This invention provides analogs of triptycene which are useful as anticancer drugs, as well as for other uses. The potency of these compounds is in a similar magnitude as daunomycin, a currently used anticancer drug. Each compound of the invention produces one or more desired effects (blocking nucleoside transport, inhibiting nucleic acid or protein syntheses, decreasing the proliferation and viability of cancer cells, inducing DNA fragmentation or retaining their effectiveness against multidrug-resistant tumor cells).

40 Claims, 23 Drawing Sheets
OTHER PUBLICATIONS

W. Marsh et al., Relation and characterization of Adriamycin–resistant HL60 cells which are not defective in the initial intracellular accumulation of drug, Cancer Res. 46:4053–4057, 1986.
* cited by examiner
Fig. 1
Fig. 2
Fig. 3
Fig. 4
Fig. 5
Fig. 6
Fig. 7
Fig. 8
Fig. 9
Fig. 10
Fig. 11
Fig. 12

Drug Treatments (μM) Maintained (+) or Removed (-)

Control
TT2(10)
TT2(25)
HHO (25)

DNA Synthesis (% of control)

Nucleoside Transport (% of control)
Fig. 13
Fig. 14
Fig. 15

Cellular Transport of $^3$H-Adenosine (A) and $^3$H-Thymidine (B) (% of control)

DAU (open) and TT2 (closed) Concentrations (μM)

Fig. 15
Fig. 16
Fig. 17
Drug Treatments (µM)

Mitotic Index

Control 0.256 0.64 1.6 4 1 0.2

TT2 (□) and DAU (■) HIO VCR

Fig. 18
DNA Fragmentation After 24h (%)

0.1024 0.256 0.64 1.6 4 10 25

TT1 (○), TT2 (●), DAU (■) and CPT (▲) Concentrations (μM)

Fig. 19
DNA Fragmentation After 24h (%) vs. TT (code number) Treatments (7 μM)

Fig. 20
Fig. 21
Fig. 22

DNA Fragmentation After 24h (%)  

- Control  
- TTI(10)  
- TT2(10)  
- DAU(1.6)  
- CPT(1.6)  

Drug Treatments (µM) Maintained For 24h (□) or Removed After 3 (□) or 8h (■)
Fig. 23
TRIPTYCENE ANALOGS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to U.S. provisional application Ser. No. 60/238,856, filed on Oct. 6, 2000, which is hereby incorporated by reference to the extent not inconsistent with the disclosure herewith.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with Government support under Grant No. NAGW-1197 awarded by NASA and Grant No. 86842 awarded by the National Institute of Health. The Government has certain rights in the invention.

BACKGROUND OF THE INVENTION


SUMMARY OF THE INVENTION

This invention provides analogs of triptycene which are useful as anticancer drugs, as well as for other uses. The potency of these compounds is in a similar magnitude as daunomycin, a currently used anticancer drug. Each compound of the invention produces one or more desired effects (blocking nucleoside transport, inhibiting nucleic acid or protein syntheses, decreasing the proliferation and viability of cancer cells, inducing DNA fragmentation or retaining their effectiveness against multidrug-resistant tumor cells).

More specifically, the invention provides triptycene analogs having the following formula:

wherein \( X \) is selected from the group consisting of: H, R, SR and \( \text{NR}^2 \); \( Y \) is selected from the group consisting of: halogen (preferably Cl, Br, I), R, \( \text{NR}_2 \), SR and H; \( R \) and \( R^1 \)–\( R^2 \) are independently selected from the group consisting of: H, halogen, OR, and hydrocarbyl (preferably lower alkyl, alky, phenyl, aryl, substituted alkyl, substituted alky, substituted phenyl, substituted phenyl, \( \text{CH}_2=\text{(CH}_3\text{)}_n\text{CO}_2\text{H} \), \( \text{CH}_2=\text{(CH}_3\text{)}_n\text{CH}(\text{NH}_2)\text{CO}_2\text{H} \), carboxylic acid, substituted carboxylic acid, amine, substituted amine, NHR, \( \text{NR}_2 \), amino acid, \( \text{RCO}_{(\text{CH}_2)_n}\text{NH} \), where one or both of the hydrogen atoms on \( \text{CH}_2 \) can be substituted with alkyl, alky, phenyl, aryl, substituted alky, substituted phenyl, substituted carboxylic acid, amine, or substituted amine, and where \( n \) is an integer from 0 to 8); \( R^3 \)–\( R^4 \), independently of one another, are selected from the group consisting of: H, halogen (preferably bromine), OR, R, SR and \( \text{NR}_2 \); \( R^5 \), independently of other \( R^5 \)s, is selected from the group consisting of: \( =\text{O} \), \( =\text{N}–\text{OH} \), and \( =\text{CHR} \); and reduced forms thereof; wherein in reduced forms, either ring A or ring C or both is replaced with

and wherein in reduced forms, each \( R^5 \) is independently H, Cl–C8 alky or \( =\text{OR} \); and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

The numbering scheme used herein is shown in the example structure below:
Other compounds of the invention include those with formula:

wherein

- \( R^2 \) is selected from the group consisting of: \( R \), halogen, \( NR_3 \), \( SR \) and \( H \); \( R^2 \) is selected from the group consisting of: \( H \), \( R \), \( SR \) and \( NR_3 \); \( R^2 \) and \( R^3 \) are independently selected from the group consisting of: \( H \), halogen, \( OR \) and \( hydrocarbyl \); \( R^2 \) and \( R^3 \) are independently selected from the group consisting of: \( H \), halogen, (preferably bromine), \( R \), \( SR \) and \( NR_3 \); \( R^{10} \) and \( R^{10} \) are independently selected from the group consisting of: \( H \), halogen, \( OR \) and \( hydrocarbyl \); reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Other compounds of the invention include those with formula:

wherein

- \( X \) is selected from the group consisting of: \( H \), \( R \), \( SR \) and \( NR_3 \); \( R \) and \( R^{11} \) are independently selected from the group consisting of: halogen, \( NR_3 \), \( SR \), \( H \) and \( R \); \( R^2 \) and \( R^{12} \) are independently selected from the group consisting of: \( H \), halogen, \( OR \) and \( hydrocarbyl \); \( R^2 \) and \( R^{12} \) are independently selected from the group consisting of: \( H \), halogen, \( OR \) and \( hydrocarbyl \); reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Also provided are compounds of the formula:

wherein \( X \) is \( -NW(CW_2)_2Z \), where the \( Ws \) are independently selected from the group consisting of: \( H \), \( Cl-C8 \) alkyl, and \( Cl-C8 \) alkenyl and \( Z \) is selected from the group consisting of: \( R \), \( COR \), \( COOR \), \( CONR_2 \), \( OOCR \) and \( NRCOR \);

- \( Y \) is selected from the group consisting of: halogen, \( Cl-C8 \) alkyl, \( Cl-C8 \) alkenyl, \( OR \), \( NR_3 \), \( SR \), \( H \), \( COR \), \( OCOR \) and \( NRCOR \);

- \( R \) and \( R^{17} \) are independently selected from the group consisting of: \( H \), \( OR \), and \( hydrocarbyl \);

- \( R^2 \), independently of one another, are selected from the group consisting of: \( H \), \( OR \), \( SR \), and \( NR_3 \);

- \( R^2 \), independently of other \( R^5 \)s, is selected from the group consisting of: \( =O \), \( =NOH \), \( =CHR \) and reduced forms thereof; \( R^{21} \) and \( R^{22} \) are independently selected from the group consisting of: \( H \), \( R \), and \( OR \); and reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Also provided are compounds of the formula:

wherein X is selected from the group consisting of: \( H \), \( R \), \( SR \) and \( NR_3 \); \( Y \) is selected from the group consisting of: halogen, \( NR_3 \), \( SR \), \( H \) and \( R \); \( R \) and \( R^{12} \) are independently selected from the group consisting of: \( H \), halogen, \( OR \) and \( hydrocarbyl \); \( R^{11} \) and \( R^{12} \) are independently selected from the group consisting of: \( H \), halogen, \( OR \) and \( hydrocarbyl \); reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Also provided are compounds of the formula:

wherein \( X \) is \( -NW(CW_2)_2Z \), where the \( Ws \) are independently selected from the group consisting of: \( H \), \( Cl-C8 \) alkyl, and \( Cl-C8 \) alkenyl and \( Z \) is selected from the group consisting of: \( R \), \( COR \), \( COOR \), \( CONR_2 \), \( OOCR \) and \( NRCOR \);

- \( Y \) is selected from the group consisting of: halogen, \( Cl-C8 \) alkyl, \( Cl-C8 \) alkenyl, \( OR \), \( NR_3 \), \( SR \), \( H \), \( COR \), \( OCOR \) and \( NRCOR \);

- \( R \) and \( R^{17} \) are independently selected from the group consisting of: \( H \), \( OR \), and \( hydrocarbyl \);

- \( R^2 \), independently of one another, are selected from the group consisting of: \( H \), \( OR \), \( SR \), and \( NR_3 \);

- \( R^2 \), independently of other \( R^5 \)s, is selected from the group consisting of: \( =O \), \( =NOH \), \( =CHR \) and reduced forms thereof; \( R^{21} \) and \( R^{22} \) are independently selected from the group consisting of: \( H \), \( R \), and \( OR \); and reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Also provided are compounds of the formula:
wherein all but one of X, Y, R1 and R2 is independently H, C1–C6 alkyl, C1–C6 alkenyl, OR, SR or NR2 wherein each R is independently H or C1–C6 alkyl and the other R1 or R2 is a solubilizing group; and each R5 is independently H, C1–C6 alkyl or OR. The solubilizing group may be of the formula: NR(CR,),X wherein X is a sugar, R, COR, COOR, CONR,, OOCR and NRCOR; R is independently selected from the group consisting of: H, C1–C8 alkyl and C1–C8 alkenyl; n is an integer from 1 to 8.

Scheme 1 shows some of the compounds of the invention and abbreviations used herein.

TT1: X = Y = H
TT2: X = PMe, Y = H
TT13: X = OMe, Y = Br
TT14: X = NHMe, Y = Br
TT15: X = NHMe, Y = OH
TT16: X = OMe, Y = Br, R2 = NMe2
TT17: X = NHCH2CH2CO2H, Y = Br
TT18: X = NHCH2CH2CO2Et, Y = OH
TT19: X = NHCH2CH2(NH2)CO2H, Y = Br
TT20: X = NHCH2CH2(NH2)CO2H, Y = OH
TT21: X = NHCH2CH2CO2H, Y = OH
TT24A: X = OMe, Y = Br, R2 = NHMe
TT24B: X = OMe, Y = Br, R2 = NHMe
N analog 1: X = NH(CH2)2CH2(NH2)CHCO2H, Y = Br
N analog 2: Y = Br, X =

N analog 3: Y = Br, X =

N analog 4: X = OMe, Y = Br, R2 =

S analog 5: Y = Br, X = SCH2CH(CO2H)NH2

TT3:

TT5: X = R3 = OMe
TT7: X = R3 = H
TT9: X = OMe, R3 = H
Another class of compounds of this invention includes homologs of the foregoing compounds. One class of compounds is the compounds TT1-13. Another class of compounds is TT14-20. One class of compounds are those where X is selected from the group consisting of: H, OMe and CO2Me; where Y is selected from the group consisting of: H, Br, and OMe; where \( R^1=R^2=H \); and where positions 1, 4, 5 and 8 are selected from the group OH, OMe, =O, H.

Another class of compounds include those where one or more substituents contains one or more N atoms. Another class of compounds includes those where X and Y are not both members of the group containing: H, OMe, Br, CO2Me, while positions 1, 4, 5, and 8 are substituted with —OH, —OMe or =O, or mixtures of those substituents. Another class of compounds include those which include at least one amine, amino acid or amine sugar substituent.

A preferred class of compounds is those which are water soluble, where one or more substituents, particularly where X, Y, R1 and/or R2 substituents of formula I are replaced with water soluble group or groups that enhance the solubility of the compound and salts thereof.

A preferred class are those compounds of formula I where X is a water soluble group or a group that enhances the water solubility of the compound; and salts thereof. These compounds include those where X is RO2C(CH2)nNH, where n is an integer from 1 to 8 and R is as defined for I.

Another class of compounds includes those that contain a sulfur containing substituent.

This invention also provides methods for inhibiting cellular transport of nucleosides; inducing DNA fragmentation; inhibiting nucleic acid and/or protein synthesis and decreasing the proliferation and viability of cancer cells (including wild type and multi-drug resistant) or other cells in which the proliferation or viability is desired to be reduced, comprising contacting the cells with an effective amount of a compound of the invention as disclosed herein. This invention provides such compounds in suitable pharmaceutical carriers in dosages effective to provide measurable nucleoside transport blocking, nucleic acid and/or protein synthesis inhibition, DNA cleavage, and/or reduction in tumor cell (including wild type and multi-drug resistant) growth and/or viability. Preferably, the compounds used in the methods of this invention are almost or at least as effective as Daunomycin, a currently used anticancer drug.

Also provided is a method of treating cancer in a host, comprising: administering to said host an effective amount of an active compound of the invention for an effective time. Administration routes include intravenously, parenterally, and other methods known in the art. As used herein, an “effective amount” is an amount which causes a measurable effect on a desired parameter. As used herein, an “effective time” is the time required to cause a measurable or desired effect on a desired parameter.

Also provided is a method for preparation of triptycene analogs, comprising in situ oxidation and [4+2] cycloaddition of substituted benzenes or quinones and optionally substituted anthracenes; and optional oxidation of the resulting compounds. The resulting compounds will be a mixture of methoxy-substituted and carbonyl-substituted triptycene analogs. This mixture can be separated into individual compounds with methods known in the art. The one pot
synthesis may be separated into oxidation and cycloaddition steps, if desired. Synthesis of particular groups of com-
ounds of the invention are described in more detail herein. All substituents of the indicated compounds in a humidified incubator at 37°C. Using the assay methods disclosed herein and those methods described herein, the following two steps (reduction with sodium hydrosulfite and methylation) have been used in certain syntheses, but preparation of 1,4-dimethoxyanthracene (1) has not been reported in one sequence of reaction.

A new synthesis of the compound TT2 is also provided. Either TT3, TT5 or a mixture of both is oxidized to give TT2. A new method to brominate triptycene analogs is also provided. Treatment of a triptycene analog which has a methoxy group on position 2 and a hydrogen on position 3 with N-bromosuccinimide gives an analog with a bromine on position 3. This bromination reaction can be extended to other triptycene analogs. For example, if a methoxy group is on position 6 or 7, bromination will result in a bromine on position 7 or 6, respectively. If the starting compound has methoxy groups on positions 5 and 6, oxidation will give the corresponding analog with carbonyl groups on positions 5 and 6.

Triptycene analogs bearing functionalities at C12 and C13 (R5 and R6), can be made in an analogous reaction as described below starting with 6,7-disubstituted 1,4-dimethoxyanthracenes (analog of compound 1). These 6,7-disubstituted 1,4-dimethoxyanthracenes are prepared from the corresponding analogs of 6,7-disubstituted 1,4-dihydroxy-9,10-anthraquinones (by following the method described below).

Also provided are triptycene analogs prepared by the methods described herein.

Compounds containing any combination of substituents or members of the Markush groups specified above are within the scope of the invention. All substituents of the compounds of the invention may be the same, all substituents may be different, or any combination of substituents may be the same or different. Compounds having substituents with a specified function, for example those that impart water solubility to the compound form a special class of compounds of this invention.

The substituents included in the compounds of the invention and used in the methods of the invention may be any substituent not having structures or reactivity which would substantially interfere with the desired activity of the compound, as may readily be determined without undue experimentation by those skilled in the art, for example, by using the assay methods disclosed herein and those methods known to one of ordinary skill in the art.

The reduced forms of all compounds described herein are included in the disclosure. It is understood that when referring to reduced forms of structures herein, any quinone-like ring of the structure may be replaced with a hydroquinone-like ring, as known in the art.

For example, in the following structure,

![Chemical Structure](image)

in reduced forms, either ring A or ring C or both is replaced with

![Chemical Structure](image)

wherein in reduced forms, each R2 is independently a reduced form of =0, for example, OR. In the reduced forms, each R5 may also be H or C1–C8 alkyl, for example, as known in the art.

**BRIEF DESCRIPTION OF THE DRAWINGS**

FIG. 1. Comparison of the abilities of novel TIs to inhibit the growth (open) and viability (striped) of L1210 cells after 4 days in vitro. For tumor cell growth and viability, L1210 cells were respectively seeded in triplicate at initial densities of 1×10⁶ and 1.1×10⁶ cells/0.5 ml/well in RPMI 1640 medium containing 7.5% FCS and penicillin (100 IU/ml)–streptomycin (100 μg/ml), and grown for 4 days in the presence or absence (control) of 250 nM concentrations of the indicated compounds in a humidified incubator at 37°C. with 5% CO₂ in air. Cell density was monitored using a Coulter counter. Cell growth results are expressed as % of the number of vehicle-treated control cells after 4 days in culture (1,465,110±292,595; 100±6%; open control). The ability of viable cells/0.5 ml to bioreduce 0.1 ml of MTS-PMS (20:1) reagent over a 3-h incubation period at 37°C was assayed by measuring the absorbance of the water-soluble formazan products at A₅₇₀ nm. Cell viability results are expressed as % of the net absorbance of MTS formazan after bioreduction by vehicle-treated control cells (A₅₇₀ nm=1.164±0.089; 100±8%; striped control) after 4 days in culture. The blank value (A₅₇₀ nm=0.202) for cell-free medium supplemented with MTS-PMS reagent has been subtracted from the results. Bars: mean±SD (n=3). *Not different from TT7 and TT9; †P<0.025, smaller than TT1, TT7 and TT9 but not different from TT13; ‡P<0.05, greater than TT1, TT7 and TT9 but not different from TT5, TT6 and TT8; ‡‡not different from control; ‡‡‡P<0.05, smaller than control.
FIG. 2. The same experiment as in FIG. 1 with additional compounds.

FIG. 3. Comparison of the concentration-dependent inhibitions of L1210 cell growth by the TT analog TT2 ( ● ) and DAU ( □ ) at days 2 (open symbols) and 4 (closed symbols) in vitro. The protocol of the experiment was identical to that of FIG. 1. The results are expressed as % of the numbers of vehicle-treated control cells (100±5%; striped area) after 2 (213,798±8,316 cells/ml) and 4 days (1,311,274±55,991 cells/ml) in culture. Drug concentrations are plotted on a logarithmic scale. Bars: means±SD (n=3). *Not different from control; †P<0.025 and ‡P<0.01, smaller than control.

