Evidence Report:

Risk of Radiation Carcinogenesis

Human Research Program

Space Radiation Element

Approved for Public Release: Month DD, YYYY

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I. PRD Risk Title: Risk of Radiation Carcinogenesis

Description: In space, astronauts are exposed to ionizing radiation that is quantitatively and qualitatively different from terrestrial radiation. This environment includes protons and high-Z high-energy (HZE) ions together with secondary radiation, including neutrons and recoil nuclei that are produced by nuclear reactions in spacecraft materials or tissue. Space radiation exposure increases cancer morbidity and mortality risk in astronauts. This risk may be influenced by other space flight factors including microgravity, environmental contaminants, nutritional issues, and psychological and physiological stress. Current space radiation risks estimates are based on human epidemiology data for X-rays and gamma-ray exposure scaled to the types and flux-rates in space using radiation quality factors and dose-rate modification factors, and assuming linearity of response. There are large uncertainties in this approach and experimental models imply additional detriment due to the severity of the phenotypes of cancers formed for the heavy ion component of the galactic cosmic rays compared to cancers produced by terrestrial radiation. A Mars mission may not be feasible (within acceptable limits) unless uncertainties in cancer projection models are reduced allowing shielding and biological countermeasures approaches to be evaluated and improved, or unless mission durations are constrained.

II. Executive Summary

Astronauts on missions to the International Space Station (ISS), the moon or Mars are exposed to ionizing radiation with effective doses in the range from 50 to 2000 mSv (milli-Sievert) projected for possible mission scenarios (Cucinotta and Durante 2006; Cucinotta et al. 2008). The evidence of cancer risk from ionizing radiation is extensive for radiation doses above about 50 mSv. Human epidemiology studies that provide evidence for cancer risks for low linear energy transfer (LET) radiation such as X-rays or gamma-rays at doses from 50 to 2000 mSv include the survivors of the atomic-bomb explosions in Hiroshima and Nagasaki, nuclear reactor workers (Cardis et al. 1995, 2007) in the US, Canada, Europe, and Russia, and patients treated therapeutically with radiation. Ongoing studies are providing new evidence of radiation cancer risks in populations accidentally exposed to radiation from the Chernobyl accident and from Russian nuclear weapons production sites. Results from the Japanese atomic bomb survivors also continue to be analyzed and integrated into risk projection models (Preston et al. 2004, 2007; Cucinotta et al. 2013). These studies provide strong evidence for cancer morbidity and mortality risks at over 12 tissue sites, with the largest risks for adults found for leukemia and tumors of the lung, breast, stomach, colon, bladder, and liver. There is also strong evidence for sex-dependent variations in risks estimates due to the due to differences in natural incidence of cancer and the additional cancer risks for the breast and ovaries and higher risk from radiation for lung cancer in females (NCRP 2000). Human studies also provide evidence for a declining risk with increasing age at exposure; however, the magnitude of the reduction above age 30 years is uncertain (NCRP 2000; BEIR 2006). Genetic and environmental factors, including possible healthy worker effects for never-smokers (Cucinotta et al. 2011; NCRP 2012), that contribute to radiation carcinogenesis are also being explored to support identification of individuals with increased or reduced risk.

In space, astronauts are exposed to protons and high energy and charge (HZE) ions along with secondary radiation, including neutrons and recoil nuclei, produced by nuclear reactions in spacecraft or tissue. Whole body doses of 1-2 mSv/day accumulate in interplanetary space, and
approximately half this value on planetary surfaces (Cucinotta et al. 2006; NCRP 2006; Slaba et al. 2011, 2013a; Zeitlin et al. 2013). In traveling to Mars, it is estimated that every cell nucleus within an astronaut will be traversed by a proton or secondary electron every few days and an HZE ion every few months (Cucinotta et al. 1998). In spite of their lower cell nucleus hit frequency as compared to protons, the large ionization power of HZE ions makes them an important contributor to the risk. Likewise, light ions and neutrons are additional components that make up a significant percentage of the space radiation environment (Norbury and Slaba 2014). Radiation shielding is an effective countermeasure for solar particle events (SPEs), which are largely protons with energies below a few hundred MeV. The energy spectrum of the galactic cosmic rays (GCRs) peaks near 1 GeV/n, and consequently, these particles are so penetrating that shielding can only partially reduce the doses absorbed by the crew (Cucinotta et al. 2006). Current shielding approaches do not effectively mitigate exposure from GCR. Increasing shielding thickness poses significant mass constraints to spacecraft launch systems and may actually increase exposures beyond certain thicknesses due to neutron build-up and electromagnetic cascades (Slaba et al. 2013a).

Epidemiological data, largely from the Atomic bomb survivors in Japan (Preston et al. 2003, 2004, 2007), provides a basis for risk estimation for low-LET radiation. Models for cancer incidence and mortality based on these data are assumed to be scalable to other populations, dose-rates, and radiation types (Cucinotta et al. 2013). However, because there is a lack of human data for protons and HZE ions, current space risk estimates must rely entirely on experimental model systems and biophysical considerations. The scaling of mortality rates for space radiation risks to astronauts to the Atomic bomb survivors introduces many uncertainties (Cucinotta et al. 2001; Cucinotta and Durante 2006) into risk estimates, and there are important questions with regard to the correctness of any scaling approach because of qualitative differences in the biological effects of HZE ions and gamma-rays. The two scaling parameters with largest uncertainties are the radiation quality factor, which estimates the increased effectiveness of HZE nuclei compared to x-rays or gamma-rays for the same dose, and the dose- and dose-rate effectiveness factor (DDREF), which reduces estimates of cancer risk at high dose- and dose-rates when the dose- and dose-rate are low (< 0.05 Gy/hr). The mixed field GCR environment in space, predominantly composed of protons with a smaller percentage attributed to more damaging heavy ions, further complicates risk estimation.

Acceptable levels of risk are often guided by societal or ethical norms. There is continued debate on what level is acceptable for space radiation cancer risks for exploration of the moon or Mars. A historical perspective is summarized herein; however, we note that other non-cancer mortality and morbidity risks associated with space radiation exposure must also be considered for Mars or other long-duration missions (i.e. central nervous system effects and circulatory disease) as well as confounding spaceflight factors such as microgravity and immune system impacts (Crucian et al. 2015). Improvements in safety in other areas of spaceflight will also place additional pressure on radiation protection efforts to reduce astronaut risk.

Ground-based experimentation is a critical requirement to provide the scientific evidence needed to improve space radiation cancer risk estimates. Flight experiments have not been a high priority because historically these have been expensive and poorly reproducible (Sihver 2008; Durante et al. 2007, Durante and Cucinotta 2008). In general, the low dose rates on the ISS preclude collection of useful data in a reasonable time and with a reasonable sample size, and experiments in the past have yielded no major findings (Kiefer and Pross 1999; Schimmerling et
al. 2003; Durante and Kronenberg 2005). Ground-based radiobiology and physics studies are performed at the NASA Space Radiation Laboratory (NSRL) at the Brookhaven National Laboratory (Upton, NY, USA). This facility is able to simulate the high-energy protons and HZE particles in space. NSRL opened for research in October 2003, and has produced experimental data of great relevance for reducing uncertainties in risk assessment. The NSRL is currently being upgraded to a GCR simulator capable of delivering a mixed field environment consisting of protons and ions at doses and dose rates that are more representative of the space environment (Norbury et al. 2016).

Mechanistic research performed at NSRL with 2D and 3D human cell culture and models, and animal studies in murine models is being pursued to establish level of risk, provide biological knowledge required to reduce uncertainties in risk projection models, guide the extrapolation from experiment to astronauts, and pave the way for biomarker discovery and biological countermeasure development. Studies with animals are an important component of space radiation research; however, they are time consuming and expensive in light of the large number of radiation types, doses, and dose rates of concern to NASA and the need to extrapolate results across species (NCRP 2005). Systems biology models of cancer risk that could be used to extrapolate radiation quality over the broad range of nuclear types, energies, and fluence rates in space are a promising new approach to these problems.

