Test Procedures for Characterizing, Evaluating, and Managing Separator Materials Used in Secondary Alkaline Batteries

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Trade names or manufacturers' names are used in this report for identification only. This usage does not constitute an official endorsement, either expressed or implied, by the National Aeronautics and Space Administration.
SECONDARY ALKALINE BATTERY TECHNOLOGIES, SUCH AS NICKEL-CADMIUM (Ni/Cd), NICKEL-HYDROGEN (Ni/H2), AND SILVER-ZINC (Ag/Zn), ARE VITAL TO AEROSPACE APPLICATIONS. THE SEPARATORS USED IN THESE BATTERY TECHNOLOGIES PLAY AN IMPORTANT ROLE IN THE CYCLE LIFE OF THE CELLS AND IN THEIR PERFORMANCE. SEPARATORS MUST BE GOOD ELECTRICAL INSULATORS AND EFFICIENT ELECTROLYTE RESERVOIRS. THIS MANUAL PRESENTS A SERIES OF TEST PROCEDURES THAT CAN BE USED TO EVALUATE AND SELECT SEPARATOR MATERIALS FOR THESE BATTERY TECHNOLOGIES.


IN THE MID-1980'S, CELLS PRODUCED BY GATES AEROSPACE BATTERIES, AT THE FORMER GE FACILITY IN GAINESVILLE, FLORIDA, EXHIBITED PROBLEMS ATTRIBUTED TO THE QUALITY OF THE PLATES AND SEPARATOR MATERIALS. IN 1988, A GATES Ni/Cd CELL MINI-WORKSHOP WAS HELD AT THE MARSHALL SPACE FLIGHT CENTER IN HUNTSVILLE, ALABAMA, TO ADDRESS THESE PROBLEMS. AT THAT WORKSHOP, A SERIES OF TASK FORCE GROUPS WERE FORMED TO ADDRESS THE ISSUES FACING THE AEROSPACE BATTERY INDUSTRY. THE TASK FORCE GROUP ON Ni/Cd SEPARATOR ACCEPTANCE AND CRITERIA RECOMMENDED THAT STANDARD PROCEDURES BE ESTABLISHED FOR SEPARATOR TESTING TO ENSURE QUALITY AND CONSISTENCY. A TASK WAS THEN FUNDED AS PART OF THE NASA AEROSPACE FLIGHT BATTERY SYSTEMS PROGRAM, SPONSORED BY NASA HEADQUARTERS CODE Q, TO DESIGN AND DEVELOP TEST PROCEDURES FOR CHARACTERIZING, EVALUATING, AND MANAGING SEPARATOR MATERIALS AND TO ESTABLISH LIMITS AND CRITERIA FOR SEPARATOR CHARACTERISTICS. THE TASK WAS ASSIGNED TO THE NASA LEWIS RESEARCH CENTER IN CLEVELAND, OHIO. THIS PUBLICATION IS THE RESULT OF THAT EFFORT.

ACKNOWLEDGMENTS

THE SCREENING METHODS OUTLINED IN REFERENCES 1 AND 2 SERVED AS GUIDELINES FOR FORMULATING THE TEST PROCEDURES INCLUDED IN THIS MANUAL. WE EXTEND OUR APPRECIATION TO GATES AEROSPACE BATTERIES, EAGLE PICHÉ, THE AEROSPACE CORPORATION, FREUDENBERG NONWOVEN INC., AND COMSAT FOR SHARING THEIR WORK, PROCEDURES, AND EXPERIENCE IN THE SEPARATOR TESTING AREA. WE RECOGNIZE AND THANK NASA PERSONNEL INVOLVED IN THE ENHANCEMENT OF BATTERY TECHNOLOGY FOR THEIR EVALUATIONS, COMMENTS, AND FEEDBACK. FINALLY, WE ESPECIALLY ACKNOWLEDGE THE PERSONNEL OF THE ELECTROCHEMICAL TECHNOLOGY BRANCH OF THE POWER TECHNOLOGY DIVISION AT THE NASA LEWIS RESEARCH CENTER IN CLEVELAND, OHIO, FOR SHARING THEIR EXPERTISE.

*IN THE EARLY 1980'S, GATES ENERGY PRODUCTS PURCHASED THE GE BATTERY FACILITY IN GAINESVILLE, FLORIDA. THEN, IN JANUARY 1994, SAFT PURCHASED THE AEROSPACE PORTION OF THAT OPERATION, GATES AEROSPACE BATTERIES.
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INTRODUCTION

The separator test procedures (STP) presented in this manual result from a comprehensive gathering of information, data, and test procedures acquired from throughout the aerospace battery industry. They can be used to evaluate the suitability of new materials and to ensure the uniformity, consistency, and reproducibility of standard materials. As a starting point, tests specified in references 1 and 2 were evaluated. Some of the tests served as a foundation for the procedures included in this text. Others were not incorporated because of limited applicability.

The test procedures have been written to be generally applicable to materials used in alkaline cells. Procedures specific to silver-zinc batteries are covered in reference 2. This manual presents the test procedures in a logical sequence to facilitate the characterization of new materials and the evaluation of already established separators. Following the STP sequence can simplify the qualification process. The STP order was designed to allow early elimination of materials whose characteristics do not meet the requirements. General guidelines are provided to ensure the proper handling and storage of materials. Each STP suggests a particular sampling frequency and sample size; however, it is recommended that a specific sampling plan be designed to accommodate sample availability in order to ensure that samples are representative of the source material.

Separator characteristics required for a specific cell design must first be identified. The testing and measurement of the characteristics then follows. A baseline must be established for materials to be incorporated into flight cells. When establishing a baseline for a separator, multiple runs of the same material should be evaluated to ensure that the material can be manufactured consistently. The determination of baseline characteristics should be supported statistically. Separator characteristics should be within the ranges and limits established to ensure consistency and quality.

In order to have an approximate representation of cell conditions, the characterization tests should be performed on samples filled with the same electrolyte composition and concentration used in the cell. For comparability, separator materials should be tested with a standard concentration of electrolyte. Some tests were designed to measure characteristics in representative cell conditions. Where this was not possible, test results still provide consistent and reliable information for evaluating the chemical and physical characteristics of the separators.

The test procedures examine such characteristics as uniformity, dimensional stability, electrolyte retention, area resistivity, permeability to gases, resistance to chemical degradation (hydrolysis and/or electrochemical oxidation), chemical purity, tensile strength, and pore size distribution. Completing these characterization tests is important to identifying separator materials that have the desired properties necessary for long life and good performance in secondary alkaline cells.

All test procedures are presented in a similar format. In the beginning of each procedure, a short introduction covers the background, methodology, and relevance of the test. The introductory segment is followed by a reagents and equipment section. Then, step-by-step instructions describe the test performance and technique. In general, a calculations section appears after the procedure. Calculations are followed by recommendations and/or criteria. In some cases, specific criteria (ranges and limits) are given. Diagrams and/or graphs relating to the particular STP are included in each procedure.
Guidelines

Four appendixes are presented at the end of the manual. Appendix A contains an evaluation form used to summarize test results and to compare experimental results with the baseline characteristics of the material. Appendix B presents a table with characterization results for some of the materials commonly used in aerospace cells. Characterization results for separator materials FS 2536, FS 2538, Pellon 2505 from Freudenberg Nonwovens Inc., and ZYK15 from Zircar Products are presented. Appendix C contains two additional test procedures that complement the procedures presented in the manual. The first, electrolyte retention in uncompressed state using a Kubelka jar, is an alternative procedure for determining electrolyte retention that is commonly used by Freudenberg Nonwovens. The second is the electrolyte distribution test. This procedure evaluates the competition and distribution of electrolyte between cell components at different percent fill levels. It is a versatile test that provides information on area density, thickness, and compressed and uncompressed electrolyte retention as well as information on the competition for electrolyte between cell components. Finally, appendix D lists and defines the symbols used in the manual according to STP.

The tests presented in this manual are specific to the NASA Lewis Research Center and reflect equipment and safety practices required. It is not the intention of this manual to regulate or to impose a certain technique, sample size, and/or equipment for obtaining data on a particular separator property. This manual has been compiled to offer interested battery scientists and technicians consistent, uniform, and reliable test procedures for characterizing and evaluating candidate separator materials for a particular technology and application.

GUIDELINES FOR STORING, SAMPLING, HANDLING, AND DISPOSING OF SEPARATOR MATERIALS

Separator materials used in secondary alkaline batteries must be kept free from contaminants and kept out of an environment that could initiate their decomposition. Different types of separator materials may require unique storage and handling conditions and specific sampling and disposal methods. A storage and sampling plan is essential to maintaining and monitoring the integrity of the materials. Sampling from various locations throughout the material source is recommended to guarantee proper characterization.

The following guidelines address some of the most important issues to be considered when working with separator materials. The proper storage, sampling, handling, and disposal of these materials are critical.

Asbestos has been commonly used as a separator in secondary alkaline batteries. This class of material, with inhalant fibers longer than 5.0 µm and a 3:1 aspect ratio, is considered hazardous. The proper handling of hazardous materials is essential for health and safety. The use of asbestos is regulated by the U.S. Environmental Protection Agency (EPA) and the U.S. Occupational Safety and Health Administration (OSHA). Because this material is still used in aerospace battery technology, its use requires specific handling and disposal guidelines. Specific instructions particular to storing, handling, and disposing of hazardous materials are also included in the following guidelines.

General Guidelines for All Separators

1. Keep all equipment used to handle and test separator materials clean, properly labeled, and where applicable, stored in sealed plastic bags.

2. Store separator materials in a controlled environment, at low temperature or room temperature, with low humidity, in properly labeled and sealed polyethylene or polypropylene plastic bags unless otherwise specified by the vendor.

3. Always avoid contact with hands by using disposable gloves or finger cots. Avoid stressing or elongating the material.
Guidelines

4. Establish a consistent size and sampling pattern that addresses the material source available for testing. Prepare samples by cutting to the desired size, preferably with an appropriate rule die. Once cut, always keep samples in a clean, properly labeled, covered container.

5. Minimize exposure to air of samples wet with electrolyte by keeping them submerged in their respective covered containers.

6. Dispose of samples exposed to electrolyte properly, in labeled plastic bags placed in a trash container, unless otherwise specified by OSHA regulations.

Specific Guidelines for Hazardous Materials

1. EPA and OSHA regulations for storing, handling, and disposing of hazardous materials must be followed. For advice, refer to your local environmental, health, and safety personnel. (At the NASA Lewis Research Center, guidelines and requirements for storing, handling, and disposing of hazardous materials are set by the Office of Environmental Programs.)

2. NASA Lewis Research Center standard operational procedures state that anyone working with hazardous materials must be properly trained and certified. A certified operators list must be posted and renewed with the safety permit. Operators are required to become familiar with the particular material safety data sheet (MSDS) concerning the material.

3. The following sign must be posted whenever any hazardous material is in use: DO NOT ENTER WITHOUT PROPER RESPIRATORY PROTECTION: HAZARDOUS MATERIAL IN USE.

4. Where practicable, all cutting operations should be performed under an exhaust hood.

5. The qualified operator must use proper protective equipment, such as a disposable laboratory coat, safety glasses, disposable gloves, and a hood mask or respirator.

6. Any materials removed from the hood must be wet with electrolyte or water or placed in sealed containers to avoid the spread of airborne fibers in the laboratory.

7. All utensils and equipment exposed to hazardous materials should be cleaned with a damp cloth or paper towel. Include mask hood and safety glasses in the cleaning process.

8. All hazardous materials, scraps, gloves, damp cloths or paper towels, or any other materials contaminated with a hazardous material must be placed in sealed plastic bags under the hood and then disposed of in a properly labeled and sealed container.

9. The room and all personnel must be regularly monitored for the presence of airborne hazardous fibers.
STP 1—THICKNESS AND AREA WEIGHT MEASUREMENT PROCEDURE

Thickness and area weight measurements provide a means of evaluating the uniformity and consistency of separator materials. These measurements are performed in almost every procedure presented in this text. Area weight is the ratio of the sample weight to the sample area. Thickness measurements are specific to the type of instrument used. An instrument with a constant and uniform load is required for reliable and reproducible measurements. Thickness measurements are commonly obtained and reported in the dry state because readings taken in the wet state can be unreliable as some materials tend to swell when wet.