FIG. 4. Comparison of the concentration-dependent inhibitions of L1210 cell viability by the TT analog TT2 ( ● ) and DAU ( □ ) at days 2 (open symbols) and 4 (closed symbols) in vitro. Cells were seeded in triplicate at initial densities of 100,000 or 11,100 cells/0.5 ml well and respectively incubated for 2 or 4 days in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale. The ability of viable cells/0.5 ml to bioreduce 0.1 ml of MTS: PMS (20:1) reagent over a 3-h incubation period was assessed as described in FIG. 1. Cell viability results are expressed as % of the net absorbance of MTS/formazan after bioreduction by vehicle-treated control cells (100±5%; striped area) at days 2 (A950 nm=1.206±0.093) and 4 (A950 nm=1.096±0.086). Blank values (A950 nm=1.185 and 0.189 at days 2 and 4) for culture medium supplemented with MTS:PMS reagent have been subtracted from the results. Bars: means±SD (n=3). *Not different from control; †P<0.005, ‡P<0.025 and ‡P<0.05, smaller than control.

FIG. 5. Comparison of the concentration-dependent inhibitions of HL-60-S ( ● ), HL-60-RV ( □ ) and HL-60-R8 ( ▲ ) cell proliferation by DAU at days 2 (A) and 4 (B) in vitro. Cell growth results are expressed as % of the numbers of vehicle-treated control tumor cells (100±5%, striped areas) after 2 (110,903±5,878 cells/ml) and 4 days (253,462±1,166 cells/ml) in culture. Bars: means±SD (n=3). *Not different from control; †P<0.05, ‡P<0.01 and ‡P<0.025, smaller than control.

FIG. 6. Comparison of the concentration-dependent inhibitions of HL-60-S ( ● ), HL-60-RV ( □ ) and HL-60-R8 ( ▲ ) cell proliferation by TT2 at days 2 (A) and 4 (B) in vitro. The conditions of the experiments and the determination of the results were identical to those of FIG. 5. Bars: means±SD (n=3). *Not different from control; †P<0.05, ‡P<0.025 and ‡P<0.005, smaller than control.

FIG. 7. Comparison of the concentration-dependent inhibitions of HL-60-S ( ● ), HL-60-RV ( □ ) and HL-60-R8 ( ▲ ) cell viability by DAU at days 2 (A) and 4 (B) in vitro. Cell viability results are expressed as % of the net absorbance of MTS/formazan after bioreduction by vehicle-treated control cells (100±6%, striped areas) at days 2 (A950 nm=1.635±0.105) and 4 (A950 nm=1.428±0.086). Blank values (A950 nm=0.412 and 0.436 at days 2 and 4) for cell-free culture medium supplemented with MTS: PMS reagent have been subtracted from the results. Bars: means±SD (n=3). *Not different from control; †P<0.05, ‡P<0.025 and ‡P<0.01, smaller than control.

FIG. 8. Comparison of the concentration-dependent inhibitions of HL-60-S ( ● ), HL-60-RV ( □ ) and HL-60-R8 ( ▲ ) cell viability by TT2 at days 2 (A) and 4 (B) in vitro. The conditions of the experiments and the determination of the results were identical to those of FIG. 5. Bars: means±SD (n=3). *Not different from control; †P<0.05, ‡P<0.025 and ‡P<0.01, smaller than control.

FIG. 9. Comparison of the concentration-dependent inhibitions of DNA synthesis by the TT analog TT2 ( ● ) and DAU ( □ ) at days 2 (A) and 4 (B) in vitro. Cells (1.14±10^4/0.5 ml of RPMI 1640 medium) were incubated at 37°C for 90 min in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale, and then pulse-labeled for an additional 30 min to determine the rate of 3H-thymidine incorporation into DNA. DNA synthesis in vehicle-treated control cells was 27,860±1,931 cpm (100±7%; striped area). The blank value (916±52 cpm) for cells incubated and pulse-labeled at 2°C with 1 μCi of 3H-thymidine has been subtracted from the results. Bars: means±SD (n=3). *Not different from control; †P<0.05 and ‡P<0.01, smaller than control.

FIG. 10. Concentration-dependent inhibition of nucleoside transport by the TT analog TT2 in L1210 cells in vitro. Cells (1.48±10^4/0.5 ml of RPMI 1640 medium) were preincubated for 15 min at 37°C in the presence or absence (control) of the indicated concentrations of TT2 ( ● ) and DAU ( □ ), which are plotted on a logarithmic scale, before being exposed to 1 μCi of 3H-thymidine for 30 sec at 37°C. After washing thrice with PBS, intact cell pellets were harvested by centrifugation, incubated for 30 min in 1 ml of HLB containing 0.2% Triton X-100, and these lysates were mixed with scintillation cocktail and counted to estimate the cellular uptake of 3H-thymidine. Results are expressed as % of 3H-thymidine transported into vehicle-treated control cells over 30 sec (15,031±872 cpm; 100±%; striped area). Bars: means±SD (n=3). *Not different from control; †P<0.005, smaller than control.

FIG. 11. Comparison of the inhibitory effects of novel TT analogs on DNA synthesis (open) and nucleoside transport (striped) in L1210 cells in vitro. The cellular uptake of 3H-thymidine (striped) and the rate of 3H-thymidine incorporation into DNA (open) were determined in cells (1.38±10^4/0.5 ml of RPMI 1640 medium) respectively preincubated at 37°C for 15 or 90 min in the presence or absence (control) of 7 μM concentrations of the indicated compounds. For the cellular transport of nucleosides, preincubated cells were then exposed to 1 μCi of 3H-thymidine for 30 sec at 37°C. Results are expressed as % of 3H-thymidine transported into vehicle-treated control cells over 30 sec (15,539±1,767 cpm; 100±%; striped control). For DNA synthesis, preincubated cells were then pulse-labeled with 1 μCi of 3H-thymidine for an additional 30 min at 37°C. Results are expressed as % of 3H-thymidine incorporation into DNA in vehicle-treated control cells over 30 min (28,505±2,594 cpm; 100±%; open control). The blank value (1,075±171 cpm) for cells incubated and pulse-labeled at 2°C with 1 μCi of 3H-thymidine has been subtracted from the results. Bars: means±SD (n=3). *Not different from TT3, TT7, TT9 and TT11; †P<0.01, smaller than TT1 but not different from TT13; ‡P<0.025, greater than TT1; ‡P<0.05, smaller than control.

FIG. 12. Irreversibility of the inhibitory effects of the novel TT analogs TT2 on DNA synthesis (open) and nucleoside transport (striped) in L1210 cells in vitro. The reversible inhibitory effects of the tricyclic pyrene H10 are demonstrated in the same experiment. The cellular uptake of 3H-thymidine (striped) and the rate of 3H-thymidine incorporation into DNA (open) were determined in cells (1.64±10^4/0.5 ml of RPMI 1640 medium) respectively preincubated at 37°C for 15 or 60 min in the presence or absence (control) of 10 and 25 μM TT2 or 25 μM H10. The protocol of the experiments and the determination of the results were identical to those of FIGS. 9 and 10, except that, after preincubation, either the drugs were maintained in the
culture medium (+) or the cells were spun, washed, and resuspended in fresh medium in order to remove the drugs (-). After preincubation, vehicle-treated controls were similarly spun and washed. For the cellular transport of nucleosides, preincubated cells were then exposed, in the presence (+) or absence (-) of TT2 or H10, to 1 μCi of 3H-thymidine for 30 sec at 37°C. Results are expressed as % of 3H-thymidine transported into vehicle-treated control cells over 30 sec (11,054±754 cpm; 100±7%; 2 spike control; 14,486±1,260 cpm; 100±5%; 3 spike control). For DNA synthesis, preincubated cells were then pulse-labeled with 1 μCi of 3H-thymidine for an additional 30 min at 37°C in the presence (+) or absence (-) of TT2 or H10. Results are expressed as % of 3H-thymidine incorporation into DNA in vehicle-treated control cells over 30 min (34,052±1,566 cpm; 100±5%; open control; 28,811±2,506 cpm; 100±9%; open control). The blank value (1,328±86 cpm) for cells incubated and pulse-labeled at 2°C. with 1 μCi of 3H-thymidine has been subtracted from the results. Bars: means±SD (n=3). Values with similar superscripts are not significantly different from each other.

**FIG. 13.** The open columns represent the rate of 3H-thymidine incorporation into DNA over 30 minutes, whereas the striped and closed columns represent the cellular transport of 3H-adenosine and 3H-thymidine occurring over only 30 seconds in vitro.

**FIG. 14.** Comparison of the concentration-dependent inhibitions of DNA synthesis by DAU (open symbols in A) and TT2 (closed symbols in B) in HL-60-S (circles), HL-60-RV (squares) and HL-60-R8 (triangles) in vitro. DNA synthesis in vehicle-treated control cells was 14,269±1,106 cpm (100±5%, striped areas). The blank value (404±38 cpm) for control cells incubated and pulse-labeled at 2°C. with 1 μCi of 3H-thymidine has been subtracted from the results. Bars: means±SD (n=3). "Not different from control; P>0.05 and P<0.025, smaller than control.

**FIG. 15.** Comparison of the concentration-dependent inhibitions of the cellular transport of purine (A) and pyrimidine (B) nucleosides by DAU (open symbols) and TT2 (closed symbols) in HL-60-S (circles), HL-60-RV (squares) and HL-60-R8 (triangles) cells in vitro. Results are expressed as % of 3H-adenosine (26,032±2,239 cpm, 100±9%, striped area in A) and 3H-thymidine (17,934±1,396 cpm; 100±5%, striped area in B) transported into vehicle-treated control cells over 30 s. Bars: means±SD (n=3). "Not different from control; P>0.025, P<0.01 and P<0.005, smaller than control.

**FIG. 16.** Comparison of the concentration-dependent inhibitions of RNA synthesis by the TT analog (●) and DAU (-) in L1210 cells in vitro. Cells (1.2±10^6/0.5 ml of RPMI 1640 medium) were incubated at 37°C for 3 h in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale, and then pulse-labeled for an additional 1 h to determine the rate of 3H-uridine incorporation into RNA. RNA synthesis in vehicle-treated control cells was 47,117±2,304 cpm (100±7%; striped area). The blank value (2,436±298 cpm) for cells incubated and pulse-labeled at 2°C. with 2.5 μCi of 3H-uridine has been subtracted from the results. Bars: means±SD (n=3). "Not different from control; P>0.05 and P<0.025, smaller than control.

**FIG. 17.** Comparison of the concentration-dependent inhibitions of protein synthesis by the TT analog (●) and DAU (-) in L1210 cells in vitro. Cells (1.5×10^6/0.5 ml of RPMI 1640 medium) were incubated at 37°C for 3 h in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale, and then pulse-labeled for an additional 1 h to determine the rate of 3H-leucine incorporation into protein. Protein synthesis in vehicle-treated control cells was 13,812±1,070 cpm (100±5%; striped area). The blank value (2,436±298 cpm) for cells incubated and pulse-labeled at 2°C. with 2.5 μCi of 3H-leucine has been subtracted from the results. Bars: means±SD (n=3). "Not different from control; P>0.025 and P<0.01, smaller than control.

**FIG. 18.** Effects of the novel TT analog TT2 and DAU on the mitotic index of L1210 cells in vitro. The antimitotic activities of VCR and the tricyclic pyrone H10 are demonstrated in the same experiment. Cells (10^6/0.5 ml of RPMI 1640 medium) were incubated in triplicate for 24 h at 37°C in the presence or absence (control) of 0.256, 0.64, 1.6 and 4 μM TT2, 0.256, 0.64 and 1.6 μM DAU or 1 μM VCR or 1 μM H10. After fixation with MeOH:acetic acid (3:1) and staining with 0.1% crystal violet, about 500 cells/slide were scored for mitotic figures and the mitotic index was expressed as the % of mitotic cells in drug-treated cultures divided by the % of mitotic cells in non-treated controls. The mean % of mitotic cells in control at 24 h was 1.86±0.43%. Bars: means±SD (n=3). "Not different from control; P<0.05, P<0.005 and P<0.005, smaller than control; P<0.005, greater than control but smaller than VCR.

**FIG. 19.** Comparison of the concentration-dependent effects of novel TT analogs and known DNA-damaging anticancer drugs on DNA cleavage in L1210 cells in vitro. Cells (10^6/0.5 ml of RPMI 1640 medium) were prelabeled with 1 μCi of 3H-thymidine for 2 h, washed and resuspended in fresh FCS-containing medium, and incubated at 37°C. For 24 h in the presence or absence (control) of the indicated concentrations of TT1 (-), TT2 (●), DAU (●) or CPT (●), which are plotted on a logarithmic scale. After lysing the cells in HLB containing 0.2% Triton X-100, the detergent-soluble DNA fragments present in the supernatants and the intact chromatin DNA remaining in the pellets were separated by centrifugation and their radioactivity estimated by LSC. Results are expressed as [cpm in supernatant/cpm in supernatant+pellet]×100 at 24 h. For untreated controls (5.3±0.6% DNA fragmentation; striped area), the fragment is 74±1±79 cpm and the pellet is 13,293±1,446 cpm. Bars: means±SD (n=3). "Not different from control; P<0.05 and P<0.01, greater than control; P>0.05, smaller than CPT but not different from 10 μM TT1; P<0.05, greater than TT1 but not different from CPT.

**FIG. 20.** Comparison of the abilities of novel TT analogs to induce DNA cleavage in L1210 cells in vitro. Cells containing prelabeled DNA were incubated at 37°C for 24 h in the presence or absence (control) of 7 μM concentrations of the indicated compounds. The protocol of the experiment and the determination of the results were identical to those of FIG. 12. Bars: means±SD (n=3). "Not different from TT7 and TT9; P>0.05, greater than TT9 but not different from TT13; P<0.05, smaller than TT7; not different from control; P<0.01, greater than control, P<0.025, greater than TT1 but not different from TT3, TT6, TT8 and TT10; P<0.01, greater than control.

**FIG. 21.** Same experiment as FIG. 20 with additional analogs.

**FIG. 22.** Irreversibility of the effects of novel TT analogs and known DNA-damaging anticancer drugs on DNA cleavage in L1210 cells in vitro. Cells containing prelabeled DNA were incubated at 37°C for various periods of time in the presence or absence (control) of 10 μM TT1 or TT2 and 1.6 μM DAU or CPT. The protocol of the experiment and the
Butyl, n-pentyl, neopentyl, 2-methylbutyl, 1-methylbutyl, aromatic rings and combinations of such groups. The term includes, but is not limited to, methyl, ethyl, n-propyl, isopropyl, n-butyl, sec-butyl, isobutyl, tert-butyl, n-pentyl, neopentyl, 2-methylbutyl, 1-methylbutyl, 1-ethylpropyl, 1,1-dimethylpropyl, n-hexyl, 1-methylhexyl, 2-methylhexyl, 3-methylpentyl, 2-ethylpentyl, 1-ethylpentyl, 4,4-dimethylpentyl, 3,3-dimethylpentyl, 1,1-dimethylpentyl, n-octyl, 6-methylheptyl, 5-methylheptyl, 4-methylheptyl, 3-methylheptyl, 2-methylheptyl, 1-methylheptyl, 1-ethylhexyl, 1-propylpentyl, 3-ethylhexyl, 5,5-dimethylhexyl, 4,4-dimethylhexyl, 2,2-dimethylbutyl, 3,3-diethylbutyl, and 1-methyl-1-propylbutyl. Alkyl groups are optionally substituted. Lower alkyl groups are CH₃, C₂H₅, and cyclopropyl groups.

The term “cycloalkyl” refers to alkyl groups having a hydrocarbon ring, particularly to those having rings of 3 to 7 carbon atoms. Cycloalkyl groups include those with alkyl substitution on the ring. Cycloalkyl groups can include straight-chain and branched-chain portions. Cycloalkyl groups include but are not limited to cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, and cyclononyl. Cycloalkyl groups can optionally be substituted.

Aryl groups may be substituted with one, two or more simple substituents including, but not limited to, lower alkyl, e.g., methyl, ethyl, butyl; halo, e.g., chloro, bromo; nitro; sulfato; sulfonyloxy; carboxy; carbo-lower-alkoxy, e.g., carbomethoxy, carboxethoxy; amino; mono- and di-lower-alkylamino, e.g., methylamino, ethylamino, dimethylamino, methylethylamino; amidino; hydroxy; lower-alkoxy, e.g., methoxy; ethoxy; and lower-alkanoyloxy, e.g., acetoxy.

The term “unsaturated alkyl” group is used herein generally to include alkyl groups in which one or more carbon single bonds have been converted to carbon-carbon double or triple bonds. The term includes alkyl and arylalkyl groups in their most general sense. The term is intended to include groups having more than one double or triple bond, or combinations of double and triple bonds. Unsaturated alkyl groups include, without limitation, unsaturated straight-chain, branched or cycloalkyl groups. Unsaturated alkyl groups include without limitation: vinyl, allyl, propenyl, isopropenyl, butenyl, pentenyl, hexenyl, hexadienyl, heptenyl, cyclopropenyl, cyclobutenyl, cyclopentenyl, cyclopentadienyl, cyclohexenyl, cyclohexadienyl, 1-propenyl, 2-butenyl, 2-methyl-2-butenyl, ethynyl, propargyl, 3-methyl-1-pentynyl, and 2-heptyn. Unsaturated alkyl groups can optionally be substituted.

Substitution of alkyl, cycloalkyl and unsaturated alkyl groups includes substitution at one or more carbons in the group by moieties containing heteroatoms. Suitable substituents for these groups include but are not limited to OH, SH, NH₂, COH, CO₂H, OR, SR, P=O, NR, CONR, halogens, particularly chlorines and bromines where R is independently, are alkyl, unsaturated alkyl or aryl groups. Preferred alkyl and unsaturated alkyl groups are the lower alkyl, alkylalkyl or alkynyl groups having from 1 to about 3 carbon atoms.

The term “aryl” is used herein generally to refer to aromatic groups which have at least one ring having a conjugated pi electron system and includes without limitation carbocyclic aryl, aralkyl, heterocyclic aryl, biaryl groups and heterocyclic biaryl, all of which can be optionally substituted. Preferred aryl groups have one or two aromatic rings.

