

**SEWAGE TREATMENT PLANT #1 AREA, SWMU 117  
PER- AND POLYFLUOROALKYL SUBSTANCES (PFAS)  
SITE ASSESSMENT PROGRESS REPORT  
KENNEDY SPACE CENTER, FLORIDA**

**Prepared for:**



**National Aeronautics and Space Administration  
Kennedy Space Center, Florida**

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**APPENDIX A**  
**HISTORICAL ANALYTICAL RESULTS**

Table A-1. Historical Soil Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	SB0061		SB0062		SB0063			SB0064		
			1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	
			0 - 0.5	0.5 - 2	0 - 0.5	0.5 - 2	0 - 0.5	0 - 0.5*	0.5 - 2	0 - 0.5	0.5 - 2	
<b>PFAS with Screening Criteria (µg/kg)</b>												
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	23	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanefluoronic acid (PFBS)	375-73-5	1900	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	130	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	19	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	19	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	13	0.36 I	3.1	3.8	1.7	1.3	1.2	1.7	1.2	2	2
<b>PFAS without Screening Criteria (µg/kg)</b>												
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	0.96 U	0.95 U	1 U	0.98 U	1.1 U	1.4 U	1.1 U	1.1 U	1 U	1 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	0.96 U	0.95 U	1 U	0.98 U	1.1 U	1.4 U	1.1 U	1.1 U	1 U	1 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.48 U	0.67	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	0.48 U	0.48 U	0.51 U	0.49 U	0.54 U	0.71 U	0.54 U	0.53 U	0.51 U	0.51 U

Table A-1. Historical Soil Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	SB0065		SB0066		SB0067		SB0068			SB0069	
			1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21	1/26/21
			0 - 0.5	0.5 - 2	0 - 0.5	0.5 - 2	0 - 0.5	0.5 - 2	0 - 0.5	0 - 0.5*	0.5 - 2	0 - 0.5	0.5 - 2
<b>PFAS with Screening Criteria (µg/kg)</b>													
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	23	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanefulfonic acid (PFBS)	375-73-5	1900	0.52 I	0.78	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.56 U	0.55 U
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	130	37.8	14.2	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.56 U	0.55 U
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	19	0.95	0.66 U	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.49 I	0.56 U	0.55 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	19	1.4	0.61 I	0.47 U	0.49 U	0.55 U	0.6 U	0.28 I	0.52 U	0.53 U	0.56 U	0.55 U
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	13	70.6	27.4	4.1	6.4	3.9	3.6	2.3	2.3	9.5	0.56 U	0.55 U
<b>PFAS without Screening Criteria (µg/kg)</b>													
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	1.3 U	1.3 U	0.94 U	0.98 U	1.1 U	1.2 U	1 U	1 U	1.1 U	1.1 U	1.1 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	1.3 U	1.3 U	0.94 U	0.98 U	1.1 U	1.2 U	1 U	1 U	1.1 U	1.1 U	1.1 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.67 U	0.66 U	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.97	0.56 U	0.55 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	0.67 U	0.66 U	0.47 U	0.49 U	0.55 U	0.6 U	0.3 I	0.31 I	0.53 U	0.56 U	0.55 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	0.79	0.47 I	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.38 I	0.33 I
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	2.1	2.1	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.56 U	0.55 U
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	0.67 U	0.66 U	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.56 U	0.55 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	0.67 U	0.66 U	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.56 U	0.55 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	0.67 U	0.66 U	0.47 U	0.49 U	0.55 U	0.6 U	0.51 U	0.52 U	0.53 U	0.56 U	0.55 U

All results reported in microgram per kilogram (µg/kg)

1 The USEPA Regional Screening Levels (RSLs) for HFPO-DA, PFBS, PFHxS, PFNA, PFOA, and PFOS are presented in this table.

2 The Soil RSL is cited from the USEPA Regional Screening Levels and calculated with the USEPA RSL Calculator based on a hazard quotient of 0.1 (USEPA, 2022)

3 HFPO-DA is commonly referred to as GenX

\* Duplicate sample results are included in this table and labeled with asterisk

-- = No applicable screening criteria

Bolding indicates analyte was detected

Shading indicates exceedance of screening criteria

USEPA = United States Environmental Protection Agency

ft bls = feet below land surface

NA = Not Applicable; compound not analyzed

PFAS = per- and polyfluoroalkyl substances

I = Estimated result < Limit of Quantitation and ≥ Detection Limit

U = Analyte was not detected

Table A-2. Historical DPT Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	DPT0059				DPT0060			
			11/7/2018	11/7/2018	11/7/2018	11/7/2018	11/6/2018	11/6/2018	11/6/2018	11/6/2018
			8 - 12	23 - 27	33 - 37	43 - 47	8 - 12	23 - 27	33 - 37	43 - 47
<b>PFAS with Screening Criteria (ng/L)</b>										
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	6	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanedisulfonic acid (PFBS)	375-73-5	600	10	3.7 M	1.8	0.89 UM	0.48 JM	1 U	0.97 U	0.95 UM
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	39	87	22 M	15 M	2 M	1.5 JM	0.8 JM	0.86 JM	0.49 JM
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	5.9	1.3 U	1.3 UM	1.3 U	1.3 U	0.58 JM	1.5 U	1.5 U	1.4 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	6	15 M	3.7 M	7.7 M	0.59 JM	2 JM	2 JM	1.9 JM	1.4 UM
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	4	11 M	11	3.4 JM	7.3	7.8	3.2 J	2.4 J	1.6 JM
<b>PFAS without Screening Criteria (ng/L)</b>										
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	8.8 U	8.7 UM	8.7 U	8.9 UJ1	9.8 UQ	10 UQ	9.7 UQ	9.5 UQ
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	8.8 U	8.7 U	8.7 U	8.9 U	9.8 U	10 U	9.7 U	9.5 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanedisulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.88 U	0.87 U	0.87 U	0.89 U	0.98 U	1 U	0.97 U	0.95 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.3 U	1.3 U	1.3 U	1.3 U	1.5 U	1.5 U	1.5 U	1.4 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	16	4.4	3.2	1.3 UM	0.61 J	1.5 U	1.5 U	1.4 U
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	32 M	12	7.3 M	0.99 JM	0.64 JM	0.5 J	0.47 JM	0.95 U
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	2.7 U	2.6 U	2.6 U	2.7 U	2.9 U	3 U	2.9 U	2.9 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	2.7 U	2.6 U	2.6 U	2.7 U	2.9 U	3 U	2.9 U	2.9 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.3 U	1.3 UM	1.3 U	1.3 U	1.5 U	1.5 U	1.5 U	1.4 U

Table A-2. Historical DPT Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	DPT0061					DPT0062				
			11/6/2018	11/6/2018	11/6/2018	11/6/2018	11/6/2018	11/7/2018	11/7/2018	11/7/2018	11/7/2018	11/7/2018
			8 - 12	23 - 27	33 - 37	33 - 37*	43 - 47	8 - 12	23 - 27	33 - 37	47 - 47	47 - 47*
<b>PFAS with Screening Criteria (ng/L)</b>												
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	6	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanefluoronic acid (PFBS)	375-73-5	600	0.6 J	0.98 U	0.97 U	0.95 UM	0.95 U	6.1 M	1.2 JM	1.7 JM	1 U	0.94 U
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	39	1.8 JM	0.98 UM	0.97 U	0.95 U	0.95 UM	100 M	21 M	30 M	1.3 JM	0.97 JM
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	5.9	1.4 U	1.5 U	1.5 U	1.4 U	1.4 U	1.4 U	1.9 M	1.5 U	1.5 UM	1.4 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	6	1.9 JM	1.5 UM	1.5 UM	1.4 UM	1.4 UM	24 M	13 M	9.3 M	0.56 JM	1.4 UM
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	4	1.6 J	2.9 U	2.9 UM	2.8 U	2.9 UM	10 M	74	3 U	2.3 JM	2.1 JM
<b>PFAS without Screening Criteria (ng/L)</b>												
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	9.7 UQM	9.8 UQ	9.7 UQ	9.5 UQM	9.5 UQM	9.6 UM	9.6 U	10 U	10 U	9.4 UM
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	9.7 U	9.8 U	9.7 U	9.5 U	9.5 U	9.6 UM	9.6 UM	10 U	10 UM	9.4 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.97 U	0.98 U	0.97 U	0.95 U	0.95 U	0.96 U	0.96 U	1 U	1 UM	0.94 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.4 U	1.5 U	1.5 U	1.4 U	1.4 U	1.4 U	1.4 U	1.5 U	1.5 U	1.4 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	0.91 J	1.5 U	1.5 U	1.4 U	1.4 U	20	5	12	1.5 U	1.4 U
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	0.61 JM	0.98 UM	0.97 U	0.95 U	0.95 U	16 M	3.7 M	11 M	1 UM	0.94 UM
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	2.9 U	2.9 U	2.9 U	2.8 U	2.9 U	2.9 U	2.9 U	3 U	3 U	2.8 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	2.9 U	2.9 U	2.9 U	2.8 U	2.9 U	2.9 U	2.9 U	3 U	3 U	2.8 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.4 U	1.5 U	1.5 U	1.4 U	1.4 U	1.4 UM	1.4 U	1.5 U	1.5 U	1.4 U

Table A-2. Historical DPT Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	DPT0064				DPT0065			
			11/6/2018	11/6/2018	11/6/2018	11/6/2018	11/6/2018	11/6/2018	11/6/2018	11/6/2018
			8 - 12	23 - 27	33 - 37	43 - 47	13 - 17	23 - 27	33 - 77	43 - 47
<b>PFAS with Screening Criteria (ng/L)</b>										
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	6	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanefluoronic acid (PFBS)	375-73-5	600	47 M	41 M	48 M	24 M	14 M	11 M	5.2 M	3.2 M
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	39	1200 D	160 M	220	75	290	45	20	16
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	5.9	7.5	7.5	6.8	1.2 JM	60	50	17	15 M
Perfluoro-n-octanoic acid (PFOA)	335-67-1	6	160 M	78	85 M	36 M	910 D	540 D	45 M	47 M
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	4	520 D	550 D	460 D	50	420 D	15	6.3	19
<b>PFAS without Screening Criteria (ng/L)</b>										
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	9.6 UQM	9.7 UQ	9.7 UQM	9.7 UQ	10 UMQ	9.6 UQ	10 UQ	10 UQ
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	9.6 U	9.7 U	9.7 U	9.7 U	10 U	9.6 U	10 U	10 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.74 JM	1.4 J	1.6 JM	0.97 U	1 U	0.96 U	1 U	1 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.4 U	1.4 U	1.4 U	1.5 U	1.5 U	1.4 U	1.5 U	1.5 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	140	26 M	36 M	15 M	1100 D	1100 D	50	51
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	220	62 M	87 M	43	2500 DM	1200 D	110 M	87
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	2.9 U	2.9 U	2.9 U	2.9 U	3 U	2.9 U	3 U	3 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	2.9 U	2.9 U	2.9 U	2.9 U	3 U	2.9 U	3 U	3 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.4 U	1.4 U	1.4 U	1.5 U	1.5 U	1.4 U	1.5 U	1.5 U

Table A-2. Historical DPT Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	DPT0124					DPT0143				
			3/26/2019	3/26/2019	3/26/2019	3/26/2019	3/26/2019	3/19/2019	3/19/2019	3/19/2019	3/19/2019	
			8 - 12	23 - 27	33 - 37	33 - 37*	43 - 47	8 - 12	23 - 27	33 - 37	43 - 47	
<b>PFAS with Screening Criteria (ng/L)</b>												
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	6	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-1-butanefluorobutane sulfonic acid (PFBS)	375-73-5	600	0.63 JM	1 UM	0.9 UM	0.93 U	0.96 UM	1 J	0.91 UM	1.1 UM	0.93 UM	
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	39	1 JM	0.7 JM	0.9 UM	0.93 U	0.78 JM	3.5 M	3.5	14 M	4.4 M	
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	5.9	1.3 U	1.6 UM	1.4 UM	1.4 U	1.4 U	1.3 UM	1.4 U	1.6 UM	1.4 U	
Perfluoro-n-octanoic acid (PFOA)	335-67-1	6	1.3 UM	1.6 UM	1.4 UM	1.4 U	1.4 UM	4.9 M	2.4 M	9.2 M	1.9 J	
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	4	1 JM	3.1 UM	2.7 UM	2.8 UM	2 JM	4.9 M	2.7 UM	120	3.7 J	
<b>PFAS without Screening Criteria (ng/L)</b>												
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
11-chlorooicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	8.9 U	10 U	9 U	9.3 U	9.6 U	8.6 U	9.1 U	11 U	9.3 U	
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	8.9 U	10 U	9 U	9.3 U	9.6 U	8.6 U	9.1 U	11 U	9.3 U	
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.89 U	1 U	0.9 UM	0.93 U	0.96 U	0.86 U	0.91 U	0.82 J	0.93 U	
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.3 UM	1.6 U	1.4 UM	1.4 U	1.4 U	1.3 U	1.4 U	1.6 U	1.4 U	
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	1.3 UM	1.6 UM	1.4 U	1.4 U	1.4 U	2.9	1.3 JM	5	0.99 JM	
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	0.89 UM	1 UM	0.9 UM	0.93 U	0.96 UM	5.4	2.3 M	11 M	2.6 M	
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	2.7 U	3.1 UM	2.7 U	2.8 U	2.9 U	2.6 U	2.7 U	3.2 U	2.8 U	
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	2.7 U	3.1 U	2.7 U	2.8 U	2.9 U	2.6 U	2.7 U	3.2 U	2.8 U	
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.3 UM	1.6 UM	1.4 UM	1.4 U	1.4 U	1.3 UM	1.4 U	1.6 U	1.4 U	



Table A-2. Historical DPT Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	DPT0144				DPT0187			
			3/19/2019	3/19/2019	3/19/2019	3/19/2019	1/27/2021	1/27/2021	1/27/2021	1/27/2021
			8 - 12	23 - 27	33 - 37	43 - 47	8 - 12	23 - 27	33 - 37	43 - 47
<b>PFAS with Screening Criteria (ng/L)</b>										
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	6	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanedisulfonic acid (PFBS)	375-73-5	600	12	3.3	2.5 M	0.92 U	20 U	20 U	39.4	52.6
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	39	62	79 J1	26	5.2 M	68.4	505	169	182
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	5.9	4	1.4 U	1.2 JM	1.4 U	18.6 I	46.4	16.4 I	50 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	6	37 M	21 M	14 M	1.9	84.7	165	50.7	33.2 I
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	4	310 M	8.3 M	87	8.5	1580	4520	2060	1190
<b>PFAS without Screening Criteria (ng/L)</b>										
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	9.4 U	9.3 U	9.6 U	9.2 U	40 U	40 U	40 U	100 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	9.4 U	9.3 U	9.6 U	9.2 U	40 U	40 U	40 U	100 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanedisulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.94 U	0.93 U	0.88 J	0.92 U	11.1 I	20 U	20 U	50 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.4 U	1.4 U	1.4 U	1.4 U	20 U	20 U	100 U	50 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	12	12	5	0.85 J	37.3	143	27.3	50 U
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	17	12 M	10	1.8 JM	50.1	169	51.1	45.2 I
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	2.8 U	2.8 U	2.9 U	2.8 U	20 U	20 U	20 U	250 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	2.8 U	2.8 U	2.9 U	2.8 U	20 U	20 U	100 U	50 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.4 U	1.4 U	1.4 U	1.4 U	20 U	20 U	20 U	50 U

**Table A-2. Historical DPT Analytical Results**

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria <sup>1,2</sup>	DPT0188				DPT0189			
			1/27/2021	1/27/2021	1/27/2021	1/27/2021	1/27/2021	1/27/2021	1/27/2021	1/27/2021
			8 - 12	23 - 27	33 - 37	43 - 47	8 - 12	23 - 27	33 - 37	43 - 47
<b>PFAS with Screening Criteria (ng/L)</b>										
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	6	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanefluoronic acid (PFBS)	375-73-5	600	32.6	4 U	36.3	35.7	4 U	6.8 U	4 U	4 U
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	39	32 U	5.1	60.5	59	4 U	6.8 U	4 U	4 U
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	5.9	32 U	4 U	80	6.1	4 U	6.8 U	4 U	4 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	6	32 U	4 U	233	95.1	4 U	6.8 U	4 U	4 U
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	4	71.4	13.7	84.9	44.8	4 U	6.8 U	10.3	4 U
<b>PFAS without Screening Criteria (ng/L)</b>										
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	320 U	8 U	80 U	48 U	40 U	68 U	8 U	8 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	320 U	8 U	80 U	48 U	40 U	68 U	8 U	8 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	32 U	4 U	20 U	24 U	4 U	6.8 U	4 U	4 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	160 U	4 U	40 U	24 U	20 U	34 U	20 U	4 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	32 U	2.2 I	552	255	4 U	6.8 U	4 U	4 U
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	32 U	2.1 I	335	252	4 U	6.8 U	4 U	4 U
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	32 U	4 U	4 U	4.8 U	4 U	6.8 U	4 U	4 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	160 U	4 U	40 U	24 U	20 U	34 U	20 U	4 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	160 U	4 U	40 U	24 U	4 U	6.8 U	4 U	4 U

All results reported in nanogram per liter (ng/L)

1 The USEPA Regional Screening Levels (RSLs) for HFPO-DA, PFBS, PFHxS, PFNA, PFOA, and PFOS are presented in this table.

2 The Groundwater RSL is cited from the USEPA Regional Screening Levels and calculated with the USEPA RSL Calculator based on a hazard quotient of 0.1 (USEPA, 2022)

3 HFPO-DA is commonly referred to as GenX

\* Duplicate sample results are included in this table and labeled with asterisk

-- = No applicable screening criteria

Bolding indicates analyte was detected

Shading indicates exceedance of screening criteria

STP1 = Sewage Treatment Plant #1

USEPA = United States Environmental Protection Agency

ft bls = feet below land surface

NA = Not Applicable; compound not analyzed

PFAS = per- and polyfluoroalkyl substances

I / J = Estimated result < Limit of Quantitation and ≥ Detection Limit

Q = Out of holding time

U = Analyte was not detected

M = Presence of material is verified but not quantified

D = Reported value is from a dilution

Note: A data quality review was performed by Tetra Tech's data manager and the results provided in this table were found to have been generated in conformance with good analytical practices. Some minor nonconformance issues were noted in the quality control elements associated with project samples, and the appropriate data qualification was applied to the affected results as needed. Additional details on data quality are included in the analytical reports provided in the Appendices.