I. Equipment:

1. TMI Electronic Micrometer Model 49-70 (fig. 1)—a precision thickness-measuring instrument with digital readout, wide flexibility, and a motor suitable for continuous operation. Maximum thickness capability is 1.27 mm (50.0 mils).

2. Ohaus Analytical Balance Model GA110 (fig. 1) with a sensitivity of 0.0001 g.

Figure 1.—Analytical balance and thickness measurement instrument.
II. Procedure:

1. Warm up and calibrate instruments.

2. Draw a piece of nonabrasive paper or cloth through the TMI anvils. This will ordinarily pick up and remove any foreign matter.

3. Cut samples to the appropriate size for the particular separator test procedure to be performed. For area weight and thickness measurements, cut two samples, 10 cm by 10 cm (4 in. by 4 in.).

4. Weigh each sample and record data.

5. Measure sample thickness with the TMI. Take between 6 and 10 thickness readings at different locations. Repeat step 2 if necessary.

III. Calculations:

1. Average the thickness readings to obtain an accurate thickness measurement $T$ in millimeters.

2. In order to calculate area weight $AW$, divide each sample dry weight $W_d$ by its area as follows:

$$AW = \frac{W_d}{A}$$

where

- $T$ = average sample thickness, mm
- $AW$ = area weight, g/m$^2$
- $W_d$ = dry weight of sample, g
- $A$ = sample area, m$^2$
STP 2—ELECTROLYTE FILL PROCEDURE FOR SEPARATOR MATERIALS

Cell performance depends on the properties that the separator materials have when wet with electrolyte. Most test procedures included in this manual require wet samples. It is essential that separators be filled with electrolyte solution to a 100% fill level, the saturation point when all pores are filled with electrolyte, to ensure consistent evaluation of their properties. In order to achieve 100% fill, samples are evacuated and backfilled with electrolyte. This procedure outlines a standard technique of ensuring 100% fill of separator materials and/or cell components. Handling materials once 100% fill is achieved is discussed as part of the subsequent STP's.

I. Equipment and reagents:

1. 100 to 200 ml of electrolyte solution
2. Electrolyte fill system (fig. 2)
   a. Vacuum pump (minimum absolute pressure of 1.9 bars)
   b. 250-ml beaker for electrolyte reservoir
   c. Vacuum trap
   d. 250-ml, wide-mouth flask fitted with stopper and tubing
   e. Three-way stopcock valve
   f. Tubing to make the necessary connections
3. Vertical sample holder (fig. 2)

![Diagram](image-url)
II. Procedure:

1. Assemble electrolyte fill system according to figure 2.

2. Place cell component (or components) in the vertical sample holder inside the wide-mouth flask. Close the wide-mouth flask with its corresponding stopper.

3. Pour sufficient electrolyte to cover the components into the beaker serving as the electrolyte reservoir.

4. Evacuate the wide-mouth flask containing the samples for 3 to 5 min.

5. Slowly position the three-way stopcock valve to allow electrolyte to flow from the reservoir into the wide-mouth flask.

6. Reposition the stopcock valve and evacuate the wide-mouth flask for 10 to 15 min. The electrolyte will boil as the pressure is reduced.

7. Vent the wide-mouth flask to atmospheric pressure and proceed with the corresponding test procedure.
STP 3

STP 3—DIMENSIONAL STABILITY TEST PROCEDURE

Dimensional stability in separator materials is required for good cell performance. Changes in the separator dimensions, as a result of exposure to the electrolyte, can result in various cell failure mechanisms. A separator that swells when exposed to electrolyte can cause increased pressure within the cell pack. A separator that shrinks is likely to tear and/or crack and expose the plates to the possibility of a short circuit.

A dimensional stability test is one of the oldest and simplest tests for evaluating separators. This procedure employs the direct measurement of the length and width of a sample in the dry state and then in the wet state to determine dimensional changes. Measurements are taken after 10 days of exposure.

I. Equipment and reagents:

1. Analytical balance
2. TMI Electronic Micrometer Model 49–70
3. Plastic ruler
4. 30-ml-capacity, wide-mouth Teflon vial with lid
5. Electrolyte solution

II. Procedure:

1. Cut three samples, approximately 9 cm² (1.4 in.²), and label them.
2. Measure the dimensions, length and width, of the samples \( l_d \) and \( w_d \).
3. Weigh all samples and measure their thickness (refer to STP 1).

**NOTE**
Dry weight and thickness measurements are taken for information only.

4. Place the three samples and 25 ml of electrolyte in the vial, cap it, and store it at room temperature.
5. Retrieve the samples after 10 days. Measure and record the dimensions of each wet sample \( l_w \) and \( w_w \).

III. Calculations:

1. Determine the percentage of dimensional change \( (D_l \) and \( D_w) \) in the length and width of each sample by using the following equations:

\[
D_l = 100 \times \frac{l_w - l_d}{l_d}
\]

\[
D_w = 100 \times \frac{w_w - w_d}{w_d}
\]
where

\[ D_l \quad \text{change in length, percent} \]
\[ l_w \quad \text{length of sample in wet state, cm} \]
\[ l_d \quad \text{length of sample in dry state, cm} \]
\[ D_w \quad \text{change in width, percent} \]
\[ w_w \quad \text{width of sample in wet state, cm} \]
\[ w_d \quad \text{width of sample in dry state, cm} \]

2. Average and report the percentage of dimensional change \( (D_l \text{ and } D_w) \) for the sample materials.

IV. Recommendation:

Typically, dimensional changes should not exceed 1% per sample material.
Electrolyte retention is a valuable parameter for evaluating separator materials used in secondary alkaline systems. Electrolyte retention is defined as the amount of electrolyte retained as a percentage of the dry weight of the sample. The separator must retain enough electrolyte to sustain the electrochemical reactions in the cell under a variety of conditions.

Three procedures are presented for determining electrolyte retention. One procedure is used to measure electrolyte retention in the uncompressed state, and two are used to measure electrolyte retention in the compressed state. The most widely used procedure, for electrolyte retention in the uncompressed state, involves evacuating air from dry samples, backfilling them with electrolyte, and determining the amount of electrolyte retained.

Although electrolyte retention is most commonly measured in the uncompressed state, measurements made in the compressed state are more representative of cell conditions and thus provide a more realistic measurement of this property. Two procedures for measuring electrolyte retention in the compressed state are presented. In both approaches, samples filled to the 100% fill level are compressed to a predetermined thickness. The thickness of the compressed sample is controlled by incorporating shims of a specified thickness, generally representing the inter-electrode spacing. The first procedure uses a fixture designed for determining electrolyte retention in the compressed state. In the second procedure, the separator is compressed between nickel and cadmium plates representing an actual cell assembly.
STP 4A

STP 4A—Electrolyte Retention in Uncompressed State
by Using Standard Method

I. Equipment and reagents:

1. Electrolyte fill system (fig. 2)
2. Analytical balance
3. Tweezers
4. Filter paper
5. Electrolyte solution
6. TMI electronic micrometer

II. Procedure:

1. Cut four samples, 2.9 cm by 2.9 cm (1.2 in. by 1.2 in.), and label them.
2. Determine the dry weight \( W_d \) and thickness of the samples according to STP 1.

**NOTE**
Thickness measurements are taken for information only.

3. Fill separator samples with electrolyte as described in STP 2.
4. Remove each sample from electrolyte and suspend it vertically for 1 min. Touch the tip of the sample to the surface of a piece of filter paper five times to remove excess electrolyte.
5. Weigh the wet sample \( W_w \) and record data.

III. Calculations:

Determine electrolyte retention in the uncompressed state by using the following equations:

\[
ER = 100 \times \frac{W_w - W_d}{W_d}
\]
\[
ER_A = \frac{(W_w - W_d)/\rho}{A}
\]

where

- \( ER \) electrolyte retention in uncompressed state, percent
- \( W_w \) wet weight of sample, g
- \( W_d \) dry weight of sample, g
- \( ER_A \) electrolyte retention per sample area in uncompressed state, cm\(^3\)/cm\(^2\)
- \( \rho \) electrolyte density, g/cm\(^3\)
- \( A \) sample area, cm\(^2\)
IV. Recommendations:

1. Electrolyte retention in the uncompressed state can be used to evaluate consistency between separators. It provides a measurement of the affinity of a material for the electrolyte. Generally, the higher the electrolyte retention, the better the separator. This procedure measures electrolyte retention for materials in the free state; therefore, it does not reflect cell conditions. Electrolyte retention rankings between materials can change when measurements are taken in the compressed state.

2. Electrolyte retention results are reported as a percentage of the dry weight and as the amount of electrolyte retained per unit area of the material $ER_A$. The latter is a more useful value when comparing materials because it gives a direct measurement of the amount of electrolyte retained in the sample.
I. Equipment and reagents:

1. Electrolyte fill system (fig. 2)
2. Cell fixture for measuring electrolyte retention in the compressed state (fig. 3)
3. Analytical balance
4. Tweezers
5. Filter paper
6. Compression machine (fig. 4) or alternative fixture for compressing cell assembly
7. Feeler gauge
8. Electrolyte solution
9. TMI electronic micrometer

Figure 3.—Cell fixture for measuring electrolyte retention in compressed state. (a) Stainless steel plate. (b) Cell assembly. (c) Shim.
II. Procedure:

1. Cut four samples, 2.9 cm by 2.9 cm (1.2 in. by 1.2 in.), and label them. The sample size corresponds to the area compressed by the cell fixture.

2. Determine the dry weight $W_d$ and thickness of the samples according to STP 1.

**NOTE**
Thickness measurements are taken for information only.
3. Fill separator samples with electrolyte as described in STP 2.

4. Remove each sample from electrolyte and suspend it vertically for 1 min. Touch the tip of the sample to the surface of a piece of filter paper five times to remove excess electrolyte.

**NOTE**
Weighing the samples at this point will enable the determination of electrolyte retention in the uncompressed state.

5. Center the sample on the bottom plate of the fixture (fig. 3). Select a shim of appropriate thickness, generally representative of the interelectrode spacing, and place it around the sample. The standard procedure is to use 0.25-, 0.19-, and 0.15-mm (10.0-, 7.5-, and 6.0-mil) shims successively. Always go from the thickest to the thinnest shim when testing at different shim sizes.

6. Compress the wet sample to the desired thickness. Use a feeler gauge to verify the sample thickness.

7. Keep material compressed for 5 min.

8. Decompress sample as quickly as possible. Remove the sample from the cell fixture, reweigh ($W_{th}$), and record data.

9. Wipe excess electrolyte from the cell fixture. Change to the next smaller shim size and repeat steps 4 to 9.

III. Calculations:

Determine electrolyte retention for the material compressed to a specific thickness $ER_{th}$ and electrolyte retention for the material compressed at a specific thickness per sample area $ER_{th,A}$ by using the following equations:

$$ER_{th} = 100 \times \frac{W_{th} - W_d}{W_d}$$

$$ER_{th,A} = 100 \times \frac{(W_{th} - W_d)}{\rho A}$$

where

- $ER_{th}$ electrolyte retention at specified thickness, percent
- $W_{th}$ wet weight of sample at specified thickness, g
- $W_d$ dry weight of sample, g
- $ER_{th,A}$ electrolyte retention at specified thickness per sample area, cm$^3$/cm$^2$
- $\rho$ electrolyte density, g/cm$^3$
- $A$ sample area, cm$^2$
TABLE I—ELECTROLYTE RETENTION IN COMPRESSED STATE

<table>
<thead>
<tr>
<th>Separator</th>
<th>ER_{th,t} cm^3/cm^2</th>
<th>ER_{th} percent</th>
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<tbody>
<tr>
<td>FS2536, lot 13A</td>
<td>0.014</td>
<td>240</td>
</tr>
<tr>
<td>FS2438, lot 2C</td>
<td>0.014</td>
<td>260</td>
</tr>
<tr>
<td>Pellon 2505 (Gates)</td>
<td>0.017</td>
<td>360</td>
</tr>
<tr>
<td>ZYK15, lot W631</td>
<td>0.020</td>
<td>120</td>
</tr>
</tbody>
</table>

IV. Recommendations and criteria:

1. Electrolyte retention in the compressed state offers a more reliable measurement of the amount of electrolyte retained when a separator material is compressed to a representative interelectrode spacing. Generally, the higher the amount of electrolyte a separator can hold, the lower the probability of going dry during cycling. Give special attention to electrolyte retention values in the compressed state.

