Predicting Bone Mechanical State During Recovery After Long-Duration Skeletal Unloading Using QCT and Finite Element Modeling

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February 2013
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Abstract

During long-duration missions at the International Space Station, astronauts experience weightlessness leading to skeletal unloading. Unloading causes a lack of a mechanical stimulus that triggers bone cellular units to remove mass from the skeleton. A mathematical system of the cellular dynamics predicts theoretical changes to volume fractions and ash fraction in response to temporal variations in skeletal loading. No current model uses image technology to gather information about a skeletal site’s initial properties to calculate bone remodeling changes and then to compare predicted bone strengths with the initial strength. The goal of this study is to use quantitative computed tomography (QCT) in conjunction with a computational model of the bone remodeling process to establish initial bone properties to predict changes in bone mechanics during bone loss and recovery with finite element (FE) modeling. Input parameters for the remodeling model include bone volume fraction and ash fraction, which are both computed from the QCT images. A non-destructive approach to measure ash fraction is also derived. Voxel-based finite element models (FEM) created from QCTs provide initial evaluation of bone strength. Bone volume fraction and ash fraction outputs from the computational model predict changes to the elastic modulus of bone via a two-parameter equation. The modulus captures the effect of bone remodeling and functions as the key to evaluate changes in strength. Application of this time-dependent modulus to FEMs and composite beam theory enables an assessment of bone mechanics during recovery. Prediction of bone strength is not only important for astronauts, but is also pertinent to millions of patients with osteoporosis and low bone density.

Introduction

Microgravity decreases the loading on the skeletal structure, which causes bone loss in astronauts. Upon return to Earth’s 1 g environment, astronauts may encounter serious health problems including increased risk of fractures and early onset of osteoporosis (Ref. 1). Research studies have been conducted on the adaptation to skeletal reloading during recovery of spaceflight-induced bone loss (Refs. 1 and 2). They have focused on analyzing data directly before and after flight and 1 year post flight. A computational bone loss and recovery model has the potential to predict the changes that occur continuously to the skeleton over a specific temporal span.

The Digital Astronaut Project (DAP) in the NASA Human Research Program is developing a model of the bone remodeling process to help predict and assess bone loss during spaceflight and bone recovery post-flight. The mathematical formulation describes the mechanisms involved in the removal and replacement of bone by basic multicellular units (BMU) in the human adult (Ref. 3). A system of differential equations governs the rate of change in cell populations and the rate of change of bone volume fractions (BVF) in trabecular and cortical bone. Based on the biology of remodeling, the cellular dynamics is controlled by
hormones, protein receptors, ligands, and other biochemical effects acting in response to mechanical stimuli from changes in skeletal loading. The mechanical stimulus is modeled with Frost’s mechanostat theory which identifies three strain ranges: low stimulus leading to bone loss, physiologic stimulus leading to no change in bone, and overload stimulus leading to bone formation (Refs. 4 and 5). The computational model is aimed toward computing temporal changes in BVFs and ash fraction due to imbalances caused by lack of sufficient skeletal loading and osteogenic benefit of exercise induced skeletal loading. Ash fraction is a measure of the amount of ash mass in dry bone mass (Refs. 6 and 7). Current work is progressing on modeling a representative volume of trabecular bone (Refs. 3 and 8).

As yet the modeling work has not included an analysis of the changes in bone strength as a result of the bone loss and adaptation predicted by the computational model. This study combines the computational model with the FE method to analyze changes to the bone structure and mechanical integrity under normal 1 g loading conditions such as stance and fall. The FE approach improves the computational model for two reasons.

1. An evaluation of the bone structure after x days of recovery is necessary to determine how well the bone will perform under certain loading conditions and if performing daily tasks will increase the likelihood of serious bone problems such as fractures.
2. Cody et al. demonstrated how the FE method better predicts fracture load than imaging tools including dual energy x-ray absorptiometry (DXA) or QCT (Ref. 9).

The objective is to establish a methodology to utilize QCT image technology to obtain initial bone strength and to initialize the computational cellular model to obtain time histories of bone volume fraction and ash fraction to evaluate bone strength during and post flight. The proximal femur is an ideal skeletal site to simulate the remodeling process because a large degree of bone loss occurs at this site during space missions (Ref. 10 and 2). However, no QCT of the proximal femur was obtained during the time of the study. Images of the eighth thoracic (T8) vertebral body were available instead. Lumbar vertebrae would have been preferred since that is where bone loss occurs. Although the thoracic region of the spine does not exhibit significant bone loss like the lumbar region and is not the primary load bearing site in the spine, it is the most likely to fracture spontaneously (Ref. 11).

**Methods**

QCT scans of bone specimens provide initial conditions of the BVF and ash fraction for the computational model. The computation scenario simulates bone remodeling for 20 days at steady state and then 180 days in a microgravity environment followed by a year of recovery after return to a 1 g environment. The output from the model gives a time history of the BVF and ash fraction, which are the parameters used to calculate a time history of mechanical properties. Given the time history of elastic modulus, composite beam theory analysis and FE modeling are used to assess predicted changes in the biomechanics of the bone samples. Custom-written MATLAB (Mathworks Inc., Natick, Massachusetts) programs are used to process the QCT scans, to simulate bone remodeling, to create FEMs, and to analyze the results.

**QCT Image Processing**

A K$_2$HPO$_4$ calibration phantom (Mindways Software Inc., Austin, Texas) is scanned with the QCTs so that sample QCT density can be calculated by comparison with the known phantom densities. The phantom has five chambers with mineral densities of 51.8, 53.4, 58.9, 157.0, and 375.8 mg/cc. A phantom calibration curve is plotted of the tube mineral densities versus their corresponding mean intensity values. Regression equations are also plotted to determine a relation to calculate the mineral density of each voxel (Figure 1). A two-part linear regression is chosen over the second order polynomial because it better fits the calibration curve.
T8 vertebrae QCT scans (0.46 mm resolution) are from a previous study on the differences in spine morphology of humans between other hominoids (Ref. 11). The posterior elements were not scanned in any of the specimens. Each specimen QCT is converted into a three-dimensional (3-D) matrix where each cell contains an intensity value in Hounsfield Units. Since samples were scanned in a 20 percent ethanol bath, the noise from the ethanol is removed by subtracting one standard deviation of the region of interest (ROI) from all the pixels (Figure 2(a) to (b)). The same region is used to calculate the mean intensity of the background or threshold value. Any intensity above this value is considered bone (Figure 2(c)). The final step in image processing is to remove any unconnected pixels from the image (Figure 2(c) to (d)).

