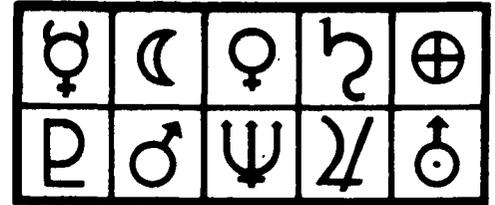


*Research Report*

N 7 2 - 1 1 2 1 0



PLANETARY QUARANTINE

SC-RR-71 0681  
September 1971

A COMPUTERIZED PROGRAM FOR STATISTICAL  
TREATMENT OF BIOLOGICAL DATA

CASE FILE  
COPY

A. L. Roark  
Planetary Quarantine Systems Studies Division  
M. C. Reynolds  
Planetary Quarantine Applied Science Division

SANDIA LABORATORIES



Issued by Sandia Corporation,  
a prime contractor to the United States Atomic Energy Commission

---

**NOTICE**

This report was prepared as an account of work sponsored by the United States Government. Neither the United States nor the United States Atomic Energy Commission, nor any of their employees, nor any of their contractors, subcontractors, or their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness or usefulness of any information, apparatus, product or process disclosed, or represents that its use would not infringe privately owned rights.

SC-RR-71 0681

A COMPUTERIZED PROGRAM FOR STATISTICAL  
TREATMENT OF BIOLOGICAL DATA

A. L. Roark  
Planetary Quarantine Systems Studies Division  
M. C. Reynolds  
Planetary Quarantine Applied Science Division  
Sandia Laboratories, Albuquerque, New Mexico 87115

Completed - September 1971

**Abstract**

Biologists frequently conduct experiments which measure the patterns of inactivation of bacterial populations after exposure to a lethal environment. This document discusses a computer program which calculates many of the quantities that have proven to be useful in the analysis of such experimental data.

---

This work was conducted under Contract No. W-12,853, Planetary Programs, Office of Space Science and Applications, NASA Headquarters, Washington, D. C.

## Contents

	<u>Page</u>
Introduction	5
Determination of Survivors	5
Statistical Methods	7
The Program	15
References	22

## Figures

	<u>Page</u>
1. Sample Assay and Dilution Procedure	6
2. Program Flow Chart	16
3. Input Format	17
4. Output Format	18
5. Example of Input Data	19
6. Example of Output Data	20
7. Graphical Representation of Program Output	21

# A COMPUTERIZED PROGRAM FOR STATISTICAL TREATMENT OF BIOLOGICAL DATA

## Introduction

In the programs now underway in the Planetary Quarantine Department, it is frequently necessary to compare subtle changes in the destruction pattern of microorganisms. The use of standard pour plate techniques<sup>1, 2</sup> for microbial assay during experimentation in some cases yields hundreds of data bits (plate counts). These must be reduced in a way that these successive samples taken during process application represent the destruction rate of microorganisms as a consequence of the process. This destruction rate is best described by a survivor curve since it relates the number of surviving organisms at any time to the sterilization process. The survivor curve is usually a y-axis plot of the logarithm of the number of organisms surviving the sterilization treatment versus the equivalent process time on the x-axis. This process time versus log of survivors or logarithmic model seems to be the most practical representation of data since essentially all thermoradiation and most heat and radiation sterilization has exhibited the logarithmic order of destruction. Consequently, the comparison of treatments can be made on the basis of the slope of the survivor curve or the D-value determined from the slope.

Based on this rationale, a computerized program has been developed to handle the statistical aspects of the data reduction. With plate counts of each successive sampling periods as an input, the program computes the mean value of the replicate plate counts, the variance, standard deviation, upper and lower .95 confidence intervals and the coefficient of variation for each sampling interval. Based on the coefficient of variation values for a sampling period, the dilution or data set exhibiting the best values are selected for each period. These best sets are then used in computing the survivor curve based on a least square fit of the logarithmic model.

## Determination of Survivors

At any specific sampling period the procedure for assay is as follows: Four replicate samples are generally used for each sampling period. Aluminum foils or 0.020" thick square planchets are used as a substrate for the test organisms. After exposure to the sterilization treatment the substrate material is placed in a beaker with 10 ml sterile water and insonated for two minutes to suspend the organisms. From this base suspension, measured amounts of the inoculum are transferred to

petri dishes or additional dilution blanks<sup>3</sup> as required to result in plate counts between 30 and 300 colonies per plate. Within this range, the counts can be accurate, and the possibility of interference of the growth of an organism with that of another is minimized.

The determination of viable population from the resultant plate count is made as follows:

Using the arrangement of dilution\*, Figure 1, the inoculum from each of the four replicate samples for a single time period is plated in duplicate. Consequently, there are eight plates for each sampling period at a single level of dilution. Sometimes as many as three dilutions are plated out with the best set of data used as the surviving population at that sampling period.

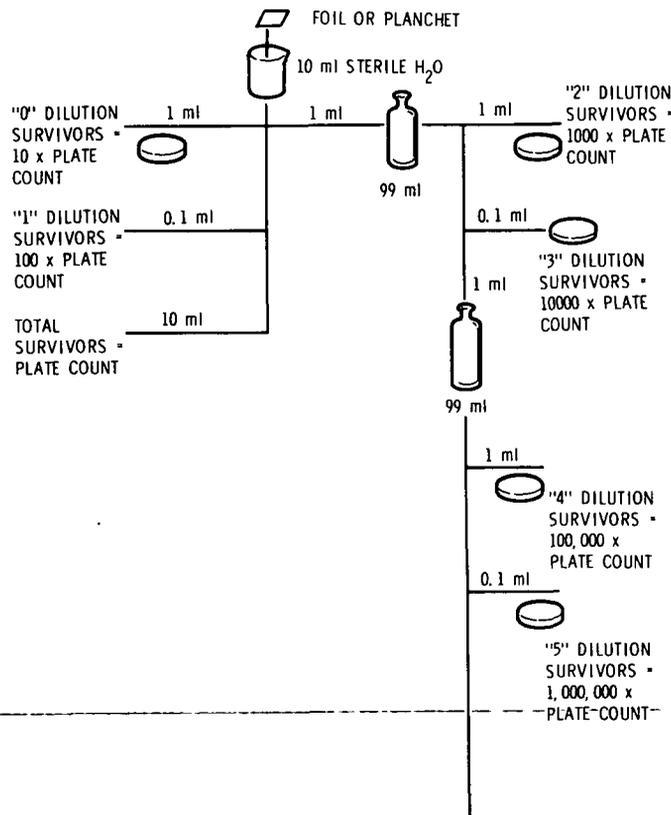


Figure 1. Sample Assay and Dilution Procedure

\*For consistency in the input data, the total survivor plate counts will be assigned on order of dilution of "1".