2. Typically, separator materials retain a minimum of 0.012 cm^3 of electrolyte per square centimeter of sample when compressed to 7.5 mils.

3. Results are reported as the electrolyte volume retained at a specified thickness per unit area of the material ER_{th,t} and as the percentage of the dry weight or electrolyte retention at a specified thickness ER_{th}. When comparing materials, ER_{th,t} is a more functional value because it is a direct measurement of the amount of electrolyte retained in the samples. As shown in table I, Zircar ZYK15 has an ER_{th} value half that for the nylon materials; yet it has a higher ER_{th,t}, indicating the presence of more electrolyte in the Zircar than in the nylon materials.
STP 5—DIRECT-CURRENT RESISTIVITY TEST PROCEDURE

Separators have two basic functions: to isolate the electrodes, and thus prevent electrical shorts, and to provide an electrolyte reservoir for ionic conductivity between the electrodes. It is this last function that involves the resistivity of the wet separator. Because of their porosity and ability to be permeated by the electrolyte, separators provide ionic conductivity in the same way as the electrolyte.

Resistivity measurements in wet separators are valuable indicators of the performance of separators in functional cells. They provide a means for determining the expected voltage drop, or IR, associated with a separator material. This test procedure employs a direct-current (DC) method to measure the ionic resistivity of the separator in the fully flooded, uncompressed state. A 50-mA current is passed through the separator and the voltage drop across the sample is measured with two reference electrodes. The lower the voltage drop across the sample, the lower the resistivity and the better the voltage performance of the cell.

I. Equipment and reagents:

1. DC power supply
2. Two voltmeters (one must be high impedance)
3. Resistivity cell (fig. 5)
4. Two reference electrodes (Ag/AgCl)—Prepare reference electrodes by submerging two 7.6-cm-long (3.0-in.-long) silver (Ag) wires in aqua regia solution for 1 min. (Other reference electrodes can be used.)
5. Two partially charged NiOOH/Ni(OH)₂ working electrodes—Partial charge is needed to avoid gas evolution.
6. Aqua regia—three parts concentrated HCl to one part concentrated HNO₃
7. TMI electronic micrometer
8. Electrolyte fill system (fig. 2)
9. Electrolyte solution
10. One 100-Ω resistor

II. Procedure:

1. Cut two samples, 2.5 cm by 2.5 cm (1.0 in. by 1.0 in.).
2. Measure and record thickness according to STP 1.
3. Fill separator samples with electrolyte as described in STP 2.
4. Insert reference electrode wires through the wells in the top of the cell fixture and into the electrolyte capillary channels. Connect the wire ends to the stainless steel bolts located on the top of the resistivity cell (fig. 5).
Figure 5.—Direct-current resistivity cell diagram and wiring. (a) Cell front view. (b) Cell end view.
5. Assemble the resistivity cell without a separator sample to obtain a blank reading. Make sure that the center holes are lined up.

6. Fill resistivity cell chambers with electrolyte and avoid air bubbles inside the cross channel and the capillaries.

7. Connect the leads from the DC power supply to the working electrodes, and connect the leads from the high-impedance voltmeter to the reference electrodes.

8. Connect the voltmeter across the 100-Ω resistor placed in series with the power supply. Adjust the power supply so that the current through the circuit is 0.05 A. A 5.0-V reading on the voltmeter means that a 0.05-A current is flowing in the cell.

9. Record the voltage drop reading. Obtain three additional readings as follows: (1) reverse the current flow, (2) reverse the polarity to the reference electrodes, and (3) reverse the current flow again. (Typical values are 60 to 100 mV for the blank \( E_b \). Unstable voltage readings may indicate the need to recondition the reference electrodes or to partially charge the working electrodes.)

10. Disconnect the wires. Remove the electrolyte from the resistivity cell, and save it for use with the samples to be tested.

11. Disassemble the cell and place the wet sample in the center of the resistivity cell so that it is centered over the cross-channel opening (fig. 5).

12. Assemble the cell fixture and repeat steps 6 to 11.

13. Repeat the blank measurements after all samples have been tested.

III. Calculations:

1. Calculate the average voltage drop for the blank \( E_b \) by using measurements from before and after sample testing.

2. Average the experimental voltage drop readings for each sample \( E_e \).

3. Calculate the voltage drop across the sample \( E_s \) by subtracting the voltage drop for the blank using

\[
E_s = E_e - E_b
\]

where

- \( E_s \) voltage drop across sample, V
- \( E_e \) average experimental voltage drop with separator sample, V
- \( E_b \) average voltage drop for blank, V

4. Determine the resistance \( R \) and the area resistivity \( AR \) for the sample by using the following equations:

\[
R = E_s/0.05A
\]

\[
AR = RA_e
\]
where

\[ R \quad \text{resistance across separator sample, } \Omega \]
\[ E \quad \text{voltage drop across sample, } V \]
\[ A \quad \text{area resistivity, } \Omega \text{-cm}^2 \]
\[ A_e \quad \text{area of separator exposed (0.95 cm}^2 \text{ depending upon fixture design), cm}^2 \]

IV. Criteria:

Area resistivity values should be less than 0.1 \( \Omega \text{-cm}^2 \).
STP 6—BUBBLE PRESSURE TEST PROCEDURE

Gases are routinely generated during normal operation of alkaline electrochemical cells. For cells to have long life and efficient operation, cell design and separator selection must accommodate the gases produced during cycling. At the end of charge and during overcharge of Ni/Cd cells, oxygen is normally evolved at the nickel electrode. In order to accommodate the oxygen generated, some cell designs require that the separator be sufficiently permeable to gas to allow oxygen to migrate to, and recombine at, the negative plate. The bubble pressure test indicates how permeable separator materials are to gases.

The apparatus used to determine bubble pressure consists of a gas regulator valve, a pressure gauge, and a fixture that distributes the gas evenly across the face of the sample (fig. 6). Air is introduced into the bottom plate of the fixture and pressure is gradually increased until a continuous flow of bubbles is observed through the separator into the water chamber (fig. 7). Bubble pressure is defined as the differential pressure required to generate a constant air bubble flux across the separator. The test is designed to identify the largest pore in a sample. Low bubble pressure indicates the presence of large pores (high permeability); high bubble pressure indicates the absence of large pores (low permeability). Because separator materials are susceptible to defects, the testing of multiple samples is recommended to ensure that experimental measurements are not the result of flaws or damaged samples.

Figure 6.—Bubble pressure apparatus.
Figure 7.—Bubble pressure fixture. (a) Cross-section view. (b) Bottom plate. (c) Top view of upper plate. (d) Bottom view of upper plate.
I. Equipment and reagents:

1. Analytical balance
2. TMI electronic micrometer
3. Bubble pressure apparatus (figs. 6 and 7)
4. Electrolyte solution
5. Deionized H₂O

II. Procedure:

1. Cut four separator samples 6.4 cm by 6.4 cm (2.5 in. by 2.5 in.).
2. Measure and record the thickness of the samples as in STP 1.
3. Weigh and record the dry weight for each sample \( W_d \).

**NOTE**
Dry weight and thickness measurements are taken only for information.

4. Fill the separator samples with electrolyte as described in STP 2.

5. Make sure the air regulator valve on the test fixture is closed. Place the wet sample on top of the porous nickel plaque on the stainless steel bottom plate. Assemble the apparatus as in figure 6.

6. Fill the recessed cavity with deionized water.

7. Record the initial pressure \( P_i \).

8. Slowly open the air regulator valve and increase the air pressure on the sample until a steady flux of bubbles is observed. The pressure at which a steady flux of bubbles is observed is called the final pressure.

9. Record the final pressure \( P_f \).

10. In order to verify that the previous recording is representative, and not the result of a flaw in the sample, slowly increase the air pressure to confirm that a uniform flux of bubbles is observed over the area of the sample with a minimum increase in pressure.

11. Close the valve, disassemble the fixture, and repeat steps 5 to 9 with the next sample.

III. Calculations:

1. Determine the bubble pressure \( P \) on the samples by using the following equation:

\[
P = P_f - P_i
\]

where

- \( P \) bubble pressure, psi
- \( P_f \) final pressure, psi
- \( P_i \) initial pressure, psi
2. Calculate the average of $P$ values for the four samples.

IV. Recommendations and criteria:

Cell design dictates the degree of permeability to gases needed in a separator. For example, some Ni/Cd cell designs require separators with high permeability to gases. Typical bubble pressure values of separators used in such Ni/Cd cells are under 2.0 psi. In contrast, some Ni/H$_2$ cell designs require separators with low permeability to gases. The asbestos separator material used in such Ni/H$_2$ cell designs has a bubble pressure higher than 30 psi.
Scanning electron microscopy (SEM) is used to investigate the morphology of separator materials. It is routinely used to establish the baseline characteristics of separators and then to screen subsequent samples of the same material for consistency and uniformity. For example, the baseline characteristics established for nylon separators include solid fiber clusters and smooth fiber surfaces with solid bonds between the fibers. Cracks, holes, irregularities in the structure, and detachment at the bond sites are unwanted characteristics, not exhibited by the baseline (fig. 8).

Two types of micrographs, surface and cross section, are taken to characterize separator materials and to screen for consistency and uniformity. Micrographs are taken at specific magnifications to allow for direct comparison of the sample with the baseline. SEM surface micrographs at low magnifications, 20X to 100X, are used to evaluate the structure and overall uniformity of the sample. Micrographs at high magnifications, 200X to 5000X, are used to investigate irregularities observed at lower magnifications and to screen for the quality of the material's constituents and for the presence of contaminants. Cross-section micrographs, taken at a magnification of 500X, are used to investigate the size and shape of the constituents and to evaluate for consistency and uniformity.

Figure 8.—Scanning electron micrographs of Pelion 2505. Surface micrographs (a) and (b) at low and high magnifications representing baseline for Pelion 2505; (c) and (d) undesirable characteristics of Pelion 2505.
I. Equipment and reagents:

1. Cambridge Scanning Electron Microscope Model 200
2. Nikon Epiphot Microscope with Sony UP 300 Photoprinter
3. Polaron Sputter Coater/SEM Unit E5100 with metal coating rings or equivalent

II. Procedure:

1. Cut and label the samples, according to your sampling plan, to fit the SEM mount for surface micrographs and the epoxy mount for cross-section micrographs.
2. For the surface micrographs, coat the samples with a metal layer by using the sputter coater.
3. Take surface micrographs with the SEM at the following magnifications: 20X, 50X, 100X, 200X, 500X, 1000X, 2000X, and 5000X.
4. For cross-section micrographs, mount the samples vertically in epoxy. Allow the epoxy to cure and polish the surface.
5. Coat the polished surface of the epoxy mount with a metal layer by using the sputter coater.
6. Take cross-section micrographs with the SEM at 500X. If difficulties are experienced in focusing the SEM, remove the metal coating and use a Nikon Epiphot Microscope at 500X magnification to take the cross-section micrographs.
7. Compare the micrographs and record any inconsistencies and/or variations between the baseline sample and the sample under investigation.

III. Recommendations and criteria:

1. Low magnifications (20X to 100X) are used to establish the general characteristics of the material. Uniformity and homogeneity are always desirable characteristics. Wherever possible, the sample should be compared with a baseline. The presence of any characteristics not exhibited by the baseline is cause for further investigation. At low magnifications, the fiber matrix of nonwoven nylon separators should look packed and solid. Signs of looseness and noncompacted fibers can indicate manufacturing irregularities and/or chemical degradation.