III. Introduction

As noted by Durante and Cucinotta (2008), cancer risk caused by exposure to space radiation is now generally considered a main hindrance to interplanetary travel for the following reasons: large uncertainties are associated with the projected cancer risk estimates; no simple and effective countermeasures are available, and significant uncertainties prevent scientists from determining the effectiveness of countermeasures. Optimizing operational parameters such as the length of space missions, crew selection for age and sex, or applying mitigation measures such as radiation shielding or use of biological countermeasures can be used to reduce risk, but these procedures have inherent limitations and are clouded by uncertainties.

Space radiation is comprised of high energy protons, neutrons and high charge (Z) and energy (E) nuclei (HZE). The ionization patterns and resulting biological insults of these particles in molecules, cells, and tissues are distinct from typical terrestrial radiation, which is largely X-rays and gamma-rays, and generally characterized as low linear energy transfer (LET) radiation. Galactic cosmic rays (GCR) are comprised mostly of highly energetic protons with a small component of high charge and energy (HZE) nuclei. Prominent HZE nuclei include He, C, O, Ne, Mg, Si, and Fe. GCR ions have median energies near 1 GeV/n, and energies as high as 10 GeV/n make important contributions to the total exposure.

Ionizing radiation is a well known carcinogen on Earth (BEIR 2006). The risks of cancer from X-rays and gamma-rays have been established at doses above 50 mSv (5 rem), although there are important uncertainties and on-going scientific debate about cancer risk at lower doses and at low dose rates (<50 mSv/h). The relationship between the early biological effects of HZE nuclei and the probability of cancer in humans is poorly understood, and it is this missing knowledge that leads to significant uncertainties in projecting cancer risks during space exploration (Cucinotta and Durante 2006; Durante and Cucinotta 2008).
A. Uncertainties in Cancer Projections

The uncertainties that occur in cancer risk projection models for space radiation include:

- Uncertainties in determining the qualitative and quantitative differences between the biological damage induced by space radiation compared to X-rays
- Uncertainties in human epidemiological data including statistical, record keeping, dosimetry, and bias resulting from mis-reporting of cancer deaths
- Uncertainties in transferring radio-epidemiology data to other populations, including cancer rates and survival data in the population of interest for space applications
- Uncertainties in transferring radio-epidemiology data to other radiation types and dose-rates of interest to space applications
- Uncertainties in the shape of the dose-response curve at low doses (i.e. linear, linear-quadratic) and the possibility of dose thresholds
- Uncertainties associated with extrapolation of experimental data from animals to humans
- Uncertainties associated with individual radiation sensitivity factors, including age, genetic, epigenetic, dietary, and “healthy worker” effects
- Uncertainties in space radiation environmental models, transport codes, geometry models, and dosimetry methods
- Uncertainties in predicting SPE occurrence, energy spectrum, and magnitude
- Possible inter-dependence of any of the uncertainties mentioned above

Quantitative methods have been developed to propagate uncertainties for several factors that contribute to cancer risk estimates (NCRP 1997; Cucinotta et al. 2013). Current estimates of levels of uncertainty represented as fold changes over the median risk projection are described in detail in Cucinotta et al. (2013). Comparison of risks for adults for terrestrial and space exposures is shown in Figure 1. Microgravity and the effects of other spaceflight stressors such as altered immune function, chronic inflammation and depressed nutritional status during a mission timeframe are unknown modifiers of radiation cancer risk and may confound the effects of radiation impacting the risk projection, and are potentially additional sources of uncertainty in current risk estimates. Several studies have been performed on the ISS and on the ground using apparatus that simulate the space environment. However these studies provide conflicting results with respect to radiation effects on chromosomal aberrations and DNA damage and repair, early endpoints relatable to the carcinogenic process (Yatagai and Ishioka 2014).

Radiation affects cells and tissues either via direct damage to cellular components or via the production of highly reactive free radicals from water (Goodhead 1994). Both of these mechanisms can result in sufficient damage to cause cellular death, DNA mutation, chromosomal aberrations, genomic instability or abnormal cellular function. The extent of damage is generally believed to be dependent upon the dose and type of particle with a linear dose-response curve (Goodhead 1994). This is true for high and moderate radiation exposure, but is extremely problematic to measure for lower doses where difficulty exists in discerning the effects of radiation exposure from those that are triggered by normal oxidative stress that cells and tissues deal with on a constant basis. The HZE nuclei are unique components of space radiation, which produce densely ionizing tracks as they pass through matter. When they traverse a biological system, they leave streaks or tracks of damage at the biomolecular level, which are fundamentally different than those left by
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low LET radiation such as gamma and X-rays. In the nucleus of a cell where genetic material is stored, the traversal of a heavy ion can produce tracks of clustered DNA damage (Cucinotta and Durante 2006) as illustrated in Figure 2.

![Figure 2](image)

**Figure 2.** Illustration of clustered DNA damage tracks produced by heavy ion traversal in the nucleus of a cell.

HZE nuclei impart damage via the primary energetic particle as well as from fragmentation events that produce a spectrum of other energetic nuclei, including protons, neutrons and heavy fragments (Wilson et al. 1995; Cucinotta et al. 1998, 2006; Durante and Cucinotta 2011). Therefore, a large penumbra of energy deposition exists that extends outward from the primary particle track (Cucinotta et al. 2000). Secondary radiation produced in shielding materials can be reduced through usage of materials with light atomic constituents such as hydrogen and carbon. However, a large percentage of secondary radiation is produced within tissue and is therefore not practically avoidable. Due to the large amount of energy deposited as these particles traverse biological structures, HZE nuclei are capable of producing the greatest amount of cellular damage and are therefore a large concern for astronaut safety. The lack of epidemiological data and sparse radiobiological data on effects for these radiation types leads to a high level of uncertainty in risk estimates for long term health effects following exposure to GCR and SPEs.
Figure 2. A comparison of particle tracks in nuclear emulsions and human cells (Cucinotta and Durante 2006). The right panel shows tracks of different ions, from protons to iron, in nuclear emulsions, clearly showing the increasing ionization density (LET = dE/dx) along the track by increasing the charge Z. The left panel shows three nuclei of human fibroblasts exposed to γ-rays, Si-, or Fe-ions, and immunostained for detection of γ-H2AX. Each green focus corresponds to a DNA DSB. While in the cell exposed to sparsely ionizing γ-rays foci of the histone variant, H2AX are uniformly distributed in the nucleus, the cells exposed to HZE particles present DNA damage along tracks (one Si- and three Fe-particles, respectively), and the spacing between DNA DSB is reduced at very high LET.

