Pharmacologic Atrial Natriuretic Peptide Reduces Human Leg Capillary Filtration

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Summary: Atrial natriuretic peptide (ANP) is produced and secreted by atrial cells. We measured calf capillary filtration rate with prolonged venous-occlusion plethysmography of supine healthy male subjects during pharmacologic infusion of ANP (48 pmol/kg/min for 15 min; n = 6) and during placebo infusion (n = 7). Results during infusions were compared to prior control measurements. ANP infusion increased plasma [ANP] from 30 ± 4 to 2,568 ± 595 pmol/L. Systemic hemoconcentration occurred during ANP infusion: mean hematocrit and plasma colloid osmotic pressure increased 4.6 and 11.3%, respectively, relative to preinfusion baseline values (p < 0.05). Mean calf filtration, however, was significantly reduced from 0.15 to 0.08 ml/100 ml/min with ANP. Heart rate increased 20% with ANP infusion, whereas blood pressure was unchanged. Calf conductance (blood flow/arterial pressure) and venous compliance were unaffected by ANP infusion. Placebo infusion had no effect relative to prior baseline control measurements. Although ANP induced systemic capillary filtration, in the calf, filtration was reduced with ANP. Therefore, pharmacologic ANP infusion enhances capillary filtration from the systemic circulation, perhaps at upper body or splanchnic sites or both, while having the opposite effect in the leg. Key Words: Extracellular fluid—Hemoconcentration—Capillary permeability—Capillary pressure.

Atrial natriuretic peptide (ANP) is a hormone produced and secreted by atrial myocytes on distention of the atria (1,2). ANP is an established vasodilator, yet may also decrease blood pressure and cardiac output by reducing intravascular volume and cardiac filling pressure, because increases in hematocrit and plasma protein concentration are commonly associated with ANP administration (3). Furthermore, ANP-induced hematocrit elevation has been observed in nephrectomized animals (4,5), and it occurs too quickly to be attributed solely to urinary loss. It seems that ANP somehow augments systemic filtration, or at least discourages reabsorption of extracellular fluid into the circulation.

Three studies have investigated effects of ANP on capillary filtration in the human forearm using prolonged venous-occlusion plethysmography. Two found that ANP increased forearm capillary filtration up to 63% (6,7), whereas the third showed only an insignificant trend toward elevation of forearm filtration (8). Using similar methods, we sought to determine whether ANP affects capillary filtration rate as measured in legs of human subjects; we hypothesized that ANP produces systemic hemoconcentration in part by increasing calf capillary filtration. We also attempted to relate effects on calf filtration rate to peripheral arteriolar or venous dilation or both, and to two systemic indices of hemoconcentration, hematocrit and plasma colloid osmotic pressure. The legs are important sites for study of capillary filtration because gravitational pressures in the circulation encourage leg filtration while upright, and because disease states such as congestive heart failure may be associated with supraphysiologic plasma [ANP] (1) and leg edema. Prolonged venous occlusion holds venous and capillary pressures relatively constant, which emphasizes effects of other Starling forces and permeability changes on capillary fluid movement. Conversely, effects of factors that alter filtration by
changing capillary pressure cannot be assessed by
this technique.

METHODS

Subjects
Ten healthy male subjects gave written, informed con-
sent to participate. All were normotensive (arterial blood
pressures ≤140/90), unmedicated, did not use tobacco,
and were not obese (life insurance tables). They were 24
to 47 years old (mean, 33 years), 180 ± 3 cm tall (mean ±
SEM), and they weighed 75.6 ± 2.9 kg. Female subjects
were not recruited because of potential reproductive risks
of radiation imposed by this research.

ANP and placebo infusions
We used synthetic 26 amino acid met-ANP provided by
Merck, Sharp, and Dohme Research Laboratories. Ly-
ophilitized ANP was reconstituted in isotonic saline such
that infusion of the resulting solution at 1.17 ml/min
would deliver 48 pmol ANP/kg/min (150 ng/kg/min) for 15
min to the subject. Placebo infusion consisted of an equal
amount of isotonic saline (17.6 ml total). This research
was approved by the UT Southwestern Human Research
and Radiation Safety Committees, and by the U.S. Food
and Drug Administration (Investigational New Drug num-
ber 30,392).