![Figure 1. A two-part linear calibration curve is chosen to calculate QCT density of the T8 vertebrae.](image1)

![Figure 2. From left to right: (a) Original bone image of HTH0439, (b) after removing noise of 20 percent ethanol, (c) after removing background, and (d) after removing unconnected regions from image. The blue box is the ROI selected to calculate the ethanol and background intensity values.](image2)
Material Properties

The non-zero cells in the 3–D matrix contain intensity values that represent voxels or 3–D pixels of bone. These intensity values are converted into QCT densities via the calibration curve. Ash densities are then determined from the QCT densities using relations based on experimental studies (Table 1). An all bone correlation for QCT density and ash density is appropriate for two reasons. First, it is difficult to distinguish between cortical and trabecular bone in the QCT images. The all bone and trabecular bone correlations are also similar for a range of QCT densities (Appendix E). No comparison is made of the cortical bone relation with the others because the sample size was not significant (Ref. 12).

<table>
<thead>
<tr>
<th>Bone</th>
<th>Correlation</th>
<th>QCT Type</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trabecular</td>
<td>$\rho_{\text{ash}} = 0.839\rho_{\text{QCT}} + 69.8$</td>
<td>CHA</td>
<td>7</td>
</tr>
<tr>
<td>Cortical</td>
<td>$\rho_{\text{ash}} = 0.290\rho_{\text{QCT}} + 80.6$</td>
<td>CHA</td>
<td>13</td>
</tr>
<tr>
<td>All</td>
<td>$\rho_{\text{ash}} = 0.887\rho_{\text{QCT}} + 63.3$</td>
<td>CHA</td>
<td>14</td>
</tr>
<tr>
<td>All</td>
<td>$\rho_{\text{ash}} = 0.953\rho_{\text{QCT}} + 45.7$</td>
<td>K$_2$HPO$_4$</td>
<td>15</td>
</tr>
</tbody>
</table>

The BVF can be calculated knowing the location of the bone voxels in the 3–D matrix. The whole specimen BVF and voxel specific ash densities are integral parameters of the computation model of the bone remodeling process.

Initializing Mechanotransduction Model

The computational bone remodeling model can initiate a simulation of bone adaptation either by starting with an initial BVF or with initial volume fractions of pure osteoid, $O_0$, unmineralized bone, and fully mineralized mature bone, $M_0$. Since the elastic modulus is used to evaluate mechanical state during recovery and is a function of ash fraction, the latter initialization is implemented because it provides a time history of ash fraction.

The osteoid and mineralized bone fractions are calculated using the definitions of BVF and ash fraction (Eqs. (1) and (2)). BVF is made of two parts: the osteoid and mature bone volume fractions. Ash fraction is a function of the osteoid and mature bone fractions as well as the osteoid density $D_o$ and the mature bone density $D_m$ (Refs. 3 and 8). Either volume fractions is calculated as a function of initial BVF and ash fraction. The other is equivalent to initial BVF minus the calculated volume fraction. Both initial values of $BVF$, $BVF_0$ and ash fraction, $\alpha_0$, are calculated using the 3–D matrix from image processing.

$$BVF_0 = O_0 + M_0$$

$$\alpha_0 = \frac{M_0D_m0.7}{M_0D_m+O_0D_o}$$

The QCT images are used to calculate the BVF as the ratio of bone volume to total volume. For a cubic volume sample, it is equal to the number of filled cells divided by the total number of cells in the 3–D matrix. BVF for a whole bone sample such as the proximal femur or vertebra is determined using built-in MATLAB functions ‘strel’ and ‘imclose’. ‘imclose’ encloses a region of numbers in a two-dimensional (2–D) matrix using the ‘strel’ function as a marker (Figure 3). The sum of the voxels within these regions equals the total number of cells enclosed in the specimen. There is a problem with this technique if the bone sample is too close to the edge of the boundaries (Figure 4). However, this small additional region does not affect the BVF value significantly.
Figure 3.—HTH561 slice 1 demonstrating the use of ‘strel’ and ‘imclose’ to determine the BVF of the vertebra. From left to right: the original layer, enclosed region using disc size 5, and enclosed region using disc size 15.

Figure 4.—HTH0439 slice 20 shows the problem when the bone image is too close to the edge of the boundaries. A smaller ‘strel’ marker cannot be used without compromising unmarked void spaces within the vertebra.

Ash mass is measured after the bone samples are ashed following removal of bone marrow and remaining fat. This experimental approach to measure ash fraction cannot be applied to this non-invasive methodology, so a theoretical equation is derived to calculate ash fraction (Appendix C). First, the bone intensity values are converted to QCT densities (Eq. (3)). Voxel ash densities are then determined from the QCT density and ash density relation for K2HPO4 phantom (Eq. (4)). Knowing the voxel specific ash densities and whole bone BVF, ash fraction is calculated for each voxel with Equation (5). An average ash fraction is then taken as the initial value for the mechanotransduction model.

\[
\rho_{QCT} \left[\text{mg cm}^{-3}\right] = \begin{cases} 
0.0495x \text{ [HU]} + 3.8975, & 0 < x \leq 1098.8 \\
1.6x \text{ [HU]} - 1716.6, & x > 1098.8 
\end{cases} 
\]  
\tag{3}

\[
\rho_{ash} \left[\text{mg cm}^{-3}\right] = 0.953 \rho_{QCT} + 45.7 
\]  
\tag{4}

\[
1.29\alpha^2 + 1.41\alpha - \frac{\rho_{ash} \left[\text{g cm}^{-3}\right]}{\text{BVF}} = 0 
\]  
\tag{5}

Ash fraction observed in human bone spans from 0 for pure osteoid to 0.7 for fully mineralized bone (Ref. 6). An intensity value associated with an ash fraction of 0.7 is determined (Eq. (6), Appendix C). Any intensity value exceeding the maximum is regarded as an anomaly, and its value set to zero.

