem.* 42, 184-207.
- Ming-Yi, S., Lee, C. and Aller, R.C. (1993) Laboratory studies of oxic and anoxic degradation of
 chlorophyll-a in Long Island Sound sediments. *Geochimica et Cosmochimica Acta* 57,
 147-157.
- Mollenhauer, G., Eglinton, T.I., Ohkouchi, N., Schneider, R.R., Müller, P.J., Grootes, P.M. and
 Rullkötter, J. (2003) Asynchronous alkenone and foraminifera records from the Benguela
 Upwelling System. *Geochimica et Cosmochimica Acta* 67, 2157-2171.
- Mollenhauer, G., Inthorn, M., Vogt, T., Zabel, M., Sinninghe Damsté, J.S. and Eglinton, T.I.
 (2007) Aging of marine organic matter during cross-shelf lateral transport in the Benguela
 upwelling system revealed by compound-specific radiocarbon dating. *Geochemistry*,
 Geophysics, Geosystems 8.
- Moodley, L., Middelburg, J.J., Herman, P.M.J., Soetaert, K. and de Lange, G.J. (2005)
 Oxygenation and organic-matter preservation in marine sediments: Direct experimental
 evidence from ancient organic carbon-rich deposits *Geology* 33, 889-892.
- Morán, X.A.G., López-Urrutia, Á., Calvo-Díaz, A. and Li, W.K.W. (2010) Increasing importance
 of small phytoplankton in a warmer ocean. *Global Change Biology* 16, 1137-1144.
- Moreau, J.W., Weber, P.K., Martin, M.C., Gilbert, B., Hutcheon, I.D. and Banfield, J.F. (2007)
 Extracellular proteins limit the dispersal of biogenic nanoparticles. *Science* 316, 1600 1603.

- Morono, Y., Terada, T., Nishizawa, M., Ito, M., Hillion, F., Takahata, N., Sano, Y. and Inagaki,
 F. (2011) Carbon and nitrogen assimilation in deep subseafloor micorbial cells. *PNAS* 108,
 18295-18300.
- Mossmann, J.-R., Aplin, A.C., Curtis, C.D. and Coleman, M.L. (1991) Geochemistry of inorganic
 and organic sulphur in organic-rich sediments from the Peru Margin. *Geochimica et Cosmochimica Acta* 55, 3581-3595.
- Müller, P.J. and Suess, E. (1979) Productivity, sedimentation rate, and sedimentary organic matter in the oceans—I. Organic carbon preservation. *Deep-Sea Research Part A* **26**, 1347-1362.
- Munhoven, G. (2007) Glacial-interglacial rain ratio changes: Implications for atmospheric CO2
 and ocean-sediment interaction. *Deep Sea Research Part II: Topical Studies in Oceanography* 54, 722-746.
- Nealson, K.H. (1997) Sediment bacteria: Who's there, what are they doing, and what's new?
 Annual Review of Earth and Planetary Sciences 25, 403-434.
- Niggemann, J., Ferdelman, T.G., Lomstein, B.A., Kallmeyer, J. and Schubert, C.J. (2007) How
 depositional conditions control input, composition, and degradation of organic matter in
 sediments from the Chilean coastal upwelling region. *Geochimica et Cosmochimica Acta* **71**, 1513-1527.
- Nobu, M.K., Dodsworth, J.A., Murugapiran, S.K., Rinke, C., Gies, E.A., Webster, G., Schwientek,
 P., Kille, P., Parkes, R.J., Sass, H., Jørgensen, B.B., Weightman, A.J., Liu, W.-T., Hallam,
 S.J., Tsiamis, G., Woyke, T. and Hedlund, B.P. (2016) Phylogeny and physiology of
 candidate phylum 'Atribacteria' (OP9/JS1) inferred from cultivation-independent
 genomics. *The ISME Journal* 10, 273-286.
- Ohkouchi, N., Eglinton, T.I., Keigwin, L.D. and Hayes, J.M. (2002) Spatial and Temporal Offsets
 Between Proxy Records in a Sediment Drift. *Science* 298, 1224.
- Ohkouchi, N., Kuroda, J. and Taira, A. (2015) The origin of Cretaceous black shales: a change in
 the surface ocean ecosystem and its triggers. *Proceedings of the Japan Academy, Series B* 91, 273-291.
- Orcutt, B.N., Bradley, J.A., Brazelton, W.J., Estes, E.R., Goordial, J.M., Huber, J.A., Jones, R.M.,
 Mahmoudi, N., Marlow, J.J., Murdock, S. and Pachiadaki, M. (2020) Impacts of deep-sea
 mining on microbial ecosystem services. *Limnology and Oceanography* n/a.
- 2269 Orcutt, B.N., LaRowe, D.E., Biddle, J.F., Cowell, F., Kirkpatrick, J.B., Lapham, L., Mills, H., 2270 Reese, B., Sylvan, J. and Wankel, S.D. (2013a) Microbial activity in the deep marine Microbiol. 2271 biosphere: Progress and prospects. Front. Extr. 4:189. doi: 2272 10.3389/fmicb.2013.00189.
- Orcutt, B.N., Wheat, C.G., Rouxel, O., Hulme, S., Edwards, K.J. and Bach, W. (2013b) Oxygen
 consumption rates in a subseafloor basaltic crust derived from a reaction transport model.
 Nat. Comm. 4, Article 2539.
- Oremland, R.S., Culbertson, C. and Simoneit, B.R.T. (1982) Methanogenic activity in sediment
 from Leg 64, Gulf of California. *Initial reports of the DSDP* 64, 759-762.
- Orsi, W. (2018) Ecology and evolution of seafloor and subseafloor microbial communities. *Nature Reveiws Microbiology*.
- Orsi, W., Biddle, J.F. and Edgcomb, V. (2013) Deep Sequencing of Subseafloor Eukaryotic rRNA
 Reveals Active Fungi across Marine Subsurface Provinces. *PLOS ONE* 8, e56335.