Plasma ANP radioimmunoassay
Atrial natriuretic peptide-like immunoreactivity in un-
extracted human plasma was measured with radioim-
unoassay (RIA) kits purchased from Research and Di-
agnostic Antibodies, Inc., as previously described (9).
Extraction is not called for by this RIA, and ANP mea-
surement in unextracted plasma is a simpler technique
with which we (9) and others (10) have had consistent and
reliable results previously. Briefly, 7-ml blood samples
were drawn into EDTA tubes after withdrawal of a 2-ml
sample to avoid dilution of blood with the saline in the
thawed plasma samples were centrifuged (20 min at 1,300
转速) with microhematocrit centrifugation. Plasma col-
loid osmotic pressure was measured in 100-μl samples
to avoid dilution of blood with the saline in the
catheter. Blood was drawn from the arm before and at the
end of control and infusion periods. The total volume of
blood drawn during a study equaled ~40 ml and was re-
placed with an equal volume of isotonic saline. Chilled
blood samples were centrifuged (20 min at 1,800 g), and
plasma aliquots were frozen at −20°C. Duplicate 100-μl
thawed plasma samples were assayed according to the
RIA kit instructions. The mean correlation coefficient for
standard curves was 0.994. Intra- and interassay coeffi-
cients of variation were 10.3 and 17.4%, respectively.
Percentage recovery by this RIA of ANP added to plasma
samples averaged 83.1 ± 7.2% (SEM) over a range from
1 to 7 fmol ANP/tube.

Resting calf blood flow and conductance
Subdiastolic venous-occlusion plethysmography was
employed with a Mercury-in-silastic strain gauge to mea-
sure calf blood flow. This method is well established and
described in the literature (11). The strain gauge is placed
around the point of maximum calf girth, and a pressure
cuff is placed proximal to the knee. Inflation of the cuff to
a subdiastolic pressure (50 mm Hg) impedes venous drainage from the calf without affecting arterial inflow.
Conductance is calculated by dividing flow by mean ar-
terial pressure and provides an index of local vasodila-
tion.

Calf capillary filtration rate, blood volume, and
vascular compliance
Calf capillary filtration rate, blood volume, and vascu-
lar compliance were measured by combining venous-
occlusion plethysmography (to measure relative calf vol-
ume changes) with blood imaging (99mTc technetium-
labeled red blood cells (12) to measure relative calf blood
volume (13,14). Two ml of centrifuge-packed erythrocy-
tes (5 min at 1,300 g) were labeled in vitro with 20 mCi
99mTc according to instructions provided with the kit
from Cadema Medical Products, Inc. (Middletown, NY,
U.S.A.). Labeling of centrifuge-packed erythrocytes
avoids undesirable and incidental labeling of plasma pro-
teins. Before injection of the labeled erythrocytes, per-
centage binding of Tc to erythrocytes was quantified by
centrifuging an aliquot of the labeled sample in a micro-
hematocrit centrifuge and then dividing gamma counts in
the packed cells by counts in cells plus supernatant. Per-
centage of Tc bound to erythrocytes was always >96% and
usually >98%, thereby confirming minimal free and
protein-bound Tc. Tc percentage binding remained at or
above these levels in vivo. A Picker Digital Dyna Gamma
Camera acquired images of both legs. Gamma counts
were corrected for background and decay. One leg, de-
termined at random, was instrumented for venous-
occlusion plethysmography, and the other served as an
unoccluded control.

Plethysmographic calf volume increase during the rapid
arterial inflow phase of venous occlusion (the first
minute, when filtration is assumed to be negligible) cali-
brates the radionuclide blood volume technique (13,14).
Baseline calf blood volume percentage is calculated by
applying the relationship between plethysmographic calf-
volume elevation and gamma counts during the first
minute of occlusion to the preclosure gamma level (14).
Prolonged venous occlusion consisted of 9 min at 30 mm
Hg. Calf vascular compliance was quantified as the in-
crease in calf blood volume at 3 min of venous occlusion.
Calf capillary filtration rate during venous occlusion
equals elevation of plethysmographic calf volume minus
the elevation of scintigraphic calf blood volume per unit
time (i.e., rate of extravascular volume elevation). Calf
filtration was quantified between 1 and 9 min of occlu-
sion.