B. Types of Cancer Caused by Radiation Exposure

Data from the Atomic Bomb survivors shows that an acute exposure to ionizing radiation increases the mortality from cancer across a wide range of tumor types typical of the spectrum observed in a population (Barcellos-Hoff et al. 2015). The tissue types that contribute to the overall cancer risk observed with low LET radiation include lung, colorectal, breast, stomach, liver, brain, ovarian, esophageal, and bladder cancers, and several types of leukemia, including acute lymphocytic leukemia, acute myeloid leukemia and chronic myeloid leukemia (NCRP 2000; Preston et al. 2003; BEIR 2006). It is not fully established if the same spectrum of tumors will occur for high LET radiation as with low LET radiation, although results thus far have not shown novel tumor types generated from HZE ions compared to low LET (Bielefeldt-Ohmann et al. 2012; Barcellos-Hoff et al. 2015). However evidence suggests that HZE ions may induce cancers with unique characteristics compared to low-LET induced cancers, with differences in incidence and latency as well as in malignant potential. There may also be distinct radiation quality effects for HZE ions in terms of the cytogenetic and molecular subtype of the induced tumor, all factors that may impact disease surveillance, progression, treatment and ultimately outcome. Further investigation is required to fully understand these differences. Relative biological effectiveness
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Factors (RBE) describe the ratio of a dose of high LET radiation to that of X-rays or gamma-rays that produce the identical biological effect. In general, the RBE values observed for solid cancer induced by HZE particles are high. However these values, as well as those observed in mice exposed to neutrons, are highly dependent on tissue type and genetic background of the animal. In contrast, the RBE values observed for leukemia are close to one, which may indicate that the mechanisms underlying tumor induction for leukemias are distinct from those controlling solid tumor formation (Bielefeldt-Ohmann et al. 2012; NCRP 1990; Fry and Storer 1987). Detailed review and discussion of HZE ion animal carcinogenesis studies can be found in recently published articles (Bielefeldt-Ohmann et al. 2012; Barcellos-Hoff et al. 2015).

C. Age, Latency, Sex, and Individual Sensitivity Issues

Because cancer is a genetic disease with important epigenetic factors, individual susceptibility issues are an important consideration for space radiation protection, and NASA’s current cancer risk prediction model considers both sex dependence and how age at exposure effects the excess relative risks for radiation induced cancers (Cucinotta et al. 2013). Females have a higher cancer risk from radiation induced cancer compared to males, largely due to the additional risks to the lung, breast and ovaries (BEIR 2006, NCRP 2000). Mechanisms responsible for increased lung cancer incidence in females have not been elucidated. In contrast, females are at a lower risk for development of radiation induced CMLs, with a recent analysis of Surveillance, Epidemiology and End Results Program (SEER) and A-bomb survivor data suggesting that this difference is due to changes in inherent risk rather than disease latency (Radivoyevitch et al. 2014).

It is generally accepted that the risk of radiation induced cancer is highest for individuals exposed in childhood and decreases with increasing age at time of exposure. At a sufficiently high age, risk would be expected to decrease with age at exposure because the distribution of latency for tumor development would extend beyond the expected lifespan at older exposure ages. Also, there may be a possible reduction in the number of cells at risk at older age due to senescence or other biological factors (Campisi 2003; Campisi; d’Adda di Fagagna 2007). Age related changes in the host tissue microenvironment may impact the ability of a tissue to support tumor growth or alter tumor properties (Nguyen et al. 2014; Beheshti et al. 2015). Shuryak et al. (2010) reported that age related risks in younger populations are dominated by initiation processes while at later ages, radiation induced cancers may also result from promotion of preexisting malignant cells. Overall, the balance between initiation and promotional effects varies by tissue target and by host age, and complicates the dependence of cancer risk on age at exposure.

Genetic and environmental factors also impact risk of cancer from radiation exposure (NCRP 2010; Barcellos-Hoff et al. 2015). Studying the mechanisms of genetic sensitivity provides important insights into understanding the radiation risks to astronauts (Durante and Cucinotta 2008). Studies of historical data sets such as the atomic-bomb survivors show that subsets of the exposed cohorts could have a higher than average radiation risk (Ponder 2001). A well-known example is ataxia-telangiectasia (AT) patients that dramatically demonstrated the importance of genetic susceptibility to radiation damage in cancer treatment. Other examples related to DNA repair genes include BRCA1&2, p53 (Ponder 2001), NBS (Pluth et al. 2008), Artemis (Wang et al. 2005), and many other so-called high-penetrance genes involved in cancer susceptibility (Ponder 2001).
Ataxia-Telangiectasia-Mutated (ATM) homozygous individuals represent only a small fraction of radiosensitive patients, although they appear to be the most sensitive. ATM heterozygotes, who are also cancer-prone, are suspected to represent a large fraction of the extreme radiosensitive patients (Thompson et al. 2005). It has been shown that cells heterozygous for ATM mutations are slightly more sensitive to radiation-induced neoplastic transformation than the wild-type (Smilenov et al. 2001). An increased sensitivity of ATM heterozygotes has been also proven in vivo, measuring the induction of cataracts in ATM homozygotes, heterozygotes, and wild-type mice exposed to 0.5-4 Gy X-rays (Worgul et al. 2002). Interestingly, although there is a clear increase in breast cancer risk in individuals who carry deleterious BRCA1/BRCA2 mutations, there is no indication that these individuals are at an increased risk for contralateral breast cancer following radiotherapy (Bernstein et al. 2013).

An important issue is how low penetrance genes impact sensitivity to radiation-induced cancer. A study on subjects exposed to high radiation doses to treat ringworm of the scalp (tinea capitis) in Israel revealed a strong familial risk of radiation-induced meningioma (Flint-Ritcher and Sadetzki 2007), suggesting that radiation carcinogenesis might be an issue for a genetically predisposed subgroup of the general population, rather than a random event (Hall 2007; Sigurdson 2012). This is also supported by identification of genetic variants associated with increased occurrence of second cancers in survivors of childhood Hodgkin’s lymphoma through the use of a genome wide association study (Best et al. 2011) and similarly, the identification of variants associated with radiation related papillary thyroid carcinoma in individuals exposed during the Chernobyl accident (Takahashi et al. 2010).

It is not known if individuals displaying hypersensitivity to low-LET radiation will also be equivalently hypersensitive to HZE nuclei, or if findings at high dose and dose-rates will hold at low doses and dose-rate. Mice heterozygous for the ATM gene are more sensitive to cataractogenesis than wild-types not only after exposure to X-rays, but also after localized irradiation with high-energy Fe-ions (Hall et al. 2006). However, there are other studies that show that high LET irradiation has a reduced dependence on genetic background compared to low LET irradiation (George et al. 2009) while other evidence suggests that variability in the susceptibility of different mouse strains for radiation induced cancers observed for low-LET radiation extends to tumors induced by heavy ions (Bielefeldt-Ohmann et al. 2012; NCRP 2014).

In contrast to hypersensitivity, certain individuals also exhibit reduced sensitivity and risk due to environmental or other personal factors. An analysis of lung cancer and other smoking-attributable cancer risks has shown significantly reduced lung cancer risk and overall cancer risk for never-smokers compared to the U.S. population. Since 90% of the astronauts are never-smokers, and the remainder former smokers, such empirical observations are applicable for space radiation risk projections. Other healthy worker effects may also be relevant for space missions and possibly modify risk projections or reduce uncertainties (Cucinotta et al. 2013). A predictive assay able to identify radiation hypersensitive, cancer-prone subjects could be useful in crew selection for long-term spaceflights. Alternatively, identifying resistant or reduced-risk individuals could substantially lower mission costs. However, as the models used currently at NASA to project space radiation risks are based on mortality data from population studies and do not include analysis of risk based on individual sensitivity, it is not currently recommended that genetic testing be performed on astronauts (NCRP 2010). Given the rapid advancement in genomics and personalized medicine, this type of assessment is likely scientifically achievable within the timeframe currently planned for a human deep exploration mission. Ultimately, for a high risk and
high cost endeavor such as a mission to Mars, screening astronauts for increased resistance to space radiation may be sought in order to reduce the costs of the missions or to support post mission disease surveillance.

D. Current NASA Permissible Exposure Limits (PELs)

Permissible Exposure Limits (PELs) for short-term and career astronaut exposures to space radiation have been approved by the NASA Chief Health and Medical Officer, and requirements and standards for mission design and crew selection have been set (NASA STD-3001 Vol 1, Rev A). This section describes the cancer risk section of the PELs.

