

# AMMPER

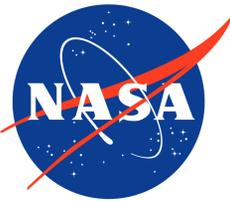
Agent-based Model for Microbial Populations Exposed to Radiation

## A user-friendly agent-based model that recapitulates simple metabolic responses of yeast to deep-space radiation

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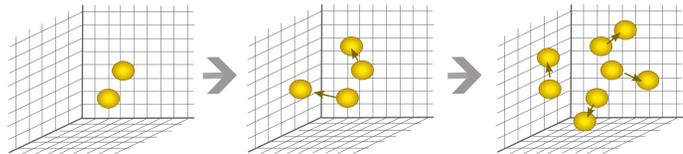


### Yeast cell replication

AMMPER models yeast cells as individual agents. They can:

- move via Brownian motion
- reproduce once per timestep
- be damaged, and repair damage.

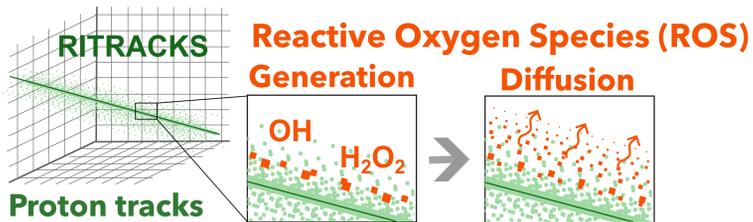
Metabolism is not explicitly simulated in AMMPER.



### Radiation and ROS generation

AMMPER includes energy deposition data for protons, generated by RITRACKS [5]. Users can choose from 3 radiation environments:

- **150 MeV proton:** parallel tracks, 2.5 - 30 Gy
- **GCRSim:** parallel tracks, 20-1000 MeV protons, 1.19 Gy
- **Deep Space:** omnidirectional tracks, 42-120 MeV protons, 4.49 mGy



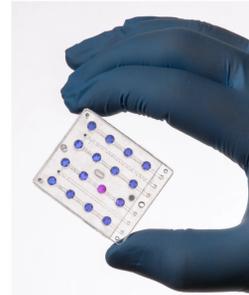
### Gamma (v 2.0)

AMMPER v2.0 includes an option for gamma radiation, simulated as a random distribution of cell damage and ROS across space.

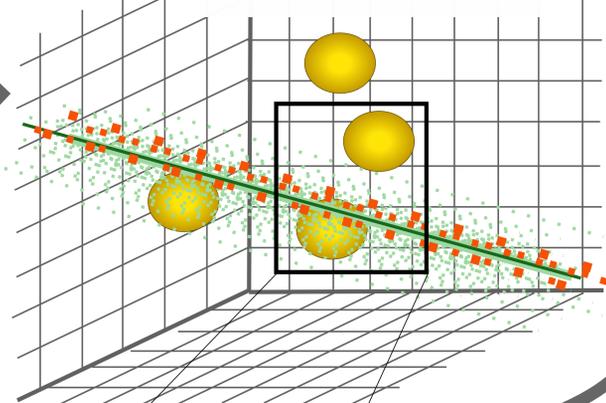
ROS (reactive oxygen species) are generated at a rate of  $2.5 \cdot \text{OH} / 100 \text{ eV}$  or  $0.7 \text{ H}_2\text{O}_2 / 100 \text{ eV}$ . AMMPER v 2.0 includes the option to simulate ROS diffusion through space.

## Overview

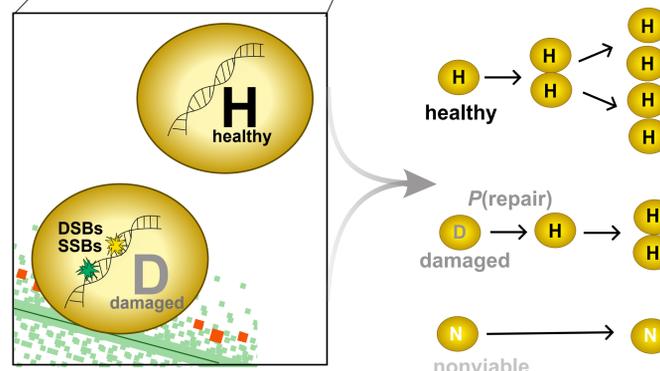
AMMPER is an agent-based model designed to predict the dynamics of yeast growth and alamarBlue reduction upon radiation exposure, to facilitate design and interpretation of NASA flight experiments such as BioSentinel [1-3] and LEIA [4].