- Orsi, W.D., Coolen, M.J.L., Wuchter, C., He, L., More, K.D., Irigoien, X., Chust, G., Johnson, C.,
 Hemingway, J.D., Lee, M., Galy, V. and Giosan, L. (2017) Climate oscillations reflected
 within the microbiome of Arabian Sea sediments. *Scientific Reports* 7, 6040.

- Ortega-Arbulú, A.-S., Pichler, M., Vuillemin, A. and Orsi, W.D. (2019) Effects of organic matter
 and low oxygen on the mycobenthos in a coastal lagoon. *Environmental Microbiology* 21, 374-388.
- Pachiadaki, M.G., Sintes, E., Bergauer, K., Brown, J.M., Record, N.R., Swan, B.K., Mathyer,
 M.E., Hallam, S.J., Lopez-Garcia, P., Takaki, Y., Nunoura, T., Woyke, T., Herndl, G.J.
 and Stepanauskas, R. (2017) Major role of nitrite-oxidizing bacteria in dark ocean carbon
 fixation. *Science* 358, 1046-1051.
- Pacton, M., Fiet, N. and Gorin, G. (2007a) Bacterial activity and sedimentary organic matter: the
 role of exopolymeric substances. *Geomicrobiology Journal* 24, 571-581.
- Pacton, M., Fiet, N., Gorin, G. and Spangenberg, J.E. (2007b) Lower Cretaceous oceanic anoxic
 event OAE1b: organic matter accumulation mediated by bacterial activity. *Geophysical Research* 9, 09956.
- Palastanga, V., Slomp, C.P. and Heinze, C. (2011) Long-term controls on ocean phosphorus and
 oxygen in a global biogeochemical model. *Global Biogeochemical Cycles* 25.
- Parkes, R.J., Cragg, B., Roussel, E., Webster, G., Weightman, A. and Sass, H. (2014) A review of
 prokaryotic populations and processes in sub-seafloor sediments, including
 biosphere:geosphere interactions. *Mar. Geol.* 352, 409-425.
- Parkes, R.J., Cragg, B.A., Fry, J.C., Herbert, R.A., Wimpenny, J.W.T., Allen, J.A. and Whitfield,
 M. (1990) Bacterial biomass and activity in deep sediment layers from the Peru Margin. *Phil. Trans. R. Soc. Lond. A* 331, 139-153.
- Parkes, R.J., Webster, G., Cragg, B.A., Weightman, A.J., Newberry, C.J., Ferdelman, T.G.,
 Kallmeyer, J. and Jørgensen, B.B. (2005) Deep sub-seafloor prokaryotes stimulate at
 interfaces over geological time. *Nature* 436, 390-394.
- Passow, U. and Carlson, C.A. (2012) The biological pump in a high CO₂ world. *Marine Ecology Progress Series* 470, 249-271.
- Pearson, A., Seewald, J.S. and Eglinton, T.I. (2005) Bacterial incorporation of relict carbon in the
 hydrothermal environment of Guaymas Basin. *Geochimica et Cosmochimica Acta* 69, 5477-5486.
- Pedersen, T.F. and Calvert, S.E. (1990) Anoxia vs. productivity: What controls the formation of
 organic-carbon-rich sediments and sedimentary rocks? *The American Association of Petroleum Geologists Bulletin* 74, 454-466.
- Petro, C., Starnawski, P., Schramm, A. and Kjeldsen, K.U. (2017) Microbial community assembly
 in marine sediments. *Aquatic Microbial Ecology* **79**, 177-195.
- Petsch, S.T., Eglinton, T.I. and Edwards, K.J. (2001) ¹⁴C Dead Living Biomass: Evidence for Microbial Assimilation of Ancient Organic Carbon During Shale Weathering. *Science* 292, 1127-1131.
- Plank, T. and Langmuir, C.H. (1998) The chemical composition of subducting sediment and its
 consequences for the crust and mantle. *Chemical Geology* 145, 325-394.
- Pörtner, H.-O., Karl, D.M., Boyd, P.W., Cheung, W., Lluch-Cota, S.E., Nojiri, Y., Schmidt, D.N.
 and Zavialov, P.O. (2014) Ocean systems, in: Field, C.B., Barros, V.R., Dokken, D.J.,
 Mach, K.J., Mastrandrea, M.D., Bilir, T.E., Chatterjee, M., Ebi, K.L., Estrada, Y.O.,
 Genova, R.C., Girma, B., Kissel, E.S., Levy, A.N., MacCracken, S., Mastrandrea, P.R.,
 White, L.L. (Eds.), *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A:*Global and Sectoral Aspects. Contribution of Working Group II to the Fifth Assessment
 Report of the Intergovernmental Panel on Climate Change. Cambridge University Press,
- 2330 New York, pp. 411-484.

- Prahl, F.G., Cowie, G.L., De Lange, G.J. and Sparrow, M.A. (2003) Selective organic matter
 preservation in "burn-down" turbidites on the Madeira Abyssal Plain. *Paleoceanography* 18.
- Prahl, F.G., de Lange, G.J., Lyle, M. and Sparrow, M.A. (1989) Post-depositional stability of long chain alkenones under contrasting redox conditions. *Nature* 341, 434-437.
- Prahl, F.G., De Lange, G.J., Scholten, S. and Cowie, G.L. (1997) A case of post-depositional
 aerobic degradation of terrestrial organic matter in turbidite deposits from the Madeira
 Abyssal Plain. *Organic Geochemistry* 27, 141-152.
- Prothero, D.R. and Schwab, F. (2004) Sedimentary Geology: An Introduction to Sedimentary
 Rocks and Straigraphy, 2nd Ed. W.H. Freeman and Company, New York.
- Puglini, M., Brovkin, V., Regnier, P. and Arndt, S. (2019) Assessing the potential for non-turbulent
 methane escape from the East Siberian Arctic Shelf. *Biogeosciences Discuss.* 2019, 1-44.