1. Career Cancer Risk Limits

Career exposure to radiation is limited to not exceed 3% risk of exposure induced death (REID) from fatal cancer. NASA policy is to assure that this risk limit is not exceeded at a 95% confidence level using a statistical assessment of the uncertainties in the risk projection calculations to limit the cumulative effective dose (in units of Sievert) received by an astronaut throughout his or her career (NASA STD-3001 Vol 1, Rev A). Methods for determining REID, including the uncertainties discussed previously, are summarized in Section VI.

2. The Principle of As Low as Reasonably Achievable (ALARA)

The ALARA principle is a requirement intended to ensure astronaut safety. An important function of ALARA is to ensure that astronauts do not approach radiation limits and that such limits are not considered as “tolerance values.” ALARA is especially important for space missions in view of the large uncertainties in cancer and other risk projection models. Mission programs and terrestrial occupational procedures resulting in radiation exposures to astronauts are required to find cost-effective approaches to implement ALARA (NASA STD-3001 Vol 1, Rev A).

IV. Evidence

The evidence and updates to projection models for cancer risk from low LET radiation are reviewed periodically by several prestigious bodies, which include the following organizations:

- The National Academy of Sciences Committee on the Biological Effects of Ionizing Radiation (BEIR)
- The United National Scientific Committee on the Effects of Atomic Radiation (UNSCEAR)
- The International Commission on Radiological Protection (ICRP)
- The National Council on Radiation Protection and Measurements (NCRP)

These committees release new reports on cancer risks applicable to low LET radiation exposures about every 10 years. Overall, the estimates of cancer risks between the different reports of these panels will agree to within a factor of two or less. However, there is continued controversy for doses below 50 mSv and for low dose-rate radiation because of debate over the linear no-
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threshold hypothesis often used in statistical analyses of these data. The BEIR VII report (BEIR 2006) and the UNSCEAR 2006 Report (UNSCEAR 2008) are the most recent major reports, and the reports are used in the summary that follows. Evidence for low LET cancer effects must be augmented by information on protons, neutrons and HZE nuclei, which is only available in experimental models. Such data has been reviewed several times in the past by the NCRP (1989, 2000, 2006, 2012). (Category IV)

A. Epidemiology data for low LET radiation

The human evidence presented in this section is Category III unless otherwise noted.

1. Life-span studies of atomic bomb survivors

The life-span study (LSS) of the survivors of the atomic-bombs in Hiroshima and Nagasaki, Japan, includes 120,321 persons that were registered in 1950. Amongst these were 82,214 from Hiroshima and 38,107 from Nagasaki that were either within 2.5 km of the hypocenters, between 2.5 and 10 km from the hypocenters, or not in the cities at the time of the bombings. The data is currently maintained by the Radiation Effects Research Foundation (RERF). RERF has an extensive list of publications using the LSS data and makes the data freely available to all researchers. The most recently published mortality data is from the LSS Report 14 and has follow-up from October 1, 1950 to December 31, 2003 (Ozasa 2012). The most recently published solid cancer incidence data has follow-up from January 1, 1958 with the establishment of the Hiroshima and Nagasaki population-based tumor registries to December 31, 1998 (Preston 2007). This incidence data was also used by BEIR VII and the UNSCEAR committees to develop cancer site specific excess risk models (BEIR 2006, UNSCEAR 2008). The most recently published leukemia incidence data has follow-up from October 1, 1950 to December 31, 2001 (Hsu 2013). There is a gap in knowledge of the earliest cancer that developed in the first few years after the war, which impacts the assessment of leukemia to an important extent and for solid cancers to a minor extent. Of these persons censures occur, leading to about 86,000 persons when the persons not in the cities are excluded and 113,000 persons when they are included in the analyses. Table 1 shows summary statistics of the number or persons and deaths for different dose groups. These comparisons show that the doses received by the LSS population overlap strongly with the doses of concern to NASA exploration mission (i.e. 50 to 2000 mSv).

<table>
<thead>
<tr>
<th>Known DS02 Weighted Colon Dose, mSv</th>
<th>Total</th>
<th>0-5</th>
<th>5-100</th>
<th>100-200</th>
<th>200-500</th>
<th>500-1000</th>
<th>1000-2000</th>
<th>&gt;2000</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. Subjects</td>
<td>86,611</td>
<td>38,509</td>
<td>29,961</td>
<td>5,974</td>
<td>6,356</td>
<td>3,424</td>
<td>1,763</td>
<td>624</td>
</tr>
<tr>
<td>Cancer Deaths</td>
<td>10,929</td>
<td>4,621</td>
<td>3,653</td>
<td>789</td>
<td>870</td>
<td>519</td>
<td>353</td>
<td>124</td>
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<tr>
<td>Non-cancer deaths</td>
<td>35,685</td>
<td>15,906</td>
<td>12,304</td>
<td>2,504</td>
<td>2,736</td>
<td>1,357</td>
<td>657</td>
<td>221</td>
</tr>
</tbody>
</table>

Figure 3 shows the dose response for the excess relative risk (ERR) for all solid cancers from Ozasa et al. (2012). The most recent publications from BEIR and UNSCEAR have used the incidence data to develop site specific cancer models. The main differences in the preferred models
from RERF, BEIR and UNSCEAR are the parameterization of the modification terms for age at exposure, attained age, and time since exposure. BEIR VII made the assumption that most individual cancer sites should have similar modification terms for age at exposure and attained age. Hence they used the parameters from the all solid cancer model for an individual site unless there was evidence of departure from the all solid cancer parameter. UNSCEAR however did not make that assumption and only included modification terms that improved the individual sites fit. UNSCEAR also characterized the age at exposure parameter differently than RERF and BEIR.

Figure 3. From Ozasa et al. (2012): Excess relative risk (ERR) for all solid cancer in relation to radiation exposure. The black circles represent ERR and 95% CI for the dose categories, together with trend estimates based on linear (L) with 95% CI (dotted lines) and linear-quadratic (LQ) models using the full dose range, and LQ model for the data restricted to dose <2 Gy.

Table 2 compares the ERR per Gy for subjects at the attained age of 70 years after exposure at age 30 for males and females calculated using the models developed by Ozasa et al. 2012, Preston et al. 2007, BEIR VII, and UNSCEAR 2006 for selected major cancer sites. Similarly, Table 3 compares the EAR per person years per Gy for subjects at the attained age of 70 years after exposure at age 30 for males and females. The National Cancer Institute has extended the results from BEIR VII to include more cancer sites in the RadRAT tool, an online calculator for estimating the lifetime risk of cancer incidence from exposure to ionizing radiation (doses below 1 Gy), for members of the U.S. population and other selected countries they have developed (Berrington de Gonzalez 2012; https://irep.nci.nih.gov/radrat).

RERF is continually updating the LSS data and it is beneficial to review the latest publications from their researchers. An update to the incidence data should be released soon. The most recent publications from RERF have focused more closely on several major cancer sites and looked at
potential confounders to the radiation effects. Most notable is the publication from Furukawa et al. (2010) that looked at the combined effects of radiation and smoking on lung cancer incidence.

Table 2. Male and Female ERR per Gy for subjects at the attained age of 70 years after exposure at age 30 using models with effect modification. Models including modifications for age at exposure and attained age are available for mortality from Ozasa et al. 2012 and for incidence from Preston et al. 2007, BEIR VII, and UNSCEAR 2006. * indicates the model does not include modifications. ** indicates that mortality data was used for the UNSCEAR all solid cancer model. NA indicates that a model is not available.