“Carbocyclic aryl” refers to aryl groups in which the aromatic ring atoms are all carbons and includes without limitation phenyl, biphenyl and naphthalene groups.
known as mercaptide groups, the sulfur analog of alkoxy -NH--, -CH=CH- and -(CH,)n, where R is an urated alkyl or aryl groups. Preferred substituents are OH, tert-butoxy, n-pentyloxy, neopentyloxy, 2-methylbutyloxy, 1-methylbutoxy, 1-ethyl propano, 1,1-dimethylpropoxy, n-hexyloxy, 1-methylpentyloxy, 2-methylpentyloxy, 3-methylpentyloxy, 4-methylpentyloxy, 3,3-dimethylbutoxy, 2,2-dimethoxybutoxy, 1,1-dimethylbutyloxy, 2-ethylbutoxy, 1-ethylbutoxy, 1,3-dimethylbutoxy, n-pentyloxy, 5-methylhexyloxy, 4-methylhexyloxy, 3,3-dimethylhexyloxy, 2,2-dimethyhexyloxy, 1,1-dimethylhexyloxy, n-octyloxy, 6-methyloctyloxy, 5-methylheptyloxy, 4-methylheptyloxy, 3,3-methylheptyloxy, 2,2-dimethyheptyloxy, 1,1-dimethylheptyloxy, 1-ethylhexyloxy, 1-propyloxy, 3-ethylhexyloxy, 5,5-dimethylhexyloxy, 4,4-dimethylhexyloxy, 2,2-diethylbutoxy, 3,3-diethylbutoxy, 1-methyl-1-propyloxy, ethoxymethyl, n-propoxymethyl, isoproxymethyl, sec-butoxyethyl, isobutoxyethyl, (1-ethyl propoxy)methyl, (2-ethoxybutoxy)methyl, (1-ethylbutoxy)methyl, (2-ethylpentayoxy)methyl, (3-ethylpentayoxy)methyl, 2-methoxyethyl, 1-methoxyethyl, 2-ethoxyethyl, 3-methoxypropyl, 2-methoxpropyl, 1-methoxpropyl, 2-ethoxypropyl, 3-(n-propoxy)propyl, 4-methoxybutyl, 2-methoxybutyl, 4-ethoxybutyl, 2-ethoxybutyl, 5-ethoxypentyl, and 6-ethoxyhexyl. Thiokalxy groups include but are not limited to the sulfur analogs of the alkyl groups specifically listed supra.

“Optional” or “optionally” means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances in which it does not. For example, “optionally substituted phenyl” means that the phenyl radical may or may not be substituted and that the description includes both unsubstituted phenyl radicals and phenyl radicals wherein there is substitution.

“Amino acids” as used herein include naturally occurring and commercially available amino acids and optical isomers thereof. Typical natural and commercially available amino acids are glycine, alanine, serine, homoserine, threonine, valine, norvaline, leucine, isoleucine, norleucine, aspartic acid, glutamic acid, lysine, ornithine, histidine, arginine, cysteine, homocysteine, methionine, phenylalanine, homophenylalanine, phenylglycine, o-, m-, and p-tyroine, tryptophan, glutamine, asparagine, proline and hydroxyproline. “Amino acid” as used herein includes amino acid residues and amino acid side chains. An “amino acid residue” is an amino acid radical —NHCH(R)C(O)O(—), wherein R is an amino acid side chain, except for the amino acid residues of proline and hydroxyproline which are —N(CH2 —CH2 —CH2)C(O)O(—), and —N(CH2 —CHOCH2)C(O)O(—), respectively. An amino acid side chain is a radical found on the α-carbon of an α-amino acid, wherein the groups is either hydrogen (side chain of glycine), methyl (side chain of alanine), or is a radical bonded to the α-carbon by a methylene (—CH2—), or phenyl group.

“Maintaining reaction components in contact” means keeping the components together in such a way that they can react with each other. Preferably, the reaction components are suspended or dissolved in a carrier fluid which is a liquid medium. “Maintaining reaction components in contact” means keeping the components together in such a way that they can react with each other.
groups attached, and either an aldehyde or ketone group. Cyclc saccharides are saccharides that are in a ring form. Disaccharides are compounds wherein two monosaccharide groups are linked. Polysaccharides are compounds wherein more than two monosaccharide groups are linked. Specific examples of saccharides useful in this invention include glucose, ribose and glucosamine, among others.

Substituents which impart water solubility include but are not limited to alcohols; polyalcohols; straight chain or cyclic saccharides; amines and polyamines; sulfate groups; phosphate groups; ascorbate groups; alkyl chains optionally substituted with —CH3 at any position; glycols, including polyethylene glycols, and polyethers. Substituents which impart water solubility are also referred to as solubilizing groups.

This invention is also directed to pharmaceutically acceptable salts of the various formulæs and structures disclosed herein. Acid addition salts are prepared by contacting compounds having appropriate basic groups therein with an acid whose anion is generally considered suitable for human or animal consumption. Pharmacologically acceptable acid addition salts include but are not limited to the hydrochloride, hydrobromide, hydroiodide, sulfate, phosphate, acetate, propionate, lactate, maleate, malate, succinate, and tartrate salts. All of these salts can be prepared by conventional means by reacting, for example, the selected acid with the selected basic compound. Base addition salts are analogously prepared by contacting compounds having appropriate acidic groups therein with a base whose cation is generally considered to be suitable for human or animal consumption. Pharmacologically acceptable base addition salts, include but are not limited to ammonium, amine and amide salts.

Pharmaceutically acceptable esters of compounds of this invention are prepared by conventional methods, for example by reaction with selected acids. Pharmaceutically acceptable esters include but are not limited to carboxylic acid esters RCOO—D (where D is a caticonic form of a compound of this invention and where R is H, alkyl or aryl groups).

Effective dosages of the compounds of this invention may be easily determined by those skilled in the art following the teachings hereof and principles known to the art.

The compounds of these inventions may be administered in the form of pharmaceutical preparations including the compounds of these inventions in suitable pharmaceutical carriers to form solutions, lotions, creams, and other dosage forms known to the art. Combinations of such compounds with pharmaceutical carriers are also provided by this invention. Combinations of triptycene analogs described herein along with other compounds which may include other triptycene analogs described herein are also provided by this invention.

This invention is also directed to prodrugs and analogs which on being metabolized will result in any of the effective triptycene analogs of this invention. For example, alkoxy or acetate groups can be metabolized to hydroxins. Labile substituents may be protected employing conventional and pharmaceutically acceptable protecting groups removable on metabolism. Pharmaceutically active compounds may be derivatized by conventional methods to provide for extended metabolic half-life, to enhance solubility in a given carrier, to provide for or facilitate slow-release or timed-release or enhance or affect other drug delivery properties.

Pharmaceutical compositions according to the present invention comprise one or more triptycene compounds, salts or esters of this invention in association with a pharmaceutically acceptable carrier or excipient adapted for use in human or veterinary medicine. Such compositions may be prepared for use in conventional manner in admixture with one or more physiologically acceptable carriers or excipients. The compositions may optionally further contain one or more other therapeutic agents. The compounds, salts or esters of this invention are present in these pharmaceutical compositions in an amount or in a combined amount sufficient to elicit a measurable positive effect on a desired parameter, or a desired physiological effect. The triptycene compounds, salts and esters of this invention may be formulated for oral, buccal, parenteral, topical or rectal administration. In particular, they may be presented in unit dose forms. The compositions may take such forms as suspensions, solutions, or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, e.g. sterile, pyrogen-free water, before use.

The pharmaceutical compositions according to the invention may also contain other active ingredients, such as antimicrobial agents, or preservatives.

The invention further provides a process for preparing a pharmaceutical composition which comprises bringing a triptycene analog of the invention into association with a pharmaceutically acceptable excipient or carrier. The carrier or excipient being selected as is known in the art for compatibility with the desired means of administration, for compatibility with the selected compounds and to minimize detrimental effects to the patient.

The magnitude of a prophylactic or therapeutic dose of a particular compound will, of course, vary with the nature of the severity of the condition to be treated, the particular triptycene analog and its route of administration. It will also vary according to the age, weight and response of the individual patient, all as will be readily ascertainable to those skilled in the art.

The compounds of the present invention are preferably formulated prior to administration. The present pharmaceutical formulations are prepared by known procedures using well-known and readily available ingredients. In making the compositions of the present invention, the active ingredient will usually be mixed with a carrier, or diluted by a carrier, or enclosed within a carrier which may be in the form of a capsule, sachet, paper or other container. When the carrier serves as a diluent, it may be a solid, semi-solid or liquid material which acts as a vehicle, excipient or medium for the active ingredient. The compositions can be in the form of tablets, pills, powders, lozenges, sachets, cachets, elixirs, suspensions, emulsions, solutions, syrups, aerosols (as a solid or in a liquid medium), ointments containing for example up to 10% by weight of the active compound, soft and hard gelatin capsules, suppositories, sterile injectable solutions and sterile packaged powders.

Some examples of suitable carriers, excipients, and diluents include lactose, dextrose, sucrose, sorbitol, mannitol, starches, gum acacia, calcium phosphate, alginites, tragacanth, gelatin, calcium silicate, microcrystalline cellulose, polyvinylpyrrolidone, cellulose, water, syrup, methyl cellulose, methyl and propylhydroxybenzoates, talc, magnesium stearate and mineral oil. The formulations can additionally include lubricating agents, wetting agents, emulsifying and suspending agents, preserving agents, sweetening agents or flavoring agents. The compositions of
the invention may be formulated so as to provide quick, sustained or delayed release of the active ingredient after administration to the patient by employing procedures well known in the art.

The term “unit dosage form” refers to physically discrete units suitable as unitary dosages for human subjects and other mammals, each unit containing a predetermined quantity of active material calculated to produce the desired therapeutic effect, in association with a suitable pharmaceutical carrier.

This invention is further directed to therapeutic methods employing the triptycene compounds of this invention and pharmaceutical compositions containing them in the treatment of disorders or physiological conditions involving abnormal cell growth or activity. These methods comprise a step of administering to a patient having the disorder or symptoms thereof a pharmaceutical composition comprising one or a mixture of the compounds, salts or esters of this invention where the compounds, or mixtures of compounds of this invention are present in the composition at a level or a combined level sufficient to effect a positive biological response. The present invention provides triptycene analogs that can be used in place of or in combination with currently known pharmaceuticals active against disorders such as cancer and malaria. Compounds of this invention exhibit improved properties (enhanced activity and/or decreased undesired side-effects) for treatment of such disorders as compared to previously known compounds useful for such treatments.

Table 1 lists the chemical names of the compounds depicted in Scheme 1.

### Table 1

<table>
<thead>
<tr>
<th>Nomenclature of various substituted 9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</th>
<th>benzenoanthracene-1,4,5,8-tetraene</th>
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<tr>
<td>TTT1: 9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraene</td>
<td>TTT2: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraene</td>
</tr>
<tr>
<td>TTT3: 2-Bromo-3-methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraene</td>
<td>TTT4: 2-Bromom-(methylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraene</td>
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<td>TTT5: 2,5,8-Trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</td>
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<td>TTT8: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</td>
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<td>TTT12: 2,5,8-Trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</td>
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<tr>
<td>TTT17: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</td>
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<td>TTT19: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</td>
<td>TTT20: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione</td>
</tr>
</tbody>
</table>

**EXAMPLES**

The following non-limiting examples are intended to aid in understanding the invention.

In contrast to their inactive parent compound triptycene (code name TTO), several triptycene (TT) analogs were shown to prevent L1210 leukemic cells from synthesizing macromolecules and growing in vitro. TT2 blocks the cellular transport of both purine and pyrimidine nucleosides (IC50: 6 µM) over a 30-sec period in vitro, in contrast to the quinone antitumor drug daunomycin. The concentration-dependent induction of DNA cleavage at 24 h and internumerical DNA fragmentation at 48-72 h by TT2 peaks at 4-10 µM but disappears at 25 µM. TT2 induction of DNA cleavage is inhibited by actinomycin D, cycloheximide and the caspase inhibitor 2-VAD-fmk, suggesting that TT2 activates endonucleases and triggers apoptosis.

The antitumor activity of TT2 has been compared to that of daunomycin (DAU), a clinically valuable anthracycline antibiotic which is structurally different from TT2 but also contains a quinone moiety. TT2 inhibits the proliferation (IC50: 300 nM at day 2 and 150 nM at day 4) and viability (IC50: 250 nM at day 2 and 100 nM at day 4) of L1210 cells to the same maximal degree than DAU, suggesting that the cytostatic and cytotoxic activities of TT2 are a combination of drug concentration and duration of drug exposure. Since TT2 does not increase the mitotic index of L1210 cells at 24 h like vincristine, it is unlikely to be an antimitotic drug that disrupts microtubule dynamics. Like DAU, a 1.5- to 3-h pretreatment with TT2 is sufficient to inhibit the rates of DNA, RNA and protein syntheses determined over 30- to 60-min periods of pulse-labeling in L1210 cells in vitro (IC50: 6 µM). In contrast to DAU, which is inactive, a 15-min pretreatment with TT2 has the advantage of also blocking the cellular transport of nucleosides occurring over a 30-sec period in vitro (IC50: 6 µM), suggesting that TT2 prevents the incorporation of 3H-thymidine into DNA because it rapidly blocks the uptake of [H]-thymidine by the tumor cells. After 24 h, TT2 induces as much DNA cleavage as camptothecin and DAU, two anticancer drugs producing DNA-strand breaks and known to respectively inhibit DNA topoisomerase I and II activities. Interestingly, the abilities of TT2 to block nucleoside transport, inhibit DNA synthesis and induce DNA fragmentation are irreversible upon drug removal, suggesting that this compound may rapidly interact with various molecular targets in cell membranes and nuclei to disrupt the function of the nucleoside transporters and nucleic acids and trigger long-lasting antitumor effects which persist after cessation of drug treatment. Because inhibition of nucleoside transport is highly unusual among DNA-damaging drugs, the use of bifunctional TBs with antileukemic activity in the nM range in vitro provides a considerable advantage in polychemotherapy to potentiate the action of antimitabolics and sensitize multidrug-resistant tumor cells.

**Materials and Methods**

**Cell Culture and Drug Treatments**

TT0 was purchased from Aldrich (Milwaukee, Wis.). All solutions of synthetic TBs and tricyclic pyrone H10, vincristine (VCR; a gift from Lilly Research Laboratories, Indianapolis, Ind.) and camptothecin (CPT; from Sigma Chemical Co., St Louis, Mo.) were dissolved and diluted in dimethyl sulfoxide (DMSO), whereas daunomycin (DAU) (from Sigma) solutions were prepared in 0.1 M potassium phosphate buffer, pH 7.4, containing 0.9% NaCl. Suspension cultures of murine L1210 lymphocytic leukemia cells (American Type Culture Collection, Rockville, Md.) were maintained in continuous exponential growth by twice-a-week passage in RPMI 1640 medium supplemented with 7.5% fortified bovine calf serum (FCS; HyClone Laboratories, Logan, Utah) and penicillin (100 IU/ml)-
streptomycin (100 μg/ml), and incubated in the presence or absence of drugs at 37°C in a humidified atmosphere containing 5% CO2. Since drugs were supplemented to the culture medium in 1 μl aliquots, the concentration of DMSO in the final incubation volume (0.5 ml) never exceeded 0.2% and did not affect the rates of macromolecule syntheses and growth in L1210 cells. Control cells incubated in the absence of drugs were similarly treated with vehicle only and, in every experiment, all incubates received the same volume of solvent. For drug removal, incubates were spun at 200xg for 10 min, drug-containing supernatants were discarded, and intact cells were washed thrice with 1 ml of the above RPMI 1640 culture medium and resuspended in 0.5 ml of fresh medium for further incubation.

Suspension cultures of WT, drug-sensitive, human HL-60-S promyelocytic leukemia cells, were obtained from American Type Culture Collection (Manassas, Va.), maintained in continuous exponential growth by twice-a-week passage in RPMI 1640 medium supplemented with 8.25% fortified bovine calf serum (ICS, Hyclone Laboratories, Logan, Utah) and penicillin (100 IU/ml)-streptomycin (100 μg/ml), and incubated in the presence or absence of drugs at 37°C in a humidified atmosphere containing 5% CO2. The MDR HL-60-RV and HL-60-R8 cells were developed in Melvin S. Center’s laboratory (Kansas State University) and similarly maintained in RPMI 1640 medium in the absence of drugs (Marsh W, et al, Cancer Res 46: 4053-4057, 1986; McGrath T et al, and Center M S: Biochem Biophys Res Commun 145: 1171-1176, 1987; Marsh W and Center M S: Cancer Res 47: 5080-5086, 1987; McGrath T, et al, Biochem Pharmacol 38: 3611-3619, 1989; Marquardt D, et al, Cancer Res 50: 1426-1430, 1990; Marquardt D and Center M S: J Natl Cancer Inst 83: 1089-1109, 1991; Krishnamachary N and Center M S; Cancer Res 53: 3658-3661, 1993). Every 4 weeks, these HL-60-RV and HL-60-R8 sublines were exposed to 41 nM DAU for 48 h to stabilize their MDR phenotype and then were exposed to 41 nM DAU for 48 h to stabilize their MDR phenotype. This concentration of DAU, which is not cytotoxic to MDR HL-60 sublines, was removed from the culture medium at least 48 h before experimentation. Since drugs were supplemented to the culture medium in 1 μl aliquots, the concentration of vehicle in the final incubation volume (0.5 ml) did not affect basal activity levels in control tumor cells incubated in the absence of drugs. RFs were determined by dividing the IC50 of the MDR cells by that of the sensitive WT parent cell line.

Cell Proliferation Assay

For tumor cell growth, L1210 cells were resuspended in fresh FCS-containing RPMI 1640 medium, plated at an initial density of 1x10^4 cells/0.5 ml, and incubated in 48-well Costar cell culture plates (Costar, Cambridge, Mass.). Except when otherwise specified, cells were grown for 4 days in the presence or absence of drugs and their density was monitored every 24 h using a Coulter counter (Coulter Electronics, Luton Beds, UK).

HL-60-S, HL-60-RV and HL-60-R8 cells were resuspended in fresh FCS-containing RPMI 1640 medium, seeded in triplicate at an initial density of 1.5x10^4 cells/0.5 ml and incubated at 37°C in 48-well Costar cell culture plates (Costar, Cambridge, Mass.). Tumor cells were grown for 4 days in the presence or absence (control) of drugs and their density was monitored every 24 h using a Z1 dual threshold Coulter counter (Beckman Coulter, Miami, Fla.).