- Quijada, M., Riboulleau, A., Faure, P., Michels, R. and Tribovillard, N. (2016) Organic matter
 sulfurization on protracted diagenetic timescales: The possible role OF anaerobic oxidation
 of methane. *Marine Geology* 381, 54-66.
- Ransom, B., Bennett, R.H. and Baerwald, R. (1997) TEM study of in situ organic matter on
 continental margins: occurrence and the 'monolayer' hypothesis. *Marine Geology* 138, 19.
- Ransom, B. and Helgeson, H.C. (1995) A chemical and thermodynamic model of dioctahedral 2:1
 layer clay minerals in diagenetic processes: Dehydration of dioctahedral aluminous
 smectite as a function of temperature and depth in sedimentary basins. *Am. J. Sci.* 295, 245281.
- Ransom, B., Kim, D., Kastner, M. and Wainwright, S. (1998a) Organic matter preservation on
 continental slopes: Importance of mineralogy and surface area. *Geochimica et Cosmochimica Acta* 62, 1329-1345.
- Ransom, B., Shea, K.F., Burkett, P.J., Bennett, R.H. and Baerwald, R. (1998b) Comparison of
 pelagic and nepheloid layer marine snow: implication for carbon cycling. *Marine Geology* 150, 39-50.
- Raven, M.R., Fike, D.A., Bradley, A.S., Gomes, M.L., Owens, J.D. and Webb, S.A. (2019) Paired
 organic matter and pyrite δ34S records reveal mechanisms of carbon, sulfur, and iron cycle
 disruption during Ocean Anoxic Event 2. *Earth and Planetary Science Letters* 512, 27-38.
- Raven, M.R., Fike, D.A., Gomes, M.L., Webb, S.M., Bradley, A.S. and McClelland, H.-L.O.
 (2018) Organic carbon burial during OAE2 driven by changes in the locus of organic
 matter sulfurization. *Nature Communications* 9, ART. 3409.
- Raven, M.R., Sessions, A.L., Adkins, J.F. and Thunell, R.C. (2016) Rapid organic matter
 sulfurization in sinking particles from the Cariaco Basin water column. *Geochimica et Cosmochimica Acta* 190, 175-190.
- Reardon, P.N., Walter, E.D., Marean-Reardon, C.L., Lawrence, C.W., Kleber, M. and Washton,
 N.M. (2018) Carbohydrates protect protein against abiotic fragmentation by soil minerals.
 Scientific Reports 8, 813.
- Reed, D.C., Algar, C.K., Huber, J.A. and Dick, G.J. (2014) Gene-centric approach to integrating
 environmental genomics and biogeochemical models. *Proceedings of the National Academy of Sciences* 111, 1879-1884.
- Regnier, P., Dale, A.W., Arndt, S., LaRowe, D.E., Mogollón, J.M. and Van Cappellen, P. (2011)
 Quantitative analysis of anaeorbic oxidation of methane (AOM) in marine sediments: A
 modeling perspective. *Earth-Science Reviews* 106, 105-130.

- Regnier, P., Friedlingstein, P., Ciais, P., Mackenzie, F.T., Gruber, N., Janssens, I.A., Laruelle,
 G.G., Lauerwald, R., Luyssaert, S., Andersson, A.J., Arndt, S., Arnosti, C., Borges, A.V.,
 Dale, A.W., Gallego-Sala, A., Godderis, Y., Goossens, N., Hartmann, J., Heinze, C., Ilyina,
 T., Joos, F., LaRowe, D.E., Leifeld, J., Meysman, F.J.R., Munhoven, G., Raymond, P.A.,
 Spahni, R., Suntharalingam, P. and Thullner, M. (2013) Anthropogenic perturbation of
 carbon fluxes from land to ocean. *Nature Geosci* 6, 597-607.
- Reimers, C.E., Jahnke, R.A. and McCorkle, D.C. (1992) Carbon fluxes and burial rates over the
 continental slope and rise off central California with implications for the global carbon
 cycle. *Global Biogeochemical Cycles* 6, 199-224.
- Repeta, D.J., Quan, T.M., Aluwihare, L.I. and Accardi, A. (2002) Chemical characterization of
 high molecular weight dissolved organic matter in fresh and marine waters. *Geochimica et Cosmochimica Acta* 66, 955-962.
- Rhoads, D.C. (1974a) Organism-sediment relations on the muddy sea floor. *Oceanogr. Mar. Biol. Ann. Rev.* 12, 263-300.
- Rhoads, D.C. (1974b) Organism-sediment relations on the muddy sea floor. *Oceanography and Marine Biology Annual Reviews* 12, 263-300.
- Rhoads, D.C., Boyer, L.F., Welsh, B.L. and Hampson, G.R. (1984) Seasonal Dynamics of Detritus
 in the Benthic Turbidity Zone (BTZ); Implications for Bottom-Rack Molluscan
 Mariculture. *Bulletin of Marine Science* 35, 536-549.
- Rice, D.L. (1986) Early diagenesis in bioadvective sediments: Relationships between the
 diagenesis of beryllium-7, sediment reworking rates, and the abundance of conveyor-belt
 deposit-feeders. *Journal of Marine Research* 44, 149-184.
- Richards, T.A., Jones, M.D.M., Leonard, G. and Bass, D. (2012) Marine Fungi: Their Ecology and
 Molecular Diversity. *Annual Review of Marine Science* 4, 495-522.
- Ridgwell, A. (2011) Evolution of the ocean's "biological pump". *Proceedings of the National Academy of Sciences* 108, 16485.
- Ridgwell, A. and Hargreaves, J.C. (2007) Regulation of athmospheric CO₂ by deep-sea sediments
 in an Earth system model. *Global Biogeochemical Cycles* 21, 10.1029/2006GB002764.