**Table A-3. Historical Monitoring Well Analytical Results**

Location ID	CAS No.	Screening Criteria <sup>1,2</sup>	CM S-MW0039
Date			6/21/2021
Screen Interval (ft bls)			5.5
<b>PFAS with Screening Criteria (ng/L)</b>			
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>3</sup>	13252-13-6	<b>6</b>	6.67 U
Perfluoro-1-butananesulfonic acid (PFBS)	375-73-5	<b>600</b>	<b>1.34 I</b>
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	<b>39</b>	<b>3.05 I</b>
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	<b>5.9</b>	0.98 U
Perfluoro-n-octanoic acid (PFOA)	335-67-1	<b>6</b>	<b>4.12</b>
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	<b>4</b>	<b>1.68 I</b>
<b>PFAS without Screening Criteria (ng/L)</b>			
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	0.9 U
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	0.9 U
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	0.86 U
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	1.58 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	0.9 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	1.44 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.3 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	<b>2.06 I</b>
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	<b>3.08 I</b>
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	1.14 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	1.23 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.24 U

All results reported in nanogram per liter (ng/L)

1 The USEPA Regional Screening Levels (RSLs) for HFPO-DA, PFBS, PFHxS, PFNA, PFOA, and PFOS are presented in this table.

2 The Groundwater RSL is cited from the USEPA Regional Screening Levels and calculated with the USEPA RSL Calculator based on a hazard quotient of 0.1 (USEPA, 2022)

3 HFPO-DA is commonly referred to as GenX

-- = No applicable screening criteria

Bolding indicates analyte was detected

Shading indicates exceedance of screening criteria

STP1 = Sewage Treatment Plant #1

EPA = United States Environmental Protection Agency

ft bls = feet below land surface

NA = Not Applicable; compound not analyzed

PFAS = per- and polyfluoroalkyl substances

I = Estimated result < Limit of Quantitation and ≥ Detection Limit

U = Analyte was not detected

Table A-4. Historical Surface Water Analytical Results

Location ID	CAS No.	Screening Criteria <sup>1</sup>	PFAS-SW0010	PFAS-SW0051	PFAS-SW0051	PFAS-SW0052	PFAS-SW0053	PFAS-SW0054	PFAS-SW0055	PFAS-SW0056
Date			3/11/2019	1/27/2021	1/27/2021	1/27/2021	9/28/2020	9/28/2020	9/28/2020	9/28/2020
Sample Depth (ft bls)			0 - 0.5	0 - 0.5	0 - 0.5*	0 - 0.5	0 - 0.5	0 - 0.5	0 - 0.5	0 - 0.5
<b>PFAS with Screening Criteria (ng/L)</b>										
Perfluoro-n-octanoic acid (PFOA)	335-67-1	500	65 M	4.4	4.3	64.1	3.3 I	12.9	6.7	8
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	10	520 D	45.1	39.3	347	53.1	131	45.3	57.5
<b>PFAS without Screening Criteria (ng/L)</b>										
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>2</sup>	13252-13-6	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	9.4 U	8.3 U	8.3 U	8.3 U	8.3 U	7.7 U	8.3 U	7.7 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	9.4 U	8.3 U	8.3 U	8.3 U	8.3 U	7.7 U	8.3 U	7.7 U
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-butanesulfonic acid (PFBS)	375-73-5	--	27 M	3.6 I	4.1 I	48.5	4.6	1.9 I	4.2 U	2.4 I
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	0.93 J	4.2 U	4.2 U	4.2 U	4.2 U	3.8 U	4.2 U	3.8 U
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	1.4 U	4.2 U	4.2 U	4.2 U	4.2 U	3.8 U	4.2 U	3.8 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	35	2.7 I	3.3 I	71.4	5.4	10.7	5.9	6.5
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	76	3.1 I	3.6 I	127	6.6	10.7	6	11.8
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	--	190	19.3	18	284	23.3	10.1	7.9	9
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	--	6.4	4.2 U	4.2 U	3.9 I	4.2 U	2.8 I	4.2 U	2.4 I
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	2.8 U	4.2 U	21 U	4.2 U	4.2 U	3.8 U	4.2 U	3.8 U
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	2.8 U	4.2 U	4.2 U	4.2 U	4.2 U	3.8 U	4.2 U	3.8 U
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	1.4 U	4.2 U	4.2 U	4.2 U	4.2 U	3.8 U	4.2 U	3.8 U

**Table A-4. Historical Surface Water Analytical Results**

Location ID	PFAS-SW0057	PFAS-SW0058	PFAS-SW0133	S014-SW0001
Date	9/28/2020	9/28/2020	8/4/2021	2/22/2022
Sample Depth (ft bls)	0 - 0.5	0 - 0.5	0 - 0.5	0 - 0.5
<b>PFAS with Screening Criteria (ng/L)</b>				
Perfluoro-n-octanoic acid (PFOA)	93.6	3.1 I	16.6	11.7
Perfluorooctanesulfonic acid (PFOS)	2280	19.1	158	239
<b>PFAS without Screening Criteria (ng/L)</b>				
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	NA	NA	NA	16 U
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	NA	NA	NA	16 U
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	NA	NA	NA	16 U
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	NA	NA	16 U	NA
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	NA	NA	16 U	NA
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	NA	NA	16 U	NA
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>2</sup>	NA	NA	8 U	16 U
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	NA	NA	NA	NA
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	7.7 U	7.7 U	8 U	16 U
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	7.7 U	7.7 U	8 U	16 U
Perfluoro-n-butanoic acid (PFBA)	NA	NA	NA	13.7 J
Perfluoro-1-butanefulfonic acid (PFBS)	15.6	3.8 U	14.1	8.7
Perfluoro-1-decanesulfonic acid (PFDS)	NA	NA	NA	8 U
Perfluoro-n-decanoic acid (PFDA)	4.1	3.8 U	4 U	8 U
Perfluoro-n-dodecanoic acid (PFDoA)	3.8 U	3.8 U	4 U	8 U
Perfluoro-1-heptanesulfonic acid (PFHpS)	NA	NA	NA	4.5 J
Perfluoro-n-heptanoic acid (PFHpA)	65.8	2.1 I	20.8	11.5
Perfluoro-n-hexanoic acid (PFHxA)	113	6.1	33.8	20.2
Perfluorohexanesulfonic acid (PFHxS)	520	3.2 I	99.6	94.4
Perfluoro-n-nonanoic acid (PFNA)	19	3.8 U	2.6 I	2.3 J
Perfluoro-1-nonanesulfonic acid (PFNS)	NA	NA	NA	8 U
Perfluorooctane sulfonamide (PFOSA)	NA	NA	NA	40 U
Perfluoro-1-pentanesulfonic acid (PFPeS)	NA	NA	NA	9.4
Perfluoro-n-pentanoic acid (PFPeA)	NA	NA	NA	22.8
Perfluoro-n-tetradecanoic acid (PFTeDA)	3.8 U	3.8 U	4 U	8 U
Perfluoro-n-tridecanoic acid (PFTrDA)	3.8 U	3.8 U	4 U	8 U
Perfluoro-n-undecanoic acid (PFUdA)	3.8 U	3.8 U	4 U	8 U

All results reported in nanogram per liter (ng/L)

1 The State of Florida human health Surface Water Screening Levels for PFOA and PFOS are presented in this table.

2 HFPO-DA is commonly referred to as GenX

\* Duplicate sample results are included in this table and labeled with asterisk

-- = No applicable screening criteria

Bolding indicates analyte was detected

Shading indicates exceedance of screening criteria

STP1 = Sewage Treatment Plant #1

ft bls = feet below land surface

NA = Not Applicable; compound not analyzed

PFAS = per- and polyfluoroalkyl substances

I / J = Estimated result < Limit of Quantitation and ≥ Detection Limit

M = Presence of material is verified by not quantified

U = Analyte was not detected

Table A-5. Historical Sediment Analytical Results

Location ID (PFAS-) Date Sample Depth (ft bls)	CAS No.	Screening Criteria	SD0020	SD0021		SD0022	SD0023	SD0024		SD0025	SD0026	SD0027	SD0037	
			1/27/2021	1/27/2021	1/27/2021	9/28/2020	9/28/2020	9/28/2020	9/28/2020	9/28/2020	9/28/2020	9/28/2020	9/28/2020	9/28/2020
			0 - 1	0 - 1	0 - 1*	0 - 1	0 - 1	0 - 1	0 - 1*	0 - 1	0 - 1	0 - 1	0 - 1	
<b>PFAS without Screening Criteria (µg/kg)</b>														
1H,1H,2H,2H-perfluorohexane sulfonic acid (4:2 FTS)	757124-72-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
1H, 1H, 2H, 2H-perfluorooctane sulfonic acid (6:2 FTS)	27619-97-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
1H, 1H, 2H, 2H-perfluorodecane sulfonic acid (8:2 FTS)	39108-34-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9CL-PF3ONS)	756426-58-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CL-PF3OUDS)	763051-92-9	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX] <sup>1</sup>	13252-13-6	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
N-ethylperfluoro-1-octanesulfonamide (EtFOSA)	4151-50-2	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
N-ethylperfluoro-1-octanesulfonamidoacetic acid (EtFOSAA)	2991-50-6	--	7 U	1.6 U	1.6 U	1 U	1.7 U	1.4 U	1.2 U	1.1 U	1 U	1.3 U	1.1 U	
N-methylperfluoro-1-octanesulfonamidoacetic acid (MeFOSAA)	2355-31-9	--	7 U	1.6 U	1.6 U	1 U	1.7 U	1.4 U	1.2 U	1.1 U	1 U	1.3 U	1.1 U	
Perfluoro-n-butanoic acid (PFBA)	375-22-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-1-butanefluoronic acid (PFBS)	375-73-5	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-1-decanesulfonic acid (PFDS)	335-77-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-decanoic acid (PFDA)	335-76-2	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-n-dodecanoic acid (PFDoA)	307-55-1	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-1-heptanesulfonic acid (PFHpS)	375-92-8	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-heptanoic acid (PFHpA)	375-85-9	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-n-hexanoic acid (PFHxA)	307-24-4	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluorohexanesulfonic acid (PFHxS)	355-46-4	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	<b>0.36 I</b>	0.67 U	0.57 U	
Perfluoro-n-nonanoic acid (PFNA)	375-95-1	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-1-nonanesulfonic acid (PFNS)	68259-12-1	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-octanoic acid (PFOA)	335-67-1	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluorooctanesulfonic acid (PFOS)	1763-23-1	--	3.5 U	<b>0.41 I</b>	<b>0.51 I</b>	0.51 U	<b>2.7</b>	0.69 U	<b>0.59</b>	<b>0.78</b>	<b>8.6</b>	<b>0.56 I</b>	<b>2.1</b>	
Perfluorooctane sulfonamide (PFOSA)	754-91-6	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-1-pentanesulfonic acid (PFPeS)	2706-91-4	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-pentanoic acid (PFPeA)	2706-90-3	--	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
Perfluoro-n-tetradecanoic acid (PFTeDA)	376-06-7	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-n-tridecanoic acid (PFTrDA)	72629-94-8	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	
Perfluoro-n-undecanoic acid (PFUdA)	2058-94-8	--	3.5 U	0.82 U	0.81 U	0.51 U	0.84 U	0.69 U	0.58 U	0.55 U	0.51 U	0.67 U	0.57 U	

All results reported in microgram per kilogram (µg/kg)

<sup>1</sup> HFPO-DA is commonly referred to as GenX

-- = No applicable screening criteria

Bolding indicates analyte was detected

USEPA = United States Environmental Protection Agency

ft bls = feet below land surface

NA = Not Applicable; compound not analyzed

PFAS = per- and polyfluoroalkyl substances

I = Estimated result < Limit of Quantitation and ≥ Detection Limit

U = Analyte was not detected

**APPENDIX B**

**FIELD DOCUMENTATION**

**(Included in Electronic Copy Only)**



# STANDARD OPERATING PROCEDURE

Number	SA-1.8	Page	1 of 15
Effective Date	10/12/2020	Revision	6
Applicability	EGS Operating Unit		
Prepared	Earth Sciences Department		

Subject  
 SAMPLE ACQUISITION FOR PERFLUOROALKYL  
 AND POLYFLUOROALKYL SUBSTANCES (PFAS)  
 ANALYSIS

Approved  
 T. Johnston *T.E. Johnston* 10/12/2020

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Subject SAMPLE ACQUISITION FOR PERFLUOROALKYL AND POLYFLUOROALKYL SUBSTANCES (PFAS) ANALYSIS	Number SA-1.8	Page 2 of 15
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## 1.0 PURPOSE

This Standard Operating Procedure (SOP) describes the methods and protocols to be used for collecting and handling samples to be analyzed for per- and polyfluoroalkyl substances (PFAS). PFAS are present in many consumer products including some typical sampling equipment and are widely present in the environment.

Low screening criteria and high cross-contamination potential require special precautions to be implemented to avoid compromising sample integrity. Instructions are provided herein for collection of environmental samples without contaminating them. This SOP is designed to supplement but not replace existing sampling SOPs SA-1.1, SA-1.2, SA-1.3, SA-1.7, and SA-5.1. In addition, some clients and/or projects may have specific PFAS-related sampling requirements that extend beyond the procedures described in this SOP. Such additional requirements typically are documented in work plans or similar documents.

## 2.0 SCOPE AND APPLICABILITY

This document provides information on selection of proper sampling equipment and techniques for groundwater, surface water, sediment, soil, and water supply sampling for PFAS analysis. Sampling of air or biota is not addressed in this SOP, but similar principles would apply for those media.

## 3.0 BACKGROUND

PFAS have been used since the 1940s as manufacturer-applied oil and water repellants on products such as clothing, upholstery, paper, and carpets and in making fluoropolymers for non-stick cookware. They are found in textiles and leather products, mist suppressants for metal plating, materials used in the photography industry, photolithography, semi-conductors, paper and packaging, coatings, cleaning products, pesticides, and cosmetics. They have been used in well-known consumer products including Teflon, StainMaster, Scotchgard, and GoreTex. In the 1960s, aqueous film-forming foam (AFFF) containing PFASs was developed for fighting flammable liquid fires, particularly petroleum-fueled (Class B) fires (ATSDR, 2009). The two most researched and most prevalent PFAS in the environment are perfluorooctane sulfonic acid (PFOS) and perfluorooctanoic acid (PFOA) (ATSDR, 2009).

Military uses of PFAS have been primarily related to fire fighting and electroplating. AFFF meeting MIL-F-24385 specifications was developed by various manufacturers for use in extinguishing fires at military bases, airports, commercial facilities, and fire-fighting training facilities throughout the United States. Beginning in the late 1960s the United States Department of Defense (DoD) used large quantities of AFFF for shipboard and shore facility fire-suppression systems, on fire-fighting vehicles, and at fire-training facilities. AFFF concentrate that contains PFAS may still be in use at DoD facilities, and large quantities of AFFF may have been released to the environment at some facilities.

PFAS are persistent in the environment, tend to bioaccumulate, and demonstrate toxicity in laboratory animals, enough to raise concerns about their presence in the environment. Some areas where PFAS may have been released to the environment include the following:

- Fire-fighting training areas
- Areas where fire-fighting products/materials are stored (e.g., fire stations)
- Aircraft crash sites
- Refineries
- DoD sites/military bases

Subject SAMPLE ACQUISITION FOR PERFLUOROALKYL AND POLYFLUOROALKYL SUBSTANCES (PFAS) ANALYSIS	Number SA-1.8	Page 3 of 15
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- Landfills (leaching from consumer products)
- Biosolids land applications
- Rail yards
- Textile/carpet manufacturing sites
- Septic systems
- Metal coating and plating facilities
- Water treatment systems and receiving water bodies
- Airport hangars and other facilities storing fire-fighting foams
- Chemical facilities, especially fluorochemical manufacturing, use, and disposal facilities

PFAS are ubiquitous in consumer products and are present in or on some materials used in environmental sampling (e.g., Teflon tubing, waterproof logbooks, and GoreTex field clothing). Laboratory detection limits are low for PFAS, and contact of sample material or sampling equipment with any one of the multitude of PFAS sources could result in detectable contamination. In addition, PFAS tend to adsorb to glass and some plastics, so certain glass or plastic sample collection containers are inappropriate for use in PFAS sample collection. Adsorption to sample containers or other materials (e.g., tubes or hoses) may result in a low bias for measured PFAS concentrations.

Collection and analysis of quality control blanks is an important aspect of verifying that samples have not been contaminated during sample collection and handling. Use of additional blanks or blanks of a different type than usual may be required, and the governing project planning documents should be consulted. Consult Section 7.7 of this SOP for instructions regarding collection of field reagent blanks (FRBs).

#### 4.0 DEFINITIONS

AFFF – Aqueous film-forming foam.

Chemical of Emerging Concern – A chemical of emerging concern (previously called “emerging contaminant” and also called “contaminant of emerging concern” or “CEC”) is a chemical or material that is characterized by a perceived, potential, or real threat to human health or the environment or by a lack of published health standards (U.S. EPA, 2014; U.S. Geological Service [USGS], 2019). A contaminant may also be of “emerging concern” because a new source or a new pathway for human exposure has been discovered or a new detection method or treatment technology has been developed (DoD, 2011).