\[
I_{\text{max}} = \frac{1}{m} \left( \frac{\text{BVF}_0 (1.29\alpha_0^2 + 1.41\alpha_0) - 0.0457}{0.00953} \right) - b 
\]  
\tag{6}

**Prediction of Bone Biomechanics**

Evaluation of the bone mechanics during flight and recovery is dependent on the change in each voxel’s elastic modulus. A two-parameter equation is used to predict the modulus (Eq. (7)) (Ref. 6). The time histories of BVF and ash fraction are obtained from the mechanotransduction model (Ref. 8).
\[ E(t) = 84.37 \text{ BVF}(t)^{2.58} \alpha(t)^{2.74} \text{ [GPa]} \]  

(7)

Each voxel has a constant Poisson’s ratio of 0.3 and a specific homogenous isotropic modulus dependent on changes in BVF and ash fraction. This definition of material properties leads to heterogeneous model of the vertebra (Figure 5). Prediction of structural integrity can be analyzed with an analytical method using composite beam theory or a computational approach using the FEMs.

Composite beam theory is an analytical method to estimate the stresses and deflections in beams with heterogeneous material properties. It can be used to calculate an effective stiffness or rigidity that accounts for geometry and material properties (Eq. (8)). The axial rigidity is calculated for each transverse cross-section of the vertebra. The minimum rigidity is used as an indication of bone strength because it is the weakest location. The calculation is repeated at 10 day increments during bone loss and recovery to observe the changes in axial rigidity (Ref. 16).

Axial composite rigidity = \( \sum E_i A_i \)  

(8)

FEMs are generated from the 3–D matrices to predict the effective strain under normal 1 g loading conditions during bone loss and recovery. Each voxel is converted into 8-noded linear brick elements with size of 0.46- by 0.46- by 0.75-mm. The corners of each voxel represent nodal coordinates from which element connectivity is determined. Boundary conditions are applied to simulate the forces applied to the lower thoracic region. Two nodes are fixed at the distal end of the vertebra. The remaining nodes on this surface are restricted along the z-direction, but allowed to move along the xy-plane. A compressive force of 250 N representative of relaxed standing is distributed to all nodes on the proximal surface (Ref. 17). SIMULIA Abaqus Standard Analysis (Dassault Systèmes, Waltham, Massachusetts) is used to determine strain and stress distributions at a specific time point in the simulation. An effective strain based on von Mises criterion is calculated based on the Abaqus results (Eq. (9)).

\[ \bar{\varepsilon} = \frac{\sqrt{2}}{3} \left[ (\varepsilon_1 - \varepsilon_2)^2 + (\varepsilon_2 - \varepsilon_3)^2 + (\varepsilon_3 - \varepsilon_1)^2 \right]^{\frac{1}{2}} \]  

(9)

Figure 5.—FEM of HTH0243 T8 Vertebra demonstrates the heterogeneity of the model. Each color represents a different elastic modulus value calculated based on the image intensity value.
Results

Time histories of osteoid and mature bone volume fractions show a continuous loss during complete unloading from 20 to 200 days (Figure 6). Pure osteoid bone is added to the vertebrae following return to a 1 g environment and plateaus toward the end of the simulation. Approximately 100 days after mission completion, increase in mineral content results in an increase in mature bone volume fraction. The changes in the two volume fractions correlate well with the overall change in BVF and ash fraction.

The cellular response is sensitive to the loading condition during the unloading period. A simulation of 90 percent unloading demonstrates that there is notably less bone loss (Figure 7). Only approximately 0.017 relative loss occurs in BVF compared to the 0.05 loss seen during zero applied loads. The results of 90 percent unloading suggest that some degree of loading in a microgravity environment does stimulate the BMUs to remove less bone mass.

Figure 6.—Time histories of osteoid and mature bone volume fraction, ash fraction, and BVF for complete unloading from the mechanotransduction model of bone remodeling in T8 vertebral bodies.
Vertebral axial strength exhibits an average loss of 20 percent during complete unloading (Figure 8). Recovery of mechanical strength is particularly small, so it would be difficult to completely return to pre-unloading conditions under daily 1 g normal loading. Loading at 10 percent of normal conditions helps prevent loss of bone mass and strength where only a 10 percent loss is seen in axial stiffness.
Ash fraction is relatively low. It is expected to see a higher ash fraction or mineralized bone content for the donor age range. Calculation of ash fraction is dependent on two parameters, the BVF and ash density. Initial BVF is on average about 0.75, a reasonable value for healthy vertebral bodies. The low ash fraction also affects the osteoid and mature bone volume fractions: the lower the ash fraction, the higher the osteoid volume fraction and vice versa (Table 2).

**TABLE 2.—BONE REMODELING RESULTS OF T8 VERTEBRAE**

<table>
<thead>
<tr>
<th>Specimen no.</th>
<th>Age</th>
<th>Body mass, kg</th>
<th>(a_{\text{mean}})</th>
<th>(\Delta a)</th>
<th>(BVF_0)</th>
<th>(M_0)</th>
<th>(O_0)</th>
<th>(% \Delta BVF)</th>
<th>(S_{\text{axial}}), kN</th>
<th>(% \Delta S_{\text{axial}}), kN</th>
</tr>
</thead>
<tbody>
<tr>
<td>HTH0243</td>
<td>40</td>
<td>63.50</td>
<td>0.255</td>
<td>0.017</td>
<td>0.768</td>
<td>0.215</td>
<td>0.553</td>
<td>-4.30</td>
<td>7344</td>
<td>-22.11</td>
</tr>
<tr>
<td>HTH0249</td>
<td>40</td>
<td>56.60</td>
<td>0.264</td>
<td>0.018</td>
<td>0.756</td>
<td>0.218</td>
<td>0.538</td>
<td>-4.37</td>
<td>8610</td>
<td>-19.94</td>
</tr>
<tr>
<td>HTH0339</td>
<td>38</td>
<td>81.65</td>
<td>0.241</td>
<td>0.013</td>
<td>0.719</td>
<td>0.188</td>
<td>0.531</td>
<td>-3.48</td>
<td>5709</td>
<td>-18.32</td>
</tr>
<tr>
<td>HTH0439</td>
<td>35</td>
<td>54.43</td>
<td>0.300</td>
<td>0.009</td>
<td>0.716</td>
<td>0.174</td>
<td>0.542</td>
<td>-3.21</td>
<td>2085</td>
<td>-16.07</td>
</tr>
<tr>
<td>HTH0561</td>
<td>25</td>
<td>56.25</td>
<td>0.300</td>
<td>0.017</td>
<td>0.869</td>
<td>0.273</td>
<td>0.595</td>
<td>-3.91</td>
<td>9275</td>
<td>-19.91</td>
</tr>
</tbody>
</table>