## Statistical Methods

If we consider a single microorganism of a given type, we see that its loss of viability in a lethal environment is a random event. This fact has been explained in terms of natural variations between microorganisms brought about, in part, by their past history and by the hypothesis that loss of viability is due to the occurrence of chemical reactions<sup>4</sup>. In modeling the inactivation of microorganisms, researchers have usually attempted to derive expressions for the probability of single spore survival as a function of time of exposure to a given environment.

As we have pointed out earlier, instead of looking at the inactivation of a single spore, an experimenter considers the number of survivors in a given population as a function of time. We shall let the random variable  $N(t)$  be the number of survivors at time  $t$  and let  $p(t)$  represent the probability of single spore survival at time  $t$ . The model we shall assume defines the conditional probability as

$$\text{Prob. } \{N(t) = k | N(0) = N_0\} = \binom{N_0}{k} [p(t)]^k [1 - p(t)]^{N_0 - k} \quad (1)$$

Using the definition of conditional probabilities we have

$$\text{Prob. } \{N(t) = k\} = \sum_{N_0=0}^{\infty} \text{Prob. } \{N(0) = N_0\} \text{Prob. } \{N(t) = k | N(0) = N_0\} \quad (2)$$

Combining (1) and (2) yields

$$\text{Prob. } \{N(t) = k\} = \sum_{N_0=0}^{\infty} \binom{N_0}{k} [p(t)]^k [1 - p(t)]^{N_0 - k} \text{Prob. } \{N(0) = N_0\} \quad (3)$$

We are usually interested in the expected value of the number of survivors as a function of time. Using the expression (3) it can be shown that

$$E(N(t)) = E(N(0)) p(t) \quad (4)$$

This is the basic expression for our model. In particular, the most widely used expression for the probability of single spore survival is provided by what is known

as the log model. Using this model we would have

$$p(t) = 10^{-t/D}$$

and thus (4) becomes

$$E(N(t)) = E(N(0)) 10^{-t/D} \quad (4')$$

In this model,  $D$  is assumed to be a fixed parameter for a microorganism belonging to a homogeneous population.

Let us return for a moment to the experimental method used and consider the quantities we wish to compute for each dilution and each time period. Let us define

$$\begin{aligned} x_{ij}(t_\ell) &= \text{number of colonies on plate } i \text{ of dilution } j \\ &\text{at sampling period } \ell, \\ \ell &= 1, \dots, M \\ j &= 1, \dots, K_\ell \\ i &= 1, \dots, N_{j\ell} \end{aligned}$$

where

$$\begin{aligned} M &= \text{number of sampling periods} \\ K_\ell &= \text{number of dilutions at sampling period } \ell \\ N_{j\ell} &= \text{number of plates of dilution } j \text{ for sampling period } \ell \end{aligned}$$

and

$$t_\ell = \text{time of sampling period } \ell \text{ (in any units desired).}$$

The mean of the plate counts for a particular dilution and sampling period is

$$\bar{x}_j(t_\ell) = \frac{1}{N_{j\ell}} \sum_{i=1}^{N_{j\ell}} x_{ij}(t_\ell)$$

while the variance of the distribution of plate counts can be approximated by the sample variance which is given by

$$\begin{aligned}
s_j^2(t_\ell) &= \frac{\sum_{i=1}^{N_{j\ell}} (\bar{x}_j(t_\ell) - x_{ij}(t_\ell))^2}{N_{j\ell} - 1} \\
&= \frac{\sum_{i=1}^{N_{j\ell}} x_{ij}(t_\ell)^2 - N_{j\ell} \bar{x}_j(t_\ell)^2}{N_{j\ell} - 1}
\end{aligned} \tag{5}$$

Similarly, the standard deviation is approximated by the sample standard deviation which is given by

$$s_j(t_\ell) = \sqrt{s_j^2(t_\ell)} \tag{6}$$

To be more precise, let  $\sigma_j^2(t_\ell)$  be the variance in the plate counts (this includes natural variation as well as any errors). The sample variance is a random variable which depends on the counts of the replicate plates. It can be shown that

$$\sigma_j^2(t_\ell) = E(s_j^2(t_\ell))$$

Another desirable quantity for each dilution at each time interval is the confidence interval for the mean. This confidence interval is given for a particular time period by the expression

$$[L_j(t_\ell), U_j(t_\ell)] = \left[ \bar{x}_j(t_\ell) - \frac{ks_j(t_\ell)}{\sqrt{N_{j\ell}}}, \bar{x}_j(t_\ell) + \frac{ks_j(t_\ell)}{\sqrt{N_{j\ell}}} \right] \tag{7}$$

It is well known that as the number of samples,  $N_{j\ell}$ , becomes larger the parameter  $k$  for the  $\alpha$  confidence limit should be chosen as the 100  $\alpha/2$  percentage point of the normal distribution. Thus, for the .95 confidence,  $k = 1.96$  if  $N_{j\ell}$  is large. Unfortunately, the number of plates of a given dilution at a given sampling time is usually small. In this case, the 100  $\alpha/2$  percentage point of the Student's  $t$ -distribution with  $N_{j\ell} - 1$  degrees of freedom is more appropriate for  $k$ . Thus we can approximate  $k$  to sufficient accuracy by

$$k = \chi \left[ 1 + \frac{\chi^2 + 1}{4(N_{j\ell} - 1)} + \frac{(\chi^2 + 3)(5\chi^2 + 1)}{96(N_{j\ell} - 1)^2} \right] , \quad (8)$$

where  $\chi$  is the 100  $\alpha/2$  percentage point of the normal distribution. For our .95 confidence interval we let  $\chi = 1.96$  in (8) to get our  $k$  for (7).

A good measure of the amount of spread in a particular set of data has been found to be the relative standard deviation<sup>5</sup>. This is more commonly known as the coefficient of variation. For each sampling period and each dilution it is defined to be

$$C_j(t_\ell) = \frac{s_j(t_\ell)}{\bar{x}_j(t_\ell)} .$$

In calculating the fit to the data of our "straight line" model, we wish to use the dilution at each time period which has the "tightest" data. We shall use the coefficient of variation as an index of the spread. Therefore, we let

$$X(t_\ell) = \bar{x}_J(t_\ell)$$

and

$$\sigma^2(t_\ell) = \sigma_J^2(t_\ell)$$

where  $J$  is chosen to minimize  $C_j(t_\ell)$  for  $j = 1, \dots, K_\ell$ . Let the order of this dilution (as defined in Figure 1) be  $d_\ell$ .