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<tbody>
<tr>
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<td>Female ERR/Gy</td>
<td>Male ERR/Gy</td>
<td>Female ERR/Gy</td>
</tr>
<tr>
<td>All solid cancer</td>
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<td>0.57</td>
<td>0.35</td>
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<tr>
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<td>0.52*</td>
</tr>
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</tr>
<tr>
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<td>0.73</td>
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</tr>
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</tr>
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</tr>
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<tr>
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<td>1.57</td>
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<tr>
<td>Brain</td>
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<td>NA</td>
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<td>0.62*</td>
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<tr>
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</tr>
<tr>
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<td>0.11*</td>
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</tr>
<tr>
<td>Uterus</td>
<td>0.22*</td>
<td>0.10*</td>
<td></td>
<td></td>
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<tr>
<td>Thyroid</td>
<td>NA</td>
<td>NA</td>
<td>0.49</td>
<td>0.65</td>
</tr>
</tbody>
</table>

2. Other Human Studies

The BEIR VII report (BEIR 2006) and the UNSCEAR 2006 report (UNSCEAR 2008) contain extensive reviews of data sets from human populations, including nuclear reactor workers and patients treated with radiation. The report from Cardis (Cardis et al. 2007) described a multinational study for reactor workers in several countries. Pooled analysis from several studies have been performed at specific cancer sites, including breast and thyroid (BEIR 2006). These studies require adjustments for photon energy, dose-rate, and country of origin as well as adjustments made in single population studies. The preferred models from the BEIR VII committee implement the results from the pooled analyses for breast and thyroid cancer. Preston et al. (2010) reanalyzed the Mayak workers data and the data for people who lived along the Techa River using Bayesian methods to estimate site-specific risk estimates for the two studies separately. The two studies were compared to results from similar methods applied to the LSS data. These types of analysis lend confidence to risk assessments as well as show limitations of such data sets. Of special interest to NASA is the age at exposure dependence of low LET cancer
Table 3. Male and Female EAR per $10^4$ person years (PY) per Gy for subjects at the attained age of 70 years after exposure at age 30 using models with effect modification. Models including modifications for age at exposure and attained age are available for mortality from Ozasa 2012 and for incidence from Preston 2007, BEIR VII, and UNSCEAR 2006. * indicates the model does not include modifications. ** indicates that mortality data was used for the UNSCEAR all solid cancer model. NA indicates that a model is not available.

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<td>Male 4 PY/Gy</td>
<td>Female 4 PY/Gy</td>
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<tr>
<td>All solid cancer</td>
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<td>27.66</td>
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<td>NA</td>
<td>0.58*</td>
<td>0.58*</td>
</tr>
<tr>
<td>Stomach</td>
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<td>5.27</td>
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<td>9.70</td>
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</tr>
<tr>
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<td>NA</td>
<td>-0.01*</td>
<td>-0.01*</td>
</tr>
<tr>
<td>Lung</td>
<td>7.30</td>
<td>5.70</td>
<td>6.00</td>
<td>9.10</td>
</tr>
<tr>
<td>Breast</td>
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<td>5.30</td>
<td></td>
</tr>
<tr>
<td>Bladder</td>
<td>1.71</td>
<td>0.69</td>
<td>3.80</td>
<td>2.60</td>
</tr>
<tr>
<td>Brain</td>
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<td>NA</td>
<td>0.51*</td>
<td>0.51*</td>
</tr>
<tr>
<td>Ovary</td>
<td>0.20</td>
<td></td>
<td>0.56*</td>
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</tr>
<tr>
<td>Prostate</td>
<td>NA</td>
<td>0.34*</td>
<td>0.17</td>
<td></td>
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<tr>
<td>Uterus</td>
<td>NA</td>
<td></td>
<td>0.56*</td>
<td></td>
</tr>
<tr>
<td>Thyroid</td>
<td>NA</td>
<td>NA</td>
<td>0.50</td>
<td>1.90</td>
</tr>
</tbody>
</table>

Risk of Radiation Carcinogenesis

In considering radiation risks for astronauts, it is useful to consider historical recommendations that NASA has received from external advisory committees. These recommendations have formed the basis for dose limits and risk projection models (Cucinotta et al. 2002). Early radiation effects usually are related to a significant fraction of cell loss, exceeding the threshold for impairment of function in a tissue. These are “deterministic” effects, so called because the statistical fluctuations in the number of affected cells are very small compared to the number of cells required to reach the threshold (ICRP 1991). Maintaining dose limits can ensure that no occurrence of early effects
take place; these are expected to be accurately understood. Late effects can result from changes in a very small number of cells, so that statistical fluctuations can be large and some level of risk is incurred even at low doses. Referring to them as “stochastic” effect recognizes the predominance of statistical effects in their manifestation.

Recommendations by NAS/NRC in 1967 (NAS/NRC 1967) noted that radiation protection in manned spaceflight is philosophically distinct from protection practices of terrestrial workers because of the high-risk nature of space missions. The report by NAS from 1967 did not recommend “permissible doses” for space operations, noting the possibility that such limits may place the mission in jeopardy and instead made estimates of what the likely effects would be for a given dose of radiation.

In 1970, the NAS Space Science Board made recommendations of guidelines for career doses to be used by NASA for long-term mission design and manned operations. At that time, NASA employed only male astronauts and the typical age of astronauts was 30-40 years. A “primary reference risk” was proposed equal to the natural probability of cancer over a period of 20 years following the radiation exposure (using the period from 35 to 55 years of age) and was essentially a doubling dose. The estimated doubling dose of 382 rem (3.82 Sv), which ignored a dose-rate reduction factor, was rounded to 400 rem (4 Sv). The NAS panel noted that their recommendations were not risk limits, but rather a reference risk, and that higher risk could be considered for planetary missions or a lower level of risk for a possible space station (NAS/NRC 1970). Ancillary reference risks were described to consider monthly, annual, and career exposure patterns. However, the NAS recommendations were implemented by NASA as dose limits used operationally for all missions until 1989.

At the time of the 1970 NAS report, the major risk from radiation was believed to be leukemia. Since that time the maturation of the data from the Japanese atomic bomb (AB) survivors has led to estimates of higher levels of cancer risk for a given dose of radiation, including the observation that the risk of solid tumors following radiation exposure occurs with a higher probability than leukemia, although with a longer latency period before expression. Figure 4 illustrates the changing estimates of cancer risks since 1970 for an average adult worker. Along with the maturation of the AB data, re-evaluation of the dosimetry of the AB survivors, scientific assessments of the dose response models, and dose-rate dependencies have contributed to the large increase in the risk estimate over this time period (1970-1997). The possibility of future changes in risk estimates can, of course, not be safely predicted today, and it is possible that such changes could potentially impact NASA mission operations. Thus protection against uncertainties is an ancillary condition to the ALARA principle, suggesting conservatism as workers approach dose limits.

By the early 1980’s, several major changes had occurred leading to the need for a new approach to define dose limits for astronauts. At that time NASA requested the NCRP to re-evaluate dose limits to be used for LEO operations. Considerations included the increases in estimates of radiation-induced cancer risks, the criteria for risk limits, and the role of the evolving makeup of the astronaut population from male test pilots to a larger diverse population (~100) of astronauts, including mission specialists, female astronauts, and career astronauts of higher ages that often participate in several missions. In 1989, the NCRP Report No. 98 recommended age and sex dependent career dose limits using as a common risk limit of a 3% increase in cancer mortality. The limiting level of 3% excess cancer fatality risk was based on several criteria, including comparison to dose limits for ground radiation workers and to rates of occupational death in the
Risk of Radiation Carcinogenesis

less-safe industries. It was noted that astronauts face many other risks, and adding an overly large radiation risk was not justified. It also should be noted that the average years of life loss from radiation induced cancer death, about 15 years for workers over age 40-y, and 20 years for workers between 20 and 40 years, is less than that of other occupational injuries. A comparison of radiation-induced cancer deaths to cancer fatalities in the US population is also complex because the smaller years of life loss in the general population where most cancer deaths occurring above age 70.