Cubes of trabecular bone were sampled from the T8 vertebrae. The sample size is chosen to be 1 cm cubes to allow for comparison with Keller’s measurements of trabecular bone in the lumbar vertebrae (Table 3). The mean ash fraction of the trabeculae in the thoracic vertebrae is approximately half that measured in lumbar vertebrae (Ref. 7).

**TABLE 3.—RESULTS OF TRABECULAR REGION WITHIN T8 VERTEBRAE USING A SAMPLE SIZE OF A 1 cm CUBE**

<table>
<thead>
<tr>
<th>Specimen no.</th>
<th>Age</th>
<th>Body mass, kg</th>
<th>(a_{\text{mean}})</th>
<th>(BVF_0)</th>
<th>(M_0)</th>
<th>(O_0)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HTH0243</td>
<td>40</td>
<td>63.50</td>
<td>0.255</td>
<td>0.640</td>
<td>0.166</td>
<td>0.474</td>
</tr>
<tr>
<td>HTH0249</td>
<td>40</td>
<td>56.60</td>
<td>0.264</td>
<td>0.727</td>
<td>0.196</td>
<td>0.530</td>
</tr>
<tr>
<td>HTH0339</td>
<td>38</td>
<td>81.65</td>
<td>0.241</td>
<td>0.740</td>
<td>0.180</td>
<td>0.560</td>
</tr>
<tr>
<td>HTH0439</td>
<td>35</td>
<td>54.43</td>
<td>0.228</td>
<td>0.636</td>
<td>0.145</td>
<td>0.491</td>
</tr>
<tr>
<td>HTH0561</td>
<td>25</td>
<td>56.25</td>
<td>0.266</td>
<td>0.850</td>
<td>0.231</td>
<td>0.619</td>
</tr>
</tbody>
</table>
Discussion

The current model predicts sudden changes in the BVF and osteoid volume fractions at 200 days when a person returns to a 1 g from a 0 g environment. This prediction of rapid recovery is unlikely to accurately describe the biological changes in volume fractions because the BMUs must adapt and respond to skeletal loading. A time delay where neither bone loss nor gain occurs should be considered when a transition is made in between different gravitational environments. As research studies discover more detail about cellular dynamics, continuous development of the computational model will provide a more thorough understanding of the bone remodeling process in response to changes in mechanical stimuli.

The mean ash fraction values are relatively low compared to values in Keller’s experimental study (Ref. 7). Keller measured the ash fraction of the trabecular bone of the lumbar vertebrae and the cortical bone of the diaphysis of the femur. He reported a mean ash fraction of 0.594 for combined data from both types of bone. This is significantly higher than the ash fractions calculated for the T8 vertebrae. Since Keller’s specimens came from much older donors (ages 46, 67, 70, 77, 84 years), it is possible that the calculated ash fractions are acceptable. The T8 specimens came from patients in the 25 to 40 year old range, where a higher rate of bone remodeling is likely. The skeletal site may also contribute to the low ash fraction. The lumbar is the weight bearing region of the spine, suggesting that the bone strength and accordingly its ash fraction maybe on average higher than that in the thoracic region.

The T8 mean ash fractions indicate a higher osteoid volume fraction or a higher rate of bone remodeling at the time of scanning. The data were, therefore, re-analyzed to see if there was a problem in the image processing procedure. An intensity histogram shows one predominating region of intensity values around 1300 HU (Figure 9(a)). The first bar in the 0 to 200 HU is ignored because it is only counting the number of voxels in the background. The minimum intensity value of the entire bone specimen is found to be 1141.9 HU, which confirms the assumption of the first bar in bone intensity histogram.

An intensity histogram of a whole bone specimen should indicate two distinct parts—one for osteoid near the minimum intensity and one for fully mineralized bone near the maximum intensity. However, there is a single large grouping near the minimum intensity for all T8 specimens (Figure 9(a)). Hence, the ash fraction histogram only shows one distinct region with a low ash fraction corresponding to more osteoid (Figure 9(d)). Placing the intermediate calculations of QCT and ash density histograms shows that this is consistent throughout the calculation process from intensity values to ash fraction (Figure 9). There is no indication that there is a problem in the image processing functions.

The image processing procedure is further analyzed to see if the ROI size affects BVF and ash fraction. ROI size does affect the bone threshold value—the smaller the ROI, the smaller the threshold (Table 4). Higher threshold corresponds to higher ash fraction, but it removes more voxels from the images. It also has the potential to decrease the BVF significantly as seen in specimens HTH0243, HTH0339, and HTH0439 (Appendix D, Figure 12).

**TABLE 4.—ASH FRACTION AND BVF ARE DEPENDENT ON THE IMAGE PROCESSING TYPE**
[Method A uses a fixed square ROI of 25 pixel length. Method B is a freehand ROI of the background.]