We are now prepared to again consider the problem of applying our model to the data. Let

$$Y(t_\ell) = X(t_\ell) \times 10^{d_\ell + 1} . \quad (9)$$

Then  $Y(t_\ell)$  is an estimate of  $E(N(t_\ell))$ . In our model we wish to use  $Y(t_\ell)$ ,  $\ell=1, \dots, M$  to determine  $E(N(0))$  and  $D$  as accurately as possible and to obtain some measurements of the statistical variations. Taking the natural log on both sides of (4') we obtain

$$\log E(N(t)) = \log (E(N(0)) + \gamma t) , \quad (10)$$

where

$$\gamma = \frac{2.303}{D} . \quad (11)$$

With this model in mind, let us consider the equation

$$y(t) = \alpha + \beta t + \epsilon \quad (12)$$

where  $\epsilon$  is a random variable representing the variation of the measured values about the line  $\alpha + \beta t$ .

Comparing (10) and (12) we see that we are assuming that

$$\alpha = \log (E(N(0)))$$

or

$$E(N(0)) = e^{\alpha} \quad (13)$$

and

$$\beta = \frac{2.303}{D} = \gamma . \quad (14)$$

The random variable  $\epsilon$  in (12) represents the variation of the mean of the plate counts from the log model. This is assumed to be independent of time. This is consistent with assuming that the distribution of the variation in plate counts from the log model is independent of time. Let

$$y_{\ell} = \log Y(t_{\ell}) .$$

Then  $y_{\ell}$  is a sampled value of the random variable  $\log E(N(t_{\ell}))$ . Let us assume that  $\epsilon$  is normally distributed and that

$$E(\epsilon) = 0.$$

For later convenience, let the variance of the distribution of  $\epsilon$  be represented by  $\sigma_{\epsilon}^2$ .

We are now prepared to calculate  $\alpha$  and  $\beta$ . The following definitions will prove valuable:

$$1. \quad \bar{t} = \frac{\sum_{\ell=1}^M t_{\ell}}{M} \quad (15)$$

$$2. \quad \bar{y} = \frac{\sum_{\ell=1}^M y_{\ell}}{M} \quad (16)$$

$$3. \quad b = \frac{\sum_{\ell=1}^M (t_{\ell} - \bar{t})(y_{\ell} - \bar{y})}{\sum_{\ell=1}^M (t_{\ell} - \bar{t})^2} = \frac{\sum_{\ell=1}^M (t_{\ell} - \bar{t}) y_{\ell}}{\sum_{\ell=1}^M (t_{\ell} - \bar{t})^2} \quad (17)$$

$$4. \quad a = \bar{y} - b\bar{t} \quad (18)$$

The quantities  $a$  and  $b$  depend on the samples used. The Gauss-Markoff theorem<sup>6</sup> tells us that  $a$  and  $b$  coincide with the maximum likelihood estimates of  $\alpha$  and  $\beta$  and that they are unbiased, i. e.,

$$E(a) = \alpha$$

and

$$E(b) = \beta.$$

Letting

$$Z(t) = a + bt$$

and defining the standard error of estimate,  $S_{y|t}$ , by

$$S_{y|\bar{t}}^2 = \frac{\sum_{\ell=1}^M (Z(t_{\ell}) - y_{\ell})^2}{M - 2}, \quad (19)$$

the Gauss-Markoff theorem also tells us that  $S_{y|t}^2$  is an unbiased estimate of  $\sigma_{\epsilon}^2$ , i. e.,<sup>7</sup>

$$E(S_{y|t}^2) = \sigma_{\epsilon}^2.$$

In addition to the Gauss-Markoff theorem, our assumptions on  $\epsilon$  imply that  $a$  and  $b$  are also the minimum variance unbiased estimates of  $\alpha$  and  $\beta$  respectively from among the class of all linear estimates. The standard deviation in  $b$ , which is given by

$$\sigma_b = \sqrt{\sigma_\epsilon^2 / \sum_{l=1}^M (t_l - \bar{t})^2}$$

can be approximated by the standard error in the slope,

$$S_b = \frac{S_{y|t}}{\sqrt{\sum_{l=1}^M (t_l - \bar{t})^2}}$$

Similarly, the standard deviation of the distribution of  $a$ ,

$$\sigma_a = \sigma_\epsilon \sqrt{\frac{1}{M} + \frac{\bar{t}}{\sum_{l=1}^M (t_l - \bar{t})^2}}$$

can be approximated by the standard error in  $a^*$ ,

$$S_a = S_{y|t} \sqrt{\frac{1}{M} + \frac{\bar{t}}{\sum_{l=1}^M (t_l - \bar{t})^2}}$$

where  $S_{y|t}$  is given by (19).

It is also desirable in many applications to have a measure of how closely the variation in the log of the means of plate counts can be explained on the basis of only the variation of time in the lethal environment. The correlation coefficient,  $r$ , is defined by

---

\*The quantity  $S_a$  shall be called the standard error in the estimated intercept.

$$r = \frac{\sum_{\ell=1}^M (t_{\ell} - \bar{t})(y_{\ell} - \bar{y})}{\sqrt{\sum_{\ell=1}^M (t_{\ell} - \bar{t})^2 \sum_{\ell=1}^M (y_{\ell} - \bar{y})^2}} = \frac{\sum_{\ell=1}^M t_{\ell} y_{\ell}}{\sqrt{\sum_{\ell=1}^M t_{\ell}^2 \sum_{\ell=1}^M y_{\ell}^2}}$$

Feller<sup>8</sup> proves the following statements can be made concerning  $r$ :

1.  $|r| \leq 1$
2.  $r = \pm 1$  implies that there exists constants  $\rho$  and  $\theta$  such that  $y = \rho t + \theta$  (except for a set of lines which have zero probability of occurring).

In addition, it can be shown that if  $y$  and  $t$  are independent, then  $r = 0$ . The converse of this statement is not true, however.