FRB – Field Reagent Blank. A blank sample prepared in the field by transferring laboratory-supplied, chemically preserved, “PFAS-free” deionized water to an empty, laboratory-supplied, collection bottle. FRBs are typically analyzed only for PFAS and are treated as site samples in all respects, including shipment to the sampling site, exposure to sampling site conditions, storage, preservation, and all PFAS analytical procedures. The purpose of FRBs is to indicate whether PFAS measured in corresponding site samples may have been introduced during sample preservation, collection, and handling. Note that the term “FRB” may be inappropriate when sampling groundwater sources not used for drinking water. In those cases Field Blank (FB) may be more appropriate. Consult project-specific planning documents for direction.

PFAS – Per- and Polyfluoroalkyl Substances. A reference term currently in use, replacing “PFCs” in recent scientific and other technical literature. The term is inclusive of both perfluorinated chemicals like PFOA and PFOS and polyfluoroalkyl substances like fluorinated telomers. In all environmental matrices, these chemicals are CECs.

PFCs – Perfluorinated Compounds or Chemicals. PFCs are a family of man-made chemicals that have been used for commercial, industrial, and military applications because they resist thermal degradation and repel oil, stains, grease, and water.

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PFOA – Perfluorooctanoic Acid. PFOA is used as an aqueous dispersion agent (surfactant) and in the manufacture of fluoropolymers (including Teflon) used in industrial components such as electrical wire casings, fire- and chemical-resistant tubing, and plumbing seal tape. PFOA is used in surface treatment products (e.g., paints) to impart oil, stain, grease, and water resistance. PFOA can also be produced by the breakdown of some fluorinated telomers.

PFOS – Perfluorooctane Sulfonic Acid. PFOS was a key ingredient in Scotchgard and used in the manufacture of Class B AFFF used per DoD military specifications. Phase out of AFFF by 3M occurred in 2002.

**5.0 SAFETY PRECAUTIONS**

Sample acquisition activities shall be conducted in accordance health and safety requirements identified in the project-specific Health and Safety Plan (HASP), Accident Prevention Plan (APP), and corporate health and safety policies. Alteration may be necessary to allow sample collection without cross contamination as dictated by site-specific conditions.

**Caution**

The use of personal protective equipment (PPE) containing PFAS (e.g., some insect repellants, sunscreens, traffic safety vests, etc.) should be avoided if possible or, if deemed necessary to control hazards, should be carefully considered as they can pose a potential cross-contamination risk for samples. Extra care (e.g., changing outer gloves) must be exercised to ensure that PFAS is not transferred directly or indirectly from PPE to samples or sample containers.

The Tetra Tech Project Manager (PM), in coordination with the Tetra Tech EGS Operating Unit Health and Safety Group, shall ensure that the development of project-specific plans balances the need to control exposure to safety hazards as well as address PFAS contamination risks.

**6.0 PERSONNEL RESPONSIBILITIES, QUALIFICATIONS, AND TRAINING**

Personnel implementing this SOP must read and understand this entire SOP prior to collection of samples designated for PFAS analysis.

Project Manager (PM) – The PM along with the management team are responsible for determining sampling objectives, initial sampling locations, and field procedures used in the collection of samples of environmental media. Additionally, in consultation with other project personnel (geologist, hydrogeologist, etc.), the PM is responsible for selecting and detailing the specific sampling techniques and equipment to be used and for providing detailed input in this regard to the project planning documents. The PM has the overall responsibility for ensuring that sampling activities are properly conducted by appropriately trained staff.

Site Safety and Health Officer (SSHO) – The SSHO (or a qualified designee) is responsible for providing the technical support necessary to implement the project HASP, APP, or equivalent. The SSHO or SSHO designee may also be required to advise the Field Operations Leader (FOL) on safety-related matters regarding sampling, such as measures to mitigate potential hazards, hazardous objects, or conditions.

Field Operations Leader (FOL) – This individual is primarily responsible for faithful execution of the planning document that governs sampling. This is accomplished through management of a field sampling team for the proper acquisition of samples. The FOL is responsible for the supervision of onsite activities; ensuring proper instrument calibration, care, and maintenance; sample collection and handling; the completion and

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accuracy of all field documentation; and making sure that custody of all samples obtained is maintained according to sample collection protocols. When appropriate and as directed by the FOL, such responsibilities may be performed by other qualified personnel (e.g., field support personnel) where credentials and time permit. The FOL is ultimately responsible for ensuring compliance with Occupational Safety and Health Administration (OSHA) regulations during these operations in accordance with the project Health and Safety Plan. The FOL person shall have significant hands-on experience with sample collection, handling, and documentation under the circumstances required for the current project.

Field Support Personnel – These individuals are responsible for the proper acquisition of samples in accordance with this SOP or other project-specific documents. In addition, these personnel are responsible for completion of all required paperwork (e.g., sample log sheets, field notebook, boring logs, container labels, custody seals, and chain-of-custody forms) associated with the collection of the samples.

General field personnel qualifications for groundwater sample collection and onsite water quality testing include the following:

- OSHA 40-hour; applicable refresher training; and, if applicable, supervisory training (HAZWOPER operations only).
- Capability of performing field work under the expected physical and environmental (i.e., weather) conditions.
- Familiarity with sampling procedures, sample handling, sample documentation, and sample packaging and shipping as documented in project specific planning documents and this SOP.

## 7.0 PROCEDURE

The following sampling procedure establishes requirements for collection of samples designated for PFAS analysis while minimizing potential cross contamination of the samples and other materials or loss of PFAS.

### 7.1 Selection of Equipment

It is important to research available equipment and materials at the project planning stage to avoid last-minute problems in the field, for example, ensuring compatibility of high-density polyethylene (HDPE) tubing with fittings for use in a pump, or ensuring that equipment (e.g., a bladder pump) does not contain Teflon.

#### 7.1.1 Sampling Equipment:

**NOTE:** PFAS cross-contamination of groundwater samples can be minimized through decontamination or conditioning of equipment left in a well. Use of dedicated equipment also is helpful so that handling during decontamination is unnecessary. Pre-labeling sample bottles before sample collection minimizes the number of glove changes required to prevent contamination of samples by PFAS that may be on the labels.

- Decontamination – All reusable equipment used in sample acquisition should be adequately decontaminated prior to use. Consult Safety Data Sheets (SDSs) to verify that soaps or detergents used in decontamination do not contain fluorosurfactants. Be aware that low levels of contaminants such as PFAS may be present in a product but not listed on the SDS.
- Unless project requirements indicate otherwise, use sampling equipment made of stainless steel, acetate, silicone, or HDPE. This applies to tubing, pumps and pump components, tape for plumbing

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fittings, trowels, mixing bowls, or other equipment that could contact the sample media. Gasket and O-ring components of sampling equipment may also contain fluoropolymers.

**NOTE:** PFAS on purchased or rented items is likely to occur predominantly in newly manufactured or rented items treated with chemicals containing PFAS. Therefore, all rental equipment that will make direct contact with the material being sampled must be thoroughly decontaminated prior to use, especially if the equipment items are new. Be cognizant of the potential for continued leaching of PFAS or other chemicals — even after decontamination.

- During sample handling, mobilization, and demobilization, avoid using sampling equipment that includes or contains polyvinylidene fluoride (PVDF) or contains “fluor” in the name such as:
  - Polytetrafluoroethylene (PTFE)
  - Teflon (DuPont brand name for PTFE)
  - Fluorinated ethylene propylene (FEP)
  - Ethylene tetrafluoroethylene (ETFE).
- Use products that are not made of low-density polyethylene products (LDPE) if contamination from those products can be transferred to environmental samples or QC samples.
- For collecting drinking water samples to be analyzed for PFAS, use polypropylene sample bottles with a polypropylene screw cap; for all other samples, use HDPE containers with unlined plastic screw caps.

#### 7.1.2 Non-Sampling Field Equipment:

- Non-waterproof loose-leaf paper or notebooks are acceptable. Avoid using waterproof field books or paper during sampling activities. Do not use plastic clipboards, binders, or spiral hard-cover notebooks that may be coated; use Masonite or aluminum clipboards instead.
- Use ballpoint pens or pencils for note taking and sample bottle labeling.
- Avoid using:
  - Post-it notes or similar removable notes.
  - Sharpies or similar indelible markers.
  - Aluminum foil.

#### 7.1.3 Field Personnel Clothing and Protective Gear:

- Wear clothing that has been washed at least six times without fabric softener to remove possible stain-resistant coatings. Clothing made of natural fibers such as cotton is preferred to other fabrics. Protective clothing must be washed in accordance with manufacturer recommendations to ensure that the protective properties necessary to control safety hazards (e.g., fire-retardant clothing) are not compromised.
- Wear non-powdered nitrile gloves at all times while collecting and handling samples, and change gloves often. Anecdotal evidence indicates that changing gloves is one of the most effective methods of reducing or eliminating sample contamination potential; therefore, change to a new pair of gloves prior to collecting each sample.
- As necessary, use sunscreens and insect repellants that are made with 100-percent natural ingredients and that the Air Force Civil Engineer Center has identified as acceptable for use. ***These products must be used in accordance with manufacturer recommendations and in combination with***

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***controls in the project-specific HASP, APP, and corporate health and safety policies. Multiple re-applications of these products per work shift may be required to ensure their effectiveness.***

- Sunscreens: Alba Organics Natural Sunscreen, Yes To Cucumbers, Aubrey Organics, Jason Natural Sun Block, Kiss My Face, baby sunscreens that are “free” or “natural.”
- Sunscreen and insect repellent: Avon Skin So Soft Bug Guard Plus – SPF 30 Lotion
- Insect Repellent: Jason Natural Quit Bugging Me, Repel Lemon Eucalyptus Insect Repellent, Herbal Armor, California Baby Natural Bug Spray, BabyGanics.

**NOTE:** The suitability of these items has not been independently verified. Products containing N-diethyl-meta-toluamide (DEET), picaridin, and IR3535 and some oil of lemon eucalyptus (OLE) and para-menthane-diol products are known to provide longer-lasting protection than others. One of the products recommended by the Air Force Civil Engineer Center and listed above, Repel Lemon Eucalyptus Insect Repellent, contains OLE and is most likely to be effective.

An independent study (Bartlett and Davis, 2018) of three insect repellents (Sawyer do-it-yourself permethrin treatment for clothing, Off! Deep Woods spray for clothing/skin, and Insect Shield pretreated clothing) determined that these products were free of PFAS compounds (17 PFAS compounds were tested), thus they may be suitable for use on a project-specific basis.

- During wet weather, use rain gear made from polyurethane or wax-coated materials.
- Avoid unnecessary contact with upholstery in vehicles because many such fabrics may be treated with stain-resistant materials that could contain PFAS. Typically, rental vehicles are newer and more likely to pose a contamination risk to samples. Well-washed towels or rags may be placed on the seats to prevent contact with car seats and other materials that could transfer PFAS to clothing worn by samplers. If practical, cover clothing and skin that has been in contact with such upholstery with non-fluorinated clothing.
- Avoid wearing:
  - Water-resistant (e.g., Gore-Tex or similar material) clothing or footwear (e.g., boots) immediately prior to or during sample collection and management.
  - Coated Tyvek or similar coated PPE suits.
  - Cosmetics, shampoos, hair conditioners, moisturizers, hand cream, or other similar personal care products on the day of sampling.

#### **7.1.4 Sample Containers and Shipping Materials:**

- Collect samples in clean, laboratory-supplied, plastic bottles only, typically polypropylene for drinking water or HDPE for other matrices.
- Confirm that Teflon-lined caps are not used in sample containers; unlined polypropylene screw caps must be used. It is best to segregate sample containers with Teflon components (e.g., Teflon-lined septa) from PFAS sample containers.

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- Use of commercially available plastic bags (e.g., 3-mil-thick trash can liners) for lining coolers to prevent leakage and to separate potential melt water from chain-of-custody forms is allowed.
- Avoid the use of glass or LDPE sample containers, which are believed to result in loss of PFAS through adsorption to the container inner walls.
- Avoid using Blue Ice or similar items to cool samples and avoid placing such items in sample coolers for shipping. Use commercially available (e.g., from convenience stores or supermarkets) double-bagged ice instead.

### **Caution**

Samples designated for PFAS analysis must be cooled to achieve a storage temperature of less than 6 °C. Cooling to this temperature may take several hours, and sample temperatures may not achieve 6 °C by the time they arrive at the laboratory. If sample temperatures upon arrival at the laboratory are not less than 10 °C, the laboratory may conclude that the sample integrity is compromised and may reject the samples. Therefore, place samples on ice as soon after collection as possible. On warm days, or when a representative from a nearby laboratory picks up the samples, take extra care (e.g., use more ice or delay shipment, if necessary) to ensure that sample temperatures will not exceed 10 °C when the samples arrive at the laboratory.

## **7.2 Other Precautions for Sample Handling**

- Wash hands thoroughly before sampling and after handling fast food, carryout food, snacks, food wrappers, or other items that may contain PFAS. Do not carry pre-packaged food items such as candy bars or microwave popcorn into sampling areas.
- Assume that shipping tape used for securing coolers could contain PFAS; therefore, take care not to transfer PFAS from tape to samples.
- Minimize exposure of samples to light. This can be done by placing the collected samples into a cooler (with ice) and closing the cooler lid.
- If in doubt about a particular product or item that comes into contact with environmental media to be sampled or is near to sampling operations, consider collecting and analyzing a rinsate blank using laboratory-supplied PFAS-free water to test the item for contamination potential. Consult the Tetra Tech PM in these cases to verify whether collection of additional blanks is warranted.
- Support personnel that are within 3 meters of the sample processing area are considered subject to the same restrictions related to precautionary measures for clothing and food as applied to sampling personnel.

These precautions must be observed during sampling activities, especially during water sample collection (groundwater, water supply, and surface water), given the high solubilities of PFAS in water. Examples of how these precautions may be applied to sampling of specific media are provided in the following sections.

## **7.3 Groundwater Sample Acquisition**

The precautions and requirements identified in Sections 7.1 and 7.2 must be observed for groundwater sampling. Do not proceed any further without reviewing each of those precautions and requirements.

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- Collect groundwater samples for PFAS analyses in accordance with this SOP, SOP SA-1.1, and/or project- or client-specific requirements.
- If non-dedicated non-disposable equipment is used between sampling locations, it should be decontaminated with Alconox or Liquinox, unless 1,4-dioxane (a potential component of Liquinox) is also a contaminant of concern. In that case, Liquinox should not be used. Products such as Decon 90, which contains fluorosurfactants, shall not be used. Alconox or Liquinox rinses should be followed by a potable water rinse then deionized water rinse. The final decontamination rinse for DoD projects must be with water certified by the laboratory to be PFAS-free.
- If sampling for multiple analytes using PFAS-appropriate equipment, collect samples for PFAS analysis last to ensure adequate purging and conditioning of sampling equipment. If practical to do so, suitable PPE (especially gloves) may also be changed out for PFAS sampling. For example, purge and sample a monitoring well for volatile organic compounds (VOCs), semivolatile organic compounds (SVOCs), and metals using a peristaltic pump with HDPE and silicone tubing, then collect the sample for PFAS analysis. If either the proper sampling sequence or proper equipment is unclear, consult the FOL or Tetra Tech PM and record the actual sequence in the field notes.
- If sampling wells that have or had dedicated Teflon or FEP tubing that potentially contained PFAS, remove the dedicated tubing and, using silicone or HDPE tubing, remove at least one well volume from the target sampling interval prior to sampling. Accomplish this removal in a manner that is rigorous enough to remove the entire water column from the well and not just a limited vertical interval of the water column. This will minimize the potential for collecting a sample that was in contact with the Teflon/FEP tubing.
- The use of detergents must be avoided during decontamination of drilling or other heavy equipment. All equipment must be scrubbed with a plastic brush or steam cleaned with potable water, and rinsed thoroughly in potable water to clean away any debris or material on exposed surfaces.
- Sample(s) representing any water collected at the point of use (e.g., a water truck or tank on site) used by the driller for drilling purposes may require analysis for PFAS. Check the governing planning document for guidance and see Section 7.8 of this SOP for guidance on waste management.
- Collect drinking water samples to be analyzed for PFAS in clean polypropylene sample bottles with a polypropylene screw cap; for all other samples, use clean, laboratory-supplied, HDPE bottles with unlined plastic screw caps.
- Pre-label all sample bottles with the correct sample identifier prior to sampling and, for each bottle at the time of sampling:
  - Verify that the correct bottle is being used.
  - Fill the bottle to slightly below the shoulder of the bottle.
  - Tightly close the bottle with the appropriate cap.
  - Repeat this process for subsequent sample bottles.



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#### 7.4 **Soil Sample Acquisition**

The precautions and requirements identified in Sections 7.1 and 7.2 must be observed for soil sampling. Do not proceed any further without reviewing each of those precautions and requirements.

- Collect soil samples for PFAS analyses in accordance with this SOP, SOP SA-1.3, and/or project- or client-specific requirements. Review client-specific (e.g., DoD component) guidance or previously approved Sampling and Analysis Plans (SAPs).
- Soil sampling equipment should not be constructed of or contain Teflon or other materials likely to contain or be coated with PFAS. Acceptable materials for sampling include stainless steel, acetate plastic, and HDPE.
- If non-dedicated non-disposable equipment is used between sampling locations, it should be decontaminated with Alconox or Liquinox, unless 1,4-dioxane (a potential component of Liquinox) is also a contaminant of concern. In that case, Liquinox should not be used. Products such as Decon 90, which contains fluorosurfactants, shall not be used. Alconox or Liquinox rinses should be followed by a potable water rinse then deionized water rinse. The final decontamination rinse for DoD projects must be with water certified by the laboratory to be PFAS-free.
- Collect samples in laboratory-provided HDPE containers specifically designated for PFAS analysis. Do not use glass jars typically used for soil sample collection because some PFAS may irreversibly adsorb to the glass and could create a negative bias (i.e., artificially low concentrations) in the measured PFAS concentrations.
- Pre-label all sample containers with the correct sample identifier prior to sampling and, for each container at the time of sampling:
  - Verify that the correct container is being used.
  - Fill the container to the appropriate level.
  - Tightly close the container with the proper cap.
  - Repeat this process for subsequent sample containers.