<table>
<thead>
<tr>
<th>Specimen no.</th>
<th>A: Threshold</th>
<th>B: Threshold</th>
<th>A: BVF₀</th>
<th>B: BVF₀</th>
<th>A: α&lt;sub&gt;mean&lt;/sub&gt;</th>
<th>B: α&lt;sub&gt;mean&lt;/sub&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>HTH0243</td>
<td>1127.9</td>
<td>1204.7</td>
<td>0.768</td>
<td>0.527</td>
<td>0.272</td>
<td>0.412</td>
</tr>
<tr>
<td>HTH0249</td>
<td>1114.6</td>
<td>1139.0</td>
<td>0.756</td>
<td>0.709</td>
<td>0.280</td>
<td>0.308</td>
</tr>
<tr>
<td>HTH0339</td>
<td>1099.1</td>
<td>1221.1</td>
<td>0.719</td>
<td>0.353</td>
<td>0.257</td>
<td>0.535</td>
</tr>
<tr>
<td>HTH0439</td>
<td>1092.7</td>
<td>1160.6</td>
<td>0.716</td>
<td>0.513</td>
<td>0.241</td>
<td>0.364</td>
</tr>
<tr>
<td>HTH0561</td>
<td>1140.9</td>
<td>1167.9</td>
<td>0.869</td>
<td>0.829</td>
<td>0.300</td>
<td>0.321</td>
</tr>
</tbody>
</table>
Figure 9.—Histogram of values in HTH0561 specimen demonstrates that most of the vertebra is not highly mineralized. These plots are made before the restriction is placed on the intensity values to maintain the limit of 0.7 for the ash fraction of fully mineralized bone. X-axes units are as follows: intensity [HU], $\rho_{\text{QCT}}$ [mg/cc], and $\rho_{\text{ash}}$ [g/cc].

Limitations of this model are as follows.

1. The voxels are assigned homogeneous isotopic material properties and a constant Poisson’s ratio. It is known, however, that bone is an anisotropic material. A precise model of bone would account for the material properties in all directions.

2. Bone remodeling is assumed to be homogenous throughout the bone specimen. However, osteoclast resorption and osteoblast formation do not occur on all surfaces simultaneously and affects bone surfaces before changing the interstitial tissue. A model of remodeling locations should be applied for a more accurate simulation (Refs. 18 and 19).

3. Observations show that more cortical bone is added to the proximal femur during recovery. Architectural changes are not simulated, but maybe an important factor in bone strength. Additional work must be completed to characterize how the remodeling process affects cortical and trabecular structure.
Axial rigidity provides a prediction of vertebral bone strength. This approach is limited because it assumes that minimum rigidity is an indicator of strength and does not consider deformations through the whole bone. Validation of the axial rigidity results cannot be made even though the T8 vertebrae are a subset of samples used in a previous study (Ref. 11). There is an error in the ash density and QCT density in the published paper. The coefficient of QCT density should be 0.95 not 9.5. It is unknown if this error persisted throughout the study. Hernandez et al. also did not use a two-parameter modulus equation to calculate the axial rigidity. Significant variations have been observed when using different equations to calculate elastic modulus (Appendix E, Figure 14).

The present work is successful in establishing a methodology to determine computational model input parameters from QCT images, to predict time-dependent changes to elastic modulus via the outputs of the cellular dynamics model, and to evaluate bone strength during bone loss and recovery using the initial bone strength as a baseline. An equation has been derived to obtain ash fraction values from QCT images, a good alternative for non-invasive studies and applications. This theoretical model combining the bone remodeling process with strength analysis may help provide guidance on normal 1 g activities to avoid during recovery and may help develop a bone loss prevention plan, particularly if used in conjunction with NASA’s probabilistic risk assessment of bone fracture risk (Ref. 20).

An extension of this study is to extend the simulation duration to include extreme long-duration space missions. NASA plans to send astronauts to Mars for a three year mission. However, American astronauts have spent at most 6 months in microgravity. The computational model would need to have built in ability to handle aging changes over longer time simulations and validated to be able to predict the amount of bone loss for 1000 days in a microgravity environment. An additional consideration for a Mars bone loss study is whether or not the 0.38 g environment will provide sufficient skeletal stimulation to help prevent bone resorption.

Prediction of bone recovery also contributes a better understanding of bone diseases here on Earth. Osteoporosis affects 10 million individuals in the United States alone, and 34 million more are estimated to have low bone density, placing them at an elevated risk for bone diseases and fractures (Ref. 21). An attribute of osteoporosis is marked reduction in bone mass due to an imbalance in the bone remodeling cycle. Modeling the physiological mechanics of bone may help to provide guidance for low bone density patients on what activities to avoid. Also, due to associated balance deficits, many elderly individuals tend to slip and fall on their hips, causing impact loading and fractures. That and the fact that astronauts lose bone mineral density in the proximal femur provide justification for extending this study to that anatomical location.
Appendix A.—Acronyms and Definitions

A.1   Acronyms

BMC  bone mineral content
BMU  basic multicellular units
BVF  bone volume fraction
DAP  Digital Astronaut Project
DXA  dual energy x-ray absorptiometry
FE   finite element
FEM  finite element models
HU   Hounsfield units
QCT  quantitative computed tomography
ROI  region of interest
T8   eighth thoracic vertebra

A.2  Definitions

apparent ash density $\rho_{\text{ash}}$ inorganic mass / total volume
ash fraction $\alpha$ ash mass / dry bone mass
BMC  mass of mineral measured from CT scans
BVF or BV/TV bone volume / total volume
dry apparent density $\rho_{\text{app}}$ (organic + inorganic mass) / total volume
Hounsfield units units of CT image numbers
mineralized bone organic + inorganic + water
QCT density $\rho_{\text{QCT}}$ density determined from calibration phantom; only quantifies inorganic mineral component
Appendix B.—Remodeling QCT FEM Outline

Note: This process is automated in the remodelingMain.m. Refer to Appendix F for a README of the MATLAB functions.