2. High magnifications (200X to 2000X) are used to establish the physical characteristics of the constituents. Wherever possible, the sample should be compared with a baseline. The presence of any characteristics not exhibited by the baseline is cause for further investigation. At high magnifications, nonwoven nylon materials should show fibers that look smooth, uniform, and solid. Similarly, the joints and bonds between fibers should look solid, without cracks or holes. The presence of cracks, holes, and/or irregularities on the surface and/or joints of nylon fibers can indicate chemical degradation.
STP 8—PROCEDURE FOR DETERMINING SUSCEPTIBILITY OF SEPARATOR MATERIALS TO HYDROLYSIS

Stability in alkaline environments is a desirable property for separator materials used in alkaline cells. However, under certain conditions, the nylon materials commonly used in Ni/Cd cells are susceptible to hydrolysis and decomposition. Studies on the susceptibility of nylon to hydrolysis have shown that degradation occurs during cell storage and cycling and that temperature and oxygen generation play important roles in the degradation (ref. 3). Reaction products from hydrolysis undergo oxidation, which ultimately results in carbonate formation. As carbonate forms, electrolyte concentration and cell voltage decrease, cadmium migration increases, and the overcharge protection is reduced. Consequently, cell performance decreases, possibly producing cell failure.

This test procedure uses weight loss and examination with scanning electron microscopy (SEM) to evaluate the susceptibility of separator samples to hydrolysis. Samples are exposed to the electrolyte solution for 20 days at 80 °C with sampling at 5-day intervals. Weight loss data and SEM micrographs are taken and compared with the baseline established for the material. At low magnifications, 20X through 100X, SEM micrographs of the samples that have undergone degradation typically show less integrity. Micrographs of these same samples, taken at high magnifications, 200X through 5000X, show degradation signs at the fiber level that are evident as cracks and the rippling of the fiber surface (fig. 9).

I. Equipment and reagents:

1. Oven
2. Twelve wide-mouth Teflon vials with 30-ml capacity
3. Analytical balance
4. TMI electronic micrometer
5. pH paper (0–14)
6. Desiccator
7. Electrolyte solution
8. 50-ml graduated cylinder
9. Deionized H₂O

II. Procedure:

1. Establish the baseline characteristics of the material by using SEM according to STP 7.

2. Cut and label 12 samples per material (3.0 cm by 3.0 cm; 1.2 in. by 1.2 in.).

3. Measure and record the thickness and dry weight \( W_d \) of each sample according to STP 1.

**NOTE**
Thickmess measurements are taken only for information.

4. Place each sample in a properly labeled, 30-ml-capacity, wide-mouth Teflon vial with 25 ml of electrolyte solution. Cap vials tightly and place them in an oven at 80 °C.

Perform steps 5 to 9 at 5-day intervals over a 20-day period.
Figure 9.—Effect of hydrolysis conditions on FS 2536 separator exposed to 31% KOH at 80 °C. These micrographs illustrate the progressiveness of the degradation process in nylon 2536 as a function of exposure time. (a) No exposure. (b) After 5 days. (c) After 10 days. (d) After 15 days. (e) After 20 days.

5. Remove three vials from the oven. Wash and dry each sample as follows:

   a. Rinse with 70 °C electrolyte solution.
   b. Rinse with deionized H₂O until the pH is 7.0 or below.
   c. Place in a 40 °C oven for 3 hr.
   d. Remove from the oven and place in a desiccator overnight.

6. Weigh (W₅) and record data for each sample.
7. Record any physical differences from initial conditions.

8. Select the sample showing the greatest degradation (i.e., the highest weight loss) and characterize with SEM according to STP 7.

9. Compare SEM micrographs with those of the baseline for the sample and note any differences.

III. Calculations:

1. Calculate the percent weight loss due to hydrolysis $\text{WL}_H$ for each sample after 5, 10, 15, and 20 days of electrolyte exposure by using the following equation:

$$\text{WL}_H = 100 \times \frac{(W_d - W_x)}{W_d}$$

where

- $\text{WL}_H$ weight loss due to hydrolysis, percent
- $W_d$ dry weight of sample, g
- $W_x$ hydrolysis weight of sample, g

2. Average the $\text{WL}_H$ values for the three samples at each five-day interval.

IV. Recommendations and criteria:

1. Exposure for 10 days is generally sufficient to determine if a material is sensitive to hydrolysis. Exposure for 15 or 20 days can indicate how progressive and/or severe decomposition can be.

2. Nylon materials have the highest susceptibility to degradation by hydrolysis among presently used separators. The $\text{WL}_H$ values for nyons should not exceed the following recommended percentages (table II), which are representative for nylon materials commonly used in aerospace applications.

Typically, non-nylon separators exhibit much lower weight loss due to hydrolysis $\text{WL}_H$ than those presented in table II.

<table>
<thead>
<tr>
<th>Length of hydrolysis, days</th>
<th>Weight loss due to hydrolysis, $\text{WL}_H$, percent</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>2</td>
</tr>
<tr>
<td>10</td>
<td>4</td>
</tr>
<tr>
<td>15</td>
<td>8</td>
</tr>
<tr>
<td>20</td>
<td>16</td>
</tr>
</tbody>
</table>

3. Materials showing excessive weight loss after 10 days of exposure to hydrolysis conditions should not be considered for use in cells.

4. Analysis of the electrolyte solution to which the samples were exposed can demonstrate the presence of hydrolysis reaction byproducts and additional information concerning the degradation process.
STP 9—ELECTROCHEMICAL OXIDATION TEST PROCEDURE

Separator degradation in secondary alkaline cells can occur in a number of ways. A common degradation process is the chemical and electrochemical oxidation of materials as a result of oxygen generated at the end of charge and during overcharge. Materials with minimal susceptibility to electrochemical oxidation are desirable for good cell performance.

Even without direct oxidation of the separator fibers, oxidation of the hydrolysis reaction products may occur. Hydrolysis and direct oxidation of the separator lead to deterioration of the material. Furthermore, oxidation of the hydrolysis reaction products leads to the production of undesirable byproducts. As a result, cell performance may be diminished, leading to premature cell failure. For example, nylon separators undergoing oxidation in Ni/Cd cells lose their integrity, and their oxidation reaction products increase the carbonate concentration in the electrolyte. An increase of carbonate concentration in the electrolyte results in higher electrolyte resistance, which is detrimental to good cell performance. Oxidation reaction products also reduce the cadmium hydroxide in the cadmium electrode. The reduction of the cadmium hydroxide to cadmium metal results in loss of overcharge protection, which leads to higher internal pressure during cell overcharge. This loss can result in premature cell failure (ref. 4).

This test procedure evaluates the susceptibility of separators to electrochemical oxidation. It exposes samples to oxygen generated during electrolysis, in flooded conditions, at 70 °C. The test runs for five days. Samples are tested at a current density of 5.0 mA/cm². Three control samples are also exposed to 70 °C electrolyte to determine percent weight loss due to hydrolysis.

I. Equipment and reagents:

1. Analytical balance
2. DC power supply
3. Voltmeter
4. Two pairs of cables with alligator clips
5. Cell components (fig. 10):
   a. Polysulfone cell case with cap (cap must have holes for the nickel sheet tabs)
   b. Four nickel sheets (11.0 cm by 7.5 cm; 4.3 in. by 3.0 in.), two of them with tabs
   c. Five pieces of separator sample (same size as nickel sheet)
   d. Four polypropylene screens (same size as nickel sheet)
   e. Plexiglas shims
   f. Electrolyte solution
6. Vacuum pump with bell jar assembly
7. Temperature-controlled water bath
8. Thermometer (in degrees Centigrade)
9. Styrofoam sheets and/or pellets
10. 30-ml, wide-mouth Teflon vial with lid
11. Three 9-cm² samples

II. Procedure:

1. Fill the water bath container and set the temperature to 70 °C. Use the thermometer to check water temperature.

2. Cut and label five separator samples the same size as the nickel sheets and the screens.

3. Cut and label three 9-cm² (1.4-in.²) separator samples to serve as controls for hydrolysis.
4. Weigh all the samples \((W_d)\) and record the data.

5. Assemble the cell as follows:

\[
\text{P} - \text{Ni} - \text{Sc} - \text{SM} - \text{Ni}_1 - \text{SM} - \text{Sc} - \text{SM} - \text{SC} - \text{SM} - \text{Ni}_1 - \text{SM} - \text{Sc} - \text{Ni} - \text{P}
\]

\(1\) \(2\) \(3\) \(4\) \(5\)
where

- P: Plexiglas shim
- Ni: nickel sheet
- Niₜ: nickel sheet with tab
- Sc: screen
- SM: separator material

6. Pour the electrolyte solution into the cell case until the electrolyte level is approximately 2 cm (0.8 in.) over the cell assembly. Mark the electrolyte level on the cell case. Place the cell case in the bell jar and evacuate for 5 min. Adjust the electrolyte level to the mark.

7. Cap the cell case, fitting the nickel sheet tabs through the cap holes (see fig. 10).

8. Place the three control samples (9 cm²; 1.4 in.²) with 25 ml of electrolyte inside a Teflon vial. Cap the vial and place it in the 70 °C water bath.

9. Place the cell case in the 70 °C water bath. Mark the water level in the bath.

10. Connect the power supply and voltmeter to the nickel sheet tabs. The voltmeter is used to monitor the voltage reading. Set power supply to maintain a current density of 5.0 mA/cm² (approximately 0.4 A). Typical voltage readings are about 2.0 V.

11. Cover water surface with Styrofoam sheets or pellets to minimize water evaporation.

12. Twice a day, for 5 days, check the current, water level, and temperature in the bath and the electrolyte level in the cell case. Adjust the water level in the bath and the electrolyte level in the cell case by adding 70 °C deionized H₂O. This addition compensates for the water lost due to evaporation and the electrolysis reaction.

13. After 120 hr, disassemble the cell. Remove the control samples from the Teflon vial. Rinse all separator samples with fresh 70 °C electrolyte solution and then with deionized H₂O until the pH is neutral.

14. Dry the samples in a 40 °C oven for 3 hr and place them in a desiccator overnight.

15. Weigh the samples exposed to electrochemical oxidation (Wₒ).

16. Weigh the three control samples (Wₙ).

III. Calculations:

1. Determine the total percent weight loss (hydrolysis plus oxidation, WLT) in the samples exposed to electrochemical oxidation by using the following equation:

   \[ WL_T = 100 \times \frac{W_d - W_O}{W_d} \]

where

- \( WL_T \): total (hydrolysis plus oxidation) weight loss, percent
- \( W_d \): dry weight of sample, g
- \( W_O \): weight of sample after exposure to oxidation, g

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2. Determine the percent weight loss due to hydrolysis $W_{LH}$ in the control samples by using the following equation:

$$W_{LH} = 100 \times \frac{W_d - W_c}{W_d}$$

where

- $W_{LH}$  weight loss due to hydrolysis, percent
- $W_d$  dry weight of sample, g
- $W_c$  weight of control samples after exposure to hydrolysis, g

3. Average the $W_{LT}$ values for the five large separators.

4. Average the $W_{LH}$ values for the three 9-cm$^2$ (1.4-in.$^2$) control samples.

5. Calculate the percent weight loss due to electrochemical oxidation $W_{LO}$ by subtracting the average percent weight loss due to hydrolysis $W_{LH}$ from the average total percent weight loss $W_{LT}$.

$$W_{LO} = W_{LT} - W_{LH}$$

IV. Recommendation:

Nonwoven nylon separators used in cells should exhibit less than 3 wt % loss due to electrochemical oxidation.
Separator composition is routinely monitored to evaluate the consistency of the material and to detect possible impurities that can affect cell performance. Chemical analysis test procedures are divided into two categories: inorganic, to determine metal and anionic composition; and organic, to identify carbon-containing compounds (fig. 11).

The first portion of the inorganic content analysis involves determining the metal composition of an ashed sample. In this procedure, inductively coupled plasma atomic emission vacuum spectrometry (ICP) and atomic absorption spectrophotometry (AAS) are used to determine metal content. The ICP uses a high-temperature excitation source for atomic emission analysis of chemical elements. ICP can detect the following elements: Al, B, Ba, Be, Bi, Ca, Co, Cr, Cu, Fe, Hf, Hg, K, La, Mg, Mn, Mo, Na, Nb, Ni, P, Pb, Sb, Si, Sn, Ta, Th, Ti, V, W, Y, Zn, and Zr. Because the ICP has low sensitivity to potassium and nickel and because cadmium emission lines overlap (interfere) with other metal emission lines, AAS is used to analyze for their presence. AAS detects the amount of radiation absorbed by an element, which is proportional to its concentration in the sample solution (ref. 5).