- Ridgwell, A. and Zeebe, R.E. (2005) The role of the global carbonate cycle in the regulation and
 evolution of the Earth system. *Earth and Planetary Science Letters* 234, 299-315.
- Robador, A., Jungbluth, S.P., LaRowe, D.E., Bowers, R.M., Rappé, M.S., Amend, J.P. and Cowen,
 J.P. (2015) Activity and phylogenetic diversity of sulfate-reducing microorganisms in lowtemperature subsurface fluids within the upper oceanic crust. *Front. Extr. Microbiol.* 5,
 Article 748 doi: 710.3389/fmicb.2014.00748
- Robador, A., LaRowe, D.E., Jungbluth, S.P., Lin, H.-T., Rappé, M.S., Nealson, K.H. and Amend,
 J.P. (2016) Nanocalorimetric characterization of microbial activity in deep subsurface
 oceanic crustal fluids. *Front. Extr. Microbiol.* 7.
- Romankevich, E.A., Vetrov, A.A. and Peresypkin, V.I. (2009) Organic matter of the world ocean.
 Russ. Geol. Geophys. 50, 299-307.
- Røy, H., Kallmeyer, J., Adhikari, R.R., Pockalny, R., Jørgensen, B.B. and D'Hondt, S. (2012)
 Aerobic microbial respiration in 86-million-year-old deep-sea red clay. *Science* 336, 922925.
- Roy, M., McManus, J., Goñi, M.A., Chase, Z., Borgeld, J.C., Wheatcroft, R.A., Muratli, J.M.,
 Megowan, M.R. and Mix, A. (2013) Reactive iron and manganese distributions in seabed
 sediments near small mountainous rivers off Oregon and California (USA). *Continental Shelf Research* 54, 67-79.

- Ruardij, P. and Van Raaphorst, W. (1995) Benthic nutrient regeneration in the ERSEM ecosystem
 model of the North Sea. *Netherlands Journal of Sea Research* 33, 453-483.
- Ruppel, C.D. and Kessler, J.D. (2017) The interaction of climate change and methane hydrates.
 Reviews of Geophysics 55, 126-168.
- Russell, J.A., León-Zayas, R., Wrighton, K. and Biddle, J.F. (2016) Deep Subsurface Life from
 North Pond: Enrichment, Isolation, Characterization and Genomes of Heterotrophic
 Bacteria. *Frontiers in Microbiology* 7.
- Schippers, A. and Neretin, L.N. (2006) Quantification of microbial communities in near-surface
 and deeply buried marine sediments on the Peru continental margin using real-time PCR.
 Environmental Microbiology 8, 1251-1260.
- Schlünz, B. and Schneider, R.R. (2000) Transport of terrestrial organic carbon to the oceans by
 rivers: re-estimating flux- and burial rates. *International Journal of Earth Sciences* 88, 599 606.
- Schmidt, F., Elvert, M., Koch, B.P., Witt, M. and Hinrichs, K.-U. (2009) Molecular
 characterization of dissolved organic matter in pore watter of continental shelf sediments. *Geochim. Cosmochim. Acta* 73, 3337-3358.
- Schmidt, M.W.I., Torn, M.S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I.A., Kleber,
 M., Kögel-Knabner, I., Lehmann, J., Manning, D.A.C., Nannipieri, P., Rasse, D.P.,
 Weiner, S. and Trumbore, S.E. (2011) Persistence of soil organic matter as an ecosystem
 property. *Nature* 478, 49-56.
- Schmidtko, S., Stramma, L. and Visbeck, M. (2017) Decline in global oceanic oxygen content
 during the past five decades. *Nature* 542, 335-339.
- Schouten, S., de Graaf, W., Sinninghe Damsté, J.S., van Driel, G.B. and de Leeuw, J.W. (1994)
 Laboratory simulation of natural sulphurization: II. Reaction of multi-functionalized lipids
 with inorganic polysulphides at low temperatures. *Organic Geochemistry* 22, 825-IN814.
- Schouten, S., Middelburg, J.J., Hopmans, E.C. and Sinninghe Damsté, J.S. (2010) Fossilization
 and degradation of intact polar lipids in deep subsurface sediments: A theoretical approach.
 Geochimica et Cosmochimica Acta 74, 3806-3814.
- Schrum, H.N., Spivack, A.J., Kastner, M. and D'Hondt, S. (2009) Sulfate-reducing ammonium
 oxidation: A thermodynamically feasible metabolic pathway in subseafloor sediment. *Geology* 37, 939-942.
- Schulz, H.D. (2006) Quantification of early diagenesis: Dissolved constituents in marine pore
 water in: Schulz, H.D., Zabel, M. (Eds.), *Marine Geochemistry*, 2nd ed. Springer Verlag,
 Heidelberg, p. 574.
- Schuur, E.A.G., Vogel, J.G., Crummer, K.G., Lee, H., Sickman, J.O. and Osterkamp, T.E. (2009)
 The effect of permafrost thaw on old carbon release and net carbon exchange from tundra.
 Nature 459, 556.
- Schwarzenbach, R.P., Gschwend, P.M. and Imboden, D.M. (2005) Sorption I: General Introduction and Sorption Processes Involving Organic Matter, in: Schwarzenbach, R.P., Gschwend, P.M., Imboden, D.M. (Eds.), *Environmental Organic Chemistry*, 2nd ed, pp. 2463 275-330.
- Seewald, J.S. (2001) Aqueous geochemistry of low molecular weight hydrocarbons at elevated
 temperatures and pressures: constraints from mineral buffered laboratory experiments.
 Geochimica et Cosmochimica Acta 65, 1641-1664.

- Seewald, J.S., Seyfried, W.E. and Thornton, E.C. (1990) Organic-rich sediment alteration: an
 experimental and theoretical study at elevated temperatures and pressures. *Applied Geochemistry* 5, 193-209.