Other dependent variables
Heart rate was determined from ECG. A Narco Bio-
systems PE-300 electrosphygmomanometer indirectly
measured brachial arterial blood pressure. Mean arterial
pressure (MAP) was calculated as:

\[
\text{MAP} = \text{DBP} + \frac{[\text{SBP} - \text{DBP}]}{3}
\]

We measured hematocrit of unoccluded arm venous
blood with microhematocrit centrifugation. Plasma col-
loid osmotic pressure was measured in 100-μl samples
with a membrane-exclusion colloid osmometer modified
from Aukland and Johnsen (15). Samples were taken from
those collected for ANP analysis. Plasma colloid osmotic
pressure was not measured in the placebo group, because
hematocrit did not change in that group (see Results).

Protocol
Subjects were instructed to abstain from caffeine, med-
ications, and alcohol for at least 24 h before the study. All
experiments were performed during mid-afternoon in the

TABLE 1. Influence of ANP infusion on plasma [ANP], hemodynamics, plasma colloid osmotic pressure, and calf capillary filtration

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>ANP</th>
</tr>
</thead>
<tbody>
<tr>
<td>ANP (pmol/L)</td>
<td>30 ± 4</td>
<td>2568 ± 595a</td>
</tr>
<tr>
<td>HR (beats/min)</td>
<td>61 ± 5</td>
<td>73 ± 6a</td>
</tr>
<tr>
<td>MAP (mm Hg)</td>
<td>78 ± 3</td>
<td>77 ± 3</td>
</tr>
<tr>
<td>Calf blood volume (vol%)</td>
<td>4.7 ± 0.8</td>
<td>5.0 ± 0.7</td>
</tr>
<tr>
<td>Conductance (vol% · 10⁻⁵/mm Hg)</td>
<td>3.1 ± 0.4</td>
<td>3.8 ± 0.4</td>
</tr>
<tr>
<td>Compliance (vol% · 10⁻⁵/mm Hg)</td>
<td>3.0 ± 0.2</td>
<td>3.0 ± 0.4</td>
</tr>
<tr>
<td>Hematocrit (%)</td>
<td>43 ± 1</td>
<td>45 ± 1</td>
</tr>
<tr>
<td>Plasma colloid osmotic pressure (mm Hg)</td>
<td>21.7 ± 0.6</td>
<td>24.9 ± 1.0a</td>
</tr>
<tr>
<td>Calf filtration (vol%/min)</td>
<td>0.15 ± 0.03</td>
<td>0.08 ± 0.02a</td>
</tr>
</tbody>
</table>

Vol%, milliliters per 100 ml tissue; ANP, atrial natriuretic peptide; HR, heart rate; MAP, mean arterial pressure.
Values reported as mean ± SEM, n = 6.
a p < 0.05.

Nuclear Medicine Division of Parkland Memorial Hospital, Dallas, Texas. Subjects were catheterized in an arm vein, after which they underwent the erythrocyte radiolabeling procedure. Subjects were then instrumented while supine with their legs elevated 15° from horizontal (calves 15-20 cm above heart level, to facilitate emptying of leg veins) and knees slightly bent for comfort. Legs were supported with pillows and foam pads such that the calves were parallel to the gamma camera face and as close to it as possible without touching it. Subjects assumed this position ~50 min before the beginning of control data collection and remained so through the study. After completion of the control venous occlusion/data collection period (see Calf Capillary Filtration Rate, Blood Volume, and Vascular Compliance for details), a 20-min period was allowed to ensure recovery from prolonged venous occlusion before beginning experimental data collection. The 9-min, 50 mm Hg venous occlusion for placebo or ANP data collection began 6 min after the onset of infusion, and infusion continued during venous occlusion (total infusion time = 15 min). Subjects were awake during data collection.