The inorganic content analysis is completed with the determination of anion concentrations in the sample. Anionic content analysis for chloride (Cl\(^{-}\)), nitrate (NO\(_3\)\(^{-}\)), and carbonate (CO\(_3\)\(^{2-}\)) are performed with the ion-specific electrodes (ISE) shown in figure 12. These electrodes measure the concentration of a specific anion in water leachants obtained from a separator sample.

The organic content analysis involves an infrared (IR) spectra analysis and an ultraviolet (UV) light screening test. The infrared spectra analysis consists of a spectrophotometric evaluation of a pellet made from methanol leachables. The IR spectra of the residue is compared with a baseline to identify the functional groups present in the sample. The UV light screening test is performed on samples to detect fibers that absorb and fluoresce against UV light. Separators used in secondary alkaline cells generally do not contain fibers that fluoresce under UV light.

![Separator sample diagram]

Figure 11.—Chemical analysis performed on separator materials.
Figure 12.—Ion-specific electrode setup.
I. Equipment and reagents:

1. Analytical balance
2. Platinum dish with a platinum lid
3. Muffle furnace with a range of 400 °C to 800 °C
4. Desiccator
5. Inductively coupled plasma atomic emission vacuum spectrometer (ICP)
6. Atomic absorption spectrophotometer (AAS)
7. Deionized H₂O
8. Concentrated HF (hydrofluoric acid)
9. 8 M HClO₄ (perchloric acid)
10. 25-ml plastic volumetric flask
11. Gloves

II. Preparation of material:

1. Weigh 5 g of separator material to the nearest 0.1 g (W_d).
2. Touch the sample material to a grounded strip to drain away as much static electricity as possible.
3. Cut the material into approximately 5-cm² (0.8-in.²) pieces and place them into a tared, 7.5-cm-diameter (3-in.-diameter), platinum dish (W_p) with a platinum lid.
4. Ash the material in a muffle furnace as follows: place the platinum dish with sample in the furnace and set the temperature to 500 °C for 1 hr; then increase the temperature to 750 °C and heat for an additional hour.
5. Remove the dish from the furnace and place it in a desiccator to cool.
6. Weigh the remaining sample ash with the dish (W_sp). Repeat exposure at 750 °C for 30 min, cool, and reweigh.
7. Repeat this cycle as necessary until a constant weight (W_sp) is achieved.

III. Procedure:

1. Prepare the sample solutions as follows:
   a. Dissolve the sample ash in the platinum dish by first adding 5 ml of deionized water, then add 2 to 3 ml of concentrated HF and heat gently. After most of the sample ash is dissolved and the reaction subsides, add 1 ml of 8 M HClO₄ with gentle heating. Transfer the solution to a 25-ml plastic volumetric flask and dilute to the mark with distilled water.
   b. Prepare a blank solution containing the added reagents (2 to 3 ml of HF and 1 ml of 8 M HClO₄, diluted to 25 ml with deionized H₂O). Also prepare standard solutions containing 1.0 ppm and 5.0 ppm of the metals of interest. Standard solutions must include the same proportion of acid solutions used to dissolve the sample ash.
2. Using the ICP element analysis software, enter the weight of the ash sample, the volume the sample was diluted to (25 ml), and the concentrations of the standards used (1.0 ppm and 5.0 ppm).

3. Run the standard solutions and the blank through the ICP.

4. Run the sample solutions through the ICP. Output results are expressed as percent weight of metal (WM) in the sample ash. Run each sample three times to ensure reproducibility.

5. Using the AAS computer program, enter the same information logged in step 2. Run blanks and standard solutions through the AAS.

6. Run the sample solution through the AAS to detect elements showing low sensitivity and emission line interference with other elements in the ICP. Output results are expressed as percent weight of metal (WM) in the sample ash. Run each sample three times to ensure reproducibility.

IV. Calculations:

1. Calculate the weight of the sample ash \( W_s \), the percent weight of ash in the sample \( WA \), the weight of metal in the sample ash \( W_m \), and the percent weight of metal in the separator sample \( WS \) by using the following equations:

\[
W_s = W_{sp} - W_p
\]

\[
WA = \frac{W_s}{W_d} \times 100
\]

\[
W_m = \frac{W_s \times WM}{100}
\]

\[
WS = \frac{W_m}{W_d} \times 100
\]

where

- \( W_s \): weight of sample ash, g
- \( W_{sp} \): weight of platinum dish, lid, and sample ash, g
- \( W_p \): weight of platinum dish with lid, g
- \( WA \): weight of ash, percent
- \( W_d \): dry weight of sample, g
- \( W_m \): weight of metal in sample ash, g
- \( WM \): weight of metal in sample ash, percent
- \( WS \): weight of metal in separator sample, percent

V. Recommendations and criteria:

1. Establish the expected composition of the material to facilitate the interpretation of results.

2. Note that inorganic content varies according to the type of material. For example, nonwoven nylon materials have an average ash content of less than 0.5 wt %. In contrast, Zircar separators have an average ash content of 99.9 wt %.

3. Always consider the effects of the metal traces on the electrochemistry and performance of the cell.
I. Equipment and reagents:

1. Analytical balance
2. Boiled deionized H₂O (used at room temperature)
3. Shaker or means to agitate sample in solution
4. Chloride (Cl⁻), nitrate (NO₃⁻), and carbonate (CO₃²⁻) ion-specific electrodes (ISE)
5. Two reference electrodes: one for the Cl⁻ and one for the NO₃⁻ ISE
6. pH/ISE meter
7. Three small stir bars
8. Semilogarithmic graph paper
9. Filling solutions for Cl⁻, NO₃⁻, and CO₃²⁻ electrodes
10. Carbon dioxide (CO₂) buffer
11. Ionic strength adjustor solution (ISA)
12. 1 M boric acid solution (preservative solution)
13. 100 ml of 1000-ppm CaCO₃ standard solution
14. 100 ml of 1000-ppm NaCl standard solution
15. 100 ml of 1000-ppm NaNO₃ standard solution
16. Twelve 100-ml volumetric flasks with stoppers
17. Five 100-ml beakers for the CO₃²⁻ standards and sample
18. Ten 150-ml beakers for the Cl⁻ and NO₃⁻ standards and samples
19. Parafilm paper
20. One 500- to 1000-ml container with cap

II. Procedure:

1. Standards and sample preparations are time consuming. Because the shelf life of the standard solutions is 2 weeks, they can be prepared ahead of time as follows:

   a. Weigh 5 g of sample to the nearest 0.1 g, cut it into small pieces, and place it in a clean 500- to 1000-ml container with 500 ml of boiled deionized H₂O.

   **NOTE**
   In order to reduce CO₂ content, all deionized H₂O used in this procedure must be boiled, covered, and cooled to room temperature before use.

   b. Cap container, place it in shaker, and agitate at low speed for 24 hr.

   c. Prepare the standard sample solutions by diluting in series the corresponding 1000-ppm anion standard solution. Make 100-ml standard solutions of 10, 100, 500, and 1000 ppm by first adding 2 ml of ISA solution in the Cl⁻ and NO₃⁻ volumetric flasks and 10 ml of CO₂ buffer in the CO₃²⁻ flasks. Also, add 1 ml of preservative solution in all standard solution flasks. Perform corresponding serial dilutions. Wrap the stoppers with Parafilm paper; cap the flasks, shake them, and store them at room temperature.

2. Assemble ion-specific electrodes and reference electrodes with their respective filling solutions. Calibrate all the electrodes and have the pH/ISE meter primed.

3. Starting with a 1000-ppm standard, pour 50 to 75 ml of solution into a beaker. Stir gently, without heating, and take three voltage readings (millivolts) with the respective ISE and reference electrode. Record the data. Before switching to the next concentration, rinse the electrodes with deionized H₂O and blot them dry. Always take voltage readings...
readings from high to low concentration. Measure and record the potentials on the other two sets of anion standard solutions.

4. For the Cl\(^-\) and NO\(_3^-\) sample measurements, dispense 100 ml of leachate into a 150-ml beaker and add 2 ml of ISA. Stir gently and obtain three voltage readings for each anion.

5. For the CO\(_3^{2-}\) sample measurements, dispense 50 ml of leachate into a 100-ml beaker. Add 5.0 ml of CO\(_2\) buffer and stir gently. Obtain three voltage readings.

III. Calculations:

1. Average the potential readings (millivolts) for each anion concentration standard solution. Prepare the calibration curves on semilogarithmic paper for each standard anion. Plot the average voltage readings on the linear axis (y axis) and the standard concentration values on the logarithmic axis (x axis).

2. Perform regressions and determine the respective linear equations for each species calibration curve. Typical calibration curves for the anions of interest are shown in figure 13.

3. Average the voltage readings from the sample anions and plot the average voltage (millivolts) on the corresponding standard curve to determine the anion species concentration in the leachate \(C_l\). Use corresponding linear equations to determine \(C_l\) outside the measured range.

4. Determine the anion concentration in the separator sample \(C_s\) as follows:

\[
C_s = \frac{C_l \times 500 \text{ ml}}{5.0 \text{ g}}
\]

where

- \(C_s\) anion concentration in separator sample, ppm in \(\mu g/g\)
- \(C_l\) anion concentration in leachate, ppm in \(\mu g/ml\)

IV. Recommendations and criteria:

1. Establish the expected composition of the material to facilitate the interpretation of results.

2. Temperature fluctuations, drift, and noise limit the reproducibility when using ISE’s (ref. 6). Work in a temperature-controlled area, free from noise.

3. Prepare the standard curves every time you are testing.
Figure 13.—Ion-specific electrode calibration curves for Cl⁻, NO₃⁻, and CO₃⁻² anions. (a) Typical chloride electrode calibration curve. (b) Typical response of carbon dioxide electrode. (c) Typical nitrate electrode calibration curve.
Figure 13.—Concluded. (c) Typical nitrate electrode calibration curve.
I. Equipment and reagents:

1. Reagent-grade methanol
2. Analytical balance
3. Shaker or means to agitate the sample in solution
4. Rotary evaporator system with a rotary flask (500-ml capacity)
5. Mortar and pestle
6. Dry potassium bromide (KBr)
7. Pellet press
8. Spatula
9. Infrared spectrophotometer with multiple internal reflectance (MIR) attached (if available)
10. Oven
11. Small stirring bar

II. Procedure:

1. Weigh 5 g of sample material to the nearest 0.1 g (Wd).

2. Preweigh the rotary flask (Wf) and place the sample inside. Add 500 ml of methanol (100 ml for every gram of sample).

   **NOTE**
   The ratio of methanol to sample can be modified to maximize leaching or to minimize methanol use. A ratio of 50:1 volume to weight is the recommended minimum.

3. Insert a small stirring bar in the rotary flask to ensure proper agitation. Seal the flask and place it in the shaker; agitate at low speed.

4. Remove the sample from the methanol leachate after 24 hr of agitation. Dry the sample in an oven at 40 °C for 15 min. Weigh the dry sample and report the weight after extraction (W_e).

5. Evaporate the leachate with the rotary evaporator system, under vacuum. Do not apply heat.

   **NOTE**
   There is a possibility that no residue will be recovered. If this occurs, repeat the extraction with a larger sample.

6. Make sure no methanol remains in the rotary flask. After the methanol has evaporated, weigh the rotary flask with the residue and record the weight (W_f).

7. Remove the residue from the rotary flask with a spatula and place it in a mortar. Mix the residue with KBr and make a pellet. (A 1 to 5% mix of sample in KBr by weight is usually used.)

8. Run an infrared spectrum on the KBr sample pellet.

9. Run an infrared spectrum on a piece of sample by using the MIR attachment. The sample can be ground with KBr and pressed into a pellet if an MIR is not available.
III. Calculations:

1. Obtain an infrared baseline spectrum for the material (fig. 14).

2. Compare the residue and the sample spectra with the baseline spectrum. Note any additional or missing absorbance peaks. Identify impurities, if any, based on this information. Keep in mind that the residue can contain portions of the material being studied.