- 2470 Seiter, K., Hensen, C., Schroter, J. and Zabel, M. (2004) Organic carbon content in surface 2471 sediments - defining regional provinces. *Deep-Sea Res. I* **51**, 2001-2026.
- Seitz, K.W., Dombrowski, N., Eme, L., Spang, A., Lombard, J., Sieber, J.R., Teske, A.P., Ettema,
 T.J.G. and Baker, B.J. (2019) Asgard archaea capable of anaerobic hydrocarbon cycling. *Nature Communications* 10, 1822.
- 2475 Selley, R.C. (1998) *Elements of Petroleum Geology, 2nd Ed.* Academic Press, San Diego, CA.
- Shaffer, G., Malskær Olsen, S. and Pepke Pedersen, J.O. (2008) Presentation, calibration and
 validation of the low-order, DCESS Earth System Model (Version 1). *Geosci. Model Dev.*1, 17-51.
- Shah, S.R., Mollenhauer, G., Ohkouchi, N., Eglinton, T.I. and Pearson, A. (2008) Origins of
 archaeal tetraether lipids in sediments: Insights from radiocarbon analysis. *Geochimica et Cosmochimica Acta* 72, 4577-4594.
- Shah Walter, S.R., Jaekel, U., Osterholz, H., Fisher, A.T., Huber, J.A., Pearson, A., Dittmar, T.
 and Girguis, P.R. (2018) Microbial decomposition of marine dissolved organic matter in
 cool oceanic crust. *Nature Geoscience* 11, 334-339.
- Shebl, M.A. and Surdam, R.C. (1996) Redox reactions in hydrocarbon clastic reservoirs:
 experimental validation of this mechanism for porosity enhancement. *Chemical Geology* 132, 103-117.
- 2488 ShipboardScientificParty (2004) Leg 207 Summary. Proc. IODP 207, 1-89.
- Siegenthaler, U. and Sarmiento, J.L. (1993) Atmospheric carbon dioxide and the ocean. *Nature*365, 119-125.
- Simoneit, B.R.T. and Lonsdale, P.F. (1982) Hydrothermal petroleum in mineralized mounds at the
 seabed of Guaymas Basin. *Nature* 295, 198-202.
- Sinninghe Damsté, J.S. and De Leeuw, J.W. (1990) Analysis, structure and geochemical
 significance of organically-bound sulphur in the geosphere: State of the art and future
 research. Organic Geochemistry 16, 1077-1101.
- Sinninghe Damsté, J.S., Eglinton, T.I., De Leeuw, J.W. and Schenck, P.A. (1989a) Organic
 sulphur in macromolecular sedimentary organic matter: I. Structure and origin of sulphurcontaining moieties in kerogen, asphaltenes and coal as revealed by flash pyrolysis. *Geochimica et Cosmochimica Acta* 53, 873-889.
- Sinninghe Damsté, J.S., Kok, M.D., Köster, J. and Schouten, S. (1998) Sulfurized carbohydrates:
 an important sedimentary sink for organic carbon? *Earth and Planetary Science Letters* 164, 7-13.
- Sinninghe Damsté, J.S., Rijpstra, W.I.C., De Leeuw, J.W. and Schenck, P.A. (1989b) The
 occurrence and identification of series of organic sulphur compounds in oils and sediment
 extracts: II. Their presence in samples from hypersaline and non-hypersaline
 palaeoenvironments and possible application as source, palaeoenvironmental and maturity
 indicators. *Geochimica et Cosmochimica Acta* 53, 1323-1341.
- 2508 Siskin, M. and Katritzky, A.R. (1991) Reactivity of Organic Compounds in Hot Water: 2509 Geochemical and Technological Implications. *Science* **254**, 231-237.
- Soetaert, K., Herman, P.M.J. and Middelburg, J.J. (1996) A model of early diagenetic processes
 from the shelf to abyssal depths. *Geochim. Cosmochim. Acta* 60, 1019-1040.

- Soetaert, K., Middelburg, J.J., Herman, P.M.J. and Buis, K. (2000) On the coupling of benthic and
 pelagic biogeochemical models. *Earth-Science Reviews* 51, 173-201.
- Sørensen, J. and Jeørgensen, B.B. (1987) Early diagenesis in sediments from Danish coastal
 waters: Microbial activity and Mn-Fe-S geochemistry. *Geochimica et Cosmochimica Acta* 51, 1583-1590.
- Spang, A., Saw, J.H., Jørgensen, S.L., Zaremba-Niedzwiedzka, K., Martijn, J., Lind, A.E., van
 Eijk, R., Schleper, C., Guy, L. and Ettema, T.J.G. (2015) Complex archaea that bridge the
 gap between prokaryotes and eukaryotes. *Nature* 521, 173.
- Starnawski, P., Bataillon, T., Ettema, T.J.G., Jochum, L.M., Schreiber, L., Chen, X., Lever, M.A.,
 Polz, M.F., Jørgensen, B.B., Schramm, A. and Kjeldsen, K.U. (2017) Microbial
 community assembly and evolution in subseafloor sediment. *Proceedings of the National Academy of Sciences* 114, 2940-2945.
- Steen, A.D., Kevorkian, R.T., Bird, J.T., Dombrowski, N., Baker, B.J., Hagen, S.M., Mulligan,
 K.H., Schmidt, J.M., Webber, A.T., Royalty, T.M. and Alperin, M.J. (2019) Kinetics and
 Identities of Extracellular Peptidases in Subsurface Sediments of the White Oak River
 Estuary, North Carolina. *Applied and Environmental Microbiology* 85, e00102-00119.
- Steen, A.D., Quigley, L.N.M. and Buchan, A. (2016) Evidence for the Priming Effect in a
 Planktonic Estuarine Microbial Community. *Frontiers in Marine Science* 3.
- Steen, A.D., Vazin, J.P., Hagen, S.M., Mulligan, K.H. and Wilhelm, S.W. (2015) Substrate
 specificity of aquatic extracellular peptidases assessed by competitive inhibition assays
 using synthetic substrates. *Aquatic Microbial Ecology* **75**, 271-281.