#### 7.5 **Surface Water and Sediment Sample Acquisition**

The precautions and requirements identified in Sections 7.1 and 7.2 must be observed for surface water and sediment sampling. Do not proceed any further without reviewing each of those precautions and requirements.

- Collect surface water and sediment samples for PFAS analysis in accordance with this SOP, SOP SA-1.2, and/or project- or client-specific requirements.
- Surface water and sediment sampling equipment should not be constructed of or contain Teflon or LDPE materials. Acceptable materials for sampling include HDPE, silicone, stainless steel, and acetate plastic. Do not use glass. The bottleware should be supplied clean by the laboratory and specifically designated for PFAS analysis. If transfer bottles are required for collection of surface water samples, the transfer bottles used should be of the same material as the containers designated for submission to the laboratory.

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- For surface water sample collection, invert the capped sample bottle, with the opening pointing downward, at least 10 cm below the water surface, at least 10 cm above the bottom of the water body, and as close to the center of the channel or water body as practical. To collect the sample, uncap the bottle underneath the water surface and point the bottle upward so that gloved hands, sample container, and sampler are downstream of where the sample is being collected.
- For aquatic samples collected from the shoreline or via wading, ensure that waders are constructed of fabric that has not been treated with waterproofing coatings, and stand downstream of the sample bottle during sample collection.
- Pre-label all sample bottles with the correct sample identifier prior to sampling and, for each bottle at the time of sampling:
  - Verify that the correct container is being used.
  - Fill the container to the appropriate level.
  - Tightly close the container with the proper cap.
  - Repeat this process for subsequent sample containers.
- If non-dedicated non-disposable sampling equipment is used between sampling locations, it should be decontaminated with Alconox or Liquinox, unless 1,4-dioxane (a potential component of Liquinox) is also a contaminant of concern. In that case, Liquinox should not be used. Products such as Decon 90, which contains fluorosurfactants, shall not be used. Alconox or Liquinox rinses should be followed by a potable water rinse then deionized water rinse. The final decontamination rinse for DoD projects must be with water certified by the laboratory to be PFAS-free.
- Avoid reusing non-stainless-steel equipment (e.g., porewater observation devices consisting of slotted PVC pipe and silicone tubing) when collecting porewater samples.

## 7.6 Water Supply Sampling

This section applies to sampling from taps, spigots, faucets, or similar devices for PFAS analysis. The precautions and requirements identified in Sections 7.1 and 7.2 must be observed for water supply sampling. Do not proceed any further without reviewing each of those precautions and requirements.

### Caution

Do not use filters when collecting samples because the filters may introduce PFAS contamination or absorb/adsorb PFAS and thus reduce PFAS concentrations in the samples.

- Collect water supply samples for PFAS analysis in accordance with applicable portions of SOP SA-1.7 and/or project- or client-specific requirements.
- Water supply sampling equipment (if needed) should not be constructed of or contain Teflon or LDPE materials. Acceptable materials for sampling include HDPE, polypropylene (drinking water sampling only), small amounts of silicone (e.g., short runs of silicone tubing used in peristaltic pumps), stainless steel, and acetate plastic. Non-drinking water supply samples should be collected in clean, laboratory-supplied, HDPE bottleware specifically designated for PFAS analysis (not glass). Collect drinking water samples in clean polypropylene bottles supplied by the laboratory.
- Ensure that sample bottles used to collect chlorinated water samples contain the proper Trizma preservative (5 g/L to remove chlorine). Non-chlorinated water does not require chemical preservatives

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designed to remove chlorine; however, some projects may require the use of Trizma regardless of whether the water is chlorinated.

- Locate the sampling point. If a specific sampling point has already been designated (e.g., a kitchen tap or well head), plan to collect the sample from that point. Otherwise, identify a location in the water supply line that is as close as possible to the water's point of origination (e.g., a well or other water source) and upstream of any local water treatment unit(s) that could affect PFAS levels (e.g., water softeners, activated carbon, or reverse osmosis treatment units). If a treatment unit is in use, a post-treatment sample may also be required in some cases, per project requirements.

**NOTE:** If treatment that could affect PFAS levels (e.g., carbon filtration or reverse osmosis) is part of the water distribution system, often a spigot will be present in the plumbing line between the water source and the treatment unit, and this spigot should be used for sample collection. If no local treatment is applied to the water, a convenient sampling point for private water supplies is often the cold water tap located at the point of use. If a sample cannot be collected from the planned location, collect the sample from an alternate, but equally representative sampling point, document the deviation in the field notes, and alert the project manager.

- Remove any aerator/diffuser from the faucet, if possible. If removal is not possible, record this observation in the field notes.
- Allow the water to run freely from the tap until project-specific requirements are achieved. This will often require purging for 3 to 5 minutes or may require monitoring of water quality parameters (e.g., temperature) until they stabilize.
- Reduce the water flow rate to minimize aeration of the sample. The water stream should be no wider than the diameter of a pencil.
- Pre-label all sample bottles with the correct sample identifier prior to sampling and, for each bottle at the time of sampling:
  - Verify that the correct bottle is being used
  - Fill the sample bottle directly from the tap to a point slightly below the shoulder of the bottle.
  - Cap the bottle tightly with the proper cap.
  - Repeat this process for subsequent sample bottles.
- After collecting the sample, ensure the bottle is capped tightly and, if solid chemical preservative is included, agitate by hand until the preservative is dissolved.

## 7.7 **Field Reagent Blank Collection**

**NOTE:** If PFAS are detected in site samples, FRBs may be analyzed to assess whether PFAS in site samples could be non-site-related contamination and whether resampling is necessary. U.S. EPA Method 537.1, and modifications or derivatives thereof for PFAS analysis in drinking water typically require an FRB to be handled along with each sample set. A sample set is described as samples collected from the same sample site and at the same time, but "sample site" and "same time" may not be precisely defined. Therefore, it is important to verify that the correct number of FRBs will be collected. The intent is to be able to verify whether samples have been contaminated and to help identify the source of contamination. In general, collecting one FRB at each sampling point is recommended when sampling drinking water; fewer

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FRBs are recommended when sampling non-drinking water matrices. The actual number will depend on project needs. *Collection of an FRB at every non-drinking water sampling point **may** be required.*

- Verify the number of FRBs to be collected for the project and where those samples must be collected. This should be described in the governing project planning documents such as work plans or SAPs. If it is not, consult the PM. FRBs are commonly collected in pairs of sample bottles, similar to corresponding site samples.

**NOTE:** Chemical preservative may not be included in FRBs that are used when sampling non-drinking water matrices. In these cases the FRBs are more appropriately called Field Blanks (FBs) but the water transfers described below will be similar.

**NOTE:** It is important that FRB collection emulate as closely as possible the conditions surrounding collection of site samples. For example, when collecting FRBs from taps or spigots, collect each FRB either immediately before or after the corresponding site sample collection **without interrupting the water flow** until both the site sample and FRB have been collected.

- At the sampling site, when ready to collect an FRB, open the bottle of chemically preserved FRB reagent water provided by the laboratory and a corresponding clean empty bottle, also provided by the laboratory.
- Pour the preserved FRB reagent water into the empty sample bottle, close the cap, and label this filled bottle as the FRB.
- Pack and ship the FRB along with site samples and required documentation (e.g., chain-of-custody form) to the laboratory.

**NOTE:** Although chain-of-custody forms will indicate that FRBs must be analyzed for PFAS, analysis of an FRB will be required only if site samples contain PFAS greater than a certain concentration. If an FRB is analyzed and any PFAS concentration in the FRB exceeds one-third of the laboratory minimum reporting limit (or equivalent), all samples collected with that FRB may be considered invalid and may require recollection and reanalysis. Consult the project planning documents governing sample collection for specifics as to whether resampling is necessary. Care in collection and handling of site samples and FRBs in a way that avoids contamination cannot be overemphasized.

**NOTE:** It will be necessary to associate individual FRBs with corresponding site samples; otherwise, decisions about which samples to recollect (if recollection is indicated) could be compromised. Associations between FRBs and corresponding site samples may be accomplished by marking chain-of-custody forms with the associations, but other methods also may be useful. Consult the governing planning document or the PM for guidance, if necessary.

## 7.8 Disposal of Investigation-Derived Waste Potentially Containing PFAS

PFAS are not hazardous wastes as defined in the Resource Conservation and Recovery Act and Comprehensive Environmental Response, Compensation, and Liability Act. It may be possible to dispose of PFAS-containing solid waste as non-hazardous, but sampling solid waste material for PFAS analysis is not advised. Consult the client PM or on-site point of contact to verify their current disposal acceptance criteria and indicate on waste manifests that the waste potentially contains PFAS. Solid waste potentially containing PFAS should be identified as such on waste manifests. Wastewater potentially containing PFAS should be analyzed for PFAS to determine the appropriate disposal option. If the sum of PFOA and PFOS

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concentrations is less than the applicable criterion (e.g., 70 ng/L) the water may be disposed of without special handling if no other enforceable regulations apply; otherwise, the water should be treated to reduce the PFOA + PFOS concentration to an acceptable level or should be directed to an appropriate treatment facility for disposal. On-site treatment (e.g., granular activated carbon filtration) may be appropriate. Consult the client PM or on-site point of contact for direction regarding disposal.

**NOTE:** If aqueous investigation-derived waste (IDW) is expected to contain PFAS greater than the applicable criterion (e.g., captured residual AFFF or AFFF concentrate from an accidental release in a hangar), special actions may be necessary and the client PM should be consulted. For wastes that are dewatered and potentially contain PFAS, containerize the wastewater and analyze it for PFAS prior to disposal.

## 8.0 REFERENCES

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Bartlett and Davis, 2018. Evaluating PFAS cross contamination issues. Remediation, 2018;28:53-57.

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Department of the Navy, 2014. Perfluorinated Compounds (PFCs) Interim Guidance/Frequently Asked Questions (FAQs). December.

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NAVFAC, 2017. Interim Per-and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC Remedial Project Managers (RPMs)/September 2017 Update.

New Hampshire Department of Health and Human Services, 2014. Fact Sheet, Perfluorinated Chemicals (PFCs). May.

U.S. EPA (United States Environmental Protection Agency), 2009. Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS), Version 1.1, EPA/600/R-08/092. September.

U.S. EPA (United States Environmental Protection Agency), 2014. Emerging Contaminants – Perfluorooctane Sulfonate (PFOS) and Perfluorooctanoic Acid (PFOA). Emerging Contaminants Fact Sheet – PFOS and PFOA. EPA-50-F-14-001. March.

U.S. EPA, 2015. Perfluorooctanoic Acid (PFOA) and Fluorinated Telomers – Frequent Questions. <http://www.epa.gov/oppt/pfoa/pubs/faq.html>. Accessed April 27.

Subject SAMPLE ACQUISITION FOR PERFLUOROALKYL AND POLYFLUOROALKYL SUBSTANCES (PFAS) ANALYSIS	Number SA-1.8	Page 15 of 15
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USGS, 2019. Contaminants of Emerging Concern in the Environment,  
<https://toxics.usgs.gov/investigations/cec/index.php>. Accessed August 2.



# BORING LOG

PROJECT NAME: NASA PFAS  
 PROJECT NUMBER: 112 G09237  
 DRILLING COMPANY: Groundwater Protection  
 DRILLING RIG: Geoprobe 8146LS

BORING No.: STP1-SB0001  
 DATE: 12-9-2021  
 GEOLOGIST: Anderson, Rogers  
 DRILLER: Dave Leasing

Sample No. and Type or RQD	Depth (Ft.) or Run No.	Blows / 6" or RQD (%)	Sample Recovery / Sample Length	Lithology Change (Depth/Ft.) or Screened Interval	MATERIAL DESCRIPTION			U S C S *	Remarks	PID/FID Reading (ppm)									
					Soil Density/ Consistency or Rock Hardness	Color	Material Classification			Sample	Sampler BZ	Borehole**	Driller BZ**						
	↑																		
	①									Loose, Dark Brown to Black fine sands - organic rich - trace silt (some form but easy crumble) - Possibly finely laminated layers dark brown/black in lower 2-3 feet (7-10' BGS)									
	↓			10															
	↑									Similar to above but less black and organics. Generally a brown/red silt - trace to minor silts * less red & more brown with depth - laminated @ 16-19, gray & brown grades to gray fine sands									
	②																		
	↓			20															
	↑																		
	③									Gray, loose fine sands with minor shell frags. - crushed shells, <1-2 mm									
	↓			22	4B														
					12														

\* When rock coring, enter rock brokenness.

\*\* Include monitor reading in 6 foot intervals @ borehole. Increase reading frequency if elevated response read.

Remarks: \_\_\_\_\_

Drilling Area Background (ppm):

Converted to Well: Yes \_\_\_\_\_ No X Well I.D. #: \_\_\_\_\_



# BORING LOG

PROJECT NAME: NASA PFAS  
 PROJECT NUMBER: 112 G-09237  
 DRILLING COMPANY: Groundwater Protection  
 DRILLING RIG: Geoprobe 8140LS

BORING No.: STPI-SB001  
 DATE: 12-9-2021  
 GEOLOGIST: Anderson, Rogers  
 DRILLER: Dave Longino

Sample No. and Type or RQD	Depth (Ft.) or Run No.	Blows / 6" or RQD (%)	Sample Recovery / Sample Length	Lithology Change Depth/Ft. or Screened Interval	MATERIAL DESCRIPTION			U S C S *	Remarks	PID/FID Reading (ppm)				
					Soil Density/Consistency or Rock Hardness	Color	Material Classification			Sample	Sampler BZ	Borehole**	Driller BZ**	
	30			L2			Gray fine sand with shells - increasing shell content and sizes with depth							
	30						Gray, loose (very) f sands and crushed shells (about 1/2 size) - trace silts							
	40						- some larger shells > 2 inches							
	40			L3			- as above but some areas (2-3 inches) of clayey silts & shells							
	46			L4			Gray, dense plastic clay with trace shell, silts							
	50			L3B			Gray fine sands with clays and silts. more silt with depth, less sand. Some > 2" shells							

\* When rock coring, enter rock brokenness.

\*\* Include monitor reading in 6 foot intervals @ borehole. Increase reading frequency if elevated response read.

Remarks: \_\_\_\_\_

Drilling Area Background (ppm): Ø

Converted to Well: Yes \_\_\_\_\_ No X Well I.D. #: \_\_\_\_\_





# BORING LOG

PROJECT NAME: NASA PFAS  
 PROJECT NUMBER: 112609237  
 DRILLING COMPANY: Groundwater Protection  
 DRILLING RIG: Geoprobe 8140LS

BORING No.: STP1-SB0001  
 DATE: 12-9-2021  
 GEOLOGIST: Anderson, Roger  
 DRILLER: Dave Longino

Sample No. and Type or RQD	Depth (Ft.) or Run No.	Blows / 6" or RQD (%)	Sample Recovery / Sample Length	Lithology Change (Depth/Ft.) or Screened Interval	MATERIAL DESCRIPTION			U S C S *	Remarks	PID/FID Reading (ppm)					
					Soil Density/Consistency or Rock Hardness	Color	Material Classification			Sample	Sampler BZ	Borehole**	Driller BZ**		
	50			L3B			Gray loose silt with trace f sands & clay. - some portions clayey silt.								
	54						Light gray fine sandy shells to shelly sands. Increasing shell content with depth - trace clays & silts								
	60			L5			- fine sandy shells with trace silts - crushed shells, most < 2mm								
	66						Olive green clayey silt with shells (< 25%) - more silts than clay but similar								
	70						EOB								

\* When rock coring, enter rock brokenness.

\*\* Include monitor reading in 6 foot intervals @ borehole. Increase reading frequency if elevated response read.