3. Calculate percent weight loss on sample WL and residue weight \( W_r \) as follows:

\[
WL = 100 \times \frac{W_d - W_e}{W_d}
\]

\[
W_r = W_{fr} - W_f
\]

Figure 14.—Representative infrared spectrum for FS 2538 separator.
where

\[
\begin{align*}
WL & \quad \text{weight loss in sample, percent} \\
W_d & \quad \text{dry weight of sample, g} \\
W_e & \quad \text{weight of sample after extraction, g} \\
W_r & \quad \text{weight of residue, g} \\
W_{fr} & \quad \text{weight of rotary flask and residue, g} \\
W_f & \quad \text{weight of rotary flask, g}
\end{align*}
\]

IV. Recommendations and criteria:

1. Establish the expected composition of the material to facilitate the interpretation of results.

2. Because the baseline and sample material spectra should show identical peak locations and relative intensities, place attention on the identification of extraneous peaks.
STP 10D—Organic Chemical Analysis: Ultraviolet Light Test
To Detect Contaminant Fibers

I. Equipment:

1. Gloves

2. Eye protection (goggles or shield glasses that protect against ultraviolet (UV); for example, goggles with thick polycarbonate lenses)

3. Multiband UV lamp (254 nm/366 nm)

4. A frame to define the area to be measured (typically 25 cm by 25 cm; 10 in. by 10 in.)

II. Procedure:

1. Assemble the sample, frame, and UV lamp (fig. 15) in a clean area that can be completely darkened.

2. Wearing gloves and eye protection, place the frame on top of the sample, turn the lights off, and apply the UV light about 1 in. from the sample surface.

3. Count the number of fibers fluorescing within the framed area by using both long and short wavelengths. Repeat on the opposite side of the material. Record the total number of fibers. If sample size allows, select two more areas and repeat the procedure.

**NOTE**
DO NOT expose eyes and skin to short-wave UV light because the rays may be harmful to unprotected eyes and skin.

![Ultraviolet light test setup.](image-url)
III. Calculations:

Average the number of fibers fluorescing within the framed areas $N$ and determine the number of fluorescent fibers per sample area $F$.

$$F = \frac{N}{A}$$

where

- $F$ number of fluorescent fibers per square centimeter
- $N$ average number of fibers within framed area
- $A$ area of frame, cm$^2$

IV. Recommendations and criteria:

1. Establish the expected composition of the material to facilitate the interpretation of results.

2. None of the fibers in separators used in secondary alkaline cells fluoresce against ultraviolet light. Screening with UV light helps to identify the presence of foreign fibers. For example, some nonwoven nylon materials have been found to be contaminated with Dacron fibers by using this screening test. The source of the Dacron was fibers remaining from previous runs in the mixing chambers used to blend the nylon fibers before manufacturing. Typical values for nylon materials are less than one fluorescent fiber per 6.4 cm$^2$ of material.
Separator materials used in secondary alkaline cells require sufficient strength to withstand handling during cell assembly. They must also retain their integrity and withstand the effects of vibration and shock under service requirements. Tensile strength measurements provide a means of evaluating the strength and integrity of a material. The tensile strength test measures elongation as a function of load when the load is applied in tension. Tensile strength is defined as the peak load, in pounds, per sample cross-sectional area measured in square inches (fig. 16).

Although tensile strength is used to evaluate the integrity of a material, a minimum tensile strength is not a prerequisite for separator selection. For example, Zircar separators, which have extremely low tensile strength when measured with this method, perform well in Ni/Cd and Ni/H₂ cells. Tensile strength measurements are also used to evaluate the quality and uniformity of separators. The measurements can also indicate changes in the molecular structure resulting from degradation of the material. For instance, when nylon fibers hydrolyze and degrade, the tensile strength decreases.

Tensile strength measurements are highly dependent on test conditions, which must be consistent and controlled to obtain reproducible and comparable results. Some materials exhibit directional characteristics that result in variations of tensile strength as a function of manufacturing direction. For such materials, measurements are taken from samples cut parallel to and perpendicular to the machine direction. Testing is performed on the material in the dry state.

I. Equipment and reagents:

1. Analytical balance
2. TMI electronic micrometer
3. Rule die (10.2 cm long by 6.4 cm wide; 4 in. long by 2.5 in. wide)
4. Tensile strength machine (Satec Systems Model 50 UD with tensile test system software) with load cell of 6000 lb.

![Figure 16.—Tensile strength test results. Test identifier, FS 2536; rate, 0.3 in./min; filament direction, parallel; exposed sample area; 2.5 in. by 3 in.; tensile strength, 2280 psi.](image)

Figure 16.—Tensile strength test results. Test identifier, FS 2536; rate, 0.3 in./min; filament direction, parallel; exposed sample area; 2.5 in. by 3 in.; tensile strength, 2280 psi.
II. Procedure:

1. Cut three samples with the rule die. For materials that exhibit directional characteristics as a result of manufacturing, cut three samples along the machine direction (MD) and three across the MD. Position the rule die with the long side parallel to the direction to be evaluated.

2. Label and measure the thickness of each sample (refer to STP 1).

3. Insert a sample between the tensile strength machine holders. Keep the area exposed between tensile strength machine holders constant. Typically, the area exposed between the holders is 7.6 cm long by 6.4 cm wide (3.0 in. long by 2.5 in. wide).

4. Enter the identification, thickness, and width of the sample in the computer program. The cross-sectional area used by the software is the product of the sample thickness and width. Run the test at a constant rate of 0.75 cm/min (0.30 in./min).

5. Repeat step 4 with all samples. The output is presented as tensile strength plots showing minimum and maximum elasticity points and the breaking point, tensile strength TS, in pounds per square inch (psi).

III. Calculations:

1. Obtain the tensile strength values TS for each sample from the output plot (fig. 16).
2. Report averaged TS values along MD and across MD for every material.
Mercury porosimetry is one of the most common methods used for determining pore size distribution and void volume in porous materials. The test provides valuable information concerning separator characteristics and helps to evaluate for consistency among separator lots. The test measures the amount of mercury taken up by the sample as a function of pressure.

The mercury porosimetry technique employs a penetration instrument (fig. 17) that includes a pressure source and a detector to determine the amount of mercury introduced into the pores as pressure is increased. From the volume of mercury penetrating the pores as a function of pressure, properties such as specific surface area, density, percent porosity, average pore size, and pore distribution can be obtained (ref. 7).

The results are calculated by assuming that sample materials are noncompressible, have cylindrical pores, and are nonreactive to mercury. However, separator materials are generally compressible and contain noncylindrical pores. These characteristics will affect the actual porosity and pore size distribution values for the sample. Because testing is performed on the material in the dry state, results are not representative of cell conditions.

The mercury porosimetry test is performed by an outside contractor. Typical mercury porosimetry reports include tabulated data and plots. Tabulated data contain information calculated from the amount of mercury introduced into the pores of the sample as a function of pressure. Tabulated data include the following information:

1. Total intrusion volume: maximum volume of mercury introduced into the sample pores
2. Total pore area: surface area within the pores, assuming cylindrical geometry (fig. 18)
3. Median pore diameter (volume or area): diameter value obtained from the area or volume distribution curves
4. Average pore diameter: four times the pore volume divided by the pore area, assuming cylindrical geometry

![Figure 17.—Schematic diagram of mercury porosimetry penetration instrument.](image-url)
5. Bulk density: actual density of the sample, calculated from the sample weight and volume

6. Skeletal density: density of the material, calculated from the total intrusion volume and the bulk density

7. Porosity: percent volume of pores in the sample, calculated from the total intrusion volume and the actual sample volume

8. Experimental raw data: pressure, pore diameter, incremental volume, cumulative volume, cumulative pore area, and incremental pore area

Mercury porosimetry plots display the raw data organized in five different graphs. The graphs show relationships of cumulative, incremental, and differential intrusions and cumulative and incremental pore areas versus pore diameters found in the sample. Figure 19 is a plot of pore size distribution or incremental intrusion versus pore diameter for Zircar separator material.
Figure 19 — Effect of pore diameter on incremental intrusion volume for Zircar
### APPENDIX A

**EVALUATION FORM FOR SEPARATOR MATERIALS**

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>STP number</th>
<th>Baseline value</th>
<th>Experimental value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Area weight, $A_W$, g/m²</td>
<td>STP 1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Thickness, $T$, mm</td>
<td>STP 1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dimensional stability, $D_I$ and $D_w$, after 10 days, percent</td>
<td>STP 3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Electrolyte retention:</td>
<td>STP 4</td>
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</tr>
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<td>Uncompressed, ER, percent</td>
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<td></td>
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</tr>
<tr>
<td>$ER_1$, cm³/cm²</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Compressed, ER, percent</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>$ER_2$, percent</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$ER_2$, cm³/cm²</td>
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</tr>
<tr>
<td>Area resistivity, $AR$, Ω·cm²</td>
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<tr>
<td>Bubble pressure, $P$, psi</td>
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</tr>
<tr>
<td>SEM screening (observations)</td>
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<td></td>
</tr>
<tr>
<td>Hydrolysis, $WL_{H}$, percent</td>
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</tr>
<tr>
<td>5 days</td>
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<td>10 days</td>
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<td>15 days</td>
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<td></td>
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<tr>
<td>20 days</td>
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<tr>
<td>Electrochemical oxidation, $WL_{EO}$, percent</td>
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<td>Inorganic chemical analysis:</td>
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<tr>
<td>Ash, WA, percent</td>
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<tr>
<td>Metals found in ash, WM, percent</td>
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</tr>
<tr>
<td>$Cl^-$ concentration, ppm</td>
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<td></td>
</tr>
<tr>
<td>$NO_3^-$ concentration, ppm</td>
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<td></td>
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</tr>
<tr>
<td>$CO_3^{2-}$ concentration, ppm</td>
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<td>Organic chemical analysis:</td>
<td>STP 10</td>
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<td>Infrared spectra functional group, cm⁻¹</td>
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<tr>
<td>Methanol extraction, $WL$</td>
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<td>Ultraviolet screening, $F$, fibers/cm²</td>
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<tr>
<td>Tensile strength, $TS$, psi</td>
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<td></td>
</tr>
<tr>
<td>Along machine direction</td>
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<td></td>
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<tr>
<td>Across machine direction</td>
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<td>Mercury porosimetry:</td>
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<tr>
<td>Total intrusion volume, ml/g</td>
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<td></td>
</tr>
<tr>
<td>Total pore area, m²/g</td>
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<td></td>
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</tr>
<tr>
<td>Median pore diameter by volume, μm</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Median pore diameter by area, μm</td>
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<td></td>
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</tr>
<tr>
<td>Average pore diameter, $4V/A$, μm</td>
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<td></td>
</tr>
<tr>
<td>Bulk density, g/ml</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Apparent (skeletal) density, g/ml</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Porosity, percent</td>
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APPENDIX B
CHARACTERIZATION RESULTS FOR SELECTED SEPARATOR MATERIALS