Decreasing concentrations of cells, such as 1x10^4 and 1.1x10^4 cells/0.5 ml/well, were initially plated at time 0 in order to collect control samples with approximately equal cell densities after 2 and 4 days in culture, respectively. The viability of TT-treated cells was assessed from their ability to bioreduce the 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulphophenyl)-2H-tetrazolium (MTS) reagent (Promega, Madison, Wis.) in the presence of phenazine methosulfate (PMS; Sigma) into a water-soluble formazan product which absorbs at 490 nm. At the appropriate time after drug treatment, cell samples (about 10^4/0.5 ml/well for controls) were further incubated at 37°C for 3 h in the dark in the presence of 0.1 ml of MTS-PMS (2:1) reagent and their relative cell viability was estimated by recording the Abs. at 490 nm, using a Cambridge model 750 automatic microplate reader (Packard, Downers Grove, Ill.). Blank values for culture medium supplemented with MTS-PMS reagent in the absence of cells were subtracted from the results.

Macromolecule Synthesis

For nucleic acid and protein syntheses, L1210 cells were resuspended in fresh FCS-containing RPMI 1640 medium at a density of about 1.1-1.6x10^6 cells/0.5 ml. To estimate the rate of DNA synthesis, HL-60-S, HL-60-RV and HL-60-R8 cells were resuspended in fresh FCS-containing RPMI 1640 medium at a density of 5x10^5 cells/0.5 ml. Except when otherwise specified, the cells were incubated at 37°C for 90 min in the presence or absence of drugs and then pulse-labeled for an additional 30 min with 1 μCi of [methyl-3H] thymidine (50 Ci/mmol; Amersham Corp., Arlington Heights, Ill.) to estimate the rate of DNA synthesis. For RNA and protein syntheses, cells were incubated at 37°C for 3 h in the presence or absence of drugs and then pulse-labeled for an additional 1 h with 2 μCi of (5,6-3H)uridine (46 Ci/mmol; ICN Biomedicals, Irvine, Calif.) or 2.5 μCi of [4,5-3H]leucine (100 Ci/mmol; Moravek Biochemicals, Brea, Calif.), respectively. The incubations were terminated by the addition of 0.5 ml of 10% trichloroacetic acid (TCA). After holding on ice for 15 min, the acid-insoluble material was recovered over Whatman GF/A glass microfibre filters and washed thrice with 2 ml of 5% TCA and twice with 2 ml of 100% EtOH. After drying the filters, the radioactivity was determined by liquid scintillation counting (LSC) in 10 ml of Bio-Safe NA (Research Products International Corp., Mount Prospect, Ill.).

Nucleoside Transport

L1210 cells (about 1.5x10^6 cells/0.5 ml) were preincubated for 15 min at 37°C in the presence or absence of TTs and then exposed to 1 μCi of [3H]thymidine for only 30 sec to assess the cellular uptake of nucleoside over such very short period of time. HL-60-S, HL-60-RV and HL-60-R8 cells (1.2x10^5 cells/0.5 ml) were preincubated for 15 min at 37°C in the presence or absence (control) of drugs and then exposed to 1 μCi of [2,8-3H]adenosine (30 Ci/mmol; American Radiolabeled Chemicals, St. Louis, Mo.) or [methyl-3H]thymidine (50 Ci/mmol; Amersham, Arlington Heights, Ill.) for only 30 s to, respectively, assess the cellular uptake of purine and pyrimidine nucleosides over such very short period of time. Reactions were diluted with 2 ml of ice-cold Ca^2+/Mg^2+-free Dulbecco’s phosphate buffered saline (PBS) and the unincorporated radiolabel was removed by centrifugation at 2000g for 10 min. After washing thrice with 2 ml of ice-cold PBS, intact cell pellets were harvested by centrifugation and incubated for 30 min in 1 ml of hypotonic lysis buffer (HLB) containing 10 mM Tris-HCl, pH 8.0, 1 mM EDTA and 0.2% Triton X-100. Cell lysates were mixed
with 9 ml of Bio-Safe II (Research Products International) and counted to estimate the cellular uptake of $^3$H-thymidine. Drug inhibition was expressed as % of $^3$H-thymidine or $^3$H-adenosine transported into vehicle-treated control cells over similar 30-sec period.

Mitotic Index

L1210 cells (about 1x10$^5$/0.5 ml of FCS-containing RPMI 1640 medium) were incubated in triplicate for 24 h at 37°C in the presence or absence of TTs or known antimitotic drugs, and collected by centrifugation at 200g for 10 min to determine their mitotic index. For hypotonic treatment, cells were resuspended in 1 ml of 75 mM KCl for 20 min at 4°C. After fixation in 1 ml of MeOH:acetic acid (3:1), the final cell pellets were collected by centrifugation, resuspended in 75 μl of MeOH:acetic acid (3:1), dispersed onto glass slides, air dried, and stained by spreading 40 μl of 0.1% crystal violet under a coverslip. The % of cells in mitosis was determined microscopically by counting 500 cells/slide. The mitotic index was calculated as the % of mitotic cells in drug-treated cultures divided by the % of mitotic cells in non-treated controls.

DNA Cleavage

Drug-induced DNA cleavage was determined by intact chromatin precipitation, using L1210, HL-60-S, HL-60-RV and HL-60-R8 cells which were prelabelled with 1 μCi of $^3$H-thymidine for 2 h at 37°C, washed with 3x1 ml of ice-cold PBS, collected by centrifugation, and resuspended in fresh FCS-containing RPMI 1640 medium at a density of about 1x10$^6$ cells/0.5 ml for L1210 and 0.5x10$^6$ cells/ml for HL cells. Except when otherwise specified, such cells containing prelabelled DNA were then exposed for 24 h to TTs and drugs known to induce DNA fragmentation. After centrifugation at 200g for 10 min to discard the drugs and wash the cells, the intact cell pellets were lysed for 30 min in 0.5 ml of HLB, centrifuged at 12,000g for 30 min to collect the supernatants, and resuspended in 0.5 ml of HLB. After another similar centrifugation, the radioactivities in the pooled supernatants (detergent-soluble low molecular weight DNA fragments) and the pellet (intact chromatin DNA) were determined by LSC. % DNA fragmentation= [cpm in supernatant/cpm in supernatant+pellet]x100. Before being counted in 6 ml of Bio-Safe NA, the intact pelleted chromatin was incubated for 2 h at 60°C in the presence of 0.6 ml of NCS tissue solubilizer (Amersham).

Results

Drugs

The TT quinones under study were synthesized in the laboratory by an in-situ oxidation of substituted dihydroxybenzences followed by [4+2]cycloaddition with 1,4-dimethoxyanthracene and then oxidation. This method of synthesis is reported below. The chemical structures of the TTs tested for their antileukemic activity in vitro are shown in Scheme 1 (serial numbers indicate the order of synthesis).

Inhibition of Tumor Cell Growth and Viability by TTs L1210

When tested at 256 nM over a 4-day period, several TTs inhibit the rate of tumor cell growth in relation with their ability to decrease tumor cell viability (FIG. 1). Although most of these drugs are much more active at higher concentrations, at this lower concentration of 256 nM, TT4 and TT7 inhibit the rate of tumor cell growth and viability (FIG. 1). The concentrations of TTs at which 50% inhibition is achieved for HL-60, L1210, and M1 cells are shown in Table 1. The IC$_{50}$ values are the concentrations of TTs at which 50% inhibition of cell growth and viability is achieved.

The concentration-dependent inhibitions of HL-60-S, HL-60-RV and HL-60-R8 cell proliferation by DAU are respectively characterized by IC$_{50}$ values of 14, 500 and 320 nM at day 2 (FIG. 5A and 9,275 and 225 nM at day 4 (FIG. 6B). Such negligible RFs (0.9-1.2 at day 2 and 1.0-1.1 at day 4) suggest that TT2 retains its antiproliferative and cytotoxic activities after 4 days are 41 and 16 nM, respectively (FIGS. 3 & 4). These relative potencies of TT2 and DAU can easily be compared using the full concentration-response curves of FIGS. 3 and 4, where the stripped areas at 100% represent the control levels of L1210 cell growth and viability after 2 and 4 days in culture. The magnitudes of the cytostatic and cytotoxic effects of both TT2 and DAU are clearly related to the combination of their increasing concentration and duration of action. For instance, 16 nM DAU and 41 nM TT2 are ineffective at day 2 but their antiproliferative (FIG. 3) and cytotoxic (FIG. 4) activities become apparent at day 4. Moreover, 41 nM DAU and 256 nM TT2 are moderately cytostatic (42 and 44% inhibition, respectively) and cytotoxic (32 and 49% inhibition, respectively) at 4 days but respectively decrease L1210 cell viability by 77 and 68% and L1210 cell proliferation by 25 and 150 nM at day 4, suggesting that DAU is about 6 times more cytostatic than TT2 in the L1210 tumor system in vitro (FIG. 3). Similarly, the inhibitions of tumor cell viability by DAU and TT2 are respectively characterized by IC$_{50}$ values of 70 and 250 nM at day 2 but 30 and 100 nM at day 4, suggesting that DAU is about 3 times more cytotoxic than TT2 in this leukemic system in vitro (FIG. 4).

The concentration-dependent inhibitions of HL-60-S, HL-60-RV and HL-60-R8 cell proliferation by DAU are respectively characterized by IC$_{50}$ values of 14, 500 and 320 nM at day 2 (FIG. 5A) and 9,275 and 225 nM at day 4 (FIG. 6B). Such RFs of 22.9-35.7 at day 2 and 25.0-30.6 at day 4 demonstrate that DAU is about 30 times less effective than TT2 in this leukemic system in vitro (FIG. 3). In contrast, the concentration-dependent inhibitions of HL-60-S, HL-60-RV and HL-60-R8 cell proliferation by TT2 are respectively characterized by similar IC$_{50}$ values of 400, 500 and 350 nM at day 2 (FIG. 6A) and 300, 260 and 290 nM at day 4 (FIG. 6B). These negligible RFs (0.9-1.3 at day 2 and 0.9-1.0 at day 4) suggest that TT2 retains its effectiveness as a cytostatic agent in MDR cells that have become 30 times less sensitive to the antiproliferative action of DAU (FIG. 6).

Since DAU decreases the viability of HL-60-S, HL-60-RV and HL-60-R8 cells with respective IC$_{50}$ values of 80, 1, 900 and 2,500 nM at day 2 (RFs: 23.8-31.3) and 15, 450 and 380 at day 4 (RFs: 25.3-30.0), this drug is about 28 times less potent as a cytostatic agent in MDR than in WT HL-60 cells (FIG. 7). In contrast, the TTs are more effective than DAU in HL-60-S (IC$_{50}$: 1,300 and 230 nM at days 2 and 4) as in HL-60-RV (IC$_{50}$: 1,100 and 340 nM at days 2 and 4) and HL-60-R8 (IC$_{50}$: 1,600 and 350 nM at days 2 and 4) cell lines (FIG. 8). Such negligible RFs (0.9-1.2 at day 2 and 1.5 at day 4) demonstrate that the cytotoxic potency of TT2 persists in MDR tumor sublines that have become 28 times less sensitive to the cytotoxic activity of DAU (FIG. 8).
Inhibition of DNA Synthesis and Nucleoside Transport by TTs
L1210

A 2-h treatment with TT2 is sufficient to inhibit, in a concentration-dependent manner, the rate of DNA synthesis determined over a 30-min period of pulse-labeling in L1210 cells in vitro (FIG. 9). DNA synthesis is totally inhibited by 25 μM TT2 but, as compared to DAU which becomes effective against DNA synthesis at 0.256 μM, concentrations greater than 1.6 μM must be used to demonstrate the inhibitory effect of TT2 on DNA synthesis. Hence, the concentration-dependent inhibitions of DNA synthesis by DAU (IC50: 1 μM) and TT2 (IC50: 6 μM) suggest that, under these experimental conditions, DAU prevents L1210 cells from synthesizing DNA about 6 times more effectively than TT2 (FIG. 9).

A critical finding is that, in contrast to DAU which serves as a negative control in the assay, a 15-min treatment with TT2 is sufficient to block, in a concentration-dependent manner, the cellular transport of 3H-thymidine occurring over only 30 sec in vitro (FIG. 10). Nucleoside transport is totally inhibited in L1210 cells treated with 25 μM TT2. But 10 μM DAU is totally unable to alter the cellular transport of 3H-thymidine (FIG. 10), even though such concentration of DAU inhibits maximally the incorporation of 3H-thymidine into DNA (FIG. 9). Interestingly, the concentration-response curves for the inhibitory effects of TT2 on nucleoside transport (FIG. 10) and DNA synthesis (FIG. 9) are nearly identical and share similar IC50 values of 6 μM, suggesting that the inhibition of 3H-thymidine incorporation into DNA caused by TT2 at 2 h (FIG. 9) may largely be due to the ability of this TT quinone to immediately block the uptake of 3H-thymidine by the cells (FIG. 10).

Moreover, when compared on an equal 7 μM concentration basis, the different magnitudes at which various TTs inhibit DNA synthesis at 2 h correlate with the different abilities of these compounds to block the cellular transport of nucleosides after 15 min (FIG. 11). Indeed, among all TTs tested at this concentration, TT4 and TT12 alter neither DNA synthesis nor nucleoside transport, whereas TT2 and TT13, which block the most the cellular transport of 3H-thymidine (72–75% inhibition), are also the most potent against the incorporation of 3H-thymidine into DNA (69–71% inhibition) (FIG. 11). Under similar conditions, TT1, TT3, TT7, TT9 and TT11 inhibit nucleoside transport by 44–57% and DNA synthesis by 41–58%, whereas TT5, TT6, TT8 and TT10 are less effective, inhibiting both responses in the 20–35% range only (FIG. 11).

Finally, the abilities of 10 and 25 μM TT2 to respectively inhibit nucleoside transport by 67 and 93% and DNA synthesis by 69 and 97% are both irreversible upon drug removal (FIG. 12), suggesting that, after a 15- to 60-min pretreatment, the presence of TT quinone in the culture medium is no longer required to continually block the cellular transport of 3H-thymidine and thereby prevent its incorporation into DNA. For the sake of comparison, the inhibitions of nucleoside transport and DNA synthesis caused by the novel tricyclic pyrone analog H10, which are fully reversible upon drug removal, are demonstrated as a positive control under similar experimental conditions in the L1210 tumor assay system (FIG. 12).

As shown in FIG. 13, TT16 is as potent as TT2 on both DNA synthesis and the cellular transport of adenosine and thymidine. Hence, the concentration-dependent inhibitions of DNA synthesis by DAU in HL-60-S (IC50: 0.21 μM), HL-60-RV (IC50: 2.5 μM) and HL-60-RS (IC50: 1.7 μM) cells are characterized by RF values of 8.1–11.9 (FIG. 14A). In contrast, TT2 inhibits the rate of incorporation of 3H-thymidine into DNA as effectively in HL-60-S (IC50: 1.8 μM) as in HL-60-RV (IC50: 2.1 μM) and HL-60-RS (IC50: 2.1 μM) cells and there are no significant RFs (1.2), indicating that TT2 maintains its ability to inhibit DNA synthesis in MDR tumor sublines that have become 10 times less sensitive to the action of DAU (FIG. 14B).

DAU, which fails to significantly inhibit nucleoside transport in WT HL-60-S parental cells, is also unable to do so in their two MDR sublines (FIG. 15). In contrast, TT2 is a quinone antitumor drug that has the advantage of rapidly blocking, in a concentration-dependent manner, the cellular transport of both purine and pyrimidine nucleosides in HL-60-S cells (FIG. 15). Moreover, the ability of TT2 to prevent the cellular transport of 3H-adenosine and 3H-thymidine in WT HL-60-S cells (IC50: 3.3 and 2.0 μM, respectively) fully persists (RFs: 1.0–1.2) in both the MDR HL-60-RV (IC50: 3.2 and 2.0 μM, respectively) and HL-60-RS (IC50: 3.7 and 2.3 μM, respectively) cell lines (FIG. 15). Inhibition of RNA and Protein Syntheses by TTs

Besides DNA synthesis, a 3-h treatment with TT2 can also inhibit, in a concentration-dependent manner, the rates of RNA (FIG. 16) and protein (FIG. 17) syntheses determined over 60-min periods of pulse-labeling in L1210 cells in vitro. The concentration-response curves for the inhibitions of RNA (FIG. 16) and protein (FIG. 17) syntheses by TT2 are nearly identical to that for the inhibition of DNA synthesis (FIG. 8): concentrations of TT2 greater than 1.6 μM must be used to demonstrate effectiveness, total inhibition is achieved at 25 μM, and the IC50 value in all cases is about 6 μM. Under similar conditions, DAU inhibits the respective syntheses of RNA (IC50: 0.8 μM) and protein (IC50: 2 μM) about 7.5 and 3 times more effectively than TT2 (FIGS. 16 & 17).

Effects of TTs on the Mitotic Index

Control populations of L1210 cells cultured for 24 h in the absence of drugs contain only 1.86% of mitotic cells (FIG. 18). In relation with their ability to block tubulin polymerization and cell cycle progression in M-phase, 24-h treatments with VCR and the tricyclic pyrone H10 respectively produce 16- and 7-fold increases in the mitotic index (FIG. 18). Such known microtubule de-stabilizing antitumor drugs, therefore, serve as positive controls in this antimitotic assay. In contrast, none of the concentrations of TT2 tested, even those in the μM range that are highly cytostatic and cytotoxic at 24 h, are able to raise the mitotic index of L1210 cells (FIG. 18), suggesting that TT quinones are unlikely to be antimitotic drugs that disrupt microtubule dynamics to trigger their anticancer activity. In relation with its known ability to first accumulate cells in G1, and then inhibit cell cycle traverse as its concentration increases, DAU actually decreases the % of mitotic cells by 74–91% (FIG. 18). Since the highest concentrations of TT2 tested significantly decrease the % of mitotic cells by 38–48% (FIG. 18), TTs might also prevent tumor cell cycle progression to mitosis. Induction of DNA Cleavage by TTs

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L1210 cells containing 3H-thymidine-prelabeled DNA were used to quantitatively determine whether TTs could induce DNA fragmentation over a 24-h period in vitro. CPT and DAU, two anticancer drugs known to induce DNA-strand breaks by respectively inhibiting Topo I and II activities, are used as positive controls in this DNA fragmentation assay (FIG. 19). As reported before with anthracycline quinone antibiotics, the concentration-dependent
induction of DNA cleavage caused by 24-h DAU treatments is biphasic, peaking at 45% in response to 1.6 μM DAU but declining back to control level (5%) at higher concentrations of DAU (Fig. 19). In contrast, the concentration-dependent increase of DNA cleavage produced by 24 h CPT treatments reaches 57% in response to 1.6 μM CPT but remains at a plateau of maximal stimulation (60–70%) at higher concentrations of CPT (Fig. 19). After 24 h, the maximal levels of DNA cleavage caused by 10 μM TT1 (45%) and TT2 (61%) respectively match those induced by 1.6 μM DAU and CPT (Fig. 19). Although concentrations of TTs higher than those of DAU are required to induce such peak of DNA cleavage, the shape of the concentration-response curves for the ability of TT1 and TT2 to break DNA resembles more that of DAU than that of CPT (Fig. 19), suggesting that the DNA-damaging effects of TT quinones and DAU might share some similarity.

