Presentation of Ligands on Hydroxylapatite

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Bioconjugate Chemistry

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Presentation of Ligands on Hydroxylapatite

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Conjugates of biotin with the decamer of glutamic acid (glu₁₀) and the trimer of d,l-2-amino-5-phosphonovaleric acid (I) have been synthesized, and it has been shown that they mediate the binding of avidin to hydroxylapatite. In a similar way a conjugate of methotrexate with glu₁₀ mediates the binding of dihydrofolate reductase to the mineral. The presentation of ligands on the hydroxylapatite component of bone may find applications in clinical medicine.

Peptides containing several aspartic and glutamic acid residues, oligonucleotides, and other polyamions bind strongly but reversibly to hydroxylapatite; this is the basis of hydroxylapatite chromatography (1, 2). We have recently found that oligomers of glutamic acid as short as the hexamer bind quantitatively to hydroxylapatite and are not removed by washing with water or a 0.1 M NaCl solution (3). This suggests that negatively charged polypeptides might be used as linkers to bind ligands to the mineral component of bone with controllable affinity and retention time. In this paper we show that conjugates of biotin and methotrexate with negatively charged peptides may be used to mediate the binding of avidin and dihydrofolate reductase, respectively, to hydroxylapatite.

Glu₁₀ was synthesized by the Peptide Biology Laboratory at The Salk Institute. d,l-2-Amino-5-phosphonovaleric acid (I), methotrexate (MTX), chicken liver dihydrofolate reductase (DHFR), and N-hydroxysuccinimide (N-OH-succ) were obtained from Sigma; 1,1-carboxyl-Diimidazole (CDI) and dichloroacetylcobidiium (DCC) were from Aldrich. Succinimidyl-6-(biotinamido) hexanoate (NHS-LC-Biotin II) was obtained from Pierce, ¹²⁵I-labeled streptavidin from Boehringer, and methotrexate (MTX) from Aldrich. The reaction mixture on an RPC-5 column was eluted by shaking the HA with 2 x 50 μL of 0.02 M pyrophosphate for 30 min. The conjugate was purified by HPLC on a C₁₈ column. Its identity was confirmed by LDMS (calculated for C₁₀H₉N₁₈O₃S + H⁺ 1745.6; found 1745.0).

Oligomers of d,l-2-amino-5-phosphonovaleric acid (pvl) were synthesized from the monomer (I) using carboxy-1-diimidazole (CDI) as a condensing agent (5). A solution of the monomer at pH 8 (0.05-0.1 M) was added to a 3-fold excess of solid CDI, and the resulting solution was allowed to stand for 6 h (or overnight). Products ranging from the dimer to the pentamer were identified by paper chromatography (α-PrOH/NH₃/H₂O 7:1:2), and samples of the oligomers were eluted from the paper. HPLC of the reaction mixture on an RPC-5 column gave a series of peaks that were assigned to oligomers of known length by cochromatography with the material eluted from paper.

To determine the shortest oligomer that binds to HA, 2-3 μg of the dimer, trimer, tetramer, or pentamer was separately shaken with 10 mg of HA, and any oligomer retained by the HA was eluted with K₂P₂O₇ as described above. HPLC analysis of the supernatant and K₂P₂O₇ eluate showed that trimers and longer oligomers of pvl were found only in the supernatant and therefore had been bound by the HA. Dimers were not bound to HA and were found in the supernatant fraction.

To obtain the biotin derivative of (pvl)₃, 6 μg of the tripeptide isolated from RPC-5 was first adsorbed to 10 mg of HA. The solid was separated by centrifugation and washed with H₂O to remove Tris and other components of the HPLC buffer. (Pvl)₃ was then eluted with pyrophosphate as described above. (Pvl)₃ (6-10 μg) in 20 μL of buffer containing 0.2 M pyrophosphate and 0.2 M NaHCO₃ (pH 8.4) was added to 0.1 mg of solid NHS-LC Biotin II. The reaction mixture was then allowed to stand for 1 h at room temperature. The biotinyl derivative of the tripeptide was purified and isolated using an
To recruit streptavidin to HA, 1 nmol of biotin-glulo or biotin-(pv)3 was first shaken with 1 mg of HA in 20 μL of 0.01 M Tris-ClO4 for 6 h (or overnight) at room temperature. The supernatant was removed by centrifugation, and the HA was washed with 100 μL of water. A solution of 0.1 nmol of 125I-labeled streptavidin (25 000–50 000 cpm) in 100 μL of buffer containing 1 M KCl and 0.01 M phosphate at pH 6.5 was added to the HA and shaken for 45 min. The supernatant was removed by centrifugation and the HA washed several times with 200 μL of H2O. The amounts of radioactivity found in the supernatant, wash, and HA fractions were then measured (see Table 1). In control experiments, biotin-glulo was replaced by glulo and biotin-(pv)3 was replaced by (pv). When a peptide bound to the HA was ligated to biotin about 70% of the streptavidin was recruited to the HA and 25–30% remained in the supernatant (Table 1). In the control experiments no more than 5% of the streptavidin was bound to the HA. Clearly the preadsorption of biotin conjugates of negatively charged polypeptides greatly enhances the adsorption of avidin to HA.