Let us return for a moment to the probability of single spore survival. Most microbiologists are interested in the D-value of the population. We have shown that we can approximate the D-value by

$$D = \frac{1}{b \log_{10} e} = \frac{2.303}{b}$$

In addition, the standard error in the estimated D is given by

$$S_D = \frac{2.303 S_b}{b^2} = \frac{D S_b}{b}$$

Another feature which it is sometimes desirable to have available is the confidence band about the curve representing the model. This is also easily computed<sup>6</sup>.  
Let

$$S_Z(t_{\ell}) = S_{y|t} \sqrt{\frac{1}{M} + \frac{t_{\ell} - \bar{t}}{\sum_{\ell=1}^M (t_{\ell} - \bar{t})^2}}$$

Then the upper 95% confidence line is given by

$$Z_u(t_{\ell}) = a + b t_{\ell} + k S_Z(t_{\ell})$$

and the lower by

$$Z_L(t_\ell) = a + b t_\ell - k S_Z(t_\ell)$$

where  $k$  is given by the Student's  $t$ -distribution of degree  $M-2$ . For the .95 confidence interval  $k$  is approximated by

$$k = \chi \left[ 1 + \frac{\chi^2 + 1}{4(M-2)} + \frac{(\chi^2 + 3)(5\chi^2 + 1)}{96(M-2)^2} \right]$$

where  $\chi = 1.96$ .

## The Program

The flow chart for the program is given in Figure 2. This is self-explanatory. The input is prepared in the manner illustrated in Figure 3. The output is described in Figure 4 using the notation of the previous section.

Figure 5 provides an example of the input data while Figure 6 gives the output from the use of the program on this example.

Finally, a graphical representation of the data, the model, and the .95 confidence interval is shown graphically in Figure 7.