Overall, the different TTs compared at 7 μM induce various levels of DNA cleavage at 24 h (Fig. 20) in relation with their respective cytostatic and cytotoxic activities when tested at 256 nM for 4 days (Fig. 1). Indeed, TT2 and TT13, which are the most potent against L1210 cell proliferation and viability (Fig. 1), again induce the most DNA fragmentation (54–57%) after 24 h (Fig. 20), whereas TT4 and TT12, which have no antileukemic activity in the growth and viability assays (Fig. 1), also fail to significantly raise the level of DNA cleavage (4–6%) over control (5%) (Fig. 20). TT1, TT7 and TT9, which have good antileukemic activities (Fig. 1), also produce substantial levels of DNA fragmentation (31–40%) (Fig. 20). Moreover, the extent of DNA cleavage is only 14–18% in L1210 cells treated with TT3, TT5, TT6, TT8 or TT10 and the ability of TT11 to break DNA is minimal (7%) (Fig. 20), in relation with the weak and marginal antiproliferative and cytotoxic effects of these compounds in the L1210 system in vitro (Fig. 1). Fig. 21 shows the same experiment as in Fig. 20 with additional analogs.

Finally, L1210 cells treated for only 3 and 8 h with 10 μM TT1 or TT2 have the same level of DNA fragmentation at 24 h (36–45% or 58–67%, respectively) than if they are exposed for the whole 24-h incubation period to TT1 or TT2 (40 or 63%, respectively) (Fig. 22). Under similar conditions, 1.6 μM DAU and CPT also rapidly trigger molecular events, which are irreversible and produce identical levels of DNA fragmentation at 24 h (33–41% and 51–58%, respectively), whether or not those drugs are maintained in the culture medium after 3 or 8 h (Fig. 20). It should be noted that no significant elevation of DNA cleavage is detectable in this assay after 3 and 8 h of drug exposure and that the increase of DNA fragmentation caused by TTs, DAU and CPT only appear after 12 h to reach a maximal level at 24 h (data not shown), suggesting that the irreversible events triggered by those drugs during the initial 3 h still require a substantial period of time, irrespective of the continual presence or absence of drugs, to fully induce DNA fragmentation. These results suggest that, like anticancer drugs known to induce DNA-strand breaks, TT quinones interact rapidly with cellular targets to induce long-lasting DNA-damaging effects, which develop and persist after drug removal.

Tumor cells containing 3H-thymidine prelabeled DNA were used to quantitatively determine whether TT2 would induce DNA fragmentation as effectively in WT as in MDR HL-60 cell lines that have become resistant to the DNA-damaging effects of DAU (Fig. 23). As reported before, the concentration-dependent induction of DNA cleavage caused by DAU is biphasic, peaking 67% above control level (15% DNA fragmentation) in HL-60-S cells treated for 24 h with 1.6 μM DAU but declining back to (18% above control at 4 μM), or even below (11 and 14% below control at 10 and 25 μM, respectively), control level at higher concentrations of DAU (Fig. 23A). Although TT2 is somewhat less potent than DAU, the concentration-dependent induction of DNA cleavage caused by this compound in HL-60-S cells is similarly biphasic, with a peak 37% above the control % of DNA fragmentation at 4 μM, followed by a decline toward (14% above control at 10 μM), or even below (11% below control at 25 μM), the control % of DNA fragmentation at higher concentrations (Fig. 23B). Interestingly, the ability of DAU to trigger maximal or submaximal DNA cleavage in WT cells is totally lost, drastically reduced and/or shifted toward higher concentrations in the MDR cell lines (Fig. 23A). For instance, the 0.04–0.64 μM concentrations of DAU producing submaximal stimulations of DNA cleavage (11–58% above the control level of 15%) in HL-60-S cells are almost totally unable to do so in the HL-60-RV and HL-60-R8 cell lines. Moreover, the magnitude of the peak of maximal DNA cleavage caused by DAU in WT cells is not only reduced by 60 and 79% in HL-60-R8 and HL-60-RV cells, respectively, but it also shifted from 1.6 to 10 μM in the latter MDR subline (Fig. 23A). As a result, the ability of 10–25 μM DAU to decrease DNA cleavage below control level in WT cells is totally or partially lost in the MDR cell lines. In contrast, the ability of TT2 to induce a peak of DNA cleavage at 4 μM and abolish the basal level of DNA cleavage at 25 μM in WT cells is pretty much unaltered in the two MDR sublines, although the magnitude of maximal stimulation observed in HL-60-S cells is reduced by 35 and 39% in the HL-60-R8 and HL-60-RV cells, respectively (Fig. 23B).

The comparison studies show that the different magnitudes at which equimolar concentrations of various TT analogs inhibit tumor cell proliferation match very well the different cytotoxic activities of these compounds (Fig. 1). Moreover, the different levels of DNA fragmentation induced by the various TT analogs (Fig. 20) match exactly the ranking of these compounds for their cytostatic/cytotoxic effects (Fig. 1), suggesting that the ability of TT analogs to break DNA plays a significant role in their molecular mechanism of antitumor activity. In addition, the various TT analogs inhibit DNA synthesis in relation with their effectiveness against nucleoside transport (Fig. 21), suggesting that they prevent DNA assembly because they block the cellular uptake of DNA precursors. Because of their ability to interact with both membrane and nuclear targets to block nucleoside transport, inhibit nucleic acid and protein syntheses, cleave DNA, and reduce tumor cell growth and viability in the nM range in vitro, these TT analogs represent a novel synthetic class of bifunctional anticancer drugs valuable to develop new means of polychemotherapy.

The cytostatic and cytotoxic effects of each concentration of TT2 increase with the time in culture (Figs. 3 and 4), suggesting that the effectiveness of TT analogs as inhibitors of tumor cell proliferation and viability in vitro is a combination of drug concentration and duration of drug exposure. As a result, TT2 inhibits L1210 tumor cell growth at 2 and 4 days with IC50 values of 300 and 150 nM, respectively, in relation with its ability to reduce L1210 tumor cell viability with IC50 values of 250 nM at day 2 and 100 nM at day 4. The IC50 required to reduce tumor cell viability at day 4 (100 nM) may be lower and perhaps more accurate than that observed for tumor cell growth inhibition at day 4 (150 nM) because the Coulter counter data include...
all viable and nonviable tumor cells that have previously accumulated and remain in the medium at day 4, irrespective of their present metabolic status and reproductive ability. Moreover, decreased tumor cell viability after several days of drug treatment may be a better predictor of anticancer activity than antiproliferation since growth delay may allow survivors to resume dividing and expand clonally once the drug is catabolized or eliminated and its effect is waning.

Overall, the new TT bisquinone TT2 is a potent antitumor agent, which is active in the mM range and is only 3.5–6 times less cytostatic/cytotoxic after 4 days than equimolar concentrations of the clinically proven anthracycline quinone antibiotic DAU, a very potent anticancer drug used as an arbitrary reference in our L1210 tumor cell studies in vitro (FIGS. 3 and 4). When IC50 values are compared, TT2 also inhibits the syntheses of DNA, RNA and protein in L1210 cells after 2–3 h about 3–6 times less effectively than DAU (FIGS. 9, 16 and 17) and concentrations of TT2 about 6 times higher than those of DAU are also required to induce the same peak of DNA cleavage in L1210 cells after 24 h (FIG. 19). But in addition to mimicking all the antitumor effects of DAU studied, the data demonstrate that TT2 remarkably blocks the cellular transport of nucleosides, which DAU cannot do (FIG. 10), suggesting that these novel antitumor TT bisquinones may have a more versatile mechanism of action and be advantageous in polychemotherapy to potentiate the antitumor effects of antimitabolites and circumvent multidrug resistance (MDR). For both DAU and TT2 concentrations in the 25–150 nM range are sufficient to inhibit tumor cell growth/viability (FIGS. 3 and 4), whereas higher concentrations in the 0.8–10 μM range must be used to inhibit macromolecule syntheses (FIGS. 9, 16 and 17) and maximally induce DNA fragmentation (FIG. 19). But apparent discrepancies in potencies may be due in part to different experimental conditions and cellular responses to various periods of drug exposure: the rates of nucleic acid and protein syntheses over 30–60 min are inhibited in cells treated for only 2–3h with TT2 or DAU, whereas the level of DNA cleavage and the reduction of tumor cell growth/viability are the results of 1- and 4-day long drug treatments, respectively. It should be noted that no significant DNA fragmentation can be detected within the first 8 h of TT2 or DAU treatments and that concentrations of DAU and TT2 respectively greater than 0.25 and 1.6 μM must be used to induce substantial antiproliferative and cytotoxic effects after only 24 h of drug exposure (data not shown).

The fact that, within 24 h, TT1 and TT2 can produce as much internucleosomal DNA fragmentation in L1210 cells than the known Topo I and II inhibitors CPT and DAU (FIG. 19) suggests that the ability of TT quinones to produce DNA-strand breaks may play a major role in their mechanism of antitumor activity. Since the shapes of the concentration-response curves for the effects of TT1 and TT2 on DNA cleavage resemble the biphasic response to DAU, which has already been reported and discussed before, it is tempting to speculate that the mechanisms by which TT quinones and DAU induce DNA fragmentation share some similarity and that DNA cleavage is not always necessary for high concentrations of TT quinones and DAU to be cytotoxic. The Topo II-associated DNA lesions occurring in cells exposed to anthracycline quinone antibiotics may facilitate subsequent internucleosomal DNA fragmentation by endogenous nucleases and trigger apoptosis. Since apoptosis is an active and cell cycle phase specific process, which requires the expression of specific genes, the syntheses of new RNA and proteins and the activation of endonuclease enzymes, inhibitors of such mechanisms can prevent DNA fragmentation in anthracycline-treated cells. In spite of their increasing cytotoxicity, the highest concentrations of TT1, TT2 and DAU tested in our study might inhibit RNA and protein syntheses and reduce the level of Topo II targets, inactivate endonucleases and/or arrest cell cycle traverse to such degrees that they actually block the molecular mechanisms required for internucleosomal DNA fragmentation and apoptosis and produce the paradoxical biphasic curve of DNA cleavage shown in FIG. 19. Taken together, the irreversibilities of the inhibitions of nucleoside transport/DNA synthesis and of the induction of DNA fragmentation upon TT2 removal suggest that, because TT analogs may rapidly and tightly interact with various membrane and intracellular targets, their presence is soon no longer required in the medium to disrupt the structures/functions of nucleoside transporters, nucleic acids and proteins and to trigger long-lasting antitumor events, which persist after cessation of drug treatment.

Combining drugs, which target different molecules and achieve complementary or synergistic antitumor effects, is an important strategy in cancer chemotherapy. AT1 quinone inhibiting nucleoside transport and inducing DNA cleavage might disrupt a wider spectrum of molecular targets in populations of unsynchronized tumor cells than another drug affecting a single of these events. Nucleoside transporters in mammalian plasma membranes function by equilibrative (facilitated diffusion) or Na+-dependent (concentrating) mechanisms. All nucleosides are substrates for the Na+-independent transporters but Na+-dependent transporters generally accept purine rather than pyrimidine nucleosides as substrates. Dipyridamole (DPR), 6-(4-nitrobenzy1mercapto)purine ribonucleoside (NBMPR) and dilazep are standard potent inhibitors of equilibrative nucleoside transport, whereas phosphoridzin specifically inhibits its Na+-dependent nucleoside transport. Like TT2, NBMPR and DPR also reduce 3H-thymidine incorporation into DNA. Nucleoside transport inhibitors block equally well the influx and efflux of nucleosides. L1210 cells possess 3 distinct nucleoside transporters: 2 equilibrative transporters sensitive (es) or insensitive (is) to NBMPR and one Na+-dependent transporter (cil), of low sensitivity to NBMPR and DPR.

For nucleotide synthesis, cells use purine and pyrimidine nucleosides generated either through de novo synthesis or through the utilization of salvage pathways. MDR is sometimes associated with increases in the number of nucleoside transporters and their rate of transport, resulting in the increased uptake of adenosine. By blocking the rescue effect of exogenous nucleosides, NBMPR, DPR and dilazep may potentiate or prolong the antitumor activity of antimitabolites which inhibit the de novo pathway for nucleoside synthesis. The clinical effectiveness of ADR and DAU is limited by their cumulative cardiotoxicity and ability to induce MDR. The multifactorial mechanisms of MDR to anthracycline quinone antibiotics may include altered expressions of P-glycoprotein (P-gp), Topo II and multidrug resistance-associated protein, increased DNA repair and glutathione-dependent detoxifying enzyme activities, and alterations in cell cycle progression and apoptotic pathways. DPR circumvents ADR resistance and its analog, BIBW 22, is a bifunctional modulator which reverses the MDR phenotype by interfering with both P-gp and nucleoside transport in MDR cells. As bifunctional inhibitors of nucleoside transport and inducers of DNA cleavage, TT quinones might be valuable in polychemotherapy to potentiate the antitumor activity of methotrexate and 5-Fluorouracil and sensitize MDR tumor cells that have become unresponsive to the cytotoxicity of other conventional DNA-damaging anticancer agents.
TT2 is also cytostatic (IC₅₀: 300 nM) and cytotoxic (IC₅₀: 230 nM) to WT human HL-60-S leukemic cells in the present study. Another attractive feature of TT2 identified in the present study is the finding that this new quinone antitumor drug retains its effectiveness in two MDR HL-60-RV and HL-60-R8 sublines that have developed different mechanisms of resistance to DAU and, therefore, might not be recognized by either the P-gp or the MRP.

The magnitude at which TT2 inhibits HL-60-S, HL-60-RV and HL-60-R8 cell proliferation (IC₅₀ values: 260–300 nM) matches its ability to decrease cell viability (IC₅₀ values: 230–350 nM) in the same WT and MDR tumor cell lines at day 4. Moreover, the cytostatic and cytotoxic effects of each concentration of TT2 obviously increase with the time in culture at days 2 and 4, suggesting that the effectiveness of TT2 as an inhibitor of tumor cell proliferation and viability is a combination of drug concentration and duration of drug exposure. Decreased WT and MDR tumor cell viability after TT2 treatment in vitro may be a reliable predictor of anticancer activity in vivo. However, it should be noted that DAU is about 3.5–6 times more cytostatic/cytotoxic than TT2 in L1210 cells, even though TT2 appears more potent in the L1210 than in the HL-60-S cells. Hence, TT2 inhibits tumor cell growth, viability and DNA synthesis respectively 33, 15 and 9 times less effectively than DAU in the HL-60-S leukemic system (FIGS. 5–8, 10). Moreover, concentrations of TT2 at least 2.5 times greater than those of DAU are required to induce more than 50% of DNA cleavage in HL-60-S cells at 24 h (FIG. 23). Nevertheless, the magnitudes of the inhibitory and stimulatory effects triggered by DAU and somewhat higher concentrations of TT2 are nearly identical in HL-60-S cells. But the critical finding is that the ability of TT2 to inhibit tumor cell growth, viability and DNA synthesis in HL-60-S cells persists unaffected in the HL-60-RV and HL-60-R8 cell lines (RFs: 0.9–1.5) while DAU becomes dramatically less effective against DNA synthesis (RFs: 8.1–11.9), cell proliferation (RFs: 22.9–35.7) and cell viability (RFs: 23.8–31.3) in these MDR HL-60 sublines than in their parental WT counterparts. Finally, TT2 which, in contrast to DAU, has the unique ability to rapidly block the cellular transport of both purine and pyrimidine nucleosides in L1210 (13) and HL-60-S cells, retains such additional advantage in the MDR HL-60 sublines (FIG. 15). These observations substantiate the hypothesis that, because of its bifunctional mechanism of action, TT2 is a quinone antitumor drug which may be more versatile than DAU and able to circumvent MDR.

SYNTHESIS EXAMPLES

Synthesis of TT2, TT13 and Their Analogos:

Although the preparation of 1,4-dimethoxyanthracene (1) has been reported (Criswell, et al., J. Org. Chem. 1974, 39, 770–774) by using methylation of quinizarin (3) followed by sequential reduction of the diketo function with sodium borohydride in diglyme, the difficulty in the reduction steps prompted us to investigate an easier method in the preparation of 1. Reduction of 3 with sodium borohydride in MeOH followed by quenching with HCl gave a 95% yield of 1,4-anthraquinone (4) (Scheme 2). Reduction of the quinone moiety of 4 with aqueous sodium hydrosulfite in 1,4-dioxane provided 1,4-dihydroxyanthracene (5). Methylation of 5 with sodium hydride in DMF at room temperature gave excellent yield of 1.

In Situ Oxidation Followed by [4+2]-Cycloaddition:

A short synthesis of TT2, TT3, and TT5 were discovered. Without the use of quinones as the dienophiles in the Diels-Alder reaction, such as 2-methoxyquinone (8), a one-pot oxidation followed by [4+2]cycloaddition of 2-methoxyhydroquinone (6) with anthracene 1 was found (Scheme 3). Hence, treatment of 2 equiv of 6, 1 equiv of anthracene 1, 2 equiv of silver oxide, and 0.2 equiv of zinc iodide in refluxing toluene for 24 h gave diketone 7 (51% yield; based on reacted 1), hydroquinone TT3 (26% yield; based on reacted 1), quinone TT5 (13% yield; based on reacted 1) and 55% recovery of 1.
Alternatively, 7 can be obtained from the Diels-Alder reaction of quinone 8 with anthracene 1 (Scheme 4). Hence, oxidation of 6 with 1.5 equiv of silver oxide and 1.2 equiv of potassium carbonate in benzene at 25°C for 3 h afforded quantitative yield of quinone 8 which underwent cyclodition with 1 gave a 48% yield of 7 as a mixture of endo- and exo-isomers (a ratio of 1:1) at C4a and C9a.
Selective Bromination of 2-Methoxy Triptycene Quinone:
A new bromination reaction was found (Scheme 6). Hence, when TT5 was treated with N-bromosuccinimide (NBS) in DMF at 40° C. for 12 h, a quantitative yield of the C-2 brominated product 9 was obtained. Without purification, compound 9 was directly subjected to the ceric ammonium nitrate oxidation and a 59% yield of TT13 was achieved.