Statistical analyses
Raw data were coded by one investigator and subsequently analyzed by another without access to identifying information. Paired t tests delineated effects of placebo infusion (n = 7) and ANP infusion (n = 6) relative to prior baseline control results for each subject group. Three subjects were in both placebo and ANP groups. Independent two-tailed t tests were employed to determine whether control data for the placebo and ANP subject groups were similar. In addition, correlation analysis determined significance of relationships between relevant variable pairs within protocol periods (16). ABstat procedures performed all statistical tests, with significance set at 0.05 (Anderson-Bell, Inc.). Results are expressed as means ± SEM.

RESULTS
Intravenous infusion of ANP at 48 pmol/kg/min for 15 min (a cumulative dose of 0.72 nmol/kg) elevated plasma ANP concentrations ~85-fold over preinfusion control levels (Table 1). Pharmacologic ANP infusion increased mean hematocrit 2 hematocrit units (%) and mean plasma colloid osmotic pressure 3.2 mm Hg relative to prior control values (Table 1). Mean capillary filtration rate in the calf, however, decreased 47% during ANP infusion. Individually, five of the six subjects who received ANP infusion exhibited a clear depression of calf filtration with ANP relative to preinfusion control values (Fig. 1). The single subject whose calf filtration increased with ANP had the lowest control value and was our oldest subject (47 years). Calf filtration related inversely with plasma colloid osmotic pressure during control data collection (R² = 0.846, Fig. 2), but this relationship became insignificant during ANP infusion (R² = 0.484, p = 0.125).

ANP infusion tended to increase calf blood flow per unit arterial pressure (conductance) and also tended to increase preclosure calf-blood volume, yet mean arterial pressure was unaffected by ANP (Table 1). Heart rate was elevated 20% during ANP infusion relative to prior control levels (Table 1). Calf conductance correlated positively with plasma [ANP] during preinfusion control periods in both ANP and placebo subject groups (r = 0.796, Fig. 3). Calf venous compliance was unaffected by ANP infusion (Table 1). Control data for placebo and ANP
ANP AND LEG CAPILLARY FILTRATION

FIG. 3. Relationship between calf conductance and plasma [ANP] during preinfusion control data collection. Three subjects were in both placebo and ANP groups; data points for those subjects represent averages of data from their two control periods. n = 10, R² = 0.634, p = 0.006. Vo% equals milliliters per 100 ml tissue.

infusion groups were statistically similar. Placebo infusion had no effect on any variable relative to prior control measurements (Table 2).

DISCUSSION

Hematocrit and plasma colloid osmotic pressure findings confirm that a pharmacologic ANP stimulus produces hemoconcentration, in accordance with the literature (1,3,17). Weidmann et al. (3) pointed out that ANP-induced hemoconcentration occurs more quickly than if diuresis were responsible. Our findings support their conclusion. A plasma volume reduction of ~200 ml occurred during the 15-min ANP infusion we employed (estimated from hematocrit increase, assuming blood volume of ~5 L; 5,9). To excrete this volume over such a short time would require a urine production rate of 13 ml/min, which is unlikely. We can conclude, therefore, that ANP enhanced fluid movement from plasma to interstitium in this study.

Although we expected ANP to increase leg capillary filtration in accordance with the systemic hemoconcentration we observed, the opposite actually occurred: ANP infusion reduced leg filtration. This observation disagrees with prior studies (6,7) that reported enhanced calf filtration despite significant increases in plasma protein concentration. Therefore, ANP-induced colloid osmotic pressure elevation cannot explain the depression of leg filtration we observed.

Fundamental differences may exist in upper and lower body filtration responses to ANP. It is possible that ANP enhances fluid transudation from the upper body or visceral microcirculation or both, while favoring reabsorption of interstitial fluid from the muscle and skin of the legs. This possibility seems reasonable in light of the biomechanical disadvantage cursorial organisms experience by adding weight to the leg (20). Other differences exist between human lower and upper body circulation. For example, capillary basement membranes are thicker (21), and tissues (22) and veins (23) are less compliant in the lower body than in the upper body. However, resting forearm and leg capillary hydraulic conductance are similar, as measured with supine venous occlusion plethysmography (24). Baseline calf filtration values in our study agree well with values from the literature (13,14,25,26).