Simulation space:  
64 x 64 x 64 um or  
300 x 300 x 300 um



### Cell damage and repair



DSB=Double-Strand Break; SSB=Single-Strand Break

### Each cell accumulates DNA damage

- from radiation: 35 DSBs / cell / Gy
- from ROS: 1 SSB /  $\cdot\text{OH}$  molecule
- $\text{H}_2\text{O}_2$  determines apoptosis (death) probability

### DNA repair is probabilistic

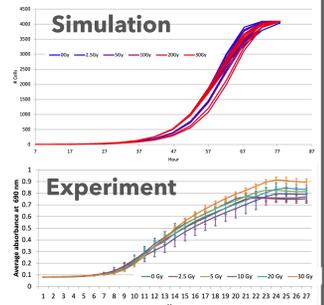
- SSB: 3 per timestep, 100% chance of success
- DSB: 1 per timestep, 80% chance of success

### Damage determines cell health state and activity

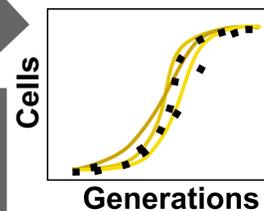
- **H Healthy:** no damage. Reproduces once per timestep. Full alamarBlue reduction rate.
- **D Damaged:** some SSBs or DSBs. No reproduction. alamarBlue reduction rate is reduced by a constant scaling parameter.
- **N Nonviable / dead:** no activity, no chance of repair

### Population growth

AMMPER tracks the number of healthy and damaged cells over time, generating growth curves for comparison with experimental data.

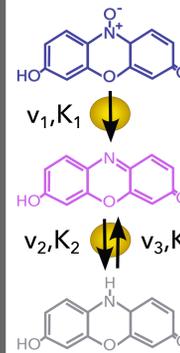


AMMPER simulation (solid line) vs Experimental data (dotted line)



As a stochastic model, it generates variability among replicate runs similar to variability among real biological replicates.

### alamarBlue dynamics (v 2.0)

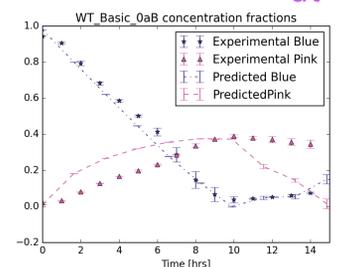


AMMPER v 2.0 simulates dynamics of alamarBlue metabolic dye. After a model run, the number of healthy and damaged cells at each timestep is used to calculate the rate of blue->pink and pink->uncolored reactions, using Michaelis-Menten kinetics.

$$\text{blue} \rightarrow \text{pink} \text{ per cell: } \frac{d[\text{pink}]}{dt} = v_1 \frac{[\text{blue}]}{[\text{blue}] + K_1}$$

$$\text{per timestep: } V_{\text{pink}} = ([\text{healthy}] + k[\text{damaged}]) \frac{d[\text{pink}]}{dt}$$

The pink-> uncolored reaction uses similar equations. Parameters  $v_1, v_2, v_3, K_1, K_2, K_3$  are estimated by fitting to experimental data using Bayesian optimization.



### Preprint and code



Read more about AMMPER v 1.0 in Singh et al. (2023) on bioRxiv! Open-source software available at [github.com/nasa/AMMPER](https://github.com/nasa/AMMPER)



### Experimental data

Empirical data from BioSentinel ground experiments [1-3] are used to:

- estimate basal growth rate
- fit alamarBlue reaction parameters
- test model outcomes

### Graphical user interface (v 2.0)

AMMPER v 2.0 includes a graphical user interface. Learn more at Madeline Marous's poster on Saturday! (UH45)



### Next steps

Stay tuned for the official release of AMMPER v 2.0, including gamma radiation, alamarBlue reduction, and the GUI. Future work includes single-cell experiments for better parameterization.

### Acknowledgments

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### References

- [1] Santa Maria et al. (2020) doi: 10.1089/ast.2019.2073
- [2] Padgen et al. (2021) doi: 10.1089/ast.2020.2305
- [3] Liddell et al. (2023) doi: 10.1089/ast.2022.0124
- [4] <https://www.nasa.gov/technology/science-instruments/what-is-the-lunar-explorer-instrument-for-space-biology-applications/>
- [5] Plante & Wu (2014) NASA software MSC-25937-1