![Figure 4](image)

Figure 4. Estimates of the risk per Sv delivered at low dose-rates for the average adult worker from 1970 to 1997.

In the 1990’s, the additional follow-up and evaluation of the AB survivor data led to further increases in the estimated cancer risk for a given dose of radiation. Recommendations from the NCRP (NCRP 2000), while keeping the basic philosophy of risk limitation in their earlier report, advocate significantly lower limits than those recommended in 1989 (NCRP 1989). Table 4 lists examples of career radiation limits for a career duration of 10 years with the doses assumed to be spread evenly over a career. The values from the previous report are also listed for comparison. Both of these reports specify that these limits do not apply to exploration missions because of the large uncertainties in predicting the risks of late effects from heavy ions.

Table 4. Career dose limits (in Sv) corresponding to 3% excess cancer mortality for 10-year careers as a function of age and sex as recommended by the National Council on Radiation Protection and Measurements (NCRP 1989; NCRP 2000).

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<th>NCRP Report No. 98</th>
<th>NCRP Report No. 132</th>
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<td></td>
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<td>Female</td>
</tr>
<tr>
<td>25</td>
<td>1.5</td>
<td>1.0</td>
</tr>
<tr>
<td>35</td>
<td>2.5</td>
<td>1.75</td>
</tr>
<tr>
<td>45</td>
<td>3.2</td>
<td>2.5</td>
</tr>
<tr>
<td>55</td>
<td>4.0</td>
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The NCRP Report No. 132 (NCRP 2000) notes that the use of comparisons to fatalities in the less-safe industries advocated by the NCRP in 1989, was no longer viable because of the large improvements made in ground-based occupational safety. Table 5 shows an update to such a comparison, and indeed the decreased rate of fatalities in the so-called less safe industries, such as mining and agriculture, would suggest a limit below the 3% fatality level today compared to the 1989. The most recent reviews of the acceptable levels of radiation risk for LEO, including a 1996 NCRP symposium (NCRP 1997), and the recent report on LEO dose limits from the NCRP (2000) instead advocate that comparisons to career dose limits for ground-based workers be used. It is also widely held that the social and scientific benefits of spaceflight continue to provide justification for the 3% risk level for astronauts participating in LEO missions.

In comparison to the NASA limits, the US nuclear industry uses age-specific limits that neglect any sex dependence. Here career limits are set at a total dose equivalent equal to the individuals Age × 0.01 Sv. It is estimated by the NCRP that ground workers that reach their dose limits would have a lifetime risk of about 3%, but note the differences in dose values corresponding to the limit due to differences in how the radiation doses are accumulated over the worker’s career. NASA’s short-term (30 day and 1-year) dose limits are several times higher than that of terrestrial workers because they are intended to prevent acute risks, while annual dose limits of 50 mSv (5 rem) followed by US terrestrial radiation workers control the accumulation of career doses.

The exposures received by radiation workers in reactors, accelerators, hospitals, etc. rarely approach dose limits with the average annual exposure of 1 to 2 mSv, which is a factor of 25 below the annual exposure limit, and significantly less than the average dose for a 6-month ISS mission (100 mSv). Similarly, transcontinental pilots, although not characterized as radiation workers in the US, receive annual exposures of about 1 to 5 mSv and enjoy long careers without approaching exposure limits recommended for terrestrial workers in the US. Under these conditions, ground-based radiation workers are estimated to be well below the career limits, even if a 95% confidence level is applied. Because space missions have been relatively short in the past, requiring minimal mitigation, the impact of dose limits when space programs actually approach such boundaries, including the application of the ALARA principle, has been unexplored.
Summary of Approaches for Setting Acceptable Levels of Risk

The various approaches to setting acceptable levels of radiation risks are summarized here (IOM 2014):

1. **Comparison to Occupational Fatalities in Less-Safe Industries**: The life-loss from attributable radiation cancer death is less than from most other occupational deaths. Also, at this time this comparison would be very restrictive on ISS operations because of continued improvements in ground based occupational safety over last 20 years.

2. **Comparison to Cancer Rates in General Population**: The life-loss from radiation-induced cancer deaths can be significantly larger than from cancer deaths in the general population, which often occur late in life >70 years.

3. **Doubling dose for 20-yrs following exposure**: Provides a roughly equivalent comparison based on life-loss from other occupational risks or background cancer fatalities during the worker’s career; however, this approach negates the role of mortality effects later in life.

4. **Use of Ground-based worker limits**: Provides a reference point equivalent to standard set on Earth and recognizes that astronauts face other risks. However, ground workers remain well below dose limits, and are largely exposed to low-LET radiation where the uncertainties of biological effects are much smaller than for space radiation.

A review of cancer and other radiation risks is provided by the NCRP Report No.153 (2006). The stated purpose of this Report is to identify and describe information needed to make radiation protection recommendations for space missions beyond LEO. The report contains a comprehensive summary of the current body of evidence for radiation-induced health risks and makes recommendations on areas requiring future experimentation. NCRP Report 23 (2014) provides a supplement to previous recommendations.

C. Past Space Missions

The doses on past space missions may be characterized using a variety of physical and biological dosimetry and radiation transport models (Wilson et al. 2004; Ballarini et al. 2006; Bernabeu and Casanova 2007; Cucinotta et al. 2008; Walker et al. 2013). Phantom torso experiments have also been performed on ISS and space shuttle (Badhwar 2000; Yasuda 2000; Cucinotta et al. 2008). The combined accuracy, or uncertainty, of these models is determined through validation against ground-based experiments (Walker et al. 2011; Sihver et al. 2012; Norman and Blattning 2013) and space-flight measurements (Sato et al. 2006; Mrigakshi et al. 2012; Slaba et al. 2013b; Wilson et al. 2014; Wilson et al. 2015) as well as verification through inter-code comparisons and benchmarking (Heinbockel et al. 2011a, 2011b; Lin et al. 2012; Slaba et al. 2013b). Such studies continue to be pursued so that computational models can be confidently used to assess exposure levels where measurements are unavailable and overall physics uncertainty can be more rigorously quantified. In addition, until 2013, cytogenetic biodosimetry was performed on US crewmembers of most ISS missions and on four astronauts who participated in Mir missions. Measureable increases in the yield of chromosome damage have been detected in the blood lymphocytes of astronauts after space missions of 3 to 6 months and this provides an alternative evaluation of organ dose equivalents that includes individual radio-sensitivity in the
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presence of confounding factors such as microgravity. Cancer risk projections can also be obtained directly from cytogenetic data using the quantitative approach derived from the European Study of Cytogenetic Biomarkers and Health, a study of several thousand persons, which has shown healthy individuals with Medium (M), and High (H) levels of chromosome aberrations in their lymphocytes have a significant increase in cancer incidence and mortality compared to the Low (L) group (Hagmar et al. 1998). This approach has been used to assess cancer risk for astronauts (George et al. 2013a). Figure 5 shows tertile risk rankings for 30 astronauts before flight and within three weeks of return from their ISS missions (panels A and B, respectively). Individual tertile rankings increased after space flight and only one individual remained in the low category. Analysis of follow up samples from 26 of the original 30 astronauts, taken at least 6 months after their respective flights (Figure 5, panel C), show that the tertile rankings remained in the high category for more than 50% of these individuals. Crewmembers with increases that remain in the high category are projected to have a significant increase in life-time cancer risk.

Figure 5. Age adjusted tertile risk rankings for astronaut assessed before (panel A), a short time after (panel B), and 6 to 18 months after (panel C) a space mission. Age adjusted yield of total chromosome damage is plotted for each individual. Solid lines indicate the cut off for the low, medium, and high groups. Individual tertile rankings increased after space flight and only one individual remained in the low category. Follow up analysis show that 50% of these individuals remained in the high category at least 6 months after the mission.