Synteses of 2-Chloro-4a,9a,10-tetrahydro-9,10-[1',2'] benzenoanthracene-1,4-dione (10), TT8, TT9 and TT7:
The in-situ oxidation and cyclization reaction is applicable to other hydroquinone substrates. For instance,
with 2 equiv of silver oxide and 1.2 equiv of potassium carbonate in benzene at 50° C. for 10 min under dark gave a quantitative yield of the corresponding quinone 16 (Scheme 9). Diels-Alder reaction of 15 and anthracene 1 at 70° C. for 14 h and then under reflux for 5 h gave 58% yield of TT11 (as a mixture of endo- and exo-isomers) and 37% yield of TT10.

Similarly, treatment of 2-methoxyhydroquinone (6) with anthracene (13) and silver oxide and zinc iodide gave a 56% yield of the 2-methoxy derivative TT8 (Scheme 8). Basic isomerization of ketone 14 with 3 equiv of KOH in 1,4-dioxane and water at 25° C. for 2 h followed by oxidation with silver oxide and sodium sulfate (anhydrous) in dried acetone at 25° C. for 3 h to give an 80% yield of quinone TT9. Quinone TT7 was prepared by the Diels-Alder reaction of benzoquinone (2) with anthracene 13 to give adduct TT6 (98% yield) which under similar isomerization and oxidation as those for the conversion of TT8 into TT9 gave TT7 (70% yield for the last two steps).

Syntheses of TT10 and TT11:
Contrary to the above results, when methyl gentisate (14) was used in the in-situ oxidation followed by [4+2] cycloaddition reaction, only small amount of a mixture of products was isolated. Compound 14 was then oxidized first

The triptycene bis-quinones such as TT1 could be treated with dienes to produce various substituted triptycene analogs such as TT4 and TT12 (Scheme 10). Hence, treatment of TT1 with 2.2 equiv of dimethyl butadiene-2,3-dicarboxylate (16) (Hamon, et al., J. Chem. Soc. Chem. Commun. 1981, 873–4) in toluene under reflux for 20 h gave a 54% yield of TT4 and 14% yield of monoadduct 17. The stereochemistry of TT4 was firmly established by a single-crystal X-ray analysis. Reduction of TT4 with 10 equiv each of sodium borohydride and cerium trichloride heptahydrate in MeOH at room temperature for 12 h afforded an 89% yield of TT12. The 1H and 13C NMR spectrum of TT12
indicated a single stereoisomer. It is anticipated that the hydride (sodium borohydride) should attack the carbonyl group from the exo face (b-face) and the stereochemistry is therefore assumed.

Syntheses of TT14, TT15, and TT16:

Bromomethoxyquinone TT13 can be converted into methylaminoquinone TT14 in 25% yield along with a 29% yield of TT15 by the treatment with methylamine in THF at room temperature for 20 min (Scheme 11). This reaction is unusual in that the nucleophile, methylamine, displaces the methoxy group of TT13 instead of the bromine. Again, unexpectedly, when TT13 was treated with dimethylamine in THF at 0°C, TT16 was isolated as the only identifiable product (Scheme 12). The regiochemistry of the dimethylamino group of TT16 is tentatively assigned.

Similar to the addition reaction of methylamine with TT13, other primary amines, such as ethyl b-alanine and L-lysine also add to TT13 to provide the corresponding amino acid adducts TT17–TT20 (Scheme 13). Hence, treatment of TT13 with ethyl b-alanine (derived from ethyl b-alanine hydrochloride with 1 equiv of sodium hydride in THF) in THF and DMF at 25°C to give a mixture of TT17 and TT18 (based on the proton NMR spectrum of the crude product). Column chromatographic separation on silica gel gave TT18 in 69% yield. The ethyl ester function of TT18 can be removed by treatment with sodium iodide in DMF with heat to give amino acid TT21 which is water soluble.
Scheme 12:

\[
\begin{align*}
&\text{TT13} \\
&\xrightarrow{\text{Me}_2\text{NH}} \text{THF, } 0^\circ\text{C, } 20 \text{ min}
\end{align*}
\]

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Scheme 13:

\[
\begin{align*}
&\text{TT13} \\
&\xrightarrow{\text{H}_2\text{NCH}_2\text{CH}_2\text{CO}_2\text{Et}} \text{THF, r.t.}
\end{align*}
\]

44

-continued

\[
\begin{align*}
&\text{TT16}
\end{align*}
\]

TT17 + TT18

\[
\begin{align*}
\text{column chromatography}
\end{align*}
\]

TT18 \xrightarrow{\text{Na, DMF, heat}}

TT21
Under similar reaction conditions, TT13 was treated with 6.6816 g (MW 174; 0.0384 mol) of sodium hydrosulfite in L-lysine (derived from L-lysine monohydrochloride and 150 mL of distilled water and 50 mL of 1,4-dioxane were equiv of NaH in THF) in THF and DMF to give adducts added. The resulting mixture was stirred at room temperature for 2-3 hours. The reaction progress is checked with TT19 and TT20 which were purified on HPLC to give TT20 (TT19 most likely hydrolyzes on the column to give TT20). Hence, a number of water-soluble drugs can be obtained by adding amino acids or amino sugars onto TT13, by reactions that are known in the art.

Preparation of 1,4-anthracenedione (4):
To a dried 100 mL-round-bottom flask equipped with stirring bar and serum cap, was added 1 g (MW 240.21; 0.0042 mol) of quinizarin (3). The compound was dried under vacuum, and then maintained under argon. Methanol (distilled over Mg), 20 mL, was added via syringe. The solid was filtered through a glass fritted funnel and washed with 30 mL of distilled water twice. The solid product was dried under vacuum to give 1.754 g (distilled over Mg), 20 mL, was added via syringe. The small amount of p-dioxane) d 8.7 (m, 2H, C 9,10 Hs), 8.05 (m, 2H, C 5,8 Hs), 7.5 (s, 2H, C 6,7 Hs), 6.6 (s, 2H, C 2,3 Hs). The diol is insoluble in chloroform but soluble in DMSO and DMF.

Preparation of 1,4-Dimethoxyanthracene (1):
To a dried flask, 0.349 g (0.0011 mol; 50% yield) of sodium borohydride was added. The resulting mixture was stirred at 0°C for 1 hour. The progress of the reaction was monitored by TLC (using hexanes:ether 1:1 as the eluant). The reaction was quenched by adding carefully 11 mL of 6 N HCl dropwise at 0°C over a period of 10 minutes. Orange solids precipitate. The solids were collected by filtration over a fritted filter funnel, and washed several times with distilled water to remove the acid. The solid was then dried under vacuum and recrystallize from acetone-ether to give 0.83 g (95% yield) of yellow crystals, mp 204–206°C.

1H NMR (CDCl3) d ppm: 7.1 (s, 2H, C 2,3 Hs), 7.7 (dd, J=6.4, 3.2 Hz, 2H, C 6,7 Hs), 8.1 (dd, J=6.4, 3.2 Hz, 2H, C 5,8 Hs), 8.6 (s, 2H, C 9,10 Hs); 13C NMR (CDCl3) d ppm: 184.68 (s, C=O), 140.06 (d), 134.83 (s), 130.22 (d), 129.59 (d), 128.87 (d), 128.37 (s).

Preparation of 1,4-Dihydroxyanthracene (5):
To a 250 mL-round-bottom flask, was added 2 g (MW 208; 0.0096 mol) of 1,4-anthracenedione (4). To it a solution of 6.6816 g (MW 174; 0.0384 mol) of sodium hydrosulfite in 50 mL of distilled water and 50 mL of 1,4-dioxane were added. The resulting mixture was stirred at room temperature for 2–3 hours. The reaction progress is checked with TLC (using CH2Cl2:hexanes:ether 1:1:0.2 as the eluant). The mixture was transferred to a 500-mL beaker slowly, cooled over an ice-water bath, and added 100 mL of distilled water. Dark green solids precipitated. The beaker was covered with aluminum foil, and placed in the refrigerator for 1 h. The solid was filtered through a glass fritted funnel and washed with 30 mL of distilled water twice. The solid was dried under vacuum, and then maintained under argon. To it 3 mL of distilled ether was added, stirred, and the ether layer containing the oil was removed and discarded. To the sodium hydride flask, 1 g (0.0048 mol) of diol 5 was added and the mixture was dried under vacuum and flame and maintained under argon. DMF (10 mL) was added via syringe and the solution was stirred at room temperature for 1.5 h. TLC of a sample from the reaction solution indicated no starting material. The solution was diluted with 20 mL of water and 6 N HCl until pH=2. The mixture was extracted with ethyl acetate three time. The combined extract was washed with water twice and brine, dried (MgSO4), concentrated to give 1.07 g (94% yield) to the crude product (proton NMR spectrum indicated >90% pure). The product was recrystallized from ether:hexane to give 0.89 g (78% yield) of black
solids, mp 255–258° C. 1H NMR (CDCl₃) δ 8.70 (s, 2H, C9,10(H1)), 7.97 (dd, J=6.1, 3.6 Hz, 2H, C12,13(H1)), 7.40 (dd, J=6.3, 3.2 Hz, 2H, C6,7(H1)), 6.55 (s, 2H, C2,3(H1)), 3.97 (s, 6H, OCH₃). 13C NMR (CDCl₃) δ 149.47 (Cα,β), 131.45 (Cα,β), 128.51 (d, C9,10), 125.47 (d, C5,8), 120.73 (d, C6,7), 100.88 (d, C2,3), 55.61 (s, OCH₃). MS: FAB (M+), m/z=238 (M), 239 (MH⁺).

2,5,8-Trimethoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzo[a]anthracene-1,4-dione (7), 1,4-Dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1',2']benzo[a]anthracene (TT3), and 2,5,8-Trimethoxy-9,10-dihydro-9,10-[1',2']benzo[a]anthracene-1,4-dione (TT4).

To a solution of 0.77 g (2 mmol) of 7, 0.002 g of silver oxide, and 0.179 g (0.56 mmol) of zinc iodide in 20 mL of toluene (distilled over CaH₂) was heated to reflux under argon for 24 h. The reaction mixture was cooled to room temperature, concentrated to give 0.77 g (100% yield) of 7T5. 1H NMR (CDCl₃) δ 7.48 (dd, J=5.5, 3 Hz, 2H, C12,13(H1)), 7.09 (dd, J=5.5, 3 Hz, 2H, C11,14(H1)), 6.63 (s, 1H), 6.16 (s, 1H), 4.17 (s, 3H, C3 OMe). 13C NMR (CDCl₃) δ 182.2 (s, C1), 182.0 (s, C4), 176.2 (s, C=O), 175.5 (s, C≡O), 156.2, 151.6, 151.4, 150.2, 141.8, 135.5, 135.4, 126.0, 126.0, 125.6, 125.4, 117.2, 61.7 (q, OMe), 43.0 (d), 42.1 (d). MS EI, m/z 422 & 422 (1:1), 344 (M–Br), 300, 287, 232, 152, 126.

2-Chloro-5,8-dimethoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzo[a]anthracene-1,4-dione (12):

A solution of 0.1 g (0.42 mmol) of 1,4-dimethoxyanthracene (1), 0.121 g (0.84 mmol) of chlorohydroquinone (11), 0.195 g (0.84 mmol) of silver oxide, and 27 mg (0.084 mmol) of zinc iodide in 10 mL of toluene was heated under reflux for 10 h. The mixture was concentrated and then transferred to a separatory funnel, washed with water and brine, dried (MgSO₄), and filtered through Celite, and the filtrate was concentrated to give 109 mg (quantitative yield) of pure TT13; mp 207–210° C.

1H NMR (CDCl₃) δ 7.48 (dd, J=5.5, 3 Hz, 2H, C12,13(H1)), 7.09 (dd, J=5.5, 3 Hz, 2H, C11,14(H1)), 6.66 (2H, C6,7(H1)), 6.23 (1H), 6.16 (1H), 4.17 (s, 3H, C3 OMe). 13C NMR (CDCl₃) δ 182.2 (s, C1), 182.0 (s, C4), 176.2 (s, C=O), 175.5 (s, C≡O), 156.2, 151.6, 151.4, 150.2, 141.8, 135.5, 135.4, 126.0, 126.0, 125.6, 125.4, 117.2, 61.7 (q, OMe), 43.0 (d), 42.1 (d). MS EI, m/z 422 & 422 (1:1), 344 (M–Br), 300, 287, 232, 152, 126.

2-Chloro-5,8-dimethoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzo[a]anthracene-1,4-dione (12):

A solution of 0.1 g (0.42 mmol) of 1,4-dimethoxyanthracene (1), 0.121 g (0.84 mmol) of chlorohydroquinone (11), 0.195 g (0.84 mmol) of silver oxide, and 27 mg (0.084 mmol) of zinc iodide in 10 mL of toluene was heated under reflux for 10 h. The mixture was concentrated and then transferred to a separatory funnel, washed with water and brine, dried (MgSO₄), and filtered through Celite, and the filtrate was concentrated to give 109 mg (quantitative yield) of pure TT13; mp 207–210° C.
2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione (TT9):

To a solution of 0.2 g (0.63 mmol) of ketone TT6 in 5 mL of p-dioxane and 1 mL of water was added 0.106 g (1.9 mmol) of KOH. The solution was stirred at room temperature for 4 h, neutralized with 1 N HCl, and extracted with methylene chloride twice. The combined extract was washed with brine, dried (MgSO₄), and column chromatographed on silica gel using a gradient mixture of hexane and ether as eluant to give 0.16 g (58% yield) of TT7. mp 219-221°C; 'H NMR (CDCl₃) d 7.42 (dd, J=5.5, 3.2 Hz, 2H, C3',6'), 6.74 (d, J=10 Hz, lH, C2H), 6.50 (d, J=10 Hz, lH, C5H), 5.70 (d, J=2.4 Hz, lH, C9H), 3.92 (s, 3H, OMe). 13C NMR (CDCl₃) d 169.7, 149.8, 140.7, 135.4, 131.5, 129.1, 126.7, 125.9, 124.6, 123.1, 121.1, 30.9. 1H NMR (CDCl₃) d 6.92 (s, 1H of one isomer, exo or endo), 6.25 (s, 1H of one isomer), 5.28 (s, 1H), 5.25 (s, 1H), 3.86 (s, 3H, OMe), 3.83 (s, 3H, OMe), 3.15 (m, 3H), 3.12 (m, 1H), 2.91-2.70 (m, 4H), 1.90-1.80 (m, 2H), 1.20-1.00 (m, 2H), 0.85-0.70 (m, 3H). 13C NMR (CDCl₃) d 169.7, 149.8, 140.7, 135.4, 131.5, 129.1, 126.7, 125.9, 124.6, 123.1, 121.1, 30.9. 1H NMR (CDCl₃) d 6.92 (s, 1H of one isomer, exo or endo), 6.25 (s, 1H of one isomer), 5.28 (s, 1H), 5.25 (s, 1H), 3.86 (s, 3H, OMe), 3.83 (s, 3H, OMe), 3.15 (m, 3H), 3.12 (m, 1H), 2.91-2.70 (m, 4H), 1.90-1.80 (m, 2H), 1.20-1.00 (m, 2H), 0.85-0.70 (m, 3H). 13C NMR (CDCl₃) d 169.7, 149.8, 140.7, 135.4, 131.5, 129.1, 126.7, 125.9, 124.6, 123.1, 121.1, 30.9. 1H NMR (CDCl₃) d 6.92 (s, 1H of one isomer, exo or endo), 6.25 (s, 1H of one isomer), 5.28 (s, 1H), 5.25 (s, 1H), 3.86 (s, 3H, OMe), 3.83 (s, 3H, OMe), 3.15 (m, 3H), 3.12 (m, 1H), 2.91-2.70 (m, 4H), 1.90-1.80 (m, 2H), 1.20-1.00 (m, 2H), 0.85-0.70 (m, 3H).
Preparation of Dimethyl Butadiene-2,3-dicarboxylate (16): (Z)-Dimethyl 2,3-dimethyl-2-butenedioate (21):

To a solution of 2 g (0.0158 mol) of 2,3-dimethylmaleic anhydride in 40 mL of methanol was added 0.2 mL of concentrated sulfuric acid. After refluxing for 7 days, the solution was diluted with 100 mL of methylene chloride, neutralized with aqueous sodium bicarbonate solution. The methylene chloride layer was washed with 30 mL of brine, the aqueous layer was extracted with methylene chloride four times (30 mL twice and 15 mL twice). The combined methylene chloride extracts were dried over anhydrous magnesium sulfate, concentrated to give 2.11 g of crude product. \(^1\)H NMR check indicated about 25% of starting material existing, the crude mixture with 0.2 mL of concentrated sulfuric acid in 40 mL of methanol was heated at reflux again for 5 days. The solution was concentrated until 30% of methanol was left, neutralized with saturated sodium bicarbonate aqueous solution, followed by addition of 30 mL of brine solution, and the mixture was extracted with ethyl acetate four times (100 mL once, 30 mL three times). The combined ethyl acetate extracts were dried over anhydrous magnesium sulfate, concentrated to give 3.26 g of crude product (100% yield). It was of satisfactory purity for the next step. \(^1\)H NMR (CDCl\(_3\)) d 3.77 (s, 6H, OMe), 1.95 (s, 6H, Me)

(Z)- and (E)-Dimethyl-2,3-bis[bromomethyl]-2-butenedioate (22a & 22b):

A mixture of 3.2384 g (0.0188 mol) of butenedioate 21, 7.37 g (0.04 mol) of N-bromosuccinimide (NBS) and 100 mg (cat.) of 2,2'-azobisobutyronitrile (AIBN) in 50 mL of carbon tetrachloride was heated under reflux and irradiated with a 275 W sunlamp for 3 h. The reaction solution was cooled, diluted with 150 mL of ether, and filtered. The filtrate was concentrated, and flash column chromatographed using a gradient mixture of ether-hexane as eluant to give 1.33 g of oily liquid trans-isomer 22b (18.4% yield) and 4.40 g of oily liquid cis-isomer 22a (71.6% yield) with a total yield of 90%. The assignment of 22a and 22b was based on the reported \(^1\)H NMR data.