Remarks: \_\_\_\_\_

Drilling Area Background (ppm): 0

Converted to Well: Yes \_\_\_\_\_ No X Well I.D. #: \_\_\_\_\_







Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: Sewage Treatment Plant Number 1 (STP1)
Project No.: 112G09581
Date: 02/21/2022

Sample ID: STP1-DPT0003
Location: 39
Sampled By: Chuck Sorden

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 7 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: PFAS QSM Table B-15, None, 2 250mL HDPE Bottles, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

MS/MSD Duplicate ID No.: STP1-FD-20220221-01 - DUP of STP1-DPT0003-010.0

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Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: Sewage Treatment Plant Number 1 (STP1)
Project No.: 112G09581
Date: 02/23/2022

Sample ID: STP1-DPT0009
Location: 29
Sampled By: Chuck Sorden

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 6 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: PFAS QSM Table B-15, None, 2 250mL HDPE Bottles, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No.: STP1-FD-20220223-02 - DUP of STP1-DPT0009-023.0

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Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: Sewage Treatment Plant Number 1 (STP1)
Project No.: 112G09581
Date: 02/24/2022

Sample ID: STP1-DPT0011
Location: 25
Sampled By: Chuck Sorden

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 6 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: PFAS QSM Table B-15, None, 2 250mL HDPE Bottles, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

MS/MSD Duplicate ID No.: STP1-FD-20220224-01 - DUP of STP1-DPT0011-042.0

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Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: Sewage Treatment Plant Number 1 (STP1)
Project No.: 112G09581
Date: 02/24/2022

Sample ID: STP1-DPT0012
Location: 26
Sampled By: Chuck Sorden

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 6 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: PFAS QSM Table B-15, None, 2 250mL HDPE Bottles, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No.: STP1-FD-20220224-02 - DUP of STP1-DPT0012-010.0

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Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: Sewage Treatment Plant Number 1 (STP1)
Project No.: 112G09581
Date: 02/24/2022

Sample ID: STP1-DPT0013
Location: 35
Sampled By: Chuck Sorden

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 6 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: PFAS QSM Table B-15, None, 2 250mL HDPE Bottles, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No.: STP1-FD-20220224-02 - DUP of STP1-DPT0012-010.0

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Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: Sewage Treatment Plant Number 1 (STP1)
Project No.: 112G09581
Date: 02/25/2022

Sample ID: STP1-DPT0015
Location: 37
Sampled By: Chuck Sorden

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 6 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: PFAS QSM Table B-15, None, 2 250mL HDPE Bottles, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

MS/MSD Duplicate ID No.:

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Project Site Name: KSC-SW3
Project No.: 112G09020
Date: 11/02/2021

Sample ID: SW3-DPT0143 (aka DPT0131)
Location: G
Sampled By: R. Seigel, K. Kercher

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 9 rows of sampling data for SW3-DPT0143 at various depths and times.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Contains 2 rows of collection information for VOCs and PFAS.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

MS/MSD Duplicate ID No.:



Project Site Name: KSC-POL
Project No.: 112G09020
Date: 10/28/2021

Sample ID: POL-DPT0136 (aka DPT1476)
Location: C
Sampled By: S. Rogers, K. Kercher

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 7 rows of sampling data for POL-DPT0136 (aka DPT1476) at location C, with depths ranging from 12' to 42'.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: SW-846 8260B VOCs, None, Two (2) - 40 mL vials delivered to Mobile Lab, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

MS/MSD Duplicate ID No.:



Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: KSC-POL
Project No.: 112G09020
Date: 10/26/2021 10/28/2021

Sample ID: POL-DPT0135 (aka DPT1475)
Location: A
Sampled By: R. Seigel, K. Kercher

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 10 rows of sampling data.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Contains 2 rows of collection information.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No.: (Empty cells for marking)



Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: KSC-POL
Project No.: 112G09020
Date: 10/26/2021

Sample ID: POL-DPT0134 (aka DPT1474)
Location: E
Sampled By: R. Seigel, K. Kercher

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 8 rows of sampling data for POL-DPT0134 (aka DPT1474) at location E, with depths ranging from 12' to 47'.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Row 1: SW-846 8260B VOCs, None, Two (2) - 40 mL vials delivered to Mobile Lab, X.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No. (with a large empty space for input).





Tetra Tech, Inc.

DPT GROUNDWATER SAMPLE LOG SHEET

Project Site Name: KSC-POL
Project No.: 112G09020
Date: 10/22/2021

Sample ID: POL-DPT0129 (aka DPT1469)
Location: I
Sampled By: R. Seigel, K. Kercher

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 10 rows of sampling data for POL-DPT0129 at various depths from 12' to 52'.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Contains 2 rows of collection data for VOCs and PFAS.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No. for marking sample collection details.



Project Site Name: KSC-POL
Project No.: 112G09020
Date: 10/20/2021

Sample ID: POL-DPT0127 (aka DPT1467)
Location: K
Sampled By: R. Seigel, K. Kercher

DPT SAMPLING DATA

Table with 8 columns: LOCATION ID, LOCATION, DATE, TIME, DEPTH, ODOR, COLOR, COMMENTS. Contains 10 rows of sampling data for various depths from 12' to 52'.

SAMPLE COLLECTION INFORMATION

Table with 4 columns: Analysis, Preservative, Container Requirements, Collected. Contains 2 rows of collection information for VOCs and PFAS.

OBSERVATIONS / NOTES

All intervals purged a minimum of five (5) screen volumes (1.5 L) prior to sample collection.
Hand auger five feet to clear utilities
Borehole abandoned via pressure grouting through boring rods
Rig, Rods, and associated tooling decon'd with pressurized steam

Circle if Applicable:

Table with 2 columns: MS/MSD, Duplicate ID No.:

01/21/2020

POL

112.6-09020

Purposed i. Chuck Sarden (CS) Geddes Tt

moon : Sunny 76°F

PPE : Level D

organic : Began PFAS/Basic Line Sample @ POL

0900 CS on Base; Gearing Equip/Supplies, Calvary

1035 Purge POL-MW00413E, well covered Teller tray; purge 3  
well before prior to sample

1110 Purge Complete

1115 Sample Collected POL-MW00413E-020.5-20200421 PFAS/8260TLL

1145 Purge POL-MW00088E-Teller present

1210 ~~Purge~~ ~~Complete~~ Purge POL-MW00088E  
Sample Collected

1235 Purge Complete on POL-MW00088E

1240 Sample Collected POL-MW00088E-020.5-20200421 PFAS/8260TLL

1255 Purge Complete on POL-MW00088E

1300 Sample Collected POL-MW00088E-027.5-20200421 PFAS/8260TLL

1325 Purge POL-MW00366E

1330 Purge ~~Complete~~ POL-MW00366E

1400 Purge Complete on POL-MW00366E

1405 Sample Collected POL-MW00366E-020.5-20200421 PFAS/8260TLL

1420 Purge Complete on POL-MW00366E

1425 Sample Collected POL-MW00366E-027.5-20200421 PFAS/8260TLL

-Mump IDW

1500 Purge POL-MW00425E

1515 Purge Complete

1520 Sample Collected POL-MW00425E-020.5-20200421 EUCO

1615 OFF.


01/22/2022

POL

112509020

Personal : Chase Screen (US) & 2/2/21  
 wear : Suny 71°F  
 PPE : Level D  
 activity : Lower Baseline / PFAS Sample

- 0800 CS on Base, getting equip/supplies
- 1010 Purging POL-MW0047E
- 1025 Purging Complete
- 1030 Sample Collected POL-MW0047E 025.0 20200922 PFAS/5200D
- 1045 Purging POL-MW0047D
- 1050 Sample Collected POL-FRB-20200922-01 PFAS
- 1100 Purging Complete
- 1105 Sample Collected POL-MW0047D 035.0 20200922 PFAS/5200D  
 - NOTE, FRB Collected Demand of active Contamination of System
- 1115 Purging POL-MW0047DD
- 1145 Purging Complete
- 1150 Sample Collected POL-MW0047DD 045.0 20200922 PFAS/5200D
- 1205 Purging POL-MW0046D
- 1220 Purging Complete
- 1225 Sample Collected POL-MW0046D 035.0 20200922 PFAS/5200D
- 1235 Purging POL-MW0046DD
- 1255 Purging Complete
- 1300 Sample Collected POL-MW0046DD 045.0 20200922 PFAS/5200D
- 1325 Purging POL-MW0045S
- 1340 Purging Complete
- 1345 Sample Collected POL-MW0045S 015.0 20200922 PFAS/5200D
- 1355 Purging POL-MW0045I
- 1430 Purging Complete
- 1435 Sample Collected POL-MW0045I 025.0 20200922 PFAS/5200D
- 1445 Purging POL-MW0045D
- 1500 Purging Complete
- 1505 Sample Collected POL-MW0045D 035.0 20200922 PFAS/5200D
- 1535 Purging POL-MW0043S
- 1550 Purging Complete
- 1555 Sample Collected POL-MW0043S 025.0 20200922 PFAS/8200D
- 1630 CS off -



112007020

POL

01/23/2021

Personnel: Chuck Sarden (C) Gregory Tr

Wanna: Sunny Hill 71.5E

PPE: Level D

Me: Sample - Composite PFAS / Baseline Sample

0840 CS on site; Getting Setup / Supplies / Calibrating

0915 Sample Collected POL-EB-20200423-01 PFAS  
- Collected through Sample Tube (3/16")

0925 Sample Collected POL-EB-20200423-02 PFAS  
- Collected through Sample Tube (1/4")

0930 Pumping POL-MW0043I

0945 Pumping Complete

0950 Sample Collected POL-MW0043I-025.0-20200423 PFAS/826.0

1000 Pumping POL-MW0043D

1015 Pumping Complete

1020 Sample Collected POL-MW0043D-035.0-20200423 PFAS/826.0

1030 Pumping POL-MW0043E

1045 Pumping Complete

1130 Pumping Complete on POL-MW0043E

1135 Sample Collected POL-MW0043E-045.0-20200423 PFAS/826.0

1200 Pumping Complete on POL-MW0043E

1205 Sample Collected POL-MW0043E-027.5-20200423 PFAS/826.0

1225 Pumping POL-MW0043E

1240 Pumping Complete

1245 Sample Collected POL-MW0043E-025.0-20200423 PFAS/826.0

1300 Pumping POL-MW0044D

1315 Pumping Complete

1320 Sample Collected POL-MW0044D-035.0-20200423 PFAS/826.0

1340 Pumping POL-MW0044E

1405 Pumping Complete

1410 Sample Collected POL-MW0044E-030.0-20200423 PFAS

1420 Pumping POL-MW0044E

1445 Pumping Complete

1450 Sample Collected POL-MW0044E-032.0-20200423 PFAS

1500 Pumping POL-MW0044E

1515 Pumping Complete

1520 Sample Collected POL-MW0044E-042.5-20200423 PFAS

1530 Pumping POL-MW0044E

1545 Pumping Complete

1550 Sample Collected POL-MW0044E-035.0-20200423 PFAS

1600 Pumping POL-MW0044E

1615 Pumping Complete

Scale: 1 square =

Rate in air Rain

01/28/2020

RD-

12409020

1270 Supply Order PD-1110000-20-2-20-3

ESS C

-Puppy Supply For Shipwa (PFA3)

1270 CE OFFER TO SELL

*[A large, diagonal scribble or signature is present across the page, starting from the bottom left and extending towards the top right.]*





PROJECT NO: 112609020		FACILITY: KSC-PO4/sh <sup>3</sup>		PROJECT MANAGER Mark Jenner		PHONE NUMBER (412) 921-8622		LABORATORY NAME AND CONTACT: Battelle — Jon Thorn																				
SAMPLERS (SIGNATURE) Chuck Sorden				FIELD OPERATIONS LEADER Chuck Sorden		PHONE NUMBER (321) 591-7580		ADDRESS 141 Longwater Dr. Suite 202																				
				CARRIER/WAYBILL NUMBER				CITY, STATE Norwell, MA																				
STANDARD TAT <input checked="" type="checkbox"/> RUSH TAT <input type="checkbox"/> <input type="checkbox"/> 24 hr. <input type="checkbox"/> 48 hr. <input type="checkbox"/> 72 hr. <input type="checkbox"/> 7 day <input type="checkbox"/> 14 day				CONTAINER TYPE PLASTIC (P) or GLASS (G) <b>P</b>		PRESERVATIVE USED <b>4°C</b>		TYPE OF ANALYSIS EPA 537 MOD PFAS																				
DATE YEAR 2020	TIME	SAMPLE ID	LOCATION ID	TOP DEPTH (FT)	BOTTOM DEPTH (FT)	MATRIX (GW, SO, SW, SD, QC, ETC.)	COLLECTION METHOD GRAB (G) COMP (C)					No. OF CONTAINERS	COMMENTS															
04/21	1115	POL-MW00415I-020.5-2020	0421	18	23	GW	G					2			COMMENTS													
	1240	POL-MW000085-009.3-2020	0421	8.6	10												COMMENTS											
	1300	POL-MW00008I-027.5-2020	0421	25	30														COMMENTS									
	1405	POL-MW000365I-020.5-2020	0421	18	23																COMMENTS							
04/21	1425	POL-MW00036I-027.5-2020	0421	25	30																		COMMENTS					
04/22	1030	POL-MW00047E-025.0-2020	0422	20	30	GW																			COMMENTS			
	1050	POL-FRB-20200422-01		-	-	QC																					COMMENTS	
	1105	POL-MW00470 <sup>470</sup> -035.0-2020	0422	30	40	GW																						
	1150	POL-MW004700-045.0-2020	0422	40	50				COMMENTS																			
	1225	POL-MW00460-035.0-2020	0422	30	40						COMMENTS																	
	1255	POL-MW004600-045.0-2020	0422	40	50								COMMENTS															
	1345	POL-MW00455-015.0-2020	0422	10	20										COMMENTS													
04/23	1435	POL-MW0045I-025.0-2020	0422	20	30	GW	G	2									COMMENTS											

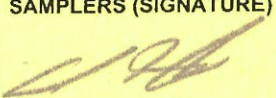
1. RELINQUISHED BY 	DATE 04/23/2020	TIME 1700	1. RECEIVED BY	DATE	TIME
2. RELINQUISHED BY	DATE	TIME	2. RECEIVED BY	DATE	TIME
3. RELINQUISHED BY	DATE	TIME	3. RECEIVED BY	DATE	TIME

COMMENTS

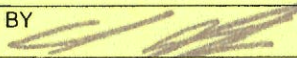


PROJECT NO: 112609020		FACILITY: KISC-POL/SW		PROJECT MANAGER Mark J. Sargent		PHONE NUMBER (413) 921-8622		LABORATORY NAME AND CONTACT: Battelle - Jan Thera			
SAMPLERS (SIGNATURE) Chris Sorden				FIELD OPERATIONS LEADER Chris Sorden		PHONE NUMBER (321) 591-7580		ADDRESS 141 Longwater Dr Suite 202			
				CARRIER/WAYBILL NUMBER				CITY, STATE Norwell, MA			
STANDARD TAT <input checked="" type="checkbox"/> RUSH TAT <input type="checkbox"/> <input type="checkbox"/> 24 hr. <input type="checkbox"/> 48 hr. <input type="checkbox"/> 72 hr. <input type="checkbox"/> 7 day <input type="checkbox"/> 14 day				CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED		TYPE OF ANALYSIS EPA 557 Method PCBs 4°C P			
DATE YEAR	TIME	SAMPLE ID	LOCATION ID	TOP DEPTH (FT)	BOTTOM DEPTH (FT)	MATRIX (GW, SO, SW, SD, QC, ETC.)	COLLECTION METHOD GRAB (G) COMP (C)				
04/21	1115	POL-MW004151-020.5-20200421	0421	18	23	GW	G	2	X		
	1240	POL-MW004155-029.3-20200421	0421	8.6	10						
	1300	POL-MW004151-022.5-20200421	0421	25	30						
	1405	POL-MW003655-020.5-20200421	0421	18	33						
04/21	1425	POL-MW003651-027.5-20200421	0421	25	30						
04/22	1030	POL-MW004750-025.0-20200422	0422	20	30	GW					
	1050	POL-FRB-20200422-01		-	-	QC					
	1105	POL-MW004750-035.0-20200422	0422	30	40	GW					
	1150	POL-MW004700-045.0-20200422	0422	40	50						
	1225	POL-MW004600-035.0-20200422	0422	30	40						
	1255	POL-MW004600-045.0-20200422	0422	40	50						
	1345	POL-MW004550-015.0-20200422	0422	10	20						
04/23	1435	POL-MW004550-025.0-20200422	0422	20	30	GW	G	2	X		
1. RELINQUISHED BY				DATE	TIME	1. RECEIVED BY				DATE	TIME
2. RELINQUISHED BY				DATE	TIME	2. RECEIVED BY				DATE	TIME
3. RELINQUISHED BY				DATE	TIME	3. RECEIVED BY				DATE	TIME
COMMENTS											



PROJECT NO: 1126-09020	FACILITY: KSC-POL/SW	PROJECT MANAGER Mark Jonnet	PHONE NUMBER (412) 921-8622	LABORATORY NAME AND CONTACT: Battelle - Jon Thom
SAMPLERS (SIGNATURE)  Chuck Sorden		FIELD OPERATIONS LEADER Chuck Sorden	PHONE NUMBER (321) 591-7580	ADDRESS 141 Longwater Dr. Ste. 202
STANDARD TAT <input checked="" type="checkbox"/> RUSH TAT <input type="checkbox"/> <input type="checkbox"/> 24 hr. <input type="checkbox"/> 48 hr. <input type="checkbox"/> 72 hr. <input type="checkbox"/> 7 day <input type="checkbox"/> 14 day			CARRIER/WAYBILL NUMBER	CITY, STATE Norwell, MA

DATE YEAR	TIME	SAMPLE ID	LOCATION ID	TOP DEPTH (FT)	BOTTOM DEPTH (FT)	MATRIX (GW, SO, SW, SD, QC, ETC.)	COLLECTION METHOD GRAB (G) COMP (C)	No. OF CONTAINERS	CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED	COMMENTS
04/23	1505	POL-MW0045I-035.0-20200423	200423	30	40	GW	G	2	X			
04/23	1555	POL-MW0043S-015.0-20200423	200423	10	20	I						
04/23	0915	POL-EB-20200423-01	-	-	-	QC						
	0925	POL-EB-20200423-02	-	-	-	QC						
	0950	POL-MW0043S-025.0-20200423	200423	20	30	GW						
	1020	POL-MW0043D-035.0-20200423	200423	30	40							
	1135	POL-MW0011S-012.5-20200423	200423	10	15							
	1205	POL-MW0011S-027.5-20200423	200423	25	30							
	1245	POL-MW0044E-025.0-20200423	200423	20	30							
	1320	POL-MW0044D-035.0-20200423	200423	30	40							
	1410	SW3-MW0027-032.0-20200423	200423	27	37							
04/23	1450	SW3-MW0028-032.0-20200423	200423	27	37	GW	G	2	X			

1. RELINQUISHED BY 	DATE 04/23/2020	TIME 1700	1. RECEIVED BY	DATE	TIME
2. RELINQUISHED BY	DATE	TIME	2. RECEIVED BY	DATE	TIME
3. RELINQUISHED BY	DATE	TIME	3. RECEIVED BY	DATE	TIME