- 2533 Stevenson, F.J. (1986) Cycles of Soil. Wiley & Sons, New York.
- Stumm, W. and Morgan, J.J. (1996) *Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters*, 3rd ed. John Wiley & Sons, New York.
- Sun, M.-Y., Aller, R.C., Lee, C. and Wakeham, S.G. (2002a) Effect of oxygen and redox
 oscillation on degradation of cell-associated lipids in surficial marine sediments. *Geochimica et Cosmochimica Acta* 66, 2003–2012.
- Sun, M.-Y., Cai, W.-J., Joye, S.B., Ding, H., Dai, J. and Hollibaugh, J.T. (2002b) Degradation of
 algal lipids in microcosm sediments with different mixing regimes. *Organic Geochemistry* 33, 445-459.
- Sun, M.-Y., Lee, C. and Aller, R.C. (1993) Laboratory studies of oxic and anoxic degradation of
 chlorophyll-a in Long Island Sound sediments. *Geochimica et Cosmochimica Acta* 57,
 147-157.
- Sun, M.-Y., Wakeham, S.G. and Lee, C. (1997) Rates and mechanisms of fatty acid degradation
 in oxic and anoxic coastal marine sediments of Long Island Sound, New York, USA. *Geochimica et Cosmochimica Acta* 61, 341-355.
- Sunagawa, S., Coelho, L.P., Chaffron, S., Kultima, J.R., Labadie, K., Salazar, G., Djahanschiri, 2548 2549 B., Zeller, G., Mende, D.R., Alberti, A., Cornejo-Castillo, F.M., Costea, P.I., Cruaud, C., d'Ovidio, F., Engelen, S., Ferrera, I., Gasol, J.M., Guidi, L., Hildebrand, F., Kokoszka, F., 2550 2551 Lepoivre, C., Lima-Mendez, G., Poulain, J., Poulos, B.T., Royo-Llonch, M., Sarmento, H., Vieira-Silva, S., Dimier, C., Picheral, M., Searson, S., Kandels-Lewis, S., Bowler, C., de 2552 2553 Vargas, C., Gorsky, G., Grimsley, N., Hingamp, P., Iudicone, D., Jaillon, O., Not, F., Ogata, H., Pesant, S., Speich, S., Stemmann, L., Sullivan, M.B., Weissenbach, J., Wincker, 2554 2555 P., Karsenti, E., Raes, J., Acinas, S.G. and Bork, P. (2015a) Structure and function of the global ocean microbiome. Science 348, 1261359. 2556

- 2557 Sunagawa, S., Coelho, L.P., Chaffron, S., Kultima, J.R., Labadie, K., Salazar, G., Djahanschiri, 2558 B., Zeller, G., Mende, D.R., Alberti, A., Cornejo-Castillo, F.M., Costea, P.I., Cruaud, C., Ovidio, F., Engelen, S., Ferrera, I., Gasol, J.M., Guidi, L., Hildebrand, F., Kokoszka, F., 2559 2560 Lepoivre, C., Lima-Mendez, G., Poulain, J., Poulos, B.T., Royo-Llonch, M., Sarmento, H., Vieira-Silva, S., Dimier, C., Picheral, M., Searson, S., Kandels-Lewis, S., Bowler, C., de 2561 2562 Vargas, C., Gorsky, G., Grimsley, N., Hingamp, P., Iudicone, D., Jaillon, O., Not, F., 2563 Ogata, H., Pesant, S., Speich, S., Stemmann, L., Sullivan, M.B., Weissenbach, J., Wincker, 2564 P., Karsenti, E., Raes, J., Acinas, S.G. and Bork, P. (2015b) Structure and function of the 2565 global ocean microbiome. Science 348, 1261359.
- Sweetman, A.K., Thurber, A.R., Smith, C.R., Levin, L.A., Mora, L.A., Wei, C., Gooday, A.J.,
 Jones, D.O.B., Yasuhara, R.M., Ingels, M., Ruhl, H.A., Frieder, C.A., Danovaro, R.,
 Würzberg, L., Baco, A., Grupe, B.M., Pasulka, A., Meyer, K.S., Dunlop, K.M., Henry, L.
 and Roberts, J.M. (2017) Major impacts of climate change on deep-sea benthic ecosystems *Elementa: Science of the Anthropocene* 5, https://doi.org/10.1525/elementa.1203.
- Tarafa, M.E., Whelan, J.K., Oremland, R.S. and Smith, R.L. (1987) Evidence of microbiological
 activity in Leg 95 (New Jersey Transect) sediments. *Initial reports of the DSDP* 95, 635 640.
- Taylor, G.T. (1995) Microbial degradation of sorbed and dissolved protein in seawater. *Limnology and Oceanography* 40, 875-885.
- Tegelaar, E.W., de Leeuw, J.W., Derenne, S. and Largeau, C. (1989) A reappraisal of kerogen
 formation. *Geochimica et Cosmochimica Acta* 53, 3103-3106.
- Teske, A., Callaghan, A.V. and LaRowe, D.E. (2014) Biosphere frontiers: Deep life in the sedimented hydrothermal system of Guaymas Basin. *Front. Extr. Microbiol.* **5**, Art. 362.
- Thamdrup, B. (2000) Bacterial Manganese and Iron Reduction in Aquatic Sediments, in: Schink,
 B. (Ed.), *Advances in Microbial Ecology*. Springer US, Boston, MA, pp. 41-84.
- Thomsen, L. and van Weering, T.C.E. (1998) Spatial and temporal variability of particulate matter
 in the benthic boundary layer at the N.W. European Continental Margin (Goban Spur).
 Progress in Oceanography 42, 61-76.