1. Determine mineral density calibration from calibration phantom image.
4. Calculate ash fraction (see derivation in Appendix C).

\[ 1.29\alpha^2 + 1.41\alpha - \frac{\rho_{ash}}{BVF} = 0 \]

(Note: \( \rho_{ash} \) calculated from QCT density and ash density relation. BVF obtained from previous step.)
5. Run InitialValues.m to get rate of formation (rof), rate of resorption (ror), initial mature bone fraction \( (M_o) \), initial osteoid fraction \( (O_o) \), and mineralization rate \( (\text{Min}_\text{rate}) \).
6. Set outputs of InitialValues.m in Set_InitialConditions.m. Run Set_InitialConditions.m.
7. Run BRequilibrium.m to get equilibrium conditions.
8. Create initial FEM from QCT. Voxel elastic modulus values determined from QCT density, ash density, and modulus relations presented in Kaneko et al. and Keller papers. Run Abaqus Standard Analysis to obtain initial bone mechanics state.
9. Run BRmodeltrabecular.m to get time histories of relative mature bone fraction, osteoid fraction, and BVF saved in timeHistories.mat file.
10. Use Hernandez’s two-parameter power law to calculate modulus time history for each voxel at 10 day increments.

\[ E(t) = 84.37 \, BVF(t)^{2.58} \, \alpha(t)^{2.74} \, \text{[GPa]} \]

11. Create FEM for x days using voxel modulus time history. Run Abaqus Standard Analysis and data processing to determine the biomechanical state at x days.
Appendix C.—Derivation of Equations

C.1 Ash Fraction

\[ BVF \approx \frac{\rho_{app}[g/cc]}{1.41+1.29\alpha} \] (Ref. 6)

\[ \rho_{apparent ash}[g/cc] = \alpha \rho_{apparent}[g/cc] \] (Ref. 7)

Combine above equations to get a function in terms of ash fraction only.

\[ BVF = \frac{\rho_{ash}[g/cc]}{\alpha} \cdot \frac{1}{1.41 + 1.29\alpha} \]

\[ \alpha(1.41 + 1.29\alpha)BVF = \rho_{ash}[g/cc] \]

\[ 1.29\alpha^2 + 1.41\alpha - \frac{\rho_{ash}[g/cc]}{BVF} = 0 \]

Note: Ash fraction equation does not output imaginary ash fractions unless the ash density is negative, which is usually not the case since calibration phantom densities are 0 mg/cc or greater.

C.2 Maximum Intensity Value Corresponding to Ash Fraction of 0.7

\[ 1.29\alpha^2 + 1.41\alpha - \frac{\rho_{ash}[g/cc]}{BVF} = 0 \]

\[ \alpha(1.41 + 1.29\alpha)BVF = \rho_{ash}[g/cc] \]

Equate with Ash Density and QCT Density Relation:

Note: \( \rho_{QCT} \) has units of mg/cc.

K\textsubscript{3}HPO\textsubscript{4} phantom

\[ BVF(1.29\alpha^2 + 1.41\alpha) = 0.000953\rho_{QCT} + 0.0457 \]

\[ \rho_{QCT} = \frac{BVF(1.29\alpha^2 + 1.41\alpha) - 0.0457}{0.000953} \]

CHA phantom

\[ BVF(1.29\alpha^2 + 1.41\alpha) = 0.000887\rho_{QCT} + 0.0633 \]

\[ \rho_{QCT} = \frac{BVF(1.29\alpha^2 + 1.41\alpha) - 0.0633}{0.000887} \]

Equate with Calibration Curve relating QCT density and Intensity Values:

K\textsubscript{3}HPO\textsubscript{4} phantom

\[ mx + b = \frac{BVF(1.29\alpha^2 + 1.41\alpha) - 0.0457}{0.000953} \]

\[ x = \frac{1}{m} \left( \frac{BVF(1.29\alpha^2 + 1.41\alpha) - 0.0457}{0.000953} \right) - b \]

CHA phantom

\[ mx + b = \frac{BVF(1.29\alpha^2 + 1.41\alpha) - 0.0633}{0.000887} \]

\[ x = \frac{1}{m} \left( \frac{BVF(1.29\alpha^2 + 1.41\alpha) - 0.0633}{0.000887} \right) - b \]
Note: The above equations are only valid for linear correlation of QCT density with image intensity values. Keyak provided a L4 vertebra QCT image. A second order polynomial better describes the calibration plot for this particular specimen (Figure 10). However, the linear relation is not that far off. The maximum root of Equation (10) is taken to be the intensity value for an ash fraction of 0.7.

\[ ax^2 + bx + c = \frac{BV F (1.29 \alpha^2 + 1.41 \alpha) - 0.0633}{0.000887} = 0 \]  

(10)
Appendix D.—Image Processing Method

Noise from the 20 percent ethanol bath and background is removed from the images to isolate the bone specimen. Two sizes of ROIs are tested to assess the effect on ash fraction and BVF (Figure 11). Method A takes a fixed region of a 25 pixel length square. The user traces a region around the bone for Method B.

The bone threshold value is smaller using Method A rather than Method B. The processed images from Method A better retain the trabeculae in these T8 vertebrae (Figure 12). Method B’s threshold removes much of the trabeculae, decreasing the BVF significantly and increasing the ash fraction. HTH0339 midlayer slice best demonstrates the difference in image processing technique.

![Figure 11.](image1)

**Figure 11.**—Two sample sizes of the background are tested to see how it affects the image: (a) uses a fixed square and (b) is done free-hand by the user.

<table>
<thead>
<tr>
<th>Specimen #</th>
<th>Original Image</th>
<th>Method A</th>
<th>Method B</th>
</tr>
</thead>
<tbody>
<tr>
<td>HTH0243</td>
<td><img src="image2" alt="Image" /></td>
<td><img src="image3" alt="Image" /></td>
<td><img src="image4" alt="Image" /></td>
</tr>
<tr>
<td>HTH0249</td>
<td><img src="image5" alt="Image" /></td>
<td><img src="image6" alt="Image" /></td>
<td><img src="image7" alt="Image" /></td>
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<tr>
<td>HTH0339</td>
<td><img src="image8" alt="Image" /></td>
<td><img src="image9" alt="Image" /></td>
<td><img src="image10" alt="Image" /></td>
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<tr>
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<td><img src="image11" alt="Image" /></td>
<td><img src="image12" alt="Image" /></td>
<td><img src="image13" alt="Image" /></td>
</tr>
<tr>
<td>HTH0561</td>
<td><img src="image14" alt="Image" /></td>
<td><img src="image15" alt="Image" /></td>
<td><img src="image16" alt="Image" /></td>
</tr>
</tbody>
</table>

**Figure 12.**—Collection of midlayer slices from all T8 vertebrae samples demonstrates the effect of varying the background sample area during image processing.
Appendix E.—Justification of Voxel Material Property Equations

Several experimental studies characterize the QCT density from image technology with ash density and elastic modulus. Appendix C summarizes these studies and validates the choice of relations used in this study.