COMMENTS



PROJECT NO: 112609020		FACILITY: RSC-PUL/K		PROJECT MANAGER M. J. Schmitt		PHONE NUMBER (412) 931-8622		LABORATORY NAME AND CONTACT: Battelle - Jon Thum							
SAMPLERS (SIGNATURE) Chris Sarda				FIELD OPERATIONS LEADER Chris Sarda		PHONE NUMBER (321) 591-7580		ADDRESS 141 Longmont Dr. Ste. 202							
				CARRIER/WAYBILL NUMBER				CITY, STATE Norwell, MA							
STANDARD TAT <input checked="" type="checkbox"/> RUSH TAT <input type="checkbox"/> <input type="checkbox"/> 24 hr. <input type="checkbox"/> 48 hr. <input type="checkbox"/> 72 hr. <input type="checkbox"/> 7 day <input type="checkbox"/> 14 day				CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED		TYPE OF ANALYSIS EPA 537 MW 906 P PEAS 406							
DATE YEAR 2020		LOCATION ID		TOP DEPTH (FT)		BOTTOM DEPTH (FT)						MATRIX (GW, SO, SW, SD, QC, ETC.)		COLLECTION METHOD GRAB (G) COMP (C)	
TIME		SAMPLE ID										COMMENTS			
01/22 1505		POL-MW00435-035.0-20200423		30 40		GW G		2		X					
01/23 1555		POL-MW00435-015.0-20200423		10 20		I									
01/23 0915		POL-EB-20200423-01		-		QC									
0925		POL-EB-20200423-02		-		QC									
0950		POL-MW00435-025.0-20200423		20 30		GW									
1030		POL-MW00430-035.0-20200423		30 40		I									
1135		POL-MW00435-015.0-20200423		10 15		I									
1205		POL-MW00435-027.5-20200423		25 30		I									
1245		POL-MW00445-025.0-20200423		20 30		I									
1320		POL-MW00440-035.0-20200423		30 40		I									
1410		SW3-MW0027-032.0-20200423		27 37		I									
01/23 1450		SW3-MW0028-032.0-20200423		27 37		GW G		2		X					
1. RELINQUISHED BY				DATE 04/23/2020		TIME 1700		1. RECEIVED BY				DATE		TIME	
2. RELINQUISHED BY				DATE		TIME		2. RECEIVED BY				DATE		TIME	
3. RELINQUISHED BY				DATE		TIME		3. RECEIVED BY				DATE		TIME	
COMMENTS															



PROJECT NO: <b>12G-09020</b>		FACILITY: <b>KSL-POL/SW</b>		PROJECT MANAGER <b>Mark Sonnet</b>		PHONE NUMBER <b>(412) 921-8022</b>		LABORATORY NAME AND CONTACT: <b>ENCO - Karlin Dymicki</b>					
SAMPLERS (SIGNATURE)  <i>Chuck Sorden</i>		FIELD OPERATIONS LEADER <b>Chuck Sorden</b>		PHONE NUMBER <b>(381) 591-7580</b>		ADDRESS <b>10775 Central Port Dr.</b>							
		CARRIER/WAYBILL NUMBER		CITY, STATE <b>O-lands, FL</b>									
STANDARD TAT <input checked="" type="checkbox"/> RUSH TAT <input type="checkbox"/> <input type="checkbox"/> 24 hr. <input type="checkbox"/> 48 hr. <input type="checkbox"/> 72 hr. <input type="checkbox"/> 7 day <input type="checkbox"/> 14 day				CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED		TYPE OF ANALYSIS <b>2601CL SOMO12 CLP-HCL G</b> <b>LITE</b>					
DATE YEAR <b>04/21 2020</b>		LOCATION ID		TOP DEPTH (FT)		BOTTOM DEPTH (FT)						MATRIX (GW, SO, SW, SD, QC, ETC.)	
TIME		SAMPLE ID										COMMENTS	
<b>1115</b>		<b>POL-MW004185-020.5-20200421</b>		<b>18 23</b>		<b>GW G 3</b>		<b>X</b>					
<b>1240</b>		<b>POL-MW00085-025.0-20200421</b>		<b>8.6 10</b>									
<b>1300</b>		<b>POL-MW00085-027.5-20200421</b>		<b>25 30</b>									
<b>1405</b>		<b>POL-MW00365-020.5-20200421</b>		<b>18 23</b>									
<b>1425</b>		<b>POL-MW00365-027.5-20200421</b>		<b>25 30</b>									
<b>1520</b>		<b>POL-MW00425-020.5-20200421</b>		<b>18 23</b>									
<b>1030</b>		<b>POL-MW006475-025.0-20200422</b>		<b>20 30</b>									
<b>1105</b>		<b>POL-MW00470-035.0-20200422</b>		<b>30 40</b>									
<b>1150</b>		<b>POL-MW00470-045.0-20200422</b>		<b>40 50</b>									
<b>1225</b>		<b>POL-MW00460-035.0-20200422</b>		<b>30 40</b>									
<b>1255</b>		<b>POL-MW00460-045.0-20200422</b>		<b>40 50</b>									
<b>1345</b>		<b>POL-MW00455-015.0-20200422</b>		<b>10 20</b>									
<b>1435</b>		<b>POL-MW00455-025.0-20200422</b>		<b>20 30</b>		<b>GW G 3</b>		<b>X</b>					
1. RELINQUISHED BY <i>Chuck Sorden</i>		DATE <b>04/24/2020</b>		TIME <b>0915</b>		1. RECEIVED BY <b>Kota Manuel</b>				DATE <b>04/24/20</b>		TIME <b>07:15</b>	
2. RELINQUISHED BY		DATE		TIME		2. RECEIVED BY				DATE		TIME	
3. RELINQUISHED BY		DATE		TIME		3. RECEIVED BY				DATE		TIME	
COMMENTS <b>12-451 0.2%</b>													



PROJECT NO: 1126-09020	FACILITY: KSC-POL/SW	PROJECT MANAGER Mark Sommer	PHONE NUMBER (412) 921-8002	LABORATORY NAME AND CONTACT: ENCL - Karina Dymalski
SAMPLERS (SIGNATURE) <i>[Signature]</i>		FIELD OPERATIONS LEADER Chuck Sordin	PHONE NUMBER (301) 591-7580	ADDRESS 10775 Central Park Dr.
CARRIER/WAYBILL NUMBER			CITY, STATE Orlando FL	

DATE YEAR	TIME	SAMPLE ID	LOCATION ID	TOP DEPTH (FT)	BOTTOM DEPTH (FT)	MATRIX (GW, SO, SW, SD, QC, ETC.)	COLLECTION METHOD GRAB (G) COMP (C)	No. OF CONTAINERS	CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED	TYPE OF ANALYSIS <i>ECOLOGICAL SAMPLES CLP-HCI-G</i>	COMMENTS
04/21	1115	POL-MW004151-020.5-20200421		18	23	GW	G	3	X				
	1246	POL-MW004152-020.5-20200421		86	10								
	1300	POL-MW004153-027.5-20200421		25	30								
	1405	POL-MW003651-020.5-20200421		18	23								
	1425	POL-MW003652-027.5-20200421		25	30								
04/21	1520	POL-MW004251-020.5-20200421		18	23								
04/22	1030	POL-MW004711-025.0-20200422		20	30								
	1105	POL-MW004710-035.0-20200422		30	40								
	1150	POL-MW004700-045.0-20200422		40	50								
	1225	POL-MW004600-035.0-20200422		30	40								
	1255	POL-MW004600-045.0-20200422		40	50								
	1345	POL-MW004550-015.0-20200422		10	20								
04/22	1435	POL-MW004551-025.0-20200422		20	30	GW	G	3	X				

1. RELINQUISHED BY <i>[Signature]</i>	DATE 04/24/2020	TIME 0915	1. RECEIVED BY Kara Maxwell	DATE 04/14/20	TIME 0915
2. RELINQUISHED BY	DATE	TIME	2. RECEIVED BY	DATE	TIME
3. RELINQUISHED BY	DATE	TIME	3. RECEIVED BY	DATE	TIME

COMMENTS: 14-451 0.2%




PROJECT NO: 112G09020	FACILITY: KSC-POL	PROJECT MANAGER <i>Mark Sommer</i> (321) 591-7580	PHONE NUMBER (412) 921-8022	LABORATORY NAME AND CONTACT: ENCO - <i>Kristin Dylnicki</i>
SAMPLERS (SIGNATURE) <i>Chuck Sorden</i>		FIELD OPERATIONS LEADER <i>Chuck Sorden</i>	PHONE NUMBER (321) 591-7580	ADDRESS 10775 Central Pkwy Dr.
CARRIER/WAYBILL NUMBER			CITY, STATE Orlando, FL	

DATE YEAR	TIME	SAMPLE ID	LOCATION ID	TOP DEPTH (FT)	BOTTOM DEPTH (FT)	MATRIX (GW, SO, SW, SD, QC, ETC.)	COLLECTION METHOD GRAB (G) COMP (C)	No. OF CONTAINERS	CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED	TYPE OF ANALYSIS	COMMENTS
04/22	1505	POL-MW00450-035.0-20200422	0422	30	40	GW	G	3	X			HCL	
04/22	1555	POL-MW00435-015.0-20200422	0422	10	20								
04/23	0950	POL-MW00435-025.0-20200423	0423	20	30								
	1020	POL-MW00430-035.0-20200423	0423	30	40								
	1135	POL-MW00116-012.5-20200423	0423	10	15								
	1205	POL-MW00112-027.5-20200423	0423	25	30								
	1245	POL-MW00441-025.0-20200423	0423	20	30								
	1320	POL-MW00440-035.0-20200423	0423	30	40								
	<del>1520</del>	<del>POL-MW00185-010.5-20200423</del>	<del>0423</del>	<del>8</del>	<del>13</del>								
	<del>1620</del>	<del>POL-MW00185-027.5-20200423</del>	<del>0423</del>	<del>25</del>	<del>30</del>								
	1520	POL-MW00180-040.5-20200423	0423	38	43								
	1550	POL-MW00185-010.5-20200423	0423	8	13								
04/23	1620	POL-MW00185-027.5-20200423	0423	25	30	GW	G	3	X				


1. RELINQUISHED BY <i>[Signature]</i>	DATE 04/24/2020	TIME 0915	1. RECEIVED BY <i>Kristin Dylnicki</i>	DATE 04/24/20	TIME 09:15
2. RELINQUISHED BY	DATE	TIME	2. RECEIVED BY	DATE	TIME
3. RELINQUISHED BY	DATE	TIME	3. RECEIVED BY	DATE	TIME

COMMENTS: L5451 0.2<sup>nd</sup>



PROJECT NO: <b>112001020</b>	FACILITY: <b>KSC-POL</b>	PROJECT MANAGER <b>Mark Sorden</b> <del>STT-758</del>	PHONE NUMBER <b>(412) 921-8622</b>	LABORATORY NAME AND CONTACT: <b>ENCO - Kevin Delmich</b>
SAMPLERS (SIGNATURE)  <b>Chuck Sorden</b>		FIELD OPERATIONS LEADER <b>Chuck Sorden</b>	PHONE NUMBER <b>(321) 591-7580</b>	ADDRESS <b>10775 Central Park Dr.</b>
CARRIER/WAYBILL NUMBER			CITY, STATE <b>Orlando, FL</b>	

DATE YEAR	TIME	SAMPLE ID	LOCATION ID	TOP DEPTH (FT)	BOTTOM DEPTH (FT)	MATRIX (GW, SO, SW, SD, QC, ETC.)	COLLECTION METHOD GRAB (G) COMP (C)	No. OF CONTAINERS	CONTAINER TYPE PLASTIC (P) or GLASS (G)		PRESERVATIVE USED	TYPE OF ANALYSIS	COMMENTS
01/23	1505	POL-MW00450-039.0-20200422		30	40	GW	G	3	X	G			
01/23	1555	POL-MW00435-015.0-20200422		10	20								
01/23	0950	POL-MW00432-025.0-20200423		20	30								
	1020	POL-MW00430-035.0-20200423		30	40								
	1155	POL-MW00415-012.5-20200423		10	15								
	1205	POL-MW00412-022.5-20200423		25	30								
	1245	POL-MW00411-025.0-20200423		20	30								
	1320	POL-MW00410-035.0-20200423		30	40								
	<del>1520</del>	<del>POL-MW00408-010.5-20200423</del>		<del>8</del>	<del>15</del>								
	<del>1520</del>	<del>POL-MW00418-027.5-20200423</del>		<del>25</del>	<del>30</del>								
	1520	POL-MW00480-040.5-20200423		38	43								
	1550	POL-MW00485-010.5-20200423		8	15								
01/23	1620	POL-MW00482-027.5-20200423		25	30	GW	G	3	X				

1. RELINQUISHED BY 	DATE <b>01/24/2020</b>	TIME <b>0915</b>	1. RECEIVED BY <b>Kota Manuel</b>	DATE <b>01/24/20</b>	TIME <b>09:15</b>
2. RELINQUISHED BY	DATE	TIME	2. RECEIVED BY	DATE	TIME
3. RELINQUISHED BY	DATE	TIME	3. RECEIVED BY	DATE	TIME

COMMENTS: **25451 0.2%**

# Calibration Report

Instrument Aqua TROLL 600 Vented  
 Serial Number 613901  
 Created 4/22/2020

Sensor **pH/ORP**  
 Serial Number 598349  
 Last Calibrated 4/22/2020

## Calibration Details

### Calibration Point 1

pH of Buffer 7.00 pH  
 pH mV 5.2 mV  
 Temperature 21.69 °C

### Pre Measurement

pH 7.05 pH  
 pH mV 5.3 mV

### Post Measurement

pH 7.00 pH  
 pH mV 5.2 mV

### Slope and Offset 1

Slope -58.5 mV/pH  
 Offset 5.2 mV

### ORP

ORP Solution Quick-Cal  
 Offset -79.2 mV  
 Temperature 21.69 °C  
 Pre Measurement 228.0 mV  
 Post Measurement 228.5 mV

Sensor **Turbidity**  
 Serial Number 684704  
 Last Calibrated 4/22/2020

## Calibration Details

TSS Conversion Factor (mg/L) 0  
 Slope 1  
 Offset -2.48 NTU

### Calibration Point 1

Pre Measurement 2.47 NTU  
 Post Measurement 0.00 NTU

<b>Sensor</b>	<b>RDO</b>
Serial Number	685525
Last Calibrated	4/22/2020

---

*Calibration Details*

Slope	1.047531
Offset	0.00 mg/L

---

*Calibration point 100%*

Concentration	8.74 mg/L
Pre Measurement	100.04 %Sat
Post Measurement	100.00 %Sat
Temperature	22.93 °C
Barometric Pressure	1,077.9 mbar

---

<b>Sensor</b>	<b>Conductivity</b>
Serial Number	613007
Last Calibrated	4/22/2020

---

*Calibration Details*

TDS Conversion Factor (ppm)	0.65
Cell Constant	1.009
Reference Temperature	25.00 °C

---

*Pre Measurement*

Actual Conductivity	7,396.2 µS/cm
Specific Conductivity	7,895.6 µS/cm

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*Post Measurement*

Actual Conductivity	7,494.0 µS/cm
Specific Conductivity	8,000.0 µS/cm

---

<b>Sensor</b>	<b>Barometric Pressure</b>
Serial Number	613901
Last Calibrated	Factory Defaults

---

<b>Sensor</b>	<b>Pressure</b>
Serial Number	609814
Last Calibrated	Factory Defaults



# Calibration Report

Instrument Aqua TROLL 600 Vented  
 Serial Number 613901  
 Created 4/21/2020

Sensor **pH/ORP**  
 Serial Number 598349  
 Last Calibrated 4/21/2020

## Calibration Details

### Calibration Point 1

pH of Buffer 7.00 pH  
 pH mV 8.1 mV  
 Temperature 22.86 °C

### Pre Measurement

pH 6.84 pH  
 pH mV 8.3 mV

### Post Measurement

pH 7.00 pH  
 pH mV 8.1 mV

### Slope and Offset 1

Slope -58.74 mV/pH  
 Offset 8.1 mV

### ORP

ORP Solution Quick-Cal  
 Offset -79.2 mV  
 Temperature 22.86 °C  
 Pre Measurement 237.0 mV  
 Post Measurement 226.7 mV

Sensor **Turbidity**  
 Serial Number 684704  
 Last Calibrated 4/21/2020

## Calibration Details

TSS Conversion Factor (mg/L) 0  
 Slope 1  
 Offset -4.77 NTU

### Calibration Point 1

Pre Measurement 2.82 NTU  
 Post Measurement 1.00 NTU

<b>Sensor</b>	<b>RDO</b>
Serial Number	685525
Last Calibrated	4/21/2020

---

*Calibration Details*

Slope	1.052618
Offset	0.00 mg/L

---

*Calibration point 100%*

Concentration	8.47 mg/L
Pre Measurement	98.80 %Sat
Post Measurement	100.00 %Sat
Temperature	23.94 °C
Barometric Pressure	1,070.0 mbar

---

<b>Sensor</b>	<b>Conductivity</b>
Serial Number	613007
Last Calibrated	4/21/2020

---

*Calibration Details*

TDS Conversion Factor (ppm)	0.65
Cell Constant	0.997
Reference Temperature	25.00 °C

---

*Pre Measurement*

Actual Conductivity	7,711.1 µS/cm
Specific Conductivity	8,039.5 µS/cm

---

*Post Measurement*

Actual Conductivity	7,673.2 µS/cm
Specific Conductivity	8,000.0 µS/cm

---

<b>Sensor</b>	<b>Barometric Pressure</b>
Serial Number	613901
Last Calibrated	Factory Defaults

---

<b>Sensor</b>	<b>Pressure</b>
Serial Number	609814
Last Calibrated	Factory Defaults