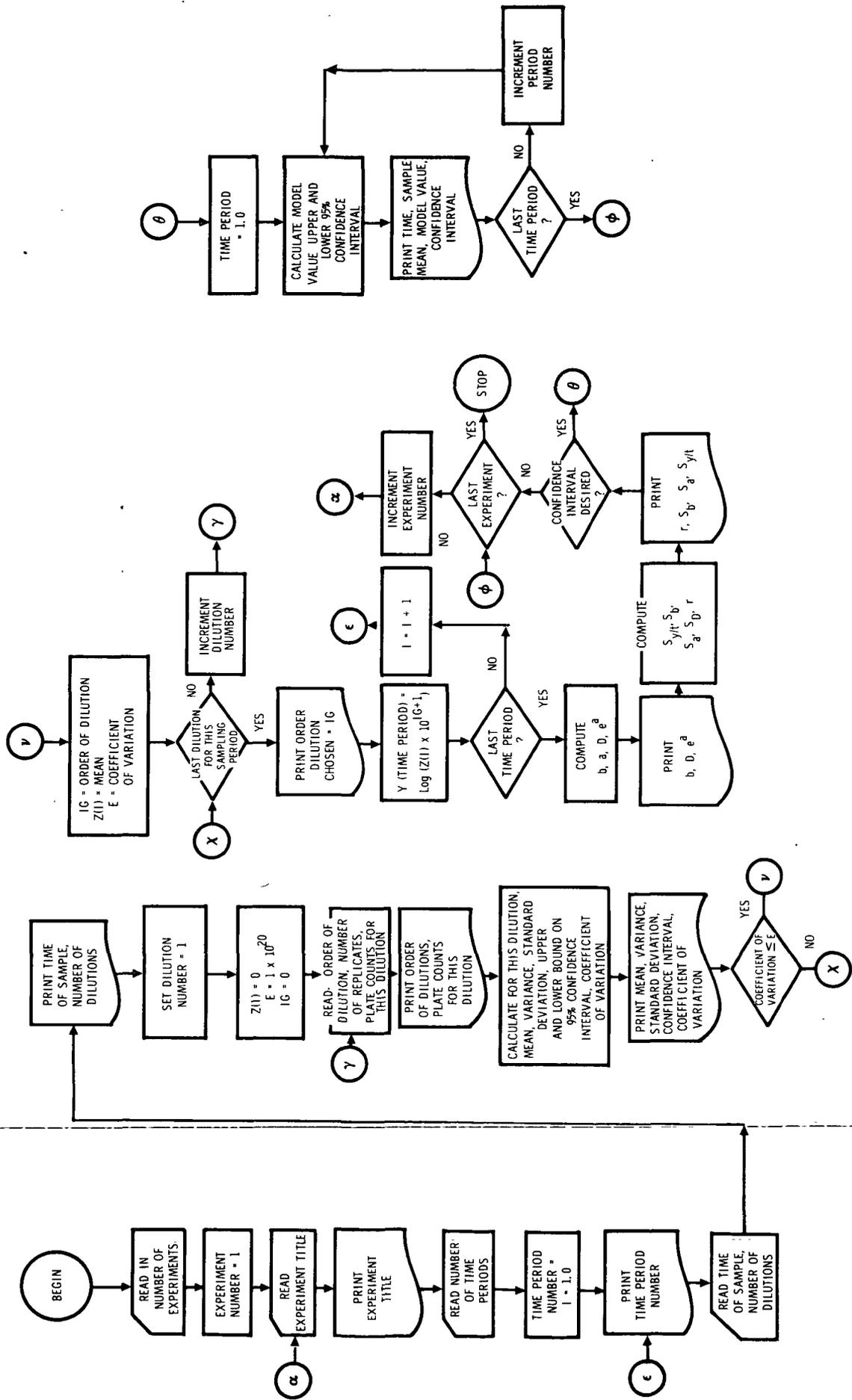


Figure 2. Program Flow Chart

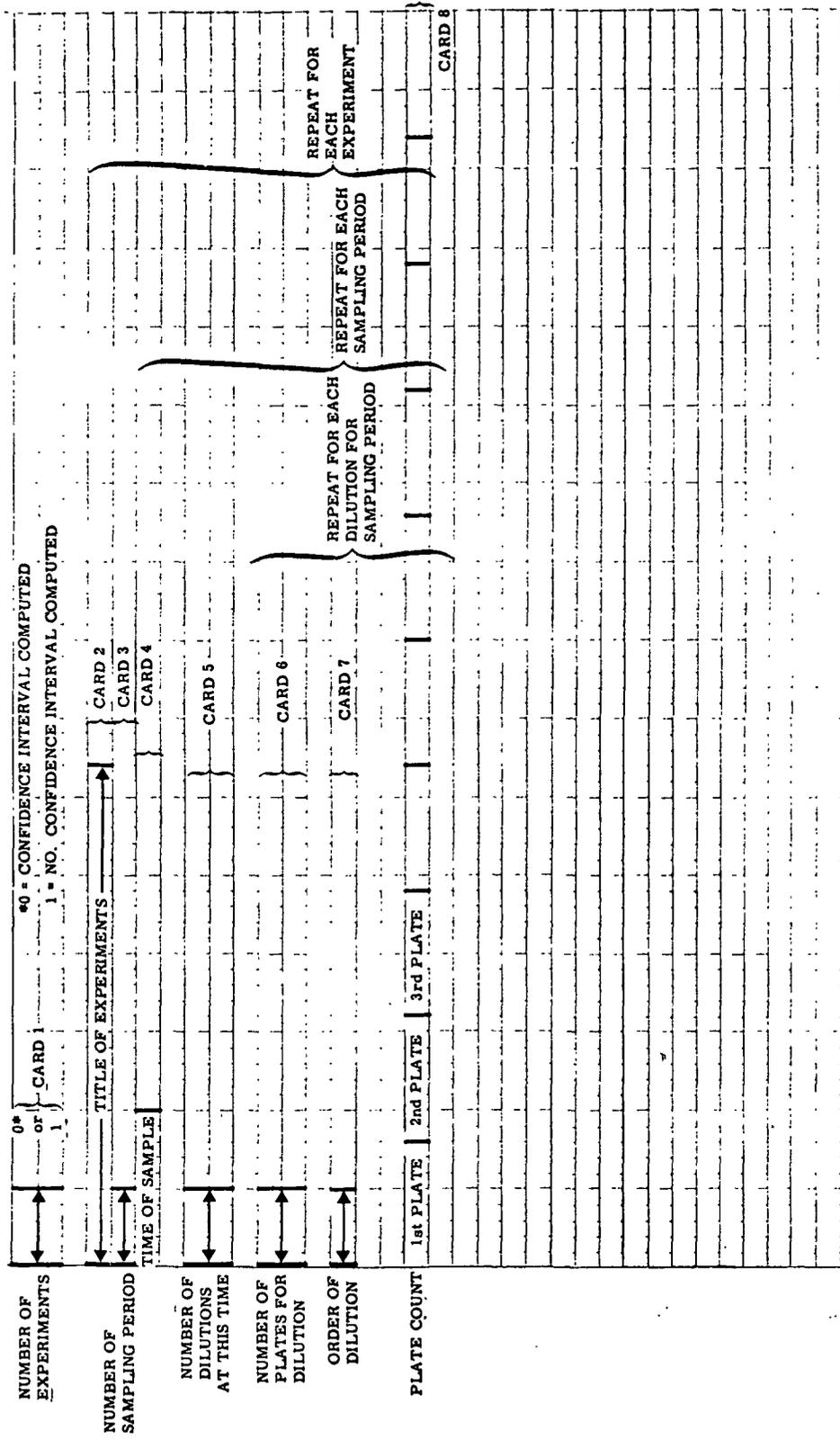


Figure 3. Input Format

TITLE OF EXPERIMENT

Repeat for each sampling time period  
 Data Set = \_\_\_\_\_  
 Time = \_\_\_\_\_ (For this time period)  
 Number of Dilutions = \_\_\_\_\_ (Number of plate counts for this time period and dilution)  
 Number of Data Points = \_\_\_\_\_  
 Order of Dilution = \_\_\_\_\_  
 Date (Plate Counts): \_\_\_\_\_  
 Mean = \_\_\_\_\_ Variance = \_\_\_\_\_ S.D. = \_\_\_\_\_ Upper .95 C.I. = \_\_\_\_\_ C.V. = \_\_\_\_\_  
 (Standard Deviation) (Upper limit of 95% Confidence Interval)(C.I.) (Coefficient of Variation)  
 Dilution Chosen = \_\_\_\_\_ (Dilution chosen for this time period on the basis of the coefficient of variation)  
 Lower .95 C.I. = \_\_\_\_\_  
 Slope = \_\_\_\_\_ (Slope of line of best fit to log of selected means) D-Value = \_\_\_\_\_ Intercept = \_\_\_\_\_ (Theoretical value of initial population)  
 Corr. Coef. = \_\_\_\_\_ Stand. Err. in Est. Slope = \_\_\_\_\_ Stand. Err. of Est. = \_\_\_\_\_

T (Time)	SAMP (Data)	MODEL (Log Model Value)	Upper (Upper Limit of 95% Confidence Band)	Lower (Lower Limit of 95% Confidence Band)
_____	_____	_____	_____	_____
_____	_____	_____	_____	_____
_____	_____	_____	_____	_____
_____	_____	_____	_____	_____
_____	_____	_____	_____	_____

\* In computer output, E + 05 means 10<sup>5</sup> e.g., 2.31E + 06 represents 2.31 x 10<sup>6</sup>

Figure 4. Output Format

```

1
24 NOVEMBER 1970
8
0.0
1
8
3
265. 282. 297. 267. 250. 277. 265. 285.
3.0
1
8
3
67. 58. 62. 57. 51. 75. 80. 72.
6.0
1
8
2
58. 63. 80. 83. 62. 66. 75. 63.
9.0
1
8
1
278. 212. 197. 201. 214. 255. 236. 231.
12.0
1
8
1
48. 37. 37. 25. 25. 21. 42. 29.
15.0
1
8
0
116. 135. 87. 62. 95. 82. 85. 84.
18.0
2
8
0
26. 16. 12. 14. 11. 10. 18. 9.
4
-1
157. 117. 92. 162.
21.0
1
4
-1
51. 50. 93. 89.