Alternative mechanisms exist to explain elevated systemic and forearm filtration induced by ANP. First, increased capillary hydrostatic pressure would oppose the reabsorptive force generated by plasma proteins (5,27,28). Second, increased capillary permeability to various extracellular fluid constituents, including protein, would favor capillary filtration (29–32). These mechanisms are not mutually exclusive and could occur separately or together in specific vascular beds. Venous occlusion hypothetically holds capillary pressure constant. If

| TABLE 2. Influence of placebo infusion on plasma [ANP], hemodynamics and calf capillary filtration |
|-----------------|----------|----------|
| Control         | Placebo |
| ANP (pmol/L)    | 21 ± 2   | 22 ± 2   |
| HR (beats/min)  | 66 ± 4   | 68 ± 4   |
| MAP (mm Hg)     | 88 ± 4   | 86 ± 4   |
| Calf blood volume (vol%) | 3.8 ± 0.5 | 3.8 ± 0.4 |
| Conductance (vol% - 10²/mm Hg) | 2.4 ± 0.3 | 2.5 ± 0.5 |
| Compliance (vol% - 10²/mm Hg) | 2.8 ± 0.2 | 2.6 ± 0.4 |
| Hematocrit (%)  | 44 ± 1   | 44 ± 1   |
| Calf filtration (vol%/min) | 0.11 ± 0.02 | 0.09 ± 0.01 |

ANP, atrial natriuretic peptide; vol%, milliliters per 100 ml tissue; HR, heart rate; MAP, mean arterial pressure.

Values reported as mean ± SEM, n = 7, α = 0.05.

ANP increases filtration by elevating capillary pressure, then it should be rendered ineffective during venous occlusion. If, however, the peptide acts by increasing capillary permeability, then ANP should necessarily increase occlusion-induced filtration (6,7). Our results, therefore, suggest that ANP actually reduces capillary permeability in the legs, while increasing it elsewhere.

ANP infusion often increases heart rate (1,17,33). Most investigators view this finding as a reflexive response to ANP-induced systemic hypotension, because no direct chronotropic (or inotropic) effects of ANP have been conclusively identified (1,3.34). The positive relationship we observed between plasma [ANP] and calf conductance during the control period (Fig. 3) suggests that ANP contributes to calf vasomotor control in supine resting conditions. An increase in calf conductance during ANP infusion, such as we saw evidence of, is indicative of arteriolar vasodilation, which should reduce blood pressure. We may not have observed a reduction in blood pressure with ANP as compared to control because the cardiac arm of the carotid baroreflex defended an already low mean arterial pressure. (The control MAP mean was 78 mm Hg.)

In summary, we confirmed that a pharmacologic ANP dose produces hemoconcentration in normal human males, and that because it occurs so quickly, this hemoconcentration cannot be solely attributed to urine production. In spite of this evidence that ANP does indeed stimulate net movement of fluid out of the systemic circulation, calf capillary filtration during venous occlusion was reduced by ANP infusion. Because venous occlusion "clamps" capillary pressure, we conclude that pharmacologic ANP administration reduces capillary permeability in legs of supine human males.

Acknowledgment: We gratefully acknowledge Merck, Sharp, and Dohme Research Laboratories for supplying ANP, Chuck Graham and Dr. Geri Meny for technical support, our research subjects for their interest and cooperation, Drs. Richard Grindeland, Shigeyo Torikoshi, and Charles Wade for comments on the manuscript, and Carolyn Donahue and Karen Hutchinson for manuscript preparation.

This research was supported by a National Aeronautics and Space Administration Graduate Student Researchers Program Fellowship to DEW (NGT-50206), by other NASA grants to CGB (NAS 9-16044) and JCB (NAG 9-267), and by grants to SFV from the Danish Medical Research Council (12-6945 and 12-7663) and the Danish Heart Foundation.

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