Biodosimetry has also been performed on five astronauts who participated in two separate ISS missions, and results show consistent increases in chromosome aberration yield in lymphocytes after the second flight as shown in Figure 6. The five plots in Figure 6 represent individual astronauts (A to E) and show a time course of aberration yields per 1000 cells plotted against time after first blood draw and supports the assumption of additivity of biological doses is generally valid for ISS crew exposures.
Figure 6. From George et al. (2013a). A time course of aberration yields per 1000 cells plotted against time after first blood draw for five astronauts (A to E) and show and supports the assumption of additivity of biological doses is generally valid for ISS crew exposures. The shaded areas indicate the time spent in space during the mission and the line on plot A signifies the timing of a short duration shuttle mission lasting 11 days. This preliminary study supports the assumption of additivity of biological doses is generally valid for ISS crew exposures.

Figure 7 shows results for the pre-flight and post-flight frequency of translocations, complex aberrations, and total exchanges. Total exchanges are increased post-flight over pre-flight values in all cases, and translocations increase in all ISS astronauts, but not for two astronauts - one returning from the Mir station and one on a Hubble repair mission. To test if the overall frequency of complex aberrations was increased by space radiation, Cucinotta et al. (2008) pooled results into two groups: all ISS data and all ISS data plus results from other NASA missions. The observed increase in complex aberrations is highly significant (P<10^-4), as is the increase in translocation frequency.
Figure 7. From Cucinotta et al. (2008) the frequency of translocations, complex aberrations, or total chromosome exchanges measured in each astronaut’s blood lymphocytes before and after their respective space missions on ISS, Mir, or STS. An increase in total exchanges was observed for all astronauts. Translocations (22 of 24) and complex aberrations (17 of 24) were increased in the majority of astronauts.

Figure 8 provides a summary of the crew doses for all NASA missions through 2007. The cancer projection model of NCRP Report No. 132 (NCRP 2000) can be applied to these effective doses and indicate REID values approaching 1% for many astronauts that have flown on ISS or the Russian space station Mir (Cucinotta et al. 2001).
V. Radiobiology Evidence for Protons and HZE nuclei

Transferring risk estimates derived from low-LET radiation to risk from exposure to the high-LET radiation environment found in space requires new knowledge on the fundamental differences in biological responses (the so-called radiation quality effects) triggered by heavy ion particle radiation versus low-LET radiation associated with Earth-based exposures. The analysis of radiation quality takes into consideration the spatial pattern of energy distribution between different types of radiation and how this relates to the biological effects. Because the energy transferred per unit distance along a heavy ion track is much higher than that deposited for the same dose of low-LET radiation, the amount and type of damage and subsequent activation of damage response systems in the cell are expected to be different and studies with protons and HZE nuclei of RBEs for molecular, cellular and tissue endpoints, including tumor induction, document the higher risk for space radiation components (NAS 1996; NCRP 2006; Cucinotta and Durante 2006). This evidence must be extrapolated to the chronic conditions in space and from the mono-energetic beams used at NSRL and other accelerators to the complex mixed radiation types in space. Sufficient proof that experimental models represent cancer processes in humans, including estimating the effectiveness of shielding and biological countermeasures, must be obtained for high risk missions where acceptable levels of cancer risks are approached or perhaps exceeded. Evidence and progress in these areas is described next.
A. Cancer Induction by Space Radiation

A necessary step for improving space radiation cancer risk assessment are studies on the molecular pathways causative of cancer initiation and progression, and to extend these studies to learn how such pathways can be disrupted by HZE ions, including both genetic and epigenetic modifications noted as the hallmarks of cancer (Figure 9) (Hanahan and Weinberg 2000, 2011). Recent studies have provided insight into what has become known as the cancer niche, which involves the tumor microenvironment comprising cancer cells, and surrounding stromal and immune cells and their interactions, all of which are necessary to support tumor growth and cancer progression (Barcellos-Hoff et al. 2013; Illa-Bochaca et al. 2014; Turley et al. 2015). This has generated a deeper understanding of tumor properties, and knowledge on how these interactions are modulated by radiation may lead to potential target for future countermeasure approaches. The goal of space radiation research is to establish a more mechanistic approach for understanding the biological impacts in order to better estimate risk and to answer questions, including whether HZE effects be can scaled from those of gamma-rays, whether risk is linear with low dose-rate, and how individual radiation sensitivity impacts risks for astronauts, a population selected for many factors related to excellence in health.

![Diagram of Space Radiation](image)

**Figure 9.** The hallmarks and emerging hallmarks of cancer (Hanahan and Weinberg 2011), enabling characteristics and possible mechanisms of radiation damage that lead to these changes observed in all human tumors.

1. The Initial Biological Events

Energy deposition by HZE ions is highly heterogeneous with a localized contribution along the trajectory of each particle and lateral diffusion of energetic electrons (δ-rays) many microns from the ion path (Goodhead 1994; Cucinotta et al. 2000). These particles are therefore characterized by a high-LET; however, they contain a low-LET component due to the high-energy
Risk of Radiation Carcinogenesis

electrons ejected by ions as they traverse tissue. Biophysical models have shown that the energy deposition events by high-LET radiation produce differential DNA lesions, including complex DNA breaks, and that there are qualitative differences between high- and low-LET radiation both in the induction and repair of DNA damage (Prise et al. 1998; Sutherland et al. 2000; Rydberg et al. 2005; Wang and Wang 2014; Saha et al. 2014). Regarding cell survival curves, the low-dose region exhibits a repair shoulder for low-LET radiation, whereas the survival curve is generally a straight line for high-LET radiation (Hall 2006). High-LET radiation has also been shown to cause much more complex chromosome rearrangements than low-LET radiation, with the aberrations involving a greater number of chromosomes and breakpoints, as well as both intra- and inter-chromosome exchanges (Durante et al. 2002; George et al. 2003; Johannes et al. 2004; Hada 2007). Furthermore, other studies have suggested that more complex clustered DNA lesions are produced by high-LET radiation compared with low-LET radiation, as evidenced by, for example, the smaller DNA fragments found following high-LET irradiation (Lobrich et al. 1996; Prise et al. 2001; Rydberg et al. 2002; Belli et al. 2002) and the demonstration that ionizing particles (e.g., protons and iron ions) yield more lesion clusters relative to double strand breaks (DSB) compared with X-rays or γ-rays (Hada 2006). This complex clustered damage is characterized by clusters containing mixtures of two or more of the various types of lesions (e.g., SSB, DSB) within a localized region of DNA. Complex damage is an uncommon consequence of endogenous damage or low-LET radiation, and it has been associated with the increased relative biological effectiveness (RBE) of densely ionizing radiation.

The repair of DSB is known to occur through direct end-joining and homologous recombination processes. Indications are 1) that for high-LET radiation, where complex DSBs occur with high frequency, little repair occurs, leading to cell death, or 2) that the mis-rejoining of unrepairable ends with other radiation-induced DSB leads to large DNA deletions and chromosome aberrations. In particular, it has been demonstrated that, regardless of radiation quality, the overall level of misrepaired damage significantly exceeds that of unrepaired damage (Loucas 2013). While the high effectiveness in cell killing provides the rationale for heavy-ion cancer therapy (hadrontherapy), residual damage in surviving cells is of concern for carcinogenesis. A comprehensive review by Sridharan et al. (2015) provides an examination of current knowledge on DNA damage and repair, associated oxidative stress and inflammation, and potential links and interplay between these early responses following HZE radiation and development of genomic instability associated with cancer development. Figure 10, from the Sridharan review, illustrates repair of simple and complex DNA lesions produced by low and high-LET radiation and their relationship to genomic instability.
Figure 10. Repair of simple and complex DNA lesions induced by low- and high-LET radiation exposure. A majority of the DNA lesions induced by low-LET irradiation are simple lesions and are repaired within hours of induction via NHEJ- and HR-mediated repair pathways, with pathway preference dependent on cell cycle. On the other hand, a majority of the high-LET radiation-induced DNA damages are clustered lesions, which may impede DNA repair pathways, causing damage to remain unrepaired for longer periods (days to weeks). In addition to radiation-induced ROS, unrepaired DNA lesions may also increase the ROS levels in cells, causing further generation of simple to complex DNA lesions. Unrepaired/misrepaired lesions in mitochondrial or nuclear DNA (dotted line) may also further enhance and perpetuate ROS levels. Ultimately, the unrepaired/misrepaired DNA lesions may promote genomic instability, leading to initiation of carcinogenesis (Sridharan et al. 2015).

2. Chromosomal Damage and Mutation

Heavy charged particles are very effective at producing chromosome exchanges with the yield of chromosome aberrations increasing linearly with dose (Hada et al. 2011; Ritter and Durante 2010). RBE values, estimated from total chromosome exchanges in human lymphocytes at the first post-irradiation mitosis, increase with LET, peaking around 100-200 keV/µm with an RBE of 35, and then decrease sharply at higher LET (George et al. 2007). The detailed RBE versus LET relationship found for chromosome exchanges is similar to studies of mutation (Kiefer et al. 2002; Liber et al. 2014), in vitro neoplastic transformation (Yang et al. 1985), and induction of solid tumors in mice (Bielefeldt-Ohmann et al. 2012). Additional studies of radiation quality
dependences of chromosome aberrations indicate that at a fixed value of LET, particles with lower charge number (Z) have a higher RBE compared to particles with a higher Z, and a saturation cross section was observed for different radiation qualities (George et al. 2013b). RBE values for mutation induction and chromosome damage indicate that low energy protons are significantly higher than unity and values are LET dependent (Belli et al. 1993; Schmid et al. 1998). Yields of chromosome damage are similar for acute exposures of higher energy protons (5 to 2500 MeV), with RBE values for total exchanges close to unity and approaching an RBE of 2 for low dose exposures due to an increased number of complex exchanges at all proton energies compared to γ-rays (George et al. 2015).

The quality of chromosome damage is different when heavy ions are compared to sparsely ionizing radiation. Multi-color fluorescence painting techniques of human chromosomes have clearly demonstrated that HZE ions and protons induce a higher percentage of complex-type chromosome exchanges when compared to acute doses of low-LET radiation, and the complexity of observed rearrangements increases with increasing LET. Most of these complex chromosomal rearrangements will ultimately lead to cell death. In fact, only a small fraction of the initial damage is transmitted in mice 2 to 4 months after the exposure to energetic iron ions. Large differences in gene expression are observed between X-rays and HZE ions reflecting differences in damage response pathways (Ding et al. 2005, 2013, 2015). Qualitative differences in the type of gene mutations have also been reported (Kronenberg et al. 1994, 1995; Hryciw et al. 2015). A low RBE for the induction of late chromosomal damage has been measured in the progeny of human lymphocytes exposed in vitro to energetic iron ions, with the interesting exception of terminal deletions, that occurred with much higher frequency in the progeny of cells exposed to heavy ions compared to gamma-rays (Durante et al. 2002). Persistent loss of heterozygosity (LOH) on multiple chromosomes and severe karyotypic instability has been observed in the progeny of mammary epithelial cells that survived X-ray or iron-ion exposure (Sudo et al. 2008), while LOH on multiple chromosomes has also been detected in vivo following exposure to high energy protons (Grygoryev et al. 2014).

3. Genomic Instability

Genomic instability in the progeny of cells irradiated with heavy ions has been observed both in vitro and in vivo in several model systems (Nagar and Morgan 2005; Mao et al. 2005; Hu et al. 2012; Werner et al. 2014). This high rate of abnormal genetic change includes single nucleotide mutations, structural alterations in chromosomes as well as numerical changes in whole chromosomes, and is considered a major driver of the carcinogenic process (Hanahan and Weinberg 2011). Sabatier et al. (1992; 2005) found that rearrangements involving telomere regions are associated with chromosomal instability in human fibroblasts many generations after exposure to accelerated heavy ions. Telomere dysfunction plays a crucial role in initiating or sustaining genomic instability (Sishc et al. 2015). Cells containing telomere-deficient chromosomes will either senesce, or undergo B/F/B cycles, promoting genetic instability. The fate of normal cells containing a single terminal deletion is not known, but it has been shown that the loss of a single telomere in cancer cells can result in instability in multiple chromosomes (Shim et al. 2014, Feldser et al. 2003; Mase and DePinho 2002). These recent results suggest that telomere instability could be an important early event in the pathway to cancer induction by HZE nuclei. Shortened telomeres
are currently being examined as biomarkers of the development of secondary malignant neoplasms (Shay 2014).

4. Cancer and Tissue Effects

The number of studies of animal carcinogenesis with HZE nuclei is growing (Bielefeldt-Ohmann et al. 2012; Rivina and Schiestl 2013; Sridharan et al. 2015; Barcellos-Hoff et al. 2015) and summarized in Table 6. In general, these animal studies demonstrate that the tumor spectra observed in irradiated animals is similar between low and high-LET irradiated animals and is dependent on the susceptibility of the specific model strain used, and HZE nuclei in general exhibit a higher carcinogenic effectiveness compared to low-LET radiation for induction of solid tumors. Relative biological effectiveness factors comparing gamma-rays to HZE ions have been measured in mice or rats for incidence for tumors of the skin (Burns et al. 1993), Harderian gland (Fry et al. 1985; Alpen et al. 1993), mammary gland (Dicello et al. 2004), lung (Delgado et al. 2014; Wang et al. 2015), blood and liver (Weil et al. 2014). RBE values reported for tumor induction range from 6-10 for lung adenocarcinomas to values as high as 25-50 for other solid cancers. RBE dependence on HZE radiation quality has been most extensively characterized in studies of mouse Harderian gland tumorigenesis. In this model, the RBE increases with LET and plateaus in the 193-953 keV µm⁻¹ range (Fry et al. 1985; Alpen et al. 1993). RBE values for acute myeloid leukemia are closer to 1 (Weil et al. 2009), with acute myeloid leukemias from both low-LET and high-LET animals exhibiting similar molecular characteristics (Steffen et al. 2013).

Table 6 High-LET Mouse Model Studies (Sridharan et al. 2015)

<table>
<thead>
<tr>
<th>Rodent model/gender/age</th>
<th>Monitoring time after exposure</th>
<th>Radiation quality</th>
<th>LET KeV/µm</th>
<th>Radiation regimen</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sprague Dawley rats, male (4 weeks)</td>
<td>~78 weeks</td>
<td>Electrons</td>
<td>0.34</td>
<td>Partial body</td>
</tr>
<tr>
<td>BALB/c mice, female (12 weeks)</td>
<td>Life time</td>
<td>Neutron</td>
<td>0.25</td>
<td>Whole body</td>
</tr>
<tr>
<td>B6CF1/AnIL mice, female (14–17 weeks)</td>
<td>~86 weeks</td>
<td>Argon</td>
<td>0.125</td>
<td>Partial body</td>
</tr>
<tr>
<td>Sprague Dawley rats, female (8.6 weeks)</td>
<td>Life time</td>
<td>Iron</td>
<td>150</td>
<td>Whole body</td>
</tr>
<tr>
<td>CBA/CAJ mice, male (8–14 weeks)</td>
<td>~157 weeks</td>
<td>γ-rays</td>
<td>0.8</td>
<td>Whole body</td>
</tr>
<tr>
<td>C57Bl/6 ApcMin/+ female and F1(AKR/JApcMin/+) male and female mice (6