```

Figure 5. Example of Input Data

24 NOVEMBER 1970

```
DATA SET = 1
TIME= 0.000
NO. DIL.= 1
NUMBER DATA POINTS= 8
ORDER OF DIL. = 3
DATA
265.00 282.00 292.00 267.00 250.00 277.00 265.00 285.00
MEAN= 272.875 VARIANCE= 135.0 S.D.= 13.6 UPPER .95 C.I.= 284.2 LOWER .95 C.I.= 261.5 CV = .0498
CIL. CHOSEN = 3

DATA SET = 2
TIME= 3.000
NO. DIL.= 1
NUMBER DATA POINTS= 8
ORDER OF DIL. = 3
DATA
67.00 58.00 62.00 57.00 51.00 75.00 80.00 72.00
MEAN= 65.250 VARIANCE= 99.4 S.D.= 10.0 UPPER .95 C.I.= 73.6 LOWER .95 C.I.= 56.9 CV = .1528
CIL. CHOSEN = 3

DATA SET = 3
TIME= 6.000
NO. DIL.= 1
NUMBER DATA POINTS= 8
ORDER OF DIL. = 2
DATA
58.00 63.00 80.00 83.00 62.00 66.00 75.00 63.00
MEAN= 69.750 VARIANCE= 86.2 S.D.= 9.3 UPPER .95 C.I.= 76.5 LOWER .95 C.I.= 61.0 CV = .1351
CIL. CHOSEN = 2

DATA SET = 4
TIME= 9.000
NO. DIL.= 1
NUMBER DATA POINTS= 8
ORDER OF DIL. = 1
DATA
278.00 212.00 197.00 201.00 214.00 255.00 236.00 231.00
MEAN= 223.000 VARIANCE= 777.7 S.D.= 27.9 UPPER .95 C.I.= 251.2 LOWER .95 C.I.= 204.3 CV = .1223
CIL. CHOSEN = 1

DATA SET = 5
TIME= 12.000
NO. DIL.= 1
NUMBER DATA POINTS= 8
ORDER OF DIL. = 1
DATA
48.00 37.00 37.00 25.00 25.00 21.00 42.00 29.00
MEAN= 33.000 VARIANCE= 99.4 S.D.= 9.5 UPPER .95 C.I.= 40.9 LOWER .95 C.I.= 25.1 CV = .2866
CIL. CHOSEN = 1

DATA SET = 6
TIME= 15.000
NO. DIL.= 1
NUMBER DATA POINTS= 8
ORDER OF DIL. = 0
DATA
116.00 135.00 87.00 62.00 95.00 82.00 85.00 84.00
MEAN= 93.250 VARIANCE= 508.5 S.D.= 22.5 UPPER .95 C.I.= 112.0 LOWER .95 C.I.= 74.5 CV = .2418
CIL. CHOSEN = 0

DATA SET = 7
TIME= 18.000
NO. DIL.= 2
NUMBER DATA POINTS= 8
ORDER OF DIL. = 0
DATA
26.00 16.00 12.00 14.00 11.00 10.00 18.00 9.00
MEAN= 14.500 VARIANCE= 30.9 S.D.= 5.6 UPPER .95 C.I.= 19.1 LOWER .95 C.I.= 9.9 CV = .3831
NUMBER DATA POINTS= 4
ORDER OF DIL. = -1
DATA
157.00 117.00 92.00 162.00
MEAN= 132.000 VARIANCE= 1116.7 S.D.= 33.4 UPPER .95 C.I.= 183.2 LOWER .95 C.I.= 80.8 CV = .2532
CIL. CHOSEN = -1

DATA SET = 8
TIME= 21.000
NO. DIL.= 1
NUMBER DATA POINTS= 4
ORDER OF DIL. = -1
DATA
51.00 50.00 93.00 89.00
MEAN= 70.750 VARIANCE= 549.6 S.D.= 23.4 UPPER .95 C.I.= 106.7 LOWER .95 C.I.= 34.8 CV = .3314
CIL. CHOSEN = -1
```

```
SLOPE= -.521 D VALUE= 4.421 INTERCEPT= 2.3104308639E+06
CORR. COEF.= .98018 STAND. ERR. IN EST. SLOPE= .01896 STAND. ERR. OF EST.= .13595 STAND. ERR. IN EST. INTER.= .19913
.95 CONF. INTERVAL
T SAMPL MODEL UPPER LOWER
0. 2.7287500000E+06 2.3104308639E+06 4.1234704598E+06 1.2945626333E+06
3.0000000000E+00 6.5250000000E+05 4.8409570870E+05 7.74220E3309E+05 3.0268975685E+05
6.0000000000E+00 6.8750000000E+04 1.0143071530E+05 1.4820424844E+05 6.3418995170E+04
9.0000000000E+00 2.2800000000E+04 2.1252388364E+04 2.9406330336E+04 1.5359414317E+04
1.2000000000E+01 3.3000000000E+03 4.4529313417E+03 6.1613954984E+03 3.211991139E+03
1.5000000000E+01 9.3250000000E+02 9.3300560831E+02 1.3632497272E+03 6.3854732390E+02
1.8000000000E+01 1.3200000000E+02 1.9548908311E+02 3.1264826143E+02 1.2223314929E+02
2.1000000000E+01 7.0750000000E+01 4.09E0077062E+01 7.3102238390E+01 2.2950431476E+01
```