# Calibration Report

Instrument Aqua TROLL 600 Vented  
 Serial Number 613901  
 Created 4/23/2020

Sensor **pH/ORP**  
 Serial Number 598349  
 Last Calibrated 4/23/2020

## Calibration Details

### Calibration Point 1

pH of Buffer 7.00 pH  
 pH mV 3.7 mV  
 Temperature 23.36 °C

### Pre Measurement

pH 7.03 pH  
 pH mV 3.7 mV

### Post Measurement

pH 7.00 pH  
 pH mV 3.7 mV

### Slope and Offset 1

Slope -58.83 mV/pH  
 Offset 3.7 mV

### ORP

ORP Solution Quick-Cal  
 Offset -64.6 mV  
 Temperature 23.36 °C  
 Pre Measurement 210.6 mV  
 Post Measurement 226.0 mV

Sensor **Turbidity**  
 Serial Number 684704  
 Last Calibrated 4/23/2020

## Calibration Details

TSS Conversion Factor (mg/L) 0  
 Slope 1  
 Offset -1.57 NTU

### Calibration Point 1

Pre Measurement 2.62 NTU  
 Post Measurement 1.00 NTU

<b>Sensor</b>	<b>RDO</b>
Serial Number	685525
Last Calibrated	4/23/2020

---

*Calibration Details*

Slope	1.053667
Offset	0.00 mg/L

---

*Calibration point 100%*

Concentration	8.49 mg/L
Pre Measurement	99.18 %Sat
Post Measurement	100.00 %Sat
Temperature	24.36 °C
Barometric Pressure	1,081.7 mbar

---

<b>Sensor</b>	<b>Conductivity</b>
Serial Number	613007
Last Calibrated	4/23/2020

---

*Calibration Details*

TDS Conversion Factor (ppm)	0.65
Cell Constant	1.003
Reference Temperature	25.00 °C

---

*Pre Measurement*

Actual Conductivity	7,792.5 µS/cm
Specific Conductivity	8,044.3 µS/cm

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*Post Measurement*

Actual Conductivity	7,749.6 µS/cm
Specific Conductivity	8,000.0 µS/cm

---

<b>Sensor</b>	<b>Barometric Pressure</b>
Serial Number	613901
Last Calibrated	Factory Defaults

---

<b>Sensor</b>	<b>Pressure</b>
Serial Number	609814
Last Calibrated	Factory Defaults





































## Tetra Tech NUS / FDEP Groundwater Sampling Sheet

SITE NAME: Paint and Oil Locker (POL)	SITE LOCATION: Kennedy Space Center (KSC), FL
LOCATION ID: POL-MW0046DD	SAMPLE ID: POL-MW0046DD- 045.0 -202004 <u>22</u> Sample depth (ddd.d)= [bottom of screen (feet bls)-Top depth] x 0.5-bottom of screen (feet bls)
DATE: 2020 / 04 / <u>22</u>	

### PURGING DATA

STATIC DEPTH TO WATER (feet btoc): <u>10.42</u>	CASING HEIGHT (feet als): <u>3</u>	STATIC DEPTH TO WATER (feet bls) = DTW (btoc) - Casing Height (feet als): <u>NA</u>	WELL SCREEN INTERVAL DEPTH (feet bls): 40 to 50
WELL DIAMETER (inches): <u>1</u>	TUBING DIAMETER (inches): <u>3/16</u>	PURGE PUMP TYPE OR BAILER: <u>Peristaltic Pump</u>	TOP DEPTH = top of screen or depth to water which ever is greatest (feet bls): BOTTOM DEPTH (feet bls): <u>50</u>

WELL VOLUME PURGE: 1 WELL VOLUME = (TOTAL WELL DEPTH - STATIC DEPTH TO WATER) X WELL CAPACITY (only fill out if applicable)

\_\_\_\_\_ Liters

EQUIPMENT VOLUME PURGE: 1 EQUIPMENT VOL. = PUMP VOLUME + (TUBING CAPACITY X TUBING LENGTH) + FLOW CELL VOLUME (only fill out if applicable)

0.78 Liters  $(0.005 \times 60) + 0.475 = 0.78$

INITIAL PUMP OR TUBING DEPTH IN WELL (feet): <u>45</u>	FINAL PUMP OR TUBING DEPTH IN WELL (feet): <u>45</u>	PURGING INITIATED AT: <u>1235</u>	PURGING ENDED AT: <u>1250</u>	TOTAL VOLUME PURGED (Liters): <u>3.0</u>
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TIME	VOLUME PURGED (Liters)	CUMUL. VOLUME PURGED (Liters)	PURGE RATE (mlpm)	DEPTH TO WATER (feet)	pH (standard units)	TEMP. (°C)	COND. (µS/cm)	DISSOLVED OXYGEN (mg/L)	TURBIDITY (NTUs)	ORP (mV)	COLOR (describe)
1245	2.0	2.0	200	10.51	6.80	25.14	2698.8	0.18	8.75	-138.7	Clear
1247	0.4	2.4	200	10.51	6.82	25.30	2683.6	0.16	8.93	-142.6	Clear
1250	0.6	3.0	200	10.51	6.84	25.22	2623	0.13	8.07	-150.6	Clear
1255	Sample Collected										

WELL CAPACITY (Liters Per Foot): 0.75" = 0.076; 1" = 0.15; 1.25" = 0.23; 2" = 0.61; 3" = 1.40; 4" = 2.46; 5" = 3.86; 6" = 5.57; 12" = 22.26  
 TUBING INSIDE DIA. CAPACITY (Liters/Ft.): 1/8" = 0.002; 3/16" = 0.005; 1/4" = 0.0098; 5/16" = 0.015; 3/8" = 0.023; 1/2" = 0.038; 5/8" = 0.09

### SAMPLING DATA

SAMPLED BY (PRINT) / AFFILIATION: C. Sorden / IT	SAMPLER(S) SIGNATURES: 	SAMPLING INITIATED AT: <u>1255</u>	SAMPLING ENDED AT: <u>1300</u>
PUMP OR TUBING DEPTH IN WELL (feet): <u>45</u>	SAMPLE PUMP FLOW RATE (mL per minute): <u>200</u>	TUBING MATERIAL CODE: <u>Teflon HDPE</u>	
FIELD DECONTAMINATION: <u>Y</u> N	FIELD-FILTERED: Y <u>N</u> FILTER SIZE: _____ µm	DUPLICATE: Y <u>N</u>	

SAMPLE CONTAINER SPECIFICATION				SAMPLE PRESERVATION			INTENDED ANALYSIS AND/OR METHOD	SAMPLING EQUIPMENT CODE
SAMPLE ID CODE	# CONTAINERS	MATERIAL CODE	VOLUME	PRESERVATIVE USED	TOTAL VOL ADDED IN FIELD (mL)	FINAL pH		
1	2	HDPE	250 mL	N/A	N/A	N/A	EPA MOD 537 PFAS	APP
2	3	CG	40 mL	HCl	N/A	<2	8260D TCL SOM01.2 CLP-LIKE	APP

REMARKS:

**MATERIAL CODES:** AG = Amber Glass; CG = Clear Glass; PE = Polyethylene; PP = Polypropylene; S = Silicone; T = Teflon; O = Other (Specify)

**SAMPLING/PURGING EQUIPMENT CODES:** APP = After Peristaltic Pump; B = Bailor; BP = Bladder Pump; ESP = Electric Submersible Pump; PP = Peristaltic Pump; RFPF = Reverse Flow Peristaltic Pump; SM = Straw Method (Tubing Gravity Drain); VT = Vacuum Trap; O = Other (Specify)





## Tetra Tech NUS / FDEP Groundwater Sampling Sheet

SITE NAME: Paint and Oil Locker (POL)	SITE LOCATION: Kennedy Space Center (KSC), FL
LOCATION ID: POL-MW0047DD	SAMPLE ID: POL-MW0047DD- 045.0 -202004 22 Sample depth (ddd.d)=[bottom of screen (feet bls)-Top depth] x 0.5-bottom of screen (feet bls)
DATE: 2020 / 04 / 22	

### PURGING DATA

STATIC DEPTH TO WATER (feet btoc): 4.45	CASING HEIGHT (feet als): 3	STATIC DEPTH TO WATER (feet bls) = DTW (btoc) - Casing Height (feet als): NA	WELL SCREEN INTERVAL DEPTH (feet bls): 40 to 50
WELL DIAMETER (inches):	TUBING DIAMETER (inches): 3/16	PURGE PUMP TYPE OR BAILER: Peristaltic Pump	TOP DEPTH = top of screen or depth to water which ever is greatest (feet bls):
BOTTOM DEPTH (feet bls): 50			

WELL VOLUME PURGE: 1 WELL VOLUME = (TOTAL WELL DEPTH - STATIC DEPTH TO WATER) X WELL CAPACITY  
 (only fill out if applicable) \_\_\_\_\_ Liters

EQUIPMENT VOLUME PURGE: 1 EQUIPMENT VOL. = PUMP VOLUME + (TUBING CAPACITY X TUBING LENGTH) + FLOW CELL VOLUME  
 (only fill out if applicable) 0.78 Liters (0.005 x 80' 60) + 0.475 = 0.78

INITIAL PUMP OR TUBING DEPTH IN WELL (feet): 45	FINAL PUMP OR TUBING DEPTH IN WELL (feet): 45	PURGING INITIATED AT: 1115	PURGING ENDED AT: 1145	TOTAL VOLUME PURGED (Liters): 6.0
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TIME	VOLUME PURGED (Liters)	CUMUL. VOLUME PURGED (Liters)	PURGE RATE (mlpm)	DEPTH TO WATER (feet)	pH (standard units)	TEMP. (°C)	COND. (µS/cm)	DISSOLVED OXYGEN (mg/L)	TURBIDITY (NTUs)	ORP (mV)	COLOR (describe)
<del>1125</del>	<del>2.0</del>	<del>2.0</del>	<del>200</del>	<del>9.58</del>	<del>6.40</del>	<del>24.92</del>	<del>2107</del>	<del>Sensor Issues</del>			
1140	5.0	5.0	200	9.58	6.96	24.59	2897.2	0.31	3.60	738.2	CLW
1142	0.4	5.4	200	9.58	6.95	24.60	2907.3	0.23	6.26	-144.8	CLW
1145	0.6	<del>3.0</del> 6.0	200	9.58	6.98	24.78	2907.4	0.19	7.30	-146.0	CLW
1150	Sample Collected										

WELL CAPACITY (Liters Per Foot): 0.75" = 0.076; 1" = 0.15; 1.25" = 0.23; 2" = 0.61; 3" = 1.40; 4" = 2.46; 5" = 3.86; 6" = 5.57; 12" = 22.26  
 TUBING INSIDE DIA. CAPACITY (Liters/Ft.): 1/8" = 0.002; 3/16" = 0.005; 1/4" = 0.0098; 5/16" = 0.015; 3/8" = 0.023; 1/2" = 0.038; 5/8" = 0.09

### SAMPLING DATA

SAMPLED BY (PRINT) / AFFILIATION: C. Sorden / TI	SAMPLER(S) SIGNATURES:	SAMPLING INITIATED AT: 1150	SAMPLING ENDED AT: 1155
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PUMP OR TUBING DEPTH IN WELL (feet): 45	SAMPLE PUMP FLOW RATE (mL per minute): 200	TUBING MATERIAL CODE: Teflon HDPE
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FIELD DECONTAMINATION: Y N	FIELD-FILTERED: Y N FILTER SIZE: _____ µm	DUPLICATE: Y N
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SAMPLE CONTAINER SPECIFICATION				SAMPLE PRESERVATION			INTENDED ANALYSIS AND/OR METHOD	SAMPLING EQUIPMENT CODE
SAMPLE ID CODE	# CONTAINERS	MATERIAL CODE	VOLUME	PRESERVATIVE USED	TOTAL VOL ADDED IN FIELD (mL)	FINAL pH		
1	2	HDPE	250 mL	N/A	N/A	N/A	EPA MOD 537 PFAS	APP
2	3	CG	40 mL	HCl	N/A	<2	8260D TCL SOM01.2 CLP-LIKE	APP

REMARKS:

**MATERIAL CODES:** AG = Amber Glass; CG = Clear Glass; PE = Polyethylene; PP = Polypropylene; S = Silicone; T = Teflon; O = Other (Specify)  
**SAMPLING/PURGING EQUIPMENT CODES:** APP = After Peristaltic Pump; B = Bailer; BP = Bladder Pump; ESP = Electric Submersible Pump; PP = Peristaltic Pump  
 RFPF = Reverse Flow Peristaltic Pump; SM = Straw Method (Tubing Gravity Drain); VT = Vacuum Trap; O = Other (Specify)



## Tetra Tech NUS / FDEP Groundwater Sampling Sheet

SITE NAME: <b>Supply Warehouse No. 3 (SW3)</b>	SITE LOCATION: <b>Kennedy Space Center (KSC), Florida</b>
LOCATION ID: <b>SW3-MW0027</b>	SAMPLE ID: <b>SW3-MW0027-032.0-2020 0423</b> Sample depth (ddd.d)=[bottom of screen (feet bls)-Top depth] x 0.5-bottom of screen (feet bls)
DATE: <b>04 / 23 / 2020</b>	

### PURGING DATA

STATIC DEPTH TO WATER (feet btoc): <b>9.90</b>	CASING HEIGHT (feet als): <b>NA 2.5</b>	STATIC DEPTH TO WATER (feet bls) = DTW (btoc) - Casing Height (feet als): <b>NA</b>	WELL SCREEN INTERVAL DEPTH (feet bls): <b>27 to 37</b>
WELL DIAMETER (inches):	TUBING DIAMETER (inches): <b>3/16</b>	PURGE PUMP TYPE OR BAILER: <b>Peristaltic Pump</b>	TOP DEPTH = top of screen or depth to water which ever is greatest (feet bls): <b>37</b>
BOTTOM DEPTH (feet bls): <b>37</b>			
WELL VOLUME PURGE: 1 WELL VOLUME = (TOTAL WELL DEPTH - STATIC DEPTH TO WATER) X WELL CAPACITY (only fill out if applicable)			
<b>4.43 Liters (37 - 7.46) x 0.15 = 4.43</b>			
EQUIPMENT VOLUME PURGE: 1 EQUIPMENT VOL. = PUMP VOLUME + (TUBING CAPACITY X TUBING LENGTH) + FLOW CELL VOLUME (only fill out if applicable)			
<b>0 Liters</b>			

INITIAL PUMP OR TUBING DEPTH IN WELL (feet): <b>32</b>	FINAL PUMP OR TUBING DEPTH IN WELL (feet): <b>32</b>	PURGING INITIATED AT: <b>1340</b>	PURGING ENDED AT: <b>1405</b>	TOTAL VOLUME PURGED (Liters): <b>10.0</b>							
TIME	VOLUME PURGED (Liters)	CUMUL. VOLUME PURGED (Liters)	PURGE RATE (mlpm)	DEPTH TO WATER (feet)	pH (standard units)	TEMP. (°C)	COND. (µS/cm)	DISSOLVED OXYGEN (mg/L)	TURBIDITY (NTUs)	ORP (mV)	COLOR (describe)
1400	8.0	8.0	400	10.11	6.53	26.35	2248.2	0.11	7.40	-112.8	Clear
1402	0.8	8.8	400	10.11	6.55	26.34	2248.4	0.09	5.14	-114.5	Clear
1405	1.2	10.0	400	10.11	6.55	26.36	2251.4	0.08	5.97	-115.7	Clear
1410	Sample	Collected									

WELL CAPACITY (Liters Per Foot): 0.75" = 0.076; 1" = 0.15; 1.25" = 0.23; 2" = 0.61; 3" = 1.40; 4" = 2.46; 5" = 3.86; 6" = 5.57; 12" = 22.26  
 TUBING INSIDE DIA. CAPACITY (Liters/Ft.): 1/8" = 0.002; 3/16" = 0.005; 1/4" = 0.0098; 5/16" = 0.015; 3/8" = 0.023; 1/2" = 0.038; 5/8" = 0.06

### SAMPLING DATA

SAMPLED BY (PRINT) / AFFILIATION: <i>Charles Sander</i> / Tetra Tech	SAMPLER(S) SIGNATURES: 	SAMPLING INITIATED AT: <b>1410</b>	SAMPLING ENDED AT: <b>1415</b>
PUMP OR TUBING DEPTH IN WELL (feet): <b>32</b>	SAMPLE PUMP SIZE FLOW RATE (mL per minute): <b>200</b>	TUBING MATERIAL CODE: <del>Teflon</del> <b>HDPE</b>	
FIELD DECONTAMINATION: <input checked="" type="radio"/> Y <input type="radio"/> N	FIELD-FILTERED: <input checked="" type="radio"/> Y <input type="radio"/> N	FILTER SIZE: _____ µm	DUPLICATE: <input checked="" type="radio"/> Y <input type="radio"/> N

SAMPLE CONTAINER SPECIFICATION				SAMPLE PRESERVATION			INTENDED ANALYSIS AND/OR METHOD	SAMPLING EQUIPMENT CODE
SAMPLE ID CODE	# CONTAINERS	MATERIAL CODE	VOLUME	PRESERVATIVE USED	TOTAL VOL ADDED IN FIELD (mL)	FINAL pH		
1	2	HDPE	250 mL	N/A	N/A	N/A	EPA MOD 537 PFAS	APP

REMARKS:

MATERIAL CODES: AG = Amber Glass; CG = Clear Glass; PE = Polyethylene; PP = Polypropylene; S = Silicone; T = Teflon; O = Other (Specify)  
 SAMPLING/PURGING EQUIPMENT CODES: APP = After Peristaltic Pump; B = Bailor; BP = Bladder Pump; ESP = Electric Submersible Pump; PP = Peristaltic Pump  
 RFPP = Reverse Flow Peristaltic Pump; SM = Straw Method (Tubing Gravity Drain); VT = Vacuum Trap; O = Other (Specify)

well central Teflon Tubing  
 well volumes purged

## Tetra Tech NUS / FDEP Groundwater Sampling Sheet

SITE NAME: <b>Supply Warehouse No. 3 (SW3)</b>	SITE LOCATION: <b>Kennedy Space Center (KSC), Florida</b>
LOCATION ID: <b>SW3-MW0028</b>	SAMPLE ID: <b>SW3-MW0028-032.0-2020 0423</b> <small>Sample depth (ddd.d)=[bottom of screen (feet bls)-Top depth] x 0.5-bottom of screen (feet bls)</small>
DATE: <b>04 / 23 / 2020</b>	

### PURGING DATA

STATIC DEPTH TO WATER (feet btoc): <b>10.04</b>	CASING HEIGHT (feet als): <b>3.0</b>	STATIC DEPTH TO WATER (feet bls) = DTW (btoc) - Casing Height (feet als): <b>7.04</b>	WELL SCREEN INTERVAL DEPTH (feet bls): <b>27 to 37</b>
WELL DIAMETER (inches): <b>1</b>	TUBING DIAMETER (inches): <b>3/16</b>	PURGE PUMP TYPE OR BAILER: <b>Peristaltic Pump</b>	TOP DEPTH = top of screen or depth to water which ever is greatest (feet bls): <b>37</b>
BOTTOM DEPTH (feet bls): <b>37</b>			
WELL VOLUME PURGE: 1 WELL VOLUME = (TOTAL WELL DEPTH - STATIC DEPTH TO WATER) X WELL CAPACITY <small>(only fill out if applicable)</small> <div style="text-align: center; font-size: 1.2em;"> <math>4.49 \text{ Liters} = (37 - 7.04) \times 0.15 = 4.49</math> </div>			
EQUIPMENT VOLUME PURGE: 1 EQUIPMENT VOL. = PUMP VOLUME + (TUBING CAPACITY X TUBING LENGTH) + FLOW CELL VOLUME <small>(only fill out if applicable)</small> <div style="text-align: center;">                 _____ Liters             </div>			

INITIAL PUMP OR TUBING DEPTH IN WELL (feet):	<b>32</b>	FINAL PUMP OR TUBING DEPTH IN WELL (feet):	<b>32</b>	PURGING INITIATED AT:	<b>1420</b>	PURGING ENDED AT:	<b>1445</b>	TOTAL VOLUME PURGED (Liters):	<b>10.0</b>		
TIME	VOLUME PURGED (Liters)	CUMUL. VOLUME PURGED (Liters)	PURGE RATE (mlpm)	DEPTH TO WATER (feet)	pH (standard units)	TEMP. (°C)	COND. (µS/cm)	DISSOLVED OXYGEN (mg/L)	TURBIDITY (NTUs)	ORP (mV)	COLOR (describe)
1440	8.0	8.0	800	10.06	6.44	26.28	2927.4	0.06	4.97	-154.4	clw
1442	6.8	8.8	400	10.06	6.44	26.24	2921.3	0.07	7.01	-157.4	clw
1443	1.2	10.0	400	10.06	6.44	26.25	2915.7	0.07	8.03	-158.1	clw
1450	Sample Collected										

WELL CAPACITY (Liters Per Foot): 0.75" = 0.076; 1" = 0.15; 1.25" = 0.23; 2" = 0.61; 3" = 1.40; 4" = 2.46; 5" = 3.86; 6" = 5.57; 12" = 22.26  
 TUBING INSIDE DIA. CAPACITY (Liters/Ft.): 1/8" = 0.002; 3/16" = 0.005; 1/4" = 0.0098; 5/16" = 0.015; 3/8" = 0.023; 1/2" = 0.038; 5/8" = 0.06

SAMPLING DATA											
SAMPLED BY (PRINT) / AFFILIATION: <i>Charles Sarda</i> /Tetra Tech				SAMPLER(S) SIGNATURES: 				SAMPLING INITIATED AT: <b>1450</b>		SAMPLING ENDED AT: <b>1455</b>	
PUMP OR TUBING DEPTH IN WELL (feet):				SAMPLE PUMP OR FLOW RATE (mL per minute): <del>100</del> <b>200</b>				TUBING MATERIAL CODE: <b>Teflon HDPE</b>			
FIELD DECONTAMINATION: Y N				FIELD-FILTERED: Y N FILTER SIZE: _____ µm Filtration Equipment Type: _____				DUPLICATE: Y N			
SAMPLE CONTAINER SPECIFICATION				SAMPLE PRESERVATION				INTENDED ANALYSIS AND/OR METHOD		SAMPLING EQUIPMENT CODE	
SAMPLE ID CODE	# CONTAINERS	MATERIAL CODE	VOLUME	PRESERVATIVE USED	TOTAL VOL ADDED IN FIELD (mL)	FINAL pH					
1	2	HDPE	250 mL	N/A	N/A	N/A	EPA MOD 537 PFAS		APP		

REMARKS:

MATERIAL CODES: AG = Amber Glass; CG = Clear Glass; PE = Polyethylene; PP = Polypropylene; S = Silicone; T = Teflon; O = Other (Specify)

SAMPLING/PURGING EQUIPMENT CODES: APP = After Peristaltic Pump; B = Bailer; BP = Bladder Pump; ESP = Electric Submersible Pump; PP = Peristaltic Pump; RFPP = Reverse Flow Peristaltic Pump; SM = Straw Method (Tubing Gravity Drain); VT = Vacuum Trap; O = Other (Specify)

well contained Teflon Tubing;  
 Replaced; purged well volume









































**APPENDIX D**  
**PHOTOGRAPHIC LOG**

**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 1**

**DATE:**  
12/09/2021

**DIRECTION:**  
North

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of east side  
of STP, facing  
north



**PHOTO 2**

**DATE:**  
12/09/2021

**DIRECTION:**  
Northwest

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of clarifiers,  
facing northwest



**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 3**

**DATE:**  
12/09/2021

**DIRECTION:**  
South

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of chlorine contact chamber



**PHOTO 4**

**DATE:**  
12/09/2021

**DIRECTION:**  
East

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of water in chlorine contact chamber





**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 5**

**DATE:**  
12/09/2021

**DIRECTION:**  
North

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of sewage treatment plant on left and aerated surge tank on right



**PHOTO 6**

**DATE:**  
12/09/2021

**DIRECTION:**  
North

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View from top of sewage treatment plant





**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 7**

**DATE:**  
12/09/2021

**DIRECTION:**  
Southwest

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View from top of  
sewage treatment  
plant



**PHOTO 8**

**DATE:**  
12/09/2021

**DIRECTION:**  
Southwest

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View from top of  
sewage treatment plant  
looking down at  
clarifiers



**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 9**

**DATE:**  
12/09/2021

**DIRECTION:**  
Southwest

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of north side of  
sewage treatment plant



**PHOTO 10**

**DATE:**  
12/09/2021

**DIRECTION:**  
South

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of southeast  
corner of north side of  
sewage treatment  
plant on right and  
aerated surge tank on  
left





**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 11**

**DATE:**  
12/09/2021

**DIRECTION:**  
South

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View of the north side  
of the aerated surge  
tank



**PHOTO 12**

**DATE:**  
12/09/2021

**DIRECTION:**  
East

**TAKEN BY:**  
S. Damphousse

**DESCRIPTION:**  
View from top of  
aerated surge tank



**Photographic Log**  
**Sewage Treatment Plant #1 – LOC 20**  
**NASA PFAS Assessments**  
**Kennedy Space Center, Florida**

**PHOTO 13**

**DATE:**  
March 2022

**DIRECTION:**  
South

**TAKEN BY:**  
C. Sorden

**DESCRIPTION:**  
Surface water sampling  
in ditch using pole to  
place tubing in water



**PHOTO 14**

**DATE:**  
March 2021

**DIRECTION:**  
East

**TAKEN BY:**  
C. Sorden

**DESCRIPTION:**  
DPT sampling along  
5<sup>th</sup> Street SE



**APPENDIX E**

**KSCRT MEETING MINUTES AND ACTION ITEM – OCTOBER 2022**

**Revision 0 Meeting Minutes for October 5<sup>th</sup> and 6<sup>th</sup>, 2022**

Attendees:

- |                            |                               |
|----------------------------|-------------------------------|
| 1. Bruce Moore/FDEP        | 11. Alex Murphy/Tetra Tech    |
| 2. Mike Deliz/NASA         | 12. Chris Pike/ Tetra Tech    |
| 3. Ryan O’Meara/NASA       | 13. Mark Speranza/Tetra Tech  |
| 4. Deda Johansen/NASA      | 14. Andrew Walters/Tetra Tech |
| 5. Anne Chrest/NASA        | 15. Jennifer Gootee/AECOM     |
| 6. Natasha Darre/NASA      | 16. Linnea King Clark/AECOM   |
| 7. Chris Adkison/NASA      | 17. Richard Smith/HGL         |
| 8. Tim Appleman/NASA       | 18. Howard Fowler/HGL         |
| 9. Michelle Moore/NEMCON   | 19. James (Jim) Montague/HGL  |
| 10. Mark Jonnet/Tetra Tech |                               |

**2210-M01 Bruce Moore/FDEP**

**Program Update**

Discussion: The #1 issue at the Florida Department of Environmental Protection (FDEP) is the staffing situation. There are fourteen positions in the federal facilities program currently available. Some hires are imminent. Environmental Administrator Laura Barrett resigned. The goal is to fill the Environmental Administrator position by the end of October. Billy Hessman joined in May as a professional geologist (PG) II position. A variety of positions are open and need to be filled. If there is an urgent matter, please call Bruce directly and he can talk in the moment about it. The routine review process may take a while.

NASA inquired if funding was the issue or just not enough people were applying. FDEP stated it has been hard to attract and retain staff. FDEP is still using the three contractors for outside review and will lean heavily on them in the short term.

samples will be collected from drainage ditches. Additional DPT groundwater samples will be collected, and more monitoring wells installed and sampled.

NASA is preparing to notify nearby residents of potential off-site migration. Information sessions are being planned for early 2023. discussed the process for getting authorization for possible sampling on neighboring property. FDEP asked whether NASA has reached out to the Department of Health yet? NASA has not. FDEP will initiate contact at the State level.

**2210-M08      Mark Jonnet/ Tetra Tech**

**Fire Station #1 (SWMU #116), Sewage Treatment Plant #1 and Sludge Disposal Area (SWMU #117) PFAS Sites Assessment Update**

**Objective:**

Present results to date for per- and polyfluorinated alkyl substances (PFAS) soil evaluation, soil cores, PFAS sediment evaluation, PFAS groundwater evaluation, PFAS surface water evaluation and path forward.

**Discussion:**

Fire Station #1(FS1) was constructed in 1964, housed crews and served as a maintenance and storage location for spent or expired fire extinguishers. It was formerly known as Fire Station #4. The Sewage Treatment Plant #1 (STP1) encompasses approximately 40 acres and includes STP1, Former Polishing Pond, Former Sludge Disposal Area, Former Spray Field, and the Paint and Oil Locker (POL) SWMU 067.

Tetra Tech presented the PFAS sampling results to date of the Fire Station #1, STP #1 and Sludge Disposal Area sites. Data for the Base Support Building (SWMU #014) was included since the PFAS plumes appear to be commingled.

Data generated by Site Assessment activities to date and prior results were screened against EPA's May 2022 tap water Regional Screening Levels (RSLs) for groundwater, resident RSL for soil, and



against the State of Florida human health Surface Water Screening Levels for surface water. There are currently no screening criteria for sediment. RSLs are available for 6 PFAS compounds: perfluorooctanoic acid (PFOA), perfluorooctane sulfonic acid (PFOS), perfluoronanoic acid (PFNA), perfluorobutanesulfonic acid (PFBS), perfluorohexane sulfonic acid (PFHxS) and hexafluoropropylene oxide dimer acid (HFPO-DA; trade name GenX).

Data summaries include all samples collected to date in the area of FS1 and STP1. Samples collected prior to 12/2021 were collected by others. Field activities were performed in accordance with FDEP Standard Operating Procedures, PFAS sampling guidelines (Michigan Department of Environmental Quality) and the KSC Sampling and Analysis Plan. Global Positioning System coordinates were collected for each sample location. Quality assurance/quality control samples were collected due to the ubiquitous nature of PFAS. Pace Analytical Services analyzed PFAS samples and reported 28 PFAS compounds.

Shallow soil samples (0-0.5 and 0.5-2 feet below land surface [ft. bls]) were collected at five locations at FS1 and nine STP1/Sludge Disposal locations. Results exceeded the residential PFOS RSL in 6 of the 28 samples. Deeper soil samples were collected from one soil boring each at FS1 and STP1. Two intervals at FS1 had PFOS detections exceeding the residential RSL. Results from the STP1 boring were less than the residential RSL. There are no soil results with a PFAS result greater than the commercial RSL. The lithologic descriptions from the soil cores were used in selection of depth intervals for direct push technology (DPT) groundwater sampling.

Sediment samples were collected from nine locations at STP1/Sludge Disposal Area and one location in the Region 1 (Industrial Area) Stormwater Pond (also called the Gator Pond). There were detections of PFOS in seven of the ten samples, and one detection of PFHxS. There are currently no State or Federal screening criteria for sediment.

For FS1, of the 126 groundwater samples from varying depths, 44 detected results were greater than the PFOA RSL, 60 detected results exceeded the PFOS RSL, 17 detected results exceeded the PFNA RSL, 13 detected results exceeded the PFBS RSL, and 38 detected



results exceeded the PFHxS RSL. Seventy-six samples were analyzed for HFPO-DA and there were no detections.

For STP1/Sludge Disposal Area, of the 218 groundwater samples from varying depths, 115 detected results exceeded the PFOA RSL, 122 detected results exceeded the PFOS RSL, 41 detected results exceeded the PFNA RSL, 2 detected results exceeded the PFBS RSL, and 86 detected results exceeded the PFHxS RSL. One hundred seventy-six samples were analyzed for HFPO-DA and there were no detections.

Twenty-four surface water samples were collected from stormwater ditches around FS1, STP1 and the Sludge Disposal Area. A sample was collected at the northwest corner of the Gator Pond in both August 2021 and March 2022. PFOA, PFOS and PFHxS were detected in all 26 samples, PFBS was detected in 24 samples, and PFNA in 18 samples. HFPO-DA was analyzed in 19 samples and detected in none. The State of Florida has surface water screening levels (SWSLs) for PFOA and PFOS. One PFOA sample result exceeded its SWSL (500 ng/L) and 24 PFOS results exceeded its SWSL (10 ng/L). The two results for the Gator Pond did not indicate a significant seasonal difference.

The Site Assessment for FS1 and STP1 will continue in phases. The upcoming phase emphasizes understanding groundwater and surface water interaction, as well as the extent of PFAS-affected groundwater. Potential human health risk by exposure to PFAS-affected soil is being managed by interim Land Use Control Implementation Plans (LUCIPs).

To show the correlation between proposed groundwater and surface water sampling locations, the image on Slide 34 displays PFOS groundwater sample locations less than 10 ft bls compared to PFOS RSL and surface water PFOS results compared to PFOS SWSL. The Team will continue plume delineation using DPT groundwater sampling based on RSLs. Monitoring wells will be installed adjacent to surface water locations with staff gauges. The co-located wells and surface water points will be sampled periodically. Groundwater level measurements and staff gauges will be read, and data evaluated to determine discharge from groundwater to surface water or from surface water to groundwater.

The image on Slide 35 displays surface water PFOS results. Future sampling will be focused on the flow path for Region 1 stormwater to discharge to the Banana River Lagoon and nearby points in the lagoon. Sample locations will include influent into the Gator Pond, effluent from the Gator Pond, associated borrow pits that are part of the stormwater management system, tributaries into Buck Creek, locations within Buck Creek, junction of Buck Creek and Banana River Lagoon with offsets north and south, isolated borrow pits northeast of Gator Pond to determine impacts, and four locations along Banana River Lagoon that will correspond to DPT locations on the shore.

Tetra Tech will email FDEP the drawing showing the proposed monitoring well locations (2210-A06).

**Results: Action Item 2210-A06**

## **DAY 2**

**2210-M09 Michelle Moore/NEMCON**

### **Meeting Minutes and Miscellaneous Items**

Team consensus was reached that Revision 1 of the meeting minutes and action/decision items for the September 2022 Team meeting will become final. Team members acknowledged and did not object to the fact that these meeting minutes may become public as part of a final report at a later date (2210-D12).

Open action items were reviewed and closed at the October 2022 KSCRT meeting:

Launch Complex 39B (LC39B) (SWMU 009) - Revisit Team consensus (Decision 1810-D13) on weir installation based on permits date expiration and Year 2 performance monitoring results. Team consensus had been reached to suspend the weir installation since chlorinated volatile organic compound (CVOC) concentrations adjacent to the pond were below their respective groundwater cleanup target levels (GCTLs) and to re-evaluate the need for the weir prior to expiration of permits from the St. Johns River Water Management District (SJRWMD) and U.S. Army Corps of Engineers (USACE) on 11 July 2023.

## KSCRT Status of Open Action Items

Action Item No.	Minutes Reference	Responsible Team Member	Action item	Status
2210-A06	2210-M08	NASA	<p><b><u>Fire Station #1 (SWMU #116), Sewage Treatment Plant #1 and Sludge Disposal Area (SWMU #117) PFAS Sites Assessment Update:</u></b> The image on Slide 35 displays surface water PFOS results. Future sampling will be focused on influent into the Gator Pond, effluent from the gator pond, associated borrow pits that are part of the stormwater management system, tributaries into Buck Creek, locations within Buck Creek, junction of Buck Creek and Banana River with offsets north and south, isolated borrow pits northeast of gator pond to determine impacts, and four locations along Banana River that will correspond to DPT locations. Tetra Tech will email FDEP the locations of the proposed monitoring well locations.</p>	Open