- Thullner, M., Dale, A.W. and Regnier, P. (2009) Global-scale quantification of mineralization
 pathways in marine sediments: A reaction-transport modeling approach. *Geochem. Geophys. Geosys.* 10, 1-24.
- Thullner, M., Regnier, P. and Van Cappellen, P. (2007) Modeling microbially induced carbon
 degradation in redox-stratified subsurface environments: Concepts and open questions.
 Geomicrobiol. J. 24, 139-155.
- Thullner, M., Van Cappellen, P. and Regnier, P. (2005) Modeling the impact of microbial activity
 on redox dynamics in porous media. *Geochim. Cosmochim. Acta* 69, 5005-5019.
- Tissot, B.P. and Welte, D.H. (1984) *Petroleum Formation and Occurrence*, 2nd ed. Springer Verlag, Berlin.
- Tjiputra, J.F., Roelandt, C., Bentsen, M., Lawrence, D.M., Lorentzen, T., Schwinger, J., Seland,
 Ø. and Heinze, C. (2013) Evaluation of the carbon cycle components in the Norwegian
 Earth System Model (NorESM). *Geosci. Model Dev.* 6, 301-325.
- Trabucho-Alexandre, J., Hay, W.W. and de Boer, P.L. (2012) Phanerozoic environments of black shale deposition and the Wilson Cycle. *Solid Earth* **3**, 29-42.
- Trembath-Reichert, E., Morono, Y., Ijiri, A., Hoshino, T., Dawson, K., Inagaki, F. and Orphan,
 V.J. (2017) Methyl-compound use and slow growth characterize microbial life in 2-km deep subseafloor coal and shale beds *PNAS* 114, E9206-E9215.

- Treseder, K.K. and Lennon, J.T. (2015) Fungal Traits That Drive Ecosystem Dynamics on Land.
 Microbiology and Molecular Biology Reviews 79, 243-262.
- Tromp, T.K., Van Cappellen, P. and Key, R.M. (1995) A global model for the early diagenesis of
 organic carbon and organic phosphorous in marine sediments. *Geochim. Cosmochim. Acta* 59, 1259-1284.
- Tully, B.J., Graham, E.D. and Heidelberg, J.F. (2018) The reconstruction of 2,631 draft metagenome-assembled genomes from the global oceans. *Scientific Data* **5**, 170203.
- Tully, B.J. and Heidelberg, J.F. (2016) Potential mechanisms for microbial energy acquisition in
 oxic deep-sea sediments. *Applied and Environmental Microbiology* 82, 4232-4243.
- Van Cappellen, P. and Ingall, E.D. (1994) Benthic phosphorus regeneration, net primary
 production, and ocean anoxia: A model of the coupled marine biogeochemical cycles of
 carbon and phosphorus. *Palaeogeog. Palaeoclim. Palaeoecol.* 9, 677-692.
- van Dongen, B.E., Schouten, S. and Sinninghe Damsté, J.S. (2006) Preservation of carbohydrates
 through sulfurization in a Jurassic euxinic shelf sea: Examination of the Blackstone Band
 TOC cycle in the Kimmeridge Clay Formation, UK. *Organic Geochemistry* 37, 1052-1073.
- van Kaam-Peters, H.M.E., Schouten, S., de Leeuw, J.W. and Sinninghe Damsté, J.S. (1997) A
 molecular and carbon isotope biogeochemical study of biomarkers and kerogen pyrolysates
 of the Kimmeridge Clay Facies: palaeoenvironmental implications. *Organic Geochemistry* 2621 27, 399-422.
- Van Kaam-Peters, H.M.E., Schouten, S., Köster, J. and Sinninghe Damstè, J.S. (1998) Controls
 on the molecular and carbon isotopic composition of organic matter deposited in a
 Kimmeridgian euxinic shelf sea: evidence for preservation of carbohydrates through
 sulfurisation. *Geochimica et Cosmochimica Acta* 62, 3259-3283.
- van Nugteren, P., Herman, P.M., Moodley, L., Middelburg, J.J. and Vos, M. (2009) Spatial
 distribution of detrital resources determines the outcome of competition between bacteria
 and a facultative detritivorous worm. *Limnology and Oceanography* 54, 1414-1419.
- Veuger, B., van Oevelen, D. and Middelburg, J.J. (2012) Fate of microbial nitrogen, carbon,
 hydrolysable amino acids, monosaccharides, and fatty acids in sediment. *Geochimica et Cosmochimica Acta* 83, 217-233.
- Vuillemin, A., Ariztegui, D., Horn, F., Kallmeyer, J., Orsi, W.D. and Team, t.P.S. (2018) Microbial
 community composition along a 50,000-year lacustrine sediment sequence. *FEMS Microbial Ecology* 94, doi.org/10.1093/femsec/fiy1029.
- Vuillemin, A., Wankel, S.D., Coskun, Ö.K., Magritsch, T., Vargas, S., Estes, E.R., Spivack, A.J.,
 Smith, D.C., Pockalny, R., Murray, R.W., D'Hondt, S. and Orsi, W.D. (2019) Archaea
 dominate oxic subseafloor communities over multimillion-year time scales. *Science Advances* 5, eaaw4108.
- Wakeham, S.G., Lee, C., Hedges, J.I., Hernes, P.J. and Peterson, M.J. (1997) Molecular indicators
 of diagenetic status in marine organic matter. *Geochimica et Cosmochimica Acta* 61, 5363 5369.
- Wallmann, K., Pinero, E., Burwicz, E.B., Haeckel, M., Hensen, C., Dale, A.W. and Ruepke, L.
 (2012) The global inventory of methane hydrate in marine sediments: a theoretical approach. *Energies* 5, 2449-2498.
- Walsh, E.A., Kirkpatrick, J.B., Rutherford, S.D., Smith, D.C., Sogin, M. and D'Hondt, S. (2015)
 Bacterial diversity and community composition from seasurface to subseafloor. *The Isme Journal* 10, 979.