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>FS 2536, lot 13A</th>
<th>FS 2538, lot 2C</th>
<th>Pellon 2505 (Gates)</th>
<th>Zircar ZYK15, lot W-631</th>
</tr>
</thead>
<tbody>
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<td>Area weight, AW, g/m²</td>
<td>88.1</td>
<td>61.7</td>
<td>570</td>
<td>215.8</td>
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<td>0.291</td>
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<td>Dimensional stability:</td>
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<td></td>
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<tr>
<td>after 10 days, percent:</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>D₁</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>D₂</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
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<tr>
<td>Electrolyte retention:</td>
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<td></td>
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<td></td>
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<tr>
<td>Uncompressed, ER, percent</td>
<td>474</td>
<td>772</td>
<td>713</td>
<td>164</td>
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<tr>
<td>ERo, cm³/cm²</td>
<td>0.032</td>
<td>0.036</td>
<td>0.028</td>
<td>0.028</td>
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<td>Compressed, ERₚ, percent</td>
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<td>303</td>
<td>400</td>
<td>122</td>
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<tr>
<td>ERp, cm³/cm²</td>
<td>0.016</td>
<td>0.017</td>
<td>0.018</td>
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<td>Bubble pressure, P, psi</td>
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<td>1.4</td>
<td>1.0</td>
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<tr>
<td>SEM screening (observations)</td>
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<td></td>
</tr>
<tr>
<td>Packed matrix with smooth and solid fibers (no cracks, no holes)</td>
<td>Packed matrix with smooth and solid fibers (no cracks, no holes)</td>
<td>Loose matrix and junctions with irregular shaped fibers (cracks and holes)</td>
<td>Packed uniform matrix with smooth and solid fibers (no cracks, no holes)</td>
<td></td>
</tr>
<tr>
<td>Hydrolysis, WLₜₜ, percent:</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>5 days</td>
<td>0.5</td>
<td>0.4</td>
<td>7.4</td>
<td>(a)</td>
</tr>
<tr>
<td>10 days</td>
<td>1.3</td>
<td>2.2</td>
<td>16.2</td>
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<td>15 days</td>
<td>2.6</td>
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<tr>
<td>20 days</td>
<td>4.6</td>
<td>7.9</td>
<td>32.1</td>
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</tr>
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<td>1.4</td>
<td>50</td>
<td>(a)</td>
</tr>
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<td>Inorganic chemical analysis:</td>
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<tr>
<td>Ash, WA, percent</td>
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<td>0.31</td>
<td>0.35</td>
<td>99.8</td>
</tr>
<tr>
<td>Metals found in ash, WM, percent</td>
<td>Na (6.6), K (5), P (3), Ti (39), other traces (&lt;1)</td>
<td>Na (5.8), K (4), P(2), Ti (40), other traces (&lt;1)</td>
<td>Na (10), K (4), Zn (12), P (3), Ti (41), other traces (&lt;1)</td>
<td>Hf (1.2), Si (4), Ti (0.13), other traces (&lt;1)</td>
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<td>Cl⁻ concentration, ppm</td>
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<td>3.6</td>
<td>28.1</td>
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<td>NO₃⁻ concentration, ppm</td>
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<td>4.1</td>
<td>15.2</td>
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<td>CO₃²⁻ concentration, ppm</td>
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<td>310</td>
<td>680</td>
<td>474</td>
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<td>Organic chemical analysis:</td>
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<tr>
<td>Infrared spectra functional group, cm⁻¹</td>
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</tr>
<tr>
<td>NH (3.3K)</td>
<td>(b)</td>
<td></td>
<td>(b)</td>
<td>(c)</td>
</tr>
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<td>CH₃–CH (3.1K–2.9K)</td>
<td>NH (3.3K)</td>
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<td>NH (3.3K)</td>
<td></td>
</tr>
<tr>
<td>O = C–NH</td>
<td>CH₃–CH (3.1K–2.9K), O = C–NH</td>
<td>COOH (1.6K–1.5K), COOH (1.6K–1.5K), C-N, C-H, C-C (1.4K–1.0K)</td>
<td>COOH (1.6K–1.5K), C-N, C-H, C-C (1.4K–1.0K)</td>
<td></td>
</tr>
<tr>
<td>COOH (1.6K–1.5K)</td>
<td>C-N, C-H, C-C (1.4K–1.0K)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Methanol extraction, WL</td>
<td>1.17</td>
<td>0.84</td>
<td>1.17</td>
<td>(c)</td>
</tr>
<tr>
<td>Ultraviolet screening, F, fibers/cm²</td>
<td>0</td>
<td>0.6</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Tensile strength, TS, psi:</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Along machine direction</td>
<td>2470</td>
<td>1630</td>
<td>1474</td>
<td>(d)</td>
</tr>
<tr>
<td>Across machine direction</td>
<td>690</td>
<td>370</td>
<td>317</td>
<td>(d)</td>
</tr>
<tr>
<td>Mercury porosimetry¹:</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total intrusion volume, ml/g</td>
<td>3.12</td>
<td>4.20</td>
<td>6.01</td>
<td>1.22</td>
</tr>
<tr>
<td>Total pore area, m²/g</td>
<td>78.79</td>
<td>56.51</td>
<td>109.80</td>
<td>42.28</td>
</tr>
<tr>
<td>Median pore diameter by volume, μm</td>
<td>42.23</td>
<td>56.71</td>
<td>49.29</td>
<td>23.31</td>
</tr>
<tr>
<td>Median pore diameter by area, μm</td>
<td>0.0055</td>
<td>0.0055</td>
<td>0.0058</td>
<td>0.0057</td>
</tr>
<tr>
<td>Average pore diameter, 4V/A, μm</td>
<td>0.1582</td>
<td>0.2075</td>
<td>0.2191</td>
<td>0.1157</td>
</tr>
<tr>
<td>Bulk density, g/ml</td>
<td>0.2594</td>
<td>0.2013</td>
<td>0.1476</td>
<td>0.7213</td>
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<tr>
<td>Apparent (skeletal) density, g/ml</td>
<td>1.3527</td>
<td>1.3080</td>
<td>1.3104</td>
<td>6.1336</td>
</tr>
<tr>
<td>Porosity, percent</td>
<td>80.82</td>
<td>84.61</td>
<td>88.74</td>
<td>88.24</td>
</tr>
</tbody>
</table>

¹Weight loss from hydrolysis and/or chemical oxidation in Zircar materials is minimal. Typically, weight loss is due to material handling.
²Figure 14 is a representative spectrum for nylon materials.
³Not applicable.
Figure 20.—Effect of pore diameter on incremental intrusion volume for selected separator materials.
APPENDIX C
ADDITIONAL TEST PROCEDURES

STP C1—Electrolyte Retention in Uncompressed State by Using Kubelka Jar

Electrolyte retention using a Kubelka jar is an alternative procedure for measuring electrolyte retention in the uncompressed state. It is commonly used by Freudenberg Nonwovens Inc., a manufacturer of nonwoven nylon separators. This procedure involves the use of a Kubelka jar, which consists of a glass reservoir attached to a graduated cylinder. Electrolyte retention is determined by difference, by knowing the amount of electrolyte originally placed in the reservoir and then measuring the amount returned to the graduated cylinder after soaking and draining the samples.

I. Equipment and reagents:

1. Analytical balance
2. TMI electronic micrometer
3. Kubelka jar (fig. 21)
4. Tweezers
5. Electrolyte solution

II. Procedure:

1. Cut and label three samples of equal area, 5 cm by 5 cm (2 in. by 2 in.).
2. Weigh the samples \(W_d\) and measure the thickness according to STP 1.
3. Fill the calibrated cylinder in the Kubelka jar with electrolyte and note the level \(V_1\). Tip the Kubelka jar slowly, causing the electrolyte solution to accumulate in the reservoir.
4. Immerse all samples in the electrolyte solution for a minimum of 1 hr (4 hr for nonwoven nylon separators) or until there are no visible dry spots on the sample.
5. Carefully turn the jar upright, allowing the excess electrolyte to flow back into the calibrated cylinder. Allow sample to drain for 1 min.
6. Record the electrolyte volume in the graduated cylinder \(V_2\).
III. Calculations:

1. Sum all three $W_d$ to obtain the total dry weight for the samples ($\Sigma W_d$).

2. Calculate the electrolyte retention in the uncompressed state ER and the electrolyte volume retained $V$ by using the following equations:

$$ V = V_1 - V_2 $$

$$ ER = 100 \times \frac{V \rho}{\Sigma W_d} $$

where

$V$ electrolyte volume retained, cm$^3$

$V_1$ initial electrolyte volume from calibrated cylinder, cm$^3$

$V_2$ final electrolyte volume from calibrated cylinder, cm$^3$

$ER$ electrolyte retention in uncompressed state, percent

$\rho$ electrolyte density, g/cm$^3$

$\Sigma W_d$ total sum of dry weights, g
STP C2—Electrolyte Distribution Test Procedure

This electrolyte distribution test procedure was designed to study the electrolyte competition between cell components. The test also provides a variety of information on separator characteristics. Measurements of area density, thickness, and electrolyte retention in the uncompressed and compressed states and information on the affinity of the cell components for the electrolyte at less than a 100% fill level can be obtained. This STP was designed specifically for Ni/Cd cell components, but it can be modified for other chemistries by using representative cell components and by evaluating and changing the cell fill procedure as necessary.

I. Equipment and reagents:

1. Electrolyte fill system (fig. 2)
2. Electrolyte distribution cell assembly (fig. 22)
3. Analytical balance
4. Oven
5. Tweezers
6. Clock
7. Feeler gauge
8. Filter paper
9. 1.0-ml graduated pipette
10. Electrolyte solution
11. One nickel and one cadmium plate, 5 cm by 5 cm (2 in. by 2 in.)
12. TMI electronic micrometer

II. Procedure to determine 100% fill level in cell components at a specified shim thickness:

1. Start with clean and dry nickel and cadmium plates and a 5-cm by 5-cm (2-in. by 2-in.) separator sample. Measure and record the thickness and weight of each component \( W_{d,x} \).

2. Fill the cell components to 100% fill level as described in STP 2.

3. Determine the appropriate shim thickness by adding the thicknesses of the electrodes (nickel and cadmium plates) and the desired interelectrode spacing (fig. 22).

4. Remove the cell components from the electrolyte, suspend each one vertically for 1 min, and touch the tip of the sample to the surface of a piece of filter paper five times to remove excess electrolyte.

**NOTE**
Weighing the separator sample at this point will enable the determination of electrolyte retention in the uncompressed state.

5. Assemble the cell components as shown in figure 22. Adjust the nuts and screws tightly. Verify the stack compression by measuring the gap between the retaining plates with a feeler gauge. Adjust the gap by adding or eliminating shims to achieve the desired thickness. Repeat check with feeler gauge.

6. Stand the cell assembly vertically for 4 hr to drain the excess electrolyte. Disassemble the cell, weigh the cell components, and record the weights \( W_{x,th,100} \).

7. Wash the nickel and cadmium plates by rinsing and vacuum filling with deionized \( H_2O \) until pH is 7.0 or below. Dry the plates in a 40 °C oven for at least 2 hr, and store them in a desiccator.
III. Calculations:

1. Determine the assembled 100% fill electrolyte volume $V_{x,th,100}$ for each component, the total electrolyte volume in the cell stack $V_{T,th,100}$, and the electrolyte distribution for each component at 100% fill $D_{x,th,100}$ by using the following equations:

$$WE_{x,th,100} = (W_{x,th,100} - W_{d,x})$$

$$V_{x,th,100} = \frac{WE_{x,th,100}}{\rho}$$

$$V_{T,th,100} = \Sigma V_{x,th,100}$$

$$D_{x,th,100} = 100 \times \frac{V_{x,th,100}}{V_{T,th,100}}$$
where

- \( W_{E,x,th,100} \): weight of electrolyte in component at specified interelectrode spacing at 100% fill, g
- \( W_{x,th,100} \): weight of component at specified interelectrode spacing at 100% fill, g
- \( W_{d,x} \): dry weight of component, g
- \( V_{x,th,100} \): electrolyte volume in component at specified interelectrode spacing at 100% fill, ml
- \( \rho \): electrolyte density, g/ml
- \( V_{T,th,100} \): total volume of electrolyte at specified interelectrode spacing in cell stack at 100% fill, ml
- \( \Sigma V_{x,th,100} \): sum of electrolyte volume of all components at specified interelectrode spacing at 100% fill, ml
- \( D_{x,th,100} \): proportion of total electrolyte for each component at specified interelectrode spacing at 100% fill, percent

Subscripts:

- \( x \): component (Ni, Cd, Sm)
- \( th \): interelectrode spacing, mils

2. Determine the corresponding electrolyte volumes for fill

\[
V_{th,\%} = \frac{V_{T,th,100} \times \%}{100}
\]

where

- \( V_{th,\%} \): electrolyte volume at specified interelectrode spacing at percent fill level, ml
- \( V_{T,th,100} \): total electrolyte volume in cell stack at specified interelectrode spacing at 100% fill, ml
- \( \% \): percent fill (20 to 90)

IV. Procedure to determine the electrolyte distribution and percent saturation at different percent fill levels for a specified thickness (typically the distribution is evaluated at 20, 40, 60, 70, 80, and 100%):

1. Start with clean and dry electrodes (nickel and cadmium plates) and a 5-cm by 5-cm (2-in. by 2-in.) separator sample. Measure and record the thickness and dry weight of all components \( W_{d,x} \).