Figure 6. Example of Output Data

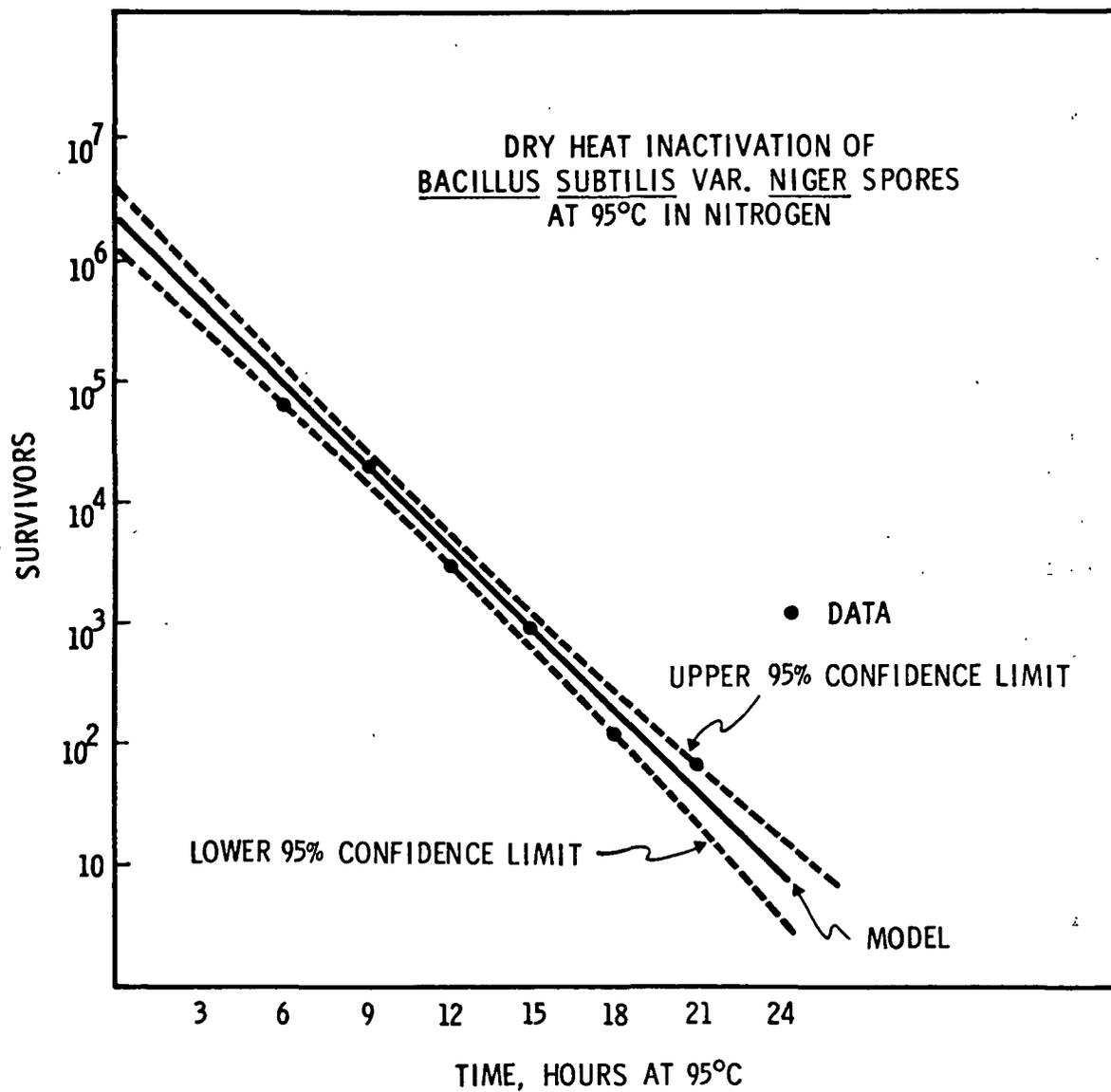


Figure 7. Graphical Representation of Program Output

## References

1. NASA Standard Procedures for the Microbial Examination of Space Hardware. NHB-5340-1, National Aeronautics and Space Administration, August 1967.
2. Pelczar, Jr., M. I., Microbiology, McGraw Hill, New York, 1965.
3. B. B. L. Manual of Products and Laboratory Procedures, 5th Edition, BioQuest, Division of Becton, Dickinson and Company, Cockeysville, Maryland, 1968.
4. Brannen, J. P., "A Rational Model for Thermal Sterilization of Microorganisms," Math. Biosci., Vol. 2 (1968), pp. 165-179.
5. Wallis, W. A., and H. V. Roberts, Statistics, a New Approach, The Free Press, Glencoe, Illinois, 1956.
6. Hillier, F. S. and G. J. Lieberman, Introduction to Operations Research, Holden-Day, Inc., San Francisco, California, 1967.
7. Hemmerle, W. J., Statistical Computations on a Digital Computer, Blaisdel Publishing Company, Waltham, Massachusetts, 1967.
8. Feller, W., An Introduction to Probability Theory and Its Applications, Vol. 1, John Wiley & Sons, Inc., New York, 1968.

DISTRIBUTION:

NASA, Code SL  
Grants and Contracts  
400 Maryland Avenue S. W.  
Washington, D. C. 20546 (25)

L. B. Hall, Code SL  
NASA Headquarters  
400 Maryland Avenue S. W.  
Washington, D. C. 20546 (25)

B. W. Colston, Director  
Space & Special Programs Division  
Office of Operations  
U. S. Atomic Energy Commission  
Albuquerque, New Mexico 87115

L. P. Daspit, Jr.  
Viking Project Quarantine Officer  
Viking Project Office, NASA  
Langley Research Center  
Hampton, Virginia 23365

University of California  
P. O. Box 808  
Livermore, California 94551  
Attn: Tech. Info. Div.  
For: Report Librarian

Los Alamos Scientific Laboratory  
P. O. Box 1663  
Los Alamos, New Mexico 87544  
Attn: Report Librarian

Richard G. Bond  
School of Public Health  
College of Medical Science  
University of Minnesota  
Minneapolis, Minnesota 55455

John H. Brewer  
Mountain View Road  
Star Route 2  
Brownwood, Texas 76801

Edward B. Kasner  
Director of Research Services  
Graduate College  
University of New Mexico  
Albuquerque, New Mexico 87106

Frank B. Engley, Jr., Chairman  
Department of Microbiology  
School of Medicine  
University of Missouri  
Columbia, Missouri 65201

Gilbert V. Levin  
Biospherics, Inc.  
4928 Wyaconda Road  
Rockville, Maryland 20853

Irving J. Pflug  
Professor of Environmental Health  
545 Space Science Center  
University of Minnesota  
Minneapolis, Minnesota 55455

Gerald J. Silverman  
Department of the Army  
U. S. Army Natick Laboratories  
Natick, Massachusetts 01760

John A. Ulrich  
Department of Microbiology  
School of Medicine  
University of New Mexico  
Albuquerque, New Mexico 87106

Samuel Schalkowsky  
Exotech Systems, Inc.  
525 School Street S. W.  
Washington, D. C. 20024

Mark A. Chatigny  
Research Engineer  
Naval Biological Laboratory  
Naval Supply Center  
University of California, Berkeley  
Oakland, California 94625

Richard G. Cornell  
Associate Professor of Statistics  
Department of Statistics  
Florida State University  
Tallahassee, Florida 32306

Dr. Richard C. Corlett  
Department of Mechanical Engineering  
University of Washington  
Seattle, Washington 98105

DISTRIBUTION (Cont)

Martin S. Favero  
Department of Health, Education  
and Welfare  
CDC-Phoenix Field Station  
4402 North 7th Street  
Phoenix, Arizona 85014

Mr. James Martin  
Viking Project Engineer  
Langley Research Center, NASA  
Langley Station  
Hampton, Virginia 23365

Q. Ussery  
Code NC3, Quality Assurance Branch  
Manned Spacecraft Center, NASA  
Houston, Texas

F. J. Beyerle  
George C. Marshall Space Flight Center  
Manufacturing Engineering Laboratory  
Code R-ME-MMC  
Huntsville, Alabama 35812

J. Gayle  
Code SOP  
Kennedy Space Center, NASA  
Cape Canaveral, Florida

Murray Schulman  
Division of Biology and Medicine  
Headquarters, AEC  
Washington, D. C. 20545

N. H. MacLeod  
Space Biology Branch  
Code 624, Bldg. 21, Room 161  
Goddard Space Flight Center  
Greenbelt, Maryland 20771