- Walsh, J.J. (1991) Importance of continental margins in the marine biogeochemical cycling of
 carbon and nitrogen. *Nature* 350, 53-55.
- Wang, G., Spivack, A.J. and D'Hondt, S. (2010) Gibbs energies of reaction and microbial
 mutualism in anaerobic deep subseafloor sediments of ODP Site 1226. *Geochim. Cosmochim. Acta* 74, 3938-3947.
- Wankel, S.D., Germanovich, L.N., Lilley, M.D., Genc, G., DiPerna, C.J., Bradley, A.S., Olson,
 E.J. and Girguis, P.R. (2011) Influence of subsurface biosphere on geochemical fluxes
 from diffuse hydrothermal fluids. *Nat. Geosci.* 4, 461-468.
- Wegener, G., Bausch, M., Holler, T., Thang, N.M., Prieto Mollar, X., Kellermann, M.Y., Hinrichs,
 K.-U. and Boetius, A. (2012) Assessing sub-seafloor microbial activity by combined stable
 isotope probing with deuterated water and 13C-bicarbonate. *Environmental Microbiology* 14, 1517-1527.
- Werne, J.P., Hollander, D.J., Lyons, T.W. and Sinninghe Damsté, J.S. (2004) Organic sulfur
 biogeochemistry: Recent advances and future research directions, in: Amend, J.P.,
 Edwards, K., Lyons, T.W. (Eds.), *Sulfur biogeochemistry Past and Present*. Geological
 Society of America Special Paper 379, Boulder, Colorado, pp. 135-150.
- Westrich, J.T. and Berner, R.A. (1984) The role of sedimentary organic matter in bacterial sulfate
 reduction: The G model tested. *Limnol. Oceanogr.* 29, 236-249.
- Wheat, C.G., Hartwell, A.M., McManus, J., Fisher, A.T., Orcutt, B.N., Schlicht, L.E.M.,
 Niedenzu, S. and Bach, W. (2019) Geology and fluid discharge at Dorado outcrop, a low
 temperature ridge-flank hydrothermal system. *Geochemistry, Geophysics, Geosystems* 20, 487-504.
- Wheat, C.G., Hulme, S.M., Fisher, A.T., Orcutt, B.N. and Becker, K. (2013) Seawater recharge
 into oceanic crust: IODP Exp. 327 Site U1363 Grizzly Bare outcrop. *Geochem. Geophys. Geosys.* 14, 1957–1972
- Whelan, J.K., Oremland, R.S., Tarafa, M.E., Smith, R., Howarth, R. and Lee, C. (1986) Evidence
 for sulfate-reducing and methane producing microorganisms in sediments from Sites 618,
 619, and 622. *Initial reports of the DSDP* 96, 767-775.
- 2676 Wignall, P.B. (1994) *Black shales*. Oxford University Press, Oxford.
- Wilson, J.D., Barker, S. and Ridgwell, A. (2012) Assessment of the spatial variability in particulate
 organic matter and mineral sinking fluxes in the ocean interior: implications for the ballast
 hypothesis. *Global Biogeochemical Cycles* 26, GB004398.
- Wirtz, K.W. (2003) Control of biogeochemical cycling by mobility and metabolic strategies of
 microbes in the sediments: an integrated model study. *FEMS Microbiology Ecology* 46,
 2682 295-306.
- Wrighton, K.C., Castelle, C.J., Varaljay, V.A., Satagopan, S., Brown, C.T., Wilkins, M.J.,
 Thomas, B.C., Sharon, I., Williams, K.H., Tabita, F.R. and Banfield, J.F. (2016) RubisCO
 of a nucleoside pathway known from Archaea is found in diverse uncultivated phyla in
 bacteria. *The Isme Journal* 10, 2702.
- Wrighton, K.C., Castelle, C.J., Wilkins, M.J., Hug, L.A., Sharon, I., Thomas, B.C., Handley, K.M.,
 Mullin, S.W., Nicora, C.D., Singh, A., Lipton, M.S., Long, P.E., Williams, K.H. and
 Banfield, J.F. (2014) Metabolic interdependencies between phylogenetically novel
 fermentters and respiratory organisms in an unconfined aquifer. *ISME J.* 8, 1452-1463.
- Wrighton, K.C., Thomas, B.C., Sharon, I., Miller, C.S., Castelle, C.J., VerBerkmoes, N.C.,
 Wilkins, M.J., Hettich, R.L., Lipton, M.S., Williams, K.H., Long, P.E. and Banfield, J.F.

- 2693 (2012) Fermentation, hydorgen, and sulfur metabolism on multiple uncultivated bacterial
 2694 phyla. *Science* 337, 1661-1665.
- Yu, T., Wu, W., Liang, W., Lever, M.A., Hinrichs, K.-U. and Wang, F. (2018a) Growth of
 sedimentary Bathyarchaeota on lignin as an energy source. *Proceedings of the National Academy of Sciences* 115, 6022-6027.
- 2698 Yu, T., Wu, W., Liang, W., Lever, M.A., Hinrichs, K.-U. and Wang, F. (2018b) Growth of 2699 sedimentary *Bathyarchaeota* on lignin as an energy source. *PNAS* **115**, 3022-6027.
- Zhao, R., Hannisdal, B., Mogollon, J.M. and Jørgensen, S.L. (2019) Nitrifier abundance and diversity peak at deep redox transition zones. *Scientific Reports* 9, 8633.
- Zhuang, G.-C., Montgomery, A., Samarkin, V.A., Song, M., Liu, J., Schubotz, F., Teske, A.,
 Hinrichs, K.-U. and Joye, S.B. (2019) Generation and Utilization of Volatile Fatty Acids
 and Alcohols in Hydrothermally Altered Sediments in the Guaymas Basin, Gulf of
 California. *Geophysical Research Letters* 46, 2637-2646.

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Figure 4









(C)





cells cm-3

