Presentation of Ligands on Hydroxylapatite

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Bioconjugate Chemistry

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Presentation of Ligands on Hydroxylapatite

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Conjugates of biotin with the decamer of glutamic acid (glu10) and the trimer of D,L-2-amino-5-phosphonovaleric acid (I) have been synthesized, and it has been shown that they mediate the binding of avidin to hydroxylapatite. In a similar way a conjugate of methotrexate with glu10 mediates the binding of dihydrofolate reductase to the mineral. The presentation of ligands on the hydroxylapatite component of bone may find applications in clinical medicine.

Peptides containing several aspartic and glutamic acid residues, oligonucleotides, and other polyamides bind strongly but reversibly to hydroxylapatite; this is the basis of hydroxylapatite chromatography (1, 2). We have recently found that oligomers of glutamic acid as short as the hexamer bind quantitatively to hydroxylapatite and are not removed by washing with water or a 0.1 M NaCl solution (3). This suggests that negatively charged polypeptides might be used as linkers to bind ligands to the mineral component of bone with controllable affinity and retention time. In this paper we show that conjugates of biotin and methotrexate with negatively charged peptides may be used to mediate the binding of avidin and dihydrofolate reductase, respectively, to hydroxylapatite.

Glu10 was synthesized by the Peptide Biology Laboratory at The Salk Institute. D,L-2-Amino-5-phosphonovaleric acid (I), methotrexate (MTX), chicken liver dihydrofolate reductase (DHFR), and N-hydroxysuccinimide (N-OH-succ) were obtained from Sigma; 1,1-carboxyldiimidazole (CDI) and dicyclohexylcarbodiimide (DCC) were from Aldrich. Succinimidyl-6-(biotinamido) hexanate (NHS-LC-Biotin II) was obtained from Pierce; streptavidin from Boehringer, 125I-labeled streptavidin from Amersham, and hydroxylapatite (HA) from Bio-Rad.

The biotin derivative of glu10 was synthesized by reacting 25 nmol of the oligomer with 190 nmol of NHS-LC-Biotin II in 20 μL of 0.1 M NaHCO3 buffer at pH 8.4 for 45 min. The product was purified on a C18 column using a 0.1% TFA/acetonitrile gradient and its identity confirmed by LDMS (calculated for C105H136N18O38 S + Na+ 1670.6; found 1670.7).

The MTX derivative of glu10 was synthesized via an N-hydroxysuccinimide intermediate (4). A solution (40 μL) containing 0.05 M MTX, 0.05 M N-OH-succ, and 0.05 M DCC in DMF was allowed to stand at room temperature for 1 h and then at 2–4 °C overnight. Ten microliters of the resulting solution was added to 5–25 nmol of glu10 in 30 μL of 0.02 M NaHCO3 at pH 8.2. The reaction mixture was shaken in the dark for 4 h and then diluted with 70 μL of water. Unreacted MTX and salts were removed by shaking the reaction mixture with 10 mg of HA overnight, removing the supernatant, and washing the HA with water. Glu10 and its MTX conjugate were eluted by shaking the HA with 2 × 50 μL of 0.02 M pyrophosphate for 30 min. The conjugate was purified by HPLC on a C18 column. Its identity was confirmed by LDMS (calculated for C170H292N18O56 S + H+ 1745.6; found 1745.0).

Oligomers of D,L-2-amino-5-phosphonovaleric acid (pvl) were synthesized from the monomer (I) using carbonyldiimidazole (CDI) as a condensing agent (5). A solution of the monomer at pH 8 (0.05–0.1 M) was added to a 3-fold excess of solid CDI, and the resulting solution was allowed to stand for 6 h (or overnight). Products ranging from the dimer to the pentamer were identified by paper chromatography (n-PrOH/NH3/H2O 7:1:2), and samples of the oligomers were eluted from the paper. HPLC of the reaction mixture on an RPC-5 column gave a series of peaks that were assigned to oligomers of known length by cochromatography with the material eluted from paper.

To determine the shortest oligomer that binds to HA, 2–3 μg of the dimer, trimer, tetramer, or pentamer was separately shaken with 10 mg of HA, and any oligomer retained by the HA was eluted with K2P2O7 as described above. HPLC analysis of the supernatant and K2P2O7 eluate showed that trimers and longer oligomers of pvl were found only in the eluate and therefore had been bound by the HA. Dimers were not bound to HA and were found in the supernatant fraction.