A relation to convert QCT density to ash density is required to compute other parameters. Kaneko et al. studied the effect of metastases on the mechanical properties and densities of cortical and trabecular bone in the human femur (Refs. 12 and 13). The QCT density and ash density correlations were plotted for all four groups in the trabecular bone study: all data, no cancer, metastatic lesions, and no lesions but died from cancer. The NC group equation does not diverge significantly from the All group equation until around 1400 mg/cc, which is a fairly high QCT density. It would also be acceptable to use the All group equation because the range of data from the NC group falls within the range of data from the All group. The cortical bone relations are also plotted, but do not yield similar trends in the All and no cancer groups. Since there is a significant disparity between the two equations, no cortical bone relation for QCT density to ash density is reported.

Keyak et al. combined the results from Kaneko et al.’s two studies and formulated a single ash density and QCT density relation (Ref. 14). This equation correlates well within the range of trabecular and cortical bone data (Figure 13). It must be noted that this equation does include data from bone samples with metastases. This equation would be beneficial to have a relation describing both types of bone.

Figure 13.—Cortical equations from Kaneko et al. 2003; trabecular equations from Kaneko et al. 2004 studying bone with and without metastases. N.S. indicates that the relation is not significant due to the small sample size. Solid lines are used to indicate the range of data; dashed lines indicate data extrapolation.
A two-part ash density and QCT density relation can be used in later projects once more knowledge is known on how to identify cortical and trabecular bone from CT scans. A crude method to identify cortical from trabecular bone is to make mashed images of one of the other and then assign these voxels identifiers, so that the two types of bone can be assigned different material properties in the Abaqus input file. However, a main issue with this method is the fact that one cannot determine if a voxel is cortical or trabecular simply from looking at the QCT scan.

Analysis of the bone biomechanics is achieved using the elastic modulus, a parameter that measures a material’s ability to deform under particular loads. Several potential modulus equations have been considered (Figure 14). This study does not use modulus relations for only cortical or trabecular bone. The final verdict was to use the two-parameter modulus equation by Hernandez et al., which is not plotted in Figure 14 (Ref. 6). Keyak et al. equation would be ideal. However, if Keyak et al. relation is used to calculate the initial conditions, a discontinuity occurs within the first 10 days. No change in the BVF and ash fraction within the first 10 days implies that the modulus should also not change. Therefore, the two-parameter equation is used for consistency. Another reason for this decision is that the Keyak et al. modulus correlation was derived from data including bone metastasis samples. Since no correlation was shown between no cancer and metastasis groups in cortical bone, it would be better to use the two-parameter modulus equation (Ref. 12).

Figure 14.—Modulus and QCT density/Ash density relations show the diversity in equations. The two-parameter Hernandez equation is not plotted here because QCT density is not its primary independent variable.
Appendix F.—Read Me for MATLAB Programs

Required Software:

- MATLAB (functions were written with version R2012.a)
- MATLAB Image Processing Toolbox 8.0
- MATLAB Image Acquisition Toolbox 4.3

Optional Software:

- ABAQUS—required for FEM (see Addition points section to comment MATLAB functions that call ABAQUS)

The main function is remodelingMain.m. It is broken down into the following sections and automatically calls the relevant functions.

- Calibration Phantom Curve
- uCT/QCT Processing and Initial Meshing
- Initial FEM and Equilibrium State
- Simulation of Bone Loss and Recovery
- Results Processing

User inputs are required during this process in particular for image processing. They are indicated in the command window with an arrow (Figure 15).

Figure 15.—Example of user inputs when running remodelingMain.m. This one requires the selection of fluid in calibration phantom chamber.
Some variables must be initialized **before** running remodelingMain.m.

- **QCTtype**—calibration phantom type: 1 for K$_2$HPO$_4$, 2 for CHA
- **boneType**—simulation type: 1 for whole bone specimen, 2 for ROI
- **voxsize**—voxel size [mm]
- **numcoarsen**—number of voxels on an edge to combine (particularly useful for large FEMs to reduce computational time)
- **sliceThickness**—distance between layers of QCT [mm]
- **cubeLength**—ROI vertical length [mm]; used for boneType option 2
- **nu_bone**—Poisson’s ratio of bone
- **bcType**—type of fixed boundary condition on distal end of FEM: 1 for pin & roller, 2 for pin

Path locations
- **mechanotransductionModelFolder**—location of mechanotransduction model MATLAB files
- **abaqusFolder**—location to save INP, Abaqus analysis files, and MATLAB workspace
- **CTFolder**—location of CT images

**F.1 Calculating Voxel Size**

If the voxel size is unknown, open a slice of the QCT in ImageJ. Under the filename in the upper left corner, there might be a line listing the image size and the number of voxels in the format 0.00- by 0.00- mm (000 by 000) (Figure 16). Use these values to calculate the voxel size.

**F.1.1 Location of CT Images**

Upon running remodelingMain.m, the user will be prompted to choose a folder containing the CT images. A single stack of QCT images should be placed in a folder with the specimen name. The program is automated to analyze multiple stacks of QCT images that use the same phantom. If the user would like to perform more than one specimen analysis, each specimen folder should be placed in a primary folder.

**F.1.2 Modifications to Bone Remodeling Computation Model**

If modifications are made to the mechanotransduction model, the user simply needs to make the changes in the respective functions. InitialValues.m and Set_InitialConditions.m from the original computation model have been incorporated into the remodelingMain.m function in “Initialize these Variables” and “Initial FEM and Equilibrium State” sections respectively.

---

**Figure 16.—One slice of the QCT.**
F.1.3 Additional Points to be Aware of

- The user must make sure that the calibration phantom curve is accurate for a particular set of QCT images. It is advised to modify the calibrationphantom.m function after plotting different curves to the plot of mineral densities versus intensity values.

- If the user does not want to use Abaqus, comment the lines with the following functions in sections “Initial FEM and Equilibrium State” and “Simulation of Bone Loss and Recovery” of remodelingMain.m:
  - cd([abaqusFolder ' \nameFolders{#} \folder{#,:}]);
  - AbaINP(inpname(#,:),meshCT,thresh,voxsizeFE,sliceThickness,in
    tE,nu,
  - bone,forceZ,sigmaZ,bctype);
  - pythonAbaqusScript(inpname(#,:),[abaqusFolder ' \n
  - nameFolders{i} \folder{#,:}]);
  - DATfile = [inpname(1,1:end-4) '.dat'];
  - effStrain(#,i) = DATProcessing(DATfile,[abaqusFolder ' \\
  - folder(#,:)]);

F.2 List of MATLAB Functions Required for Remodeling Program remodelingMain.m

Image Processing:

- calibrationPhantom.m
- coremeshQCT.m

FEMP:

- coarsen.m
- AbaINP.m
- im2h8.m
- bc3.m
- writeAbaH8.m
- pythonAbaqusScript.m

Results Processing:

- compositeRigidity.m
- DATProcessing.m

Mechanotransduction Model:

- BRequilibrium.m
- BRequilibrium_rhs.m
- BRmodeltrabecular.m
- BRmodel_trabecular_rhs.m
- depthFM
- depthR.m
- dR.m
- dA.m
• g.m
• Fa.m (updated in remodelingMain.m)

F.3 List of Current Results Outputted From remodelingMain.m

• Axial rigidity time history in kN
• Relative Osteoid Volume Fraction
• Relative Mineralized Volume Fraction
• Relative Bone Volume Fraction
• Ash Fraction
Appendix G.—Study of the Proximal Femur

Methodology to study the proximal femur during prolonged unloading and recovery is similar to the one created using the thoracic vertebra. The primary difference is the physiological boundary conditions. Loading conditions for a normal 1 g single-stance loading condition is described here. Another important condition to consider is fall onto the greater trochanter.

Keyak et al. and Hazelwood et al. studied the proximal femur in response to long-term spaceflight and marathon training respectively (Ref. 22 and 15). Both FEM methods fully constrained the distal end of the femur. However, a pin and roller boundary condition is applied to the distal end of this study’s FEMs. The primary reason is that the femur rests on top of a cartilaginous tissue, which is a viscous material allowing movement in the lateral direction. Two nodes on the distal surface are also constrained in all coordinate directions to prevent the models from moving freely. The remaining nodes are only constrained in the z-direction to simulate rollers.

NASA is interested in the continued wellbeing of the astronauts in particular when they return to Earth, so it follows to use normal 1 g loading conditions in the FEM. The boundary conditions for all FEM simulations are kept the same to evaluate the changes in mechanical response of the proximal femur to single-leg stance. Hazelwood et al. used daily loading as a basis for a study on the proximal femurs of marathon runners (Ref. 22). This study uses the same applied loads on the greater trochanter and femoral head (Figure 17 and Table 5). It was decided to include the muscle forces given the results of studies like Duda et al (Ref. 23).

<table>
<thead>
<tr>
<th>Location</th>
<th>Single-Leg Stance (1)</th>
<th>Abduction (2)</th>
<th>Adduction (3)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Greater Trochanter</td>
<td>842 N at −8°</td>
<td>1158 N at −15°</td>
<td>1126 N at 65°</td>
</tr>
<tr>
<td>Femoral Head</td>
<td>2317 N at 24°</td>
<td>1685 N at 28°</td>
<td>1548 N at 56°</td>
</tr>
</tbody>
</table>

Table 5.—SINGLE-LEG STANCE LOADS ON THE PROXIMAL FEMUR INCLUDE ABDUCTION AND ADDUCTION MUSCLE FORCES (Ref. 22)

Figure 17.—Hazelwood et al. 2007 loading condition for daily loads reduced to coordinate forces for FEM (Ref. 22). Image of femur is a CAD model of the 3rd generation composite femur made by Pacific Research Labs, Vashon Island, Washington.
MATLAB functions to apply the single-leg stance loading condition were not written because no QCT of the femur was obtained during the time of study. However, a theoretical approach was established. To determine the surface nodes, two transverse slices from the proximal end are taken at a time. Intersect of the two layers are declared as nodes of the surface of the femur (Figure 18). The process is repeated up to the diaphysis.

All the surface nodes have been identified. However, it is uncertain at this point which nodes are on the femoral head or the greater trochanter surfaces. Identification can be made by seeing which side of the centroid the nodes are located. If the nodes have a y-coordinate value greater than the centroid’s, then the nodes are placed in a node set for the femoral head. Again, QCT images of the proximal femur are integral to writing the MATLAB functions to apply boundary conditions to rounded surfaces.

Figure 18.—Illustration of how to determine which nodes are on surface of femoral head and greater trochanter. The black region seen in the overlap of the two layers indicates the nodes that are on the surface where the single-stance loading condition will be applied.
References


Predicting Bone Mechanical State During Recovery After Long-Duration Skeletal Unloading Using QCT and Finite Element Modeling

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During long-duration missions at the International Space Station, astronauts experience weightlessness leading to skeletal unloading. Unloading causes a lack of a mechanical stimulus that triggers bone cellular units to remove mass from the skeleton. A mathematical system of the cellular dynamics predicts theoretical changes to volume fractions and ash fraction in response to temporal variations in skeletal loading. No current model uses image technology to gather information about a skeletal site’s initial properties to calculate bone remodeling changes and then to compare predicted bone strengths with the initial strength. The goal of this study is to use quantitative computed tomography (QCT) in conjunction with a computational model of the bone remodeling process to establish initial bone properties to predict changes in bone mechanics during bone loss and recovery with finite element (FE) modeling. Input parameters for the remodeling model include bone volume fraction and ash fraction, which are both computed from the QCT images. A non-destructive approach to measure ash fraction is also derived. Voxel-based finite element models (FEM) created from QCTs provide initial evaluation of bone strength. Bone volume fraction and ash fraction outputs from the computational model predict changes to the elastic modulus of bone via a two-parameter equation. The modulus captures the effect of bone remodeling and functions as the key to evaluate of changes in strength. Application of this time-dependent modulus to FEMs and composite beam theory enables an assessment of bone mechanics during recovery. Prediction of bone strength is not only important for astronauts, but is also pertinent to millions of patients with osteoporosis and low bone density.

Bone mineral content; Microgravity; Mathematical model; Finite element method; Quantitative computer tomography; Simulation; Spine