2. Start with the lowest percent fill level (20%) and, using a 1.0-ml graduated pipette, dispense the appropriate volume of electrolyte (as calculated in section IV) for the desired fill level into the components. Equally distribute the electrolyte volume between the three components. The exact distribution of the electrolyte among cell components is not critical as long as sufficient time is allowed for equilibration.

3. Employing the same shim size used in section II, assemble the cell fixture (fig. 22). For the 20% fill level, allow the electrolyte to be absorbed into the components for 30 min before tightening the bolts.

4. Tighten the bolts and stand the cell fixture vertical for 1 hr.

5. Quickly disassemble the cell. Weigh and record the weights of the components \( W_{x,th,\%} \).

6. Add an additional increment of electrolyte in each component for the next percent fill level and quickly reassemble the cell.
7. Repeat steps 4, 5, and 6 until the distribution at the 100% fill level has been evaluated.

8. Wash the nickel and cadmium plates by rinsing and vacuum filling with deionized H₂O until pH is 7.0 or below. Dry electrodes in a 40 °C oven for at least 2 hr and store them in a desiccator.

V. Calculations:

1. The percent saturation of electrolyte for a particular component represents its fill level as a function of its 100% fill level, which is defined as the saturation point. Determine the weight of electrolyte in each component \( W_{E,x,th,\%} \) and the percent saturation of individual components \( S_{x,th,\%} \) at a specified thickness at each percent fill level by using the following equations:

\[
W_{E,x,th,\%} = W_{x,th,\%} - W_d,
\]

\[
S_{x,th,\%} = 100 \times \frac{W_{E,x,th,\%}}{W_{E,x,th,100}}
\]

where

- \( W_{E,x,th,\%} \): weight of electrolyte in component at specified interelectrode spacing at percent fill level, g
- \( W_{x,th,\%} \): weight of component at specified interelectrode spacing at percent fill level, g
- \( W_d \): dry weight of components, g
- \( S_{x,th,\%} \): saturation level of cell component at specified interelectrode spacing at percent fill level, percent
- \( W_{E,x,th,100} \): weight of electrolyte in component at specified interelectrode spacing at 100% fill level (from section IV, step 1), g

Subscripts:
- \( x \): component (Ni, Cd, or Sm)
- \( th \): interelectrode spacing, mils

2. Graph the results, percent fill level on the \( x \) axis versus percent saturation for each individual component on the \( y \) axis (see fig. 23).

3. Determine the electrolyte distribution \( D_{x,th,\%} \) for the individual components at a specified thickness at each percent fill level by using the following equations:

\[
W_{E,T,th,\%} = \Sigma W_{E,x,th,\%}
\]

\[
D_{x,th,\%} = 100 \times \frac{W_{E,x,th,\%}}{W_{E,T,th,\%}}
\]

where

- \( W_{E,T,th,\%} \): total weight of electrolyte in cell stack at specified interelectrode spacing at percent fill level, g
- \( \Sigma W_{E,x,th,\%} \): sum of electrolyte weight for each component at specified interelectrode spacing at percent fill level, g
Figure 23.—Electrolyte distribution and percent saturation plots for three-component Ni/Cd cell with FS 2538 separator material. (Plots show how cell components compete for the electrolyte and how the saturation states vary at different percent fill levels. The separator begins to compete for electrolyte after cell is 40% filled. (a) Electrolyte distribution. (b) Saturation of individual components.
**STP C2**

\[ D_{x,\text{th},\%} \]
proportion of total electrolyte in cell component at specified interelectrode spacing at percent fill level, percent

\[ \text{WE}_{x,\text{th},\%} \]
weight of electrolyte in component at specified interelectrode spacing at percent fill level, g

**Subscripts:**
- \( x \) component (Ni, Cd, Sm)
- \( \text{th} \) interelectrode spacing, mils

**NOTE**
\[ \sum \text{WE}_{x,\text{th},\%} \] is the summation of the experimental values and not the calculated values in section IV, step 2. Also, the saturation level and the electrolyte distribution values can be calculated by using the volume of electrolyte in the components at the particular percent fill level.

4. Graph the results, percent fill level on the \( x \) axis versus percent electrolyte distribution of individual components on the \( y \) axis (see fig. 23).

VI. Recommendations and criteria:

Recommendations and criteria from STP's 4A and 4B similarly apply in this procedure.
## APPENDIX D
### SYMBOLS

**STP 1**

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$A$</td>
<td>sample area, $m^2$</td>
</tr>
<tr>
<td>$A_W$</td>
<td>area weight, $g/m^2$</td>
</tr>
<tr>
<td>$T$</td>
<td>average sample thickness, mm</td>
</tr>
<tr>
<td>$W_d$</td>
<td>dry weight of sample, g</td>
</tr>
</tbody>
</table>

**STP 3**

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$D_w$</td>
<td>change in sample width, percent</td>
</tr>
<tr>
<td>$D_l$</td>
<td>change in sample length, percent</td>
</tr>
<tr>
<td>$l_d$</td>
<td>length of sample in dry state, cm</td>
</tr>
<tr>
<td>$l_w$</td>
<td>length of sample in wet state, cm</td>
</tr>
<tr>
<td>$w_d$</td>
<td>width of sample in dry state, cm</td>
</tr>
<tr>
<td>$w_w$</td>
<td>width of sample in wet state, cm</td>
</tr>
</tbody>
</table>

**STP’s 4A and 4B**

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$A$</td>
<td>sample area, $cm^2$</td>
</tr>
<tr>
<td>$ER$</td>
<td>electrolyte retention in uncompressed state, percent</td>
</tr>
<tr>
<td>$ER_A$</td>
<td>electrolyte retention per sample area in uncompressed state, $cm^3/cm^2$</td>
</tr>
<tr>
<td>$ER_{th}$</td>
<td>electrolyte retention at specified thickness, percent</td>
</tr>
<tr>
<td>$ER_{th,A}$</td>
<td>electrolyte retention at specified thickness per sample area, $cm^3/cm^2$</td>
</tr>
<tr>
<td>$W_d$</td>
<td>dry weight of sample, g</td>
</tr>
<tr>
<td>$W_{th}$</td>
<td>wet weight of sample at specified thickness, g</td>
</tr>
<tr>
<td>$W_w$</td>
<td>wet weight of sample, g</td>
</tr>
<tr>
<td>$\rho$</td>
<td>electrolyte density, $g/cm^3$</td>
</tr>
</tbody>
</table>

**STP 5**

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<tr>
<th>Symbol</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$A_e$</td>
<td>area of separator exposed, $cm^2$</td>
</tr>
<tr>
<td>$AR$</td>
<td>area resistivity, $\Omega \cdot cm^2$</td>
</tr>
<tr>
<td>$E_b$</td>
<td>average voltage drop for blank, V</td>
</tr>
<tr>
<td>$E_e$</td>
<td>average experimental voltage drop with separator sample, V</td>
</tr>
<tr>
<td>$E_s$</td>
<td>voltage drop across sample, V</td>
</tr>
<tr>
<td>$R$</td>
<td>resistance for separator sample, $\Omega$</td>
</tr>
</tbody>
</table>

**STP 6**

<table>
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<th>Symbol</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$P$</td>
<td>bubble pressure, psi</td>
</tr>
<tr>
<td>$P_f$</td>
<td>final pressure, psi</td>
</tr>
<tr>
<td>$P_i$</td>
<td>initial pressure, psi</td>
</tr>
</tbody>
</table>
Symbols

**STP 8**

- $W_d$: dry weight of sample, g
- $WL_H$: weight loss due to hydrolysis, percent
- $W_x$: hydrolysis weight of sample, g

**STP 9**

- $W_c$: weight of control samples after exposure to hydrolysis, g
- $W_d$: dry weight of sample, g
- $WL_H$: weight loss due to hydrolysis, percent
- $WL_O$: weight loss due to electrochemical oxidation, percent
- $WL_T$: total (hydrolysis plus oxidation) weight of sample, g
- $W_O$: weight of sample after exposure to oxidation, g

**STP’s 10A, 10B, 10C, and 10D**

- $A$: area of frame, cm$^2$
- $C_s$: anion concentration in separator sample, ppm in $\mu$g/g
- $C_l$: anion concentration in leachate, ppm in $\mu$g/ml
- $F$: number of fluorescent fibers per square centimeter, fibers/cm$^2$
- $N$: average number of fibers within framed area
- $TS$: tensile strength, psi
- $WA$: weight of ash, percent
- $W_d$: dry weight of sample, g
- $W_e$: weight of sample after extraction, g
- $W_f$: weight of rotary flask, g
- $W_{fr}$: weight of rotary flask and residue, g
- $WL$: weight loss in sample, percent
- $WM$: weight of metal in sample ash, percent
- $W_m$: weight of metal in sample ash, g
- $W_p$: weight of platinum dish with lid, g
- $W_r$: weight of residue, g
- $WS$: weight of metal in separator sample, percent
- $W_s$: weight of sample ash, g
- $W_{sp}$: weight of platinum dish, lid, and sample ash, g

**STP C1**

- $ER$: electrolyte retention in uncompressed state, percent
- $V$: electrolyte volume retained, cm$^3$
- $V_1$: initial electrolyte volume from calibrated cylinder, cm$^3$
- $V_2$: final electrolyte volume from calibrated cylinder, cm$^3$
- $\rho$: electrolyte density, g/cm$^3$
- $\Sigma W_d$: total sum of dry weights, g
Symbols

**STP C2**

\[ D_{x,\text{th},100} \]
proportion of total electrolyte for each component at specified interelectrode spacing at 100% fill, percent

\[ D_{x,\text{th},\%} \]
proportion of total electrolyte in cell component at specified interelectrode spacing at percent fill level, percent

\[ S_{x,\text{th},\%} \]
saturation level of cell component at specified interelectrode spacing at percent fill level, percent

\[ V_{T,\text{th},100} \]
total volume of electrolyte at specified interelectrode spacing in cell stack at 100% fill, ml

\[ V_{\text{th},\%} \]
electrolyte volume at specified interelectrode spacing at percent fill level, ml

\[ V_{x,\text{th},100} \]
electrolyte volume in component at specified interelectrode spacing at 100% fill, ml

\[ \Sigma V_{x,\text{th},100} \]
sum of electrolyte volume of all components at specified interelectrode spacing at 100% fill, ml

\[ W_{d,x} \]
dry weight of components, g

\[ W_{x,\text{th},100} \]
weight of component at specified interelectrode spacing at 100% fill, g

\[ W_{x,\text{th},\%} \]
weight of component at specified interelectrode spacing at percent fill level, g

\[ \text{WE}_{T,\text{th},\%} \]
total weight of electrolyte in cell stack at specified interelectrode spacing at percent fill level, g

\[ \text{WE}_{x,\text{th},100} \]
weight of electrolyte in component at specified interelectrode spacing at 100% fill, g

\[ \text{WE}_{x,\text{th},\%} \]
weight of electrolyte in component at specified interelectrode spacing at percent fill level, g

\[ \Sigma \text{WE}_{x,\text{th},\%} \]
sum of electrolyte weight for each component at specified interelectrode spacing at percent fill level, g

\[ \rho \]
electrolyte density, g/ml

\[ \% \]
percent fill (20 to 90)

**Subscripts:**

\[ x \]
component (Ni, Cd, Sm)

\[ \text{th} \]
interelectrode spacing, mils
REFERENCES


Test Procedures for Characterizing, Evaluating, and Managing Separator Materials Used in Secondary Alkaline Batteries

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Cleveland, Ohio 44135–3191

NASA TM–107292

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Secondary alkaline batteries, such as nickel-cadmium and silver-zinc, are commonly used for aerospace applications. The uniform evaluation and comparison of separator properties for these systems is dependent upon the measurement techniques. This manual presents a series of standard test procedures that can be used to evaluate, compare, and select separator materials for use in alkaline batteries. Detailed test procedures evaluating the following characteristics are included in this manual: physical measurements of thickness and area weight, dimensional stability measurements, electrolyte retention, resistivity, permeability as measured via bubble pressure, surface evaluation via SEM, chemical stability, and tensile strength.