To recruit dihydrofolate reductase to HA, 1 nmol of MTX-glulo was adsorbed to HA as described above for biotin-glulo. DHFR (0.52 nmol) in 200 μL of buffer containing 0.1 M ammonium sulfate, 0.01 M potassium phosphate, at pH 6.4, and 5% glycerol was added to the HA and shaken for 45 min. The HA was separated from the supernatant, washed with 100 μL of water, and then eluted twice with 20 μL of 0.02 M K4P2O7. In control experiments the MTX-glulo was replaced by glulo. The supernatant, the washes, and the pyrophosphate eluate were analyzed on a 6% acrylamide SDS gel using Coomassie Blue to visualize DHFR. Figure 1 shows that in the control experiments with glulo more than 75% of the DHFR was found in the supernatant (Figure 1, lane 1) and only a small amount in the pyrophosphate eluate (Figure 1, lane 3). In experiments involving MTX-glulo more than 75% of the DHFR was found in the pyrophosphate eluate (Figure 1, lane 6) and very little in the supernatant (Figure 1, lane 4). Preadsorption of MTX-glulo, therefore, greatly increases the amount of DHFR that binds to HA.

The above results show that conjugates of various ligands with anionic polypeptides adsorbed noncovalently on hydroxyapatite could be used as supports for affinity chromatography. More importantly, HA presents special opportunities in a related context, because it is the main mineral component of bone. The surface of bone is freely accessible to molecules in the extracellular fluid even if they are as large as proteins. The bisphosphonates, small molecules carrying four negative charges, have been used extensively to attach technetium to hydroxyapatite for bone scintigraphy. One example of the recruitment of an anticancer drug, methotrexate, to bone using a bisphosphonate has been reported. We believe that anionic polypeptides may prove particularly convenient as carriers of ligands to bone and may sometimes have advantages over the bisphosphonates.

**Table 1. Biotin-Mediated Binding of Streptavidin to Hydroxyapatite**

<table>
<thead>
<tr>
<th>Biotin conjugate</th>
<th>% [125I]streptavidin in supernatant</th>
<th>% [125I]streptavidin on hydroxyapatite</th>
</tr>
</thead>
<tbody>
<tr>
<td>glulo</td>
<td>97</td>
<td>3</td>
</tr>
<tr>
<td>pv3</td>
<td>97</td>
<td>3</td>
</tr>
<tr>
<td>biotin-glulo</td>
<td>25</td>
<td>75</td>
</tr>
<tr>
<td>biotin-pv</td>
<td>32</td>
<td>68</td>
</tr>
</tbody>
</table>

RPC-5 column. Its identity was confirmed by ESMS (calculated for C31H57NeO16P38 - H 893.2; found 893).

The mechanism of action of bisphosphonates on bone resorption is not fully understood, but it seems clear that it is not entirely a matter of adsorption to hydroxyapatite. The properties of osteoblasts are profoundly affected by submicromolar concentrations of bisphosphonates, suggesting that they attach to receptors, possibly pyrophosphate receptors, on the cell surface. The structures of polypeptides are completely unrelated to that of inorganic pyrophosphate, so by using them as carriers it should be possible to dissociate the direct effects of adsorption to HA from the indirect effects due to interaction with extracellular receptors on osteoblasts (or osteoclasts).

Polypeptides are uniquely convenient as carriers, because effective automated methods are already available for their synthesis, and the use of combinatorial peptide libraries is well-established. The strength of adsorption of the carriers could easily be controlled via their length, while more or less degradable carriers could be obtained by varying the ratio of D- to L-residues. In the special case of a peptide ligand, the ligand and the anionic carrier could be assembled in a single solid-phase peptide synthesis. Presentation of ligands on HA that interact directly with receptors on osteoblasts or osteoclasts, or which recruit proteins to bone, may find applications in medicine.

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**LITERATURE CITED**