To obtain the biotin derivative of (pvl)6, 6 μg of the tripeptide isolated from RPC-5 was first adsorbed to 10 mg of HA. The solid was separated by centrifugation and washed with H2O to remove Tris and other components of the HPLC buffer. (Pvl)6 was then eluted with pyrophosphate as described above. (Pvl)6 (5–10 μg) in 20 μL of buffer containing 0.2 M pyrophosphate and 0.2 M NaHCO3 (pH 8.4) was added to 0.1 mg of solid NHS-LC Biotin II. The reaction mixture was then allowed to stand for 1 h at room temperature. The biotinyl derivative of the tripeptide was purified and isolated using an
Table 1. Biotin-Mediated Binding of Streptavidin to Hydroxylapitate

<table>
<thead>
<tr>
<th></th>
<th>% 125I-streptavidin in supernatant</th>
<th>% 125I-streptavidin on hydroxylapitate</th>
</tr>
</thead>
<tbody>
<tr>
<td>glu10</td>
<td>97</td>
<td>3</td>
</tr>
<tr>
<td>pvl3</td>
<td>97</td>
<td>3</td>
</tr>
<tr>
<td>biotin-glul0</td>
<td>25</td>
<td>75</td>
</tr>
<tr>
<td>biotin-pvl3</td>
<td>32</td>
<td>68</td>
</tr>
</tbody>
</table>

RPC-5 column. Its identity was confirmed by ESMS (calculated for C31H57NeO16P3S - H 893.2; found 893).

To recruit streptavidin to HA, 1 nmol of biotin-glul0 or biotin-(pvl)3 was first shaken with 1 mg of HA in 20 μL of 0.01 M Tris-ClO4 for 6 h (or overnight) at room temperature. The supernatant was removed by centrifugation, and the HA was washed with 100 μL of water. A solution of 0.1 nmol of 125I-labeled streptavidin (25 000–50 000 cpm) in 100 μL of buffer containing 1 M KCl and 0.01 M phosphate at pH 6.5 was added to the HA and shaken for 45 min. The supernatant was removed by centrifugation and the HA washed several times with 200 μL of H2O. The amounts of radioactivity found in the supernatant, wash, and HA fractions were then measured (see Table 1). In control experiments, biotin-glul0 was replaced by glu10 and biotin-(pvl)3 was replaced by (pvl)3. When a peptide bound to the HA was ligated to biotin about 70% of the streptavidin was recruited to the HA and 25–30% remained in the supernatant (Table 1). In the control experiments no more than 5% of the streptavidin was bound to the HA. Clearly the preadsorption of biotin conjugates of negatively charged polypeptides greatly enhances the adsorption of avidin to HA.

To recruit dihydrofolate reductase to HA, 1 nmol of MTX-glul0 was adsorbed to HA as described above for biotin-glul0. DHFR (0.52 nmol) in 200 μL of buffer containing 0.1 M ammonium sulfate, 0.01 M potassium phosphate, at pH 6.4, and 5% glycerol was added to the HA and shaken for 45 min. The HA was separated from the supernatant, washed with 100 μL of water, and then eluted twice with 20 μL of 0.02 M K4P2O7. In control experiments the MTX-glul0 was replaced by glu10. The supernatant, the washes, and the pyrophosphate eluate were analyzed on a 6% acrylamide SDS gel using Coomassie Blue to visualize DHFR. Figure 1 shows that in the control experiments with glu10 more than 75% of the DHFR was found in the supernatant (Figure 1, lane 1) and only a small amount in the pyrophosphate eluate (Figure 1, lane 3). In experiments involving MTX-glul0 more than 75% of the DHFR was found in the pyrophosphate eluate (Figure 1, lane 6) and very little in the supernatant (Figure 1, lane 4). Preadsorption of MTX-glul0, therefore, greatly increases the amount of DHFR that binds to HA.

The above results show that conjugates of various ligands with anionic polypeptides adsorbed noncovalently on hydroxylapitate could be used as supports for affinity chromatography. More importantly, HA presents special opportunities in a related context, because it is the main mineral component of bone. The surface of bone is freely accessible to molecules in the extracellular fluid even if they are as large as proteins (6). The bisphosphonates, small molecules carrying four negative charges, have been used extensively to attach technetium to hydroxylapitate for bone scintigraphy (7). One example of the recruitment of an anticancer drug, methotrexate, to bone using a bisphosphonate has been reported (8). We believe that anionic polypeptides may prove particularly convenient as carriers of ligands to bone and may sometimes have advantages over the bisphosphonates.

The mechanism of action of bisphosphonates on bone resorption is not fully understood, but it seems clear that it is not entirely a matter of adsorption to hydroxylapitate. The properties of osteoblasts are profoundly affected by submicromolar concentrations of bisphosphonates, suggesting that they attach to receptors, possibly pyrophosphate receptors, on the cell surface (9). The structures of polypeptides are completely unrelated to that of inorganic pyrophosphate, so by using them as carriers it should be possible to dissociate the direct effects of adsorption to HA from the indirect effects due to interaction with extracellular receptors on osteoblasts (or osteoclasts).

Polypeptides are uniquely convenient as carriers, because effective automated methods are already available for their synthesis, and the use of combinatorial peptide libraries is well-established. The strength of adsorption of the carriers could easily be controlled via their length, while more or less degradable carriers could be obtained by varying the ratio of D- to L-residues. In the special case of a peptide ligand, the ligand and the anionic carrier could be assembled in a single solid-phase peptide synthesis. Presentation of ligands on HA that interact directly with receptors on osteoblasts or osteoclasts, or which recruit proteins to bone, may find applications in medicine.

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LITERATURE CITED


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