22a: \(^1\)H NMR (CDCl\(_3\)) d 4.50 (s, 4H, CH\(_2\), Br), 3.91 (s, 6H, OMe); \(^13\)C NMR (CDCl\(_3\)) d 165.54 (s, C=O), 136.95 (s, C=), 52.97 (s, OMe), 26.75 (t, CH\(_2\), Br)
22b: \(^1\)H NMR (CDCl\(_3\)) d 4.24 (s, 4H, CH\(_2\), Br), 3.84 (s, 6H, OMe); \(^13\)C NMR (CDCl\(_3\)) d 165.96 (s, C=O), 137.06 (s, C=), 52.99 (s, OMe), 23.95 (CH\(_2\), Br).

Dimethyl 1,3-butadiene-2,3-dicarboxylate (16):

To a solution of 6.53 g (0.02 mol) of bromoesters 22 in 20 mL of acetone was added 9.49 g (0.06 mol) of sodium thiosulfate and 9.96 g (0.06 mol) of potassium iodide. After refluxing for 2 h, the mixture was cooled, poured onto 50 g of ice, and extracted with methylene chloride three times (150 mL once, 50 mL twice). The combined methylene chloride extracts were washed with 20 mL of saturated aqueous sodium chloride, dried over anhydrous magnesium sulfate, concentrated, and column chromatographed on silica gel using a gradient mixture of hexane-ethyl acetate as eluant to give 1.15 g of pure 20 in a yield of 67.5% (based on crude 19). The melting point and IR agree with reported data. M.p. 271.5-272.5°C; IR: 3060 cm\(^{-1}\) (A-H), 1660 cm\(^{-1}\) (C=O), 1515 cm\(^{-1}\) (C-C).
under argon and heated under reflux for 22 h. The solution was directly subjected onto a silica gel column and eluted with a gradient mixture of hexane, methylene chloride, and ethyl acetate to give 0.160 g (54% yield) of TT4 and 30 mg (14% yield) of compound 17. Recrystallization of TT4 from methylene chloride gave light yellow crystals. Single crystal X-ray diffraction analysis was carried on a crystal and the structure was solved. The following Figure shows the ORTEP drawing of the compound. 1H NMR (CDCl3) δ 7.45 (m, 2H), 7.09 (m, 2H), 6.07 (s, 2H, C6,13 Hs), 3.75 (s, 12H, OMe), 3.2 (bs, 4H, 3C4a,7a,lla,14a Hs), 2.62–2.5 (m, 8H); 13C NMR (CDCl3) d 192.36 (s, C=O), 167.12 (s, C=O of ester), 154.04 (d), 141.49 (s), 132.51 (d), 126.02 (s), 125.57 (s), 52.33 (q, OMe), 45.5 (d), 43.27 (d), 25.48 (t).

Compound 17: 1H NMR (CDCl3) δ 7.42 (dd, J=5, 3 Hz, 2H), 7.01 (dd, J=5, 2H), 6.38 (s, 2H), 3.71 (s, 6H, OMe), 3.17 (bs, 2H, CH2=O), 2.60 (dd, J=17, 4 Hz, 4H), 2.45 (dd, J=17, 4 Hz), (1H). (4aS*, 5S*, 7R*, 12S*, 14R*, 7aR*, 11aS*, 14aR*)-Tetramethyl 1,4,4a,5,6,7,7a,8,11,12,13,14,14a-tetrahydroxy-6,13-[1’,2’]-benzenoanthracene-2,3,9,10-tetraoxycarbylate (TT12):

To a solution of 0.10 g (0.15 mmol) of TT14 in 7 mL of methanol (distilled over magnesium turning) under argon were added 0.52 g (1.4 mmol) of CeCl3·7H2O and 51 mg of sodium borohydride. The solution was stirred at room temperature for 2 h, diluted with methylene chloride, washed with water and brine, dried (MgSO4), and concentrated. Column chromatographic separation on silica gel using a gradient mixture of hexane and methylene chloride gave light yellow crystals. Single crystal X-ray diffraction analysis was carried on a crystal and the structure was solved. The following Figure shows the ORTEP drawing of the compound. 1H NMR (CDCl3) δ 7.48 (dd, J=5, 3 Hz, 1H), 7.45 (dd, J=5, 3 Hz, 1H), 7.07 (dd, J=5, 3 Hz, 2H), 6.64 (2 s, 2H, C6,7 Hs), 6.28 (s, 1H), 6.11 (s, 1H), 5.85 (bs, 1H, NH), 3.33 (d, J=5.6 Hz, 3H, McN). For TT15: 1H NMR (CDCl3) δ 7.45 (m, 2H, Ar=H), 7.04 (m, 2H, Ar=H), 6.41 (2d, J=6.3 Hz, 2H, C6,7 Hs), 6.23 (s, 1H), 6.1 (s, 1H), 5.9 (s, 1H, NH), 4.8 (s, 1H, OH), 3.33 (d, J=5.8 Hz, 3H, McN).

A solution of 24 mg (0.057 mmol) of TT13 and 1.8 mg (0.00325 mol) of L-lysine monohydrochloride in 5 mL of THF under argon was added 0.1315 g (0.0027 mol, 50% oil dispersion) of sodium hydroxide. The mixture was stirred at room temperature for 2 h, concentrated to dryness under vacuum and while solids, b-alanine ethyl ester and sodium chloride, were obtained.

To a solution of 64 mg (0.151 mmol) of TT13 in 2 mL of THF and 0.5 mg of DMF under argon was added 0.302 mmol each of the above mixture of b-alanine ethyl ester and sodium chloride. The solution was stirred at room temperature for 16 hours, 14.13 diluted with methylene chloride, the organic layer was washed with water twice, brine, dried (MgSO4), and concentrated. The proton NMR spectrum of the crude products indicated a mixture of TT17 and TT18. This crude product was subjected to column chromatographic separation on silica gel using a gradient mixture of hexane and ethyl acetate as eluant to give 5 mg of recovered starting material TT13 and 28 mg of the product TT16. 1H NMR (CDCl3) δ 7.45 (m, 2H, Ar=H), 7.04 (m, 2H, Ar=H), 6.22 (s, 1H), 6.15 (s, 1H), 5.4 (s, 1H, CH), 3.81 (s, 3H, OMe), 3.12 (s, 6H, Me3N).
a sequence of three reactions was carried out: Diels-Alder reaction of 1,4-dimethoxyanthracene (Criswell, T. R.; Klanderman, B. H. J. Org. Chem. 1974, 39, 770) and p-benzoquinone followed by isomerization of the adducts (endo- and exo-adducts) with potassium hydroxide and then oxidation with silver oxide. These three reactions have been successfully combined into one simple operation. Hence, heating of anthracene 2 (derived from the reduction of 1,4-anthracenedione (Perchellet, E. M.; Magill, M. J.; Huang, X.; Brantis, C. E.; Hua, D. H.; Perchellet, J. P. Anti-cancer Drugs, 1999, 10, 749) with sodium hydrosulfite followed by sodium hydride and iodomethane), 1.4 equiv of methoxyhydroquinone (3), 2.7 equiv of silver oxide, and 0.2 equiv of zinc iodide in toluene under reflux for 4 days gave a 70% yield of triptycene monoquinone 4 (Scheme 14). Presumably, silver oxide oxidizes methoxyhydroquinone to methoxy-p-benzoquinone which undergoes Diels-Alder reaction with anthracene 2 to give adducts 5 (endo- and exo-isomers). Compound 5 then undergoes oxidation with silver oxide to give quinone 4. A catalytic amount of zinc iodide was added to facilitate the Diels-Alder reaction. No other regioisomers (such as isomers with methoxy substituent attached at C4a) were detected. To verify the reaction sequence, methoxyquinone 6, obtained from the oxidation of 3 with silver oxide and potassium carbonate in benzene (98% yield), was treated with anthracene 2 in toluene at 150°C in a sealed tube to give adducts 5 (48% yield). Isomerization of ketones 5 with KOH in p-dioxane and water followed by oxidation with silver oxide gave monoquinone 4 (95% overall yield). Oxidation of monoquinone 4 with ceric ammonium nitrate afforded a 94% yield of triptycene bisquinone 7. Regioselective bromination of 7 was achieved by the treatment with N-bromosuccinimide (NBS) in DMF at 25°C for 10 h to give bromoquinone 1 (45% yield). Alternatively, compound 1 can also be obtained from the bromination of monoquinone 4 with NBS in DMF at 40°C followed by oxidation with ceric ammonium nitrate (59% overall yield). The bromination of quinones with NBS has not been reported previously, and apparently, reactivity of the methoxyquinone moiety towards NBS is greater than that of quinone and 1,4-dimethoxyphenyl moieties. Likely, the methoxy substituent of 7 or 4 (at C2) enhances nucleophilicity of adjacent double bond carbon resulting the bromination.
To synthesize new analogs, 1 was treated with primary and secondary amines (Scheme 15). Surprisingly, compound 1 reacts with primary amines such as methylamine and secondary amines such as dimethylamine to give different regio-isomeric products. Hence, addition of bromide 1 with methylamine in THF at 0°C for 20 min. gave a 66% yield of displacement product 8. On the other hand, when 1 was treated with dimethylamine in THF at 0°C, a 96% yield of regioisomers 9 and 10 (a ratio of 1:1) was isolated. Compounds 9 and 10 were separated by silica gel column chromatography. The regiochemistry of 9 and 10 has not been determined. The presence of bromine atom in the products was indicated by their mass spectra in which the M+2 peaks (6Br isotope) almost equal in intensity to the molecular (EI) or quasimolecular (CI) ions. No other by-products are identifiable in these reactions. These unusual addition reactions are unprecedented. It has been reported that p-benzoquinone undergoes addition reactions with aliphatic primary amines to give a mixture of products including mono- and di-adducts from the 1,4-addition reactions, and tetrachloro-1,4-benzoquinone and 2,3-dichloro-1,4-naphthoquinone react with secondary amines to give displacement products (addition to the double bond followed by elimination of chlorine). Contrary to these literature results, bromide 1 undergoes displacement of the methoxy group instead of bromine with primary amines and simple 1,4-addition on the unsubstituted quinone ring followed by oxidation with secondary amines, exclusively. Presumably, a less basic primary amine, methylamine, undergoes 1,4-addition reaction on the greater electron-deficient bromoquinone ring. Since the C2-bromine of 1 is an electron-withdrawing group, the anion resulted from the addition of an amine on C3 would be more stabilized than that from the addition on C2; resulting a displacement of methoxy group to provide compound 8. On the other hand, a more basic secondary amine, dimethylamine, is less affected by electronic (since it is a more reactive amine) effect of the quinone ring and is more affected by steric effect, in turn it prefers the addition on the unsubstituted (less hindered) quinone moiety to provide compounds 9 and 10.

Due to the potent anticancer activity of amine 8 (vide infra) and a need of water soluble analogs, aliphatic primary amines containing ester function were used to synthesize various triptycene quinones, and their biological activities were studied (Scheme 16).
Treatment of bromide 1 with ethyl 2-aminopropanoate in THF at −40°C for 6 h afforded a 46% yield of displacement product 11 along with starting material 1 (30% recovery). No other regioisomers were detected. Similarly, reaction of 1 with t-butyl 2-aminopropanoate at −40°C for 8 h gave a 62% yield of 12. Recrystallization of 12 in hexane:ethyl acetate (5:1) afforded single crystals which structure was unequivocally shown by X-ray analysis. Deprotection of the t-butyl ester of 12 with trifluoroacetic acid in dichloromethane gave a quantitative yield of acid 13. Sodium salt 14, a water soluble material, was obtained from the treatment of 13 with 1 equiv of sodium hydroxide.

When compound 1 was treated with dimethylamine in THF (1 M solution in THF; purchased from Aldrich Chemical Company), a mixture of 9 and 10 (1:1) was obtained (Scheme 18). However, when compound 1 was treated with dimethylamine in THF which prepared by adding dimethylamine (a gas purchased from Linde Company) gas into THF, a mixture of 9, 10, TT24A, and TT24B (4:4:1:1) was obtained. Compounds 9, 10, and TT24 were separated by column chromatography. TT24A and TT24B were inseparable, however, carbon-13 NMR spectrum indicates two isomers.

Likely, a small of amounts of methylamine is contaminated in the dimethylamine gas, this methylamine then added to the unsubstituted quinone ring of triptycene bisquinone to produce TT24. This mode of addition is different from the reaction of 1 with pure methylamine. It is suggested that dimethylamine acts as a base and remove the proton of methylamine to provide methylamine anion which is more reactive than methylamine itself, and in turn undergoes addition reaction on the unactivated C=C of the quinone moiety.

Anticancer testing of TT24 show it has an IC₅₀ value of 48 nM (or 0.048 μM). The compound is as active as daunomycin.

Anticancer and antimalarial activities of triptycene bisquinones have been evaluated. Table 2 summarizes IC₅₀ values (the concentrations of drugs required to inhibit by 50% the viability of L1210 leukemic cells at day 4) of various triptycene bisquinones in the inhibition of L1210 leukemic cell growth. In general, IC₅₀ values are in the 0.24-0.43 μM range with the exception of carboxylic acid 13 and its sodium salt 14. Possibly, the passage ability of sodium salt 14 through cell membrane decreases as water solubility increases; in turn, ability in destroying cancer cells decreases. The IC₅₀ value of daunomycin, a known anticancer
cer drug, under similar treatment conditions is 0.041 μM. Triptycene bisquinones induce DNA cleavage and inhibit nucleoside transport. In contrast, daunomycin, inhibits topoisomerases I and II, and does not block nucleoside transport. Moreover, triptycene bisquinones 7, 8, 9, and ritoavir inhibit Plasmodium falciparum 3D7 (a malaria strain) with IC₅₀ values of 9.7, 23.6, and 0.10 μM, respectively. Presumably, these reactive triptycene bisquinones undergo addition reactions with lysine, tryptophan, histidine, and cysteine residues of proteins to produce cross-linked proteins.

### General Methods
Nuclear magnetic resonance spectra were obtained at 400 MHz for ¹H and 100 MHz for ¹³C in deuterochloroform, unless otherwise indicated. Infrared spectra are reported in wavenumbers (cm⁻¹). Mass spectra (M+); 'H NMR δ 8.70 (s, 2H, C₉,10H), 7.97 (dd, J=6.1, 3.6 Hz, 2H, C₅,8H), 7.40 (dd, J=6.6, 3.2 Hz, 2H, C₆,7H), 6.55 (s, 2H, C₂,3H), 3.97 (s, 6H, OCH₃); ¹³C NMR δ 149.5 (s, C₈a,10a), 131.5 (s, C₄a,9a), 128.5 (d, C₉,10), 125.5 (d, C₅,8), 120.7 (d, C₆,7H), 109.0 (d, C₂,3), 56.6 (s, OCH₃).

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**TABLE 2**

Cytotoxicities of triptycene bisquione analogs in L1210 leukemic cell system in vitro.

A solution of 1.00 g (7.10 mmol) of methoxyhydroquinone (3), 2.50 g (10.7 mmol) of silver oxide, and 1.20 g 3-aminopropanoate hydrochloric acid salt were purchased from Aldrich Chem. Co. Davisil silica gel, grade 643 (200-425 mesh), was used for the flash column chromatographic-separation. 1,4-Dimethoxyanthracene (2).

To a cold (0°C) methanol (20 mL) solution of 1.00 g (0.0042 mol) of quinizarin under argon was added 0.638 g (0.0168 mol) of sodium borohydride. The resulting mixture was stirred at 0°C for 1 h. To it, 11 mL of 6 N HCl was added dropwise at 0°C over a period of 10 min. The lH, =CH), 3.84 (s, 2H, OMe), 3.72 (s, 2H, OMe), 3.12 (s, 2H, OMe), 3.06 (s, 2H, OMe). More polar isomer: MS, FAB, m/z 239 (M+), 238 (M+).

**TABLE 3**

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A mixture of 1.00 g (7.10 mmol) of methoxyhydroquinone (3), 2.50 g (10.7 mmol) of silver oxide, and 1.20 g 3-aminopropanoate hydrochloric acid salt were purchased from Aldrich Chem. Co. Davisil silica gel, grade 643 (200-425 mesh), was used for the flash column chromatographic-separation. 1,4-Dimethoxyanthracene (2).

To a cold (0°C) methanol (20 mL) solution of 1.00 g (0.0042 mol) of quinizarin under argon was added 0.638 g (0.0168 mol) of sodium borohydride. The resulting mixture was stirred at 0°C for 1 h. To it, 11 mL of 6 N HCl was added dropwise at 0°C over a period of 10 min. The precipitated orange solids were collected, washed several times with distilled water, dried under vacuum, and recrystallized from acetone-ether to give 0.83 g (95% yield) of 4. The filtrate was concentrated to give 0.970 g (99% yield) of 4. The mixture was filtered, washed twice with water, and dried under vacuum. To a solution of 0.70 g (2.90 mmol) of 2 and 1.00 g (7.20 mmol) of 3 in 10 mL of toluene in a sealed tube was heated to 204-206°C. The filtrate was washed with aqueous NH₄Cl, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent gave 2.21 g (70% yield) of 4: mp 108-110°C.

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A mixture of 1.00 g (7.10 mmol) of methoxyhydroquinone (3), 2.50 g (10.7 mmol) of silver oxide, and 1.20 g 3-aminopropanoate hydrochloric acid salt were purchased from Aldrich Chem. Co. Davisil silica gel, grade 643 (200-425 mesh), was used for the flash column chromatographic-separation. 1,4-Dimethoxyanthracene (2).

To a cold (0°C) methanol (20 mL) solution of 1.00 g (0.0042 mol) of quinizarin under argon was added 0.638 g (0.0168 mol) of sodium borohydride. The resulting mixture was stirred at 0°C for 1 h. To it, 11 mL of 6 N HCl was added dropwise at 0°C over a period of 10 min. The precipitated orange solids were collected, washed several times with distilled water, dried under vacuum, and recrystallized from acetone-ether to give 0.83 g (95% yield) of 4. The filtrate was concentrated to give 0.970 g (99% yield) of 4. The mixture was filtered, washed twice with water, and dried under vacuum. To a solution of 0.70 g (2.90 mmol) of 2 and 1.00 g (7.20 mmol) of 3 in 10 mL of toluene in a sealed tube was heated to 150°C for 1 day. The mixture was filtered, washed twice with water, and dried under vacuum. To a solution of 0.70 g (2.90 mmol) of 2 and 1.00 g (7.20 mmol) of 3 in 10 mL of toluene in a sealed tube was heated to 150°C for 1 day. The mixture was filtered, washed twice with water, and dried under vacuum. To a solution of 0.70 g (2.90 mmol) of 2 and 1.00 g (7.20 mmol) of 3 in 10 mL of toluene in a sealed tube was heated to 150°C for 1 day. The mixture was filtered, washed twice with water, and dried under vacuum.
of KOH. The solution was stirred at 25°C for 1 h, acidified with 1 N HCl, and extracted three times with dichloromethane. The combined extract was washed with brine, dried (MgSO₄), and concentrated to give 0.77 g (100% yield) of 1,4-dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene. MS, m/z 376 (M+). ¹H NMR d 7.43 (dd, J=5.5, 3 Hz, 1H), 7.38 (dd, J=5.5, 3 Hz, 1H), 6.96 (dd, J=5.5, 3 Hz, 2H), 6.51 (s, 2H, C₆H₂), 6.29 (s, 1H, C₃H), 6.09 (s, 1H), 5.98 (s, 1H, OH), 4.96 (bs, 1H, OMe). 3.81 (s, 3H, OMe), 3.805 (s, 3H, OMe), 3.66 (s, 3H, OMe). This compound was used in the next step.

To a mixture of 0.77 g (2.0 mmol) of 1,4-dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene and 0.60 g (4.2 mmol) of sodium sulfate (anhydrous) in 15 mL of dried acetone under argon at 25°C was added 0.56 g (4.0 mmol) of silver oxide. The mixture was heated under reflux for 6 h, cooled to 25°C, diluted with dichloromethane, and filtered through Celite. The filtrate was concentrated to give 109 mg (quantitative yield) of 2-bromo-3,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (11).

To a solution of 0.200 g (0.473 mmol) of 1 in 2 mL of THF under argon at 0°C, was added 0.24 mL (0.473 mmol) of dimethylamine (2.0 M in THF) in 1 mL of THF was stirred under argon at 0°C for 1 h. The solution was concentrated to dryness, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent to give 0.132 g (66% yield) of 6: mp. 142-152°C. MS, CI m/z 424, 422 (-1:1; M+1); ¹H NMR d 7.47-7.43 (m, 2H, Ar-H), 7.05-7.01 (m, 2H, C₆H₂), 6.64 (s, 2H, C₆H₂), 6.10 (s, 1H), 5.88 (bs, 1H, NH), 3.33 (s, J=5.6 Hz, 3H, MeN); ¹³C NMR (acetone-d₆) d 183.3, 183.3, 178.2, 170.9, 154.4, 152.3, 152.0, 148.0, 147.0, 143.7, 143.5, 136.4, 136.3, 130.7, 126.0, 125.7, 125.0, 44.3, 42.9, 33.2. Anal. Calcd for C₂₀H₁₆BrN₂O₂: C, 59.74; H, 2.86. Found: C, 59.33; H, 2.87. C₂₀H₁₆BrO₂ N₂, M-H').

2-Bromo-3-methoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (8). A solution of 0.200 g (0.473 mmol) of 1 and 0.47 mL (0.946 mmol) of methylene (2.0 M in THF) in 1 mL of THF was stirred under argon at 0°C for 1 h. The solution was concentrated to dryness, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent to give 0.132 g (66% yield) of 8: mp. 142-152°C. MS, CI m/z 424, 422 (-1:1; M+1); ¹H NMR d 7.47-7.43 (m, 2H, Ar-H), 7.05-7.01 (m, 2H, C₆H₂), 6.64 (s, 2H, C₆H₂), 6.10 (s, 1H), 5.88 (bs, 1H, NH), 3.33 (s, J=5.6 Hz, 3H, MeN); ¹³C NMR (acetone-d₆) d 183.3, 183.3, 178.2, 170.9, 154.4, 152.3, 152.0, 148.0, 147.0, 143.7, 143.5, 136.4, 136.3, 130.7, 126.0, 125.7, 125.0, 44.3, 42.9, 33.2. Anal. Calcd for C₂₀H₁₆BrN₂O₂: C, 59.74; H, 2.86. Found: C, 59.74; H, 2.76. 2-Bromo-3-methoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (9)

To a solution of 0.200 g (0.473 mmol) of 9 in 2 mL of THF under argon at 0°C, was added 0.24 mL (0.473 mmol) of dimethylamine (2.0 M in THF). After stirring at 0°C for 2 h, the reaction solution was concentrated to dryness, and column chromatographed on silica gel using a mixture of benzene and ethyl acetate (10:1 as solvent to give 0.099 g (48% yield) of 9 (less polar isomer; the regiochemistry has not been determined) and 0.098 g (48% yield) of 10 (more polar).
of 1 in 1 mL of DMF was added via cannula. The solution was stirred at 46°C for 6 h, diluted with a mixture of ethyl acetate and benzene (1:1), washed twice with water, brine, dried (Na₂SO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as the eluent to give compounds 1, 2, and TT24.

Compound TT24: 1H NMR (CDCl₃) δ 7.44-7.40 (m, 2H), 7.03 (m, 2H), 6.27 (s, 1H), 6.15 and 6.12 (2 s, 1H), 5.69 (q, J=5.6 Hz, NH), 5.26 (s, 1H), 3.86 & 3.79 (2s, 3H, OMe), 2.79 (d, J=5.6 Hz, 3H, NMe); 13C NMR (CDCl₃) δ 181.7, 181.6, 179.5, 156.2, 147.8, 147.6, 143.9, 143.8, 143.6, 142.3, 141.8, 139.6, 139.1, 131.2, 126.9, 125.52, 125.49, 125.46, 125.4, 124.8, 124.75, 124.3, 124.2, 102.3, 95.8, 95.7, 61.1, 60.4, 42.2, 41.9, 41.3, 41.0, 29.3 (2C).

Synthesis of Other N Analogs and S Analogs

As illustrated in Scheme 17, nitrogen analog 1 has been prepared. Treatment of TT13 with 1 equivalent each of L-lysine hydrochloride and sodium hydride in a 1:1 mixture of THF and DMF under argon at room temperature gave amino acid analog 1 which is a water soluble drug.


Preparation of TT24:

To 20 mL of THF solution, 1 g (0.022 mol) of dimethylamine gas (from a gas cylinder purchased from Linde Company) was added. A 1.1 M of dimethylamine in THF was used. To a cold (0°C) solution of 0.081 g (0.19 mmol) of triptycene bisquinone 1 in 2 mL of THF under argon was added 0.17 mL (0.19 mmol) of dimethylamine. The solution turn to purple immediately and was stirred at 0°C for 10 min. The solvent was removed on a rotary evaporator and then under vacuum and the crude product was column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as the eluent to give compounds 1, 2, and TT24.

Compound TT24: 1H NMR (CDCl₃) δ 7.44-7.40 (m, 2H), 7.03 (m, 2H), 6.27 (s, 1H), 6.15 and 6.12 (2 s, 1H), 5.69 (q, J=5.6 Hz, NH), 5.26 (s, 1H), 3.86 & 3.79 (2s, 3H, OMe), 2.79 (d, J=5.6 Hz, 3H, NMe); 13C NMR (CDCl₃) δ 181.7, 181.6, 179.5, 156.2, 147.8, 147.6, 143.9, 143.8, 143.6, 142.3, 141.8, 139.6, 139.1, 131.2, 126.9, 125.52, 125.49, 125.46, 125.4, 124.8, 124.75, 124.3, 124.2, 102.3, 95.8, 95.7, 61.1, 60.4, 42.2, 41.9, 41.3, 41.0, 29.3 (2C).

Similarly, glucosamine hydrochloride car, be used to prepare glucosamine analog (of 2; instead of galactosamine analog).
Syntheses of nitrogen analogs 3 and 4 are depicted in Scheme 20. Protection of N-9 of adenine (7) with (trimethylsilyl)ethyl chloromethyl ether (SEM-Cl) and sodium hydride in DMF produces compound 8. Addition of 8 with TT3 in THF at -40°C followed by removal of the SEM protecting group with tetra-n-butylammonium fluoride in THF furnishes nitrogen analog 3. Direct treatment of adenine and catalytic amount of sodium hydride in THF at -40°C gives nitrogen analog 4. The N9 hydrogen of adenine (7) is more acidic than that of C6 amine group, hence, N9 nitrogen should react with TT3.

Sulfur analogs: Uses of thiol containing amino acid such as N-protected L-cysteine provides sulfur analog (Scheme 21). Hence, addition of N-Boc cysteine and sodium hydride in DMF-THF followed by trifluoroacetic acid (removal of the Boc protecting group) will afford sulfur analog.
The above four nitrogen analogs and the sulfur analog are water soluble materials. Other nitrogen and sulfur analogs are prepared by methods known to one of ordinary skill in the art, in using substitutions which are known to one of ordinary skill in the art.

Compounds of this invention other than those particularly shown may be prepared without undue experimentation by those skilled in the art of synthetic chemistry by methods analogous to, those specifically disclosed herein or in publications and patent applications incorporated by reference or by methods known in the art.

Although the description above contains many specificities these should not be construed as limiting the scope of the invention, but merely providing illustrations of some of the presently preferred embodiments of the invention. Thus, the scope of the invention should be determined by the appended claims and their legal equivalents, rather than by the examples given. All references cited herein are incorporated by reference to the extent not inconsistent with the disclosure herein.

Literature References in Synthesis Section:
7. Hamon, D. P. G.; Spurr, P. R. Reductive elimination of bromine from 2,3-disubstituted 1,4-dibromo-2-butenes by iodide ion: a convenient route to 2,3-bis[jodomethyl]-1,3-butadiene and related compounds. *Syntheses* 1981, 873–874.

What is claimed is:
1. A triptycene analog comprising a compound of formula:

\[
R^1\, R^2\, R^3\, R^4\, R^5
\]

wherein

\[
X \text{ is selected from the group consisting of: } H, R, SR \text{ and } NR_2^+.
\]
Y is selected from the group consisting of: halogen, R, 
NR₂, SR and H;
R and R¹-² are independently selected from the group 
consisting of: H, halogen, OR, and hydrocarbyl;
R³-⁴, independently of one another, are selected from the 
group consisting of: H, bromine, R, SR, and NR₂;
R⁵, independently of other R⁵s, is selected from the group 
consisting of: ==O, ==N—OH and ==CHR; and reduced forms thereof, wherein in reduced forms, either 
ring A or ring C or both is replaced with

and wherein reduced form, each R⁵ is 
independently H, Cl–C₈ alkyl or —OR; 
and pharmaceutically acceptable salts of the foregoing, as 
well as optical isomers thereof; 
wherein when all of R⁵ are ==O, at least one of X, Y, 
R¹—R⁴ is not H.
2. The triptycene analog of claim 1 having the formula:

3. The triptycene analog of claim 1 having the formula:

4. The triptycene analog of claim 1, wherein:
X is selected from the group consisting of: H, OMe and 
CO₂Me;
Y is selected from the group consisting of: H, Br, and 
OMe;
R¹, R², R³ and R⁴ are all H; and
R⁵ is, independently of other R⁵s, selected from the group 
consisting of: OH, OMe, ==O, and H.

5. A triptycene analog having the formula:

wherein
X is selected from the group consisting of: H, R, SR and 
NR₂;
Y is selected from the group consisting of: halogen, NR₂, 
R, SR and H;
R and R¹-², are independently selected from the group 
consisting of: H, halogen, OR, and hydrocarbyl;
R³-⁴, independently of one another, are selected from the 
group consisting of: H, bromine, R, SR, and NR₂;
R⁵, independently of other R⁵s, is selected from the group 
consisting of: ==O, ==N—OH and ==CHR; 
R²¹ and R²² are independently selected from the group 
consisting of: H, R, and OR; and reduced forms thereof and pharmaceutically acceptable 
salts of the foregoing, as well as optical isomers thereof.
6. A triptycene analog having the formula:

wherein
R⁷ is selected from the group consisting of: R, halogen, 
NR₂, SR, and H;
R⁸ is selected from the group consisting of: H, R, SR and 
NR₂;
R⁷ and R⁸ are independently selected from the group 
consisting of: H, halogen, and hydrocarbyl;
R⁷ and R⁸ are independently selected from the group 
consisting of: H, bromine, R, SR, and NR₂;
R¹⁰ and R¹⁰ are, independently of one another, H, R, or 
OR;
(R⁹ and R¹⁰) and (R¹¹ and R¹²) and (R¹³ and R¹⁴) and 
(R¹⁵ and R¹⁶) are independently together ==O or are 
independently H or —OR;
R is selected from the group consisting of: H, halogen, 
OR, and hydrocarbyl; and reduced forms thereof; 
and pharmaceutically acceptable salts of the foregoing, as 
well as optical isomers thereof.
7. A method of making a compound of claim 1, comprising: heating an optionally substituted anthracene with an optionally substituted quinone with silver oxide.

8. The method of claim 7, further comprising adding zinc iodide.

9. The method of claim 7, wherein the optionally substituted anthracene has the formula:

```
   OR
   OR
```

and the optionally substituted quinone has the formula:

```
   OR
   OR
```

where R is H or hydrocarbyl.

10. A method of brominating a triptycene derivative of formula (I):

```
   R1
   R2
   R3
   R4
   R5
```

wherein X is selected from the group consisting of: H, R, SR and NR₂;
Y is selected from the group consisting of: halogen, R, NR₂, SR and H;
R and R₁-₅ are independently selected from the group consisting of: H, halogen, OR, and hydrocarbyl;
R₂-₄, independently of one another, are selected from the group consisting of: H, bromine, R, SR, and NR₂;
R₅, independently of other R₅s, is selected from the group consisting of: =0, =N—OH and =CHR; and reduced forms thereof, wherein in reduced forms, either ring A or ring C or both is replaced with

11. The method of claim 10, wherein the triptycene derivative is:

```
   Br
   MeO
```

12. A triptycene analog of claim 1, wherein at least one of X, Y, R₁ and R₂ is selected from the group consisting of: a nitrogen containing group, a water soluble group, and a sulfur containing group.

13. The compound of claim 12, wherein X is —NR₂.

14. The compound of claim 12, wherein R₂ is —NR₂.

15. The compound of claim 14, wherein R₂ is —NMe₂.

16. The compound of claim 12, wherein at least one of X, Y, R₁ and R₂ is selected from the group consisting of: amine, amino acid and amine sugar.

17. The compound of claim 12, wherein X is —NH—(CH₂)ₙ—CO₂R, where n is an integer from 0 to 8, and R is as defined in claim 12.

18. The compound of claim 17 wherein R is H.

19. The compound of claim 12, wherein one or more of X, Y, R₁ and R₂ contains an optionally substituted nitrogen containing hydrocarbyl group.

20. The compound of claim 19, wherein the optionally substituted nitrogen containing hydrocarbyl group is a fused ring structure.

21. The compound of claim 12, wherein X is a sulfur containing group.

22. The compound of claim 21, wherein the sulfur containing group also contains one or more N atoms.
23. The compound of claim 12 having the formula:

24. The compound of claim 12 having the formula:

25. The compound of claim 12 having the formula:

26. The compound of claim 12 having the formula:
31. The compound of claim 12 having the formula:

32. The triptycene analog of claim 12 having the formula:

33. A triptycene analog comprising a compound of formula:

34. The triptycene analog of claim 5, wherein at least one of X, Y, R¹ and R² is selected from the group consisting of: a nitrogen containing group, a water soluble group, and a sulfur containing group.

35. The compound of claim 34 wherein at least one of R²¹ and R²² is —CO₂R.

36. The compound of claim 12 which blocks nucleoside transport, induces DNA fragmentation, inhibits nucleic acid synthesis, inhibits protein synthesis, decreases the proliferation of cancer cells, or decreases the viability of cancer cells.

37. The triptycene analog of claim 6, wherein at least one of R³, R⁴, R⁵ and R⁶ is selected from the group consisting of: a nitrogen containing group, a water soluble group, and a sulfur containing group.

38. A method of making a nitrogen-containing compound of claim 12, comprising: reacting a triptycene derivative of formula:

39. A triptycene analog of formula:

40. And the reduced forms thereof, wherein in said reduced forms, either ring A or ring C or both is reduced to

41. Wherein all but one of X, Y, R¹ and R² is independently selected from the group consisting of: H, C₁–C₆ alkyl, C₁–C₆ alkenyl, OR, SR or NR²;

42. And the reduced forms thereof, wherein wherein each R is independently H or C₁–C₆ alkyl and the other R¹ or R² is a solubilizing group; and each R⁵ is independently H, C₁–C₆ alkyl or OR.
40. The triptycene analog of claim 39, wherein the solubilizing group is of the formula:

NR(CR₂₃)₃X wherein X is a sugar, R, COR, COOR, CONR₂, OOCR and NRCOR; R is independently

selected from the group consisting of H, C₁–C₈ alkyl and C₁–C₈ alkenyl; n is an integer from 1 to 8.

* * * * *
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Title Page:
Under heading "References Cited" under "U.S. PATENT DOCUMENTS", please add the following
-5,958,970 9/1999 Hua et al..............514/555--.

In the Specification:
Column 5, line 52, replace "TT2:X = PMe, Y = H" with "TT2:X = OMe, Y = H".

Column 6, lines 28-50, move the structures

N analog 3: Y = Br, X =

N analog 4: X = OMe, Y = Br, R₂ =

S analog 9: Y = Br, X = SCH₂CH(CO₂H)NH₂" to the bottom of column 5 after X =.
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Column 7, lines 13-27, replace the following structure

```
TT4: \( R^9 = R^{10} = R^{11} = R^{12} = R^{13} = R^{14} = R^{15} = R^{16} = O \)
TT12: \( R^{9} = R^{11} = R^{13} = R^{15} = H, R^{10} = R^{12} = R^{14} = R^{16} = OH \)
```

with

```
TT4: \( R^9 = R^{10} = R^{11} = R^{12} = R^{13} = R^{14} = R^{15} = R^{16} = O \)
TT12: \( R^{9} = R^{11} = R^{13} = R^{15} = H, R^{10} = R^{12} = R^{14} = R^{16} = OH \)
```
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Column 15, line 52, replace "combinations thereof" with -- combinations thereof. --.

Column 34, lines 57-65, Scheme 3, insert -- + -- after the structure.

Column 37, lines 23-63, Scheme 6, replace all structures shown in Scheme 6 with --

Column 40, line 24, underneath the arrow after structure 15, insert -- toluene, reflux 5 h --.

Column 41, line 32, Scheme 10, under the first structure, insert -- TT1 --.

Column 53, line 11, replace "3C4a," with -- C4a, --.
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Column 66, lines 38-64, Scheme 17, replace the entire Scheme with the following

---

![](image)

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Signed and Sealed this

Twenty-seventh Day of February, 2007

JON W. DUDAS

Director of the United States Patent and Trademark Office