Jeptha E. Campbell, Ph. D., Chief  
Division of Microbiology  
Food and Drug Administration  
DHEW, Public Health Service  
1090 Tusculum Avenue  
Cincinnati, Ohio 54226

G. Rotariu  
Process Radiation Staff  
Division of Isotopes Development  
Headquarters, AEC  
Washington, D. C. 20545

Martin G. Koesterer, Microbiologist  
Bioscience Operation  
Valley Forge Space Technology Center  
P. O. Box 8555  
Philadelphia, Pennsylvania 19101

Carl W. Bruch, Chief  
Drug Microbiology Branch  
BD415  
Food and Drug Administration  
200 C Street S. W.  
Washington, D. C. 20204

John W. Beakley  
Department of Biology  
University of New Mexico  
Albuquerque, New Mexico 87106

Loren D. Potter, Chairman  
Department of Biology  
University of New Mexico  
Albuquerque, New Mexico 87106

Loris W. Hughes  
Department of Biology  
New Mexico State University  
University Park, New Mexico

Richard W. Porter  
Corporate Engineering Staff  
General Electric Company  
570 Lexington Avenue  
New York, New York 10022

Fred L. Whipple  
Smithsonian Astrophysical Observatory  
Cambridge, Massachusetts 02138

J. J. McDade  
Environmental Research Laboratory  
Building 1710  
Dow Chemical Company  
Midland, Michigan 48640

Otto Hamberg  
Aerospace Corporation  
Building A2, Room 2019  
2350 East El Segundo Blvd.  
El Segundo, California

DISTRIBUTION (Cont)

Lawrence P. Chambers  
NASA Headquarters  
Office of Manned Space Flight  
Code MLR  
Washington, D. C. 20546

Arthur H. Neill  
Code SL  
400 Maryland Avenue S. W.  
Washington, D. C. 20546

Richard H. Green  
Sterilization Group  
Jet Propulsion Laboratory  
4800 Oak Grove Drive  
Pasadena, California 91103

Rudy Puleo  
Spacecraft Bioassay Unit  
Center for Disease Control  
USPHS  
Cape Kennedy, AFS, Florida 32900

USAEC, Division of Technical Information  
P. O. Box 62  
Oak Ridge, Tennessee 37830  
Attn: Reference Branch  
P. E. Postell

Carl Sagan  
Cornell University  
Center for Radiophysics and Space Research  
Space Science Building  
Ithaca, New York 14850

Document Library  
Lovelace Foundation for Medical  
Education and Research  
5200 Gibson Blvd. S. E.  
Albuquerque, New Mexico 87108

Martin S. Tierney  
Group J-10  
Los Alamos Scientific Laboratory  
Los Alamos, New Mexico 87544

Jack Kaye  
11607 Georgetowne Court  
Potomac, Maryland 20854

Dr. Robert Angelotti  
Office of Food Sanitation, FDA  
200 C Street S. W. (BF-201)  
Washington, D. C. 20204

Vance I. Oyama, Chief  
Life Detection Systems Branch  
NASA, Ames Research Center  
Moffett Field, California 94035

Byron W. Brown, Jr.  
Department of Preventive Medicine  
Stanford University School of Medicine  
Stanford University Medical School  
Palo Alto, California 94304

Don G. Fox  
Sterility Control Officer  
NASA Headquarters, Code SL  
400 Maryland Avenue S. W.  
Washington, D. C. 20546

A. A. Rothstein  
Manager, Planetary Quarantine  
Viking Program  
Martin Marietta Corporation  
P. O. Box 179  
Denver, Colorado 80201

Hillel S. Levinson  
U. S. Army Natick Laboratory  
Natick, Massachusetts 01760

Dr. Walter M. Urbain  
College of Agriculture  
Michigan State University  
East Lansing, Michigan 48823

H. O. Halvorson  
1901 East River Road  
Minneapolis, Minnesota 55414

A. Anellis  
U. S. Army Natick Laboratories  
Natick, Massachusetts 01760

H. W. Johnson, LTC  
U. S. Army Medical Research and  
Development Command  
Washington, D. C. 20314

DISTRIBUTION (Cont)

Donald A. Kautter  
Department of HEW  
Food and Drug Administration  
Division of Microbiology  
BF-135  
200 C Street S. W.  
Washington, D. C. 20204

Lt. Keith C. Hopkins (SAH)  
Biomedical Group  
Air Force Weapons Laboratory  
Kirtland Air Force Base  
Albuquerque, New Mexico 87115

Briggs Phillips  
Becton, Dickinson Research Center  
P. O. Box 11276  
Raleigh, North Carolina 27604

Dr. Wolf Vishniac  
Department of Biology  
University of Rochester  
Rochester, New York 14627

F. A. Leone, Program Manager  
Radiation Preservation of Foods  
Division of Isotope Development  
AEC Headquarters, Mail Sta. 255  
Washington, D. C. 20545

Dr. Orr Reynolds  
American Physiological Society  
2134 LeRoy Place N. W.  
Washington, D. C. 20008

Dr. Henry Eyring  
Chemistry Department  
University of Utah  
Salt Lake City, Utah 84112

Dr. Allan H. Brown  
Department of Biology  
University of Pennsylvania  
Philadelphia, Pa. 19104

Mr. Marvin Morris  
20 Whitman Road, Apt. 1-1  
Waltham, Mass. 02154

J. A. Hornbeck, 1  
J. M. Wiesen, 100  
W. J. Howard, 1000  
D. B. Shuster, 1200  
C. B. McCampbell, 1310  
W. A. Gardner, 1500  
H. E. Lenander, 1600  
T. M. Burford, 1700  
C. Winter, 1710  
T. M. Burford (Actg.), 1720  
J. M. Worrell, Jr., 1721  
D. P. Peterson, 1724  
R. G. Clem, 1730  
H. E. Sivinski, 1740 (35)  
A. A. Lieber, 1750  
B. H. Van Domelen, 1913  
A. M. Clogston, 5000  
L. C. Hebel, 5200  
J. V. Walker, 5220  
R. M. Jefferson, 5221  
J. E. McDonald, 5300  
L. M. Berry, 5500  
R. E. Henderson, 7000  
G. A. Fowler, 9000  
J. H. Scott, 9200  
L. Hollingsworth, 9300  
L. A. Hopkins, Jr., 9400  
D. W. Ballard, 9461  
R. S. Gillespie, 3151 (3) (For Public Release)  
L. S. Ostrander, 8232  
W. K. Cox, 3142-1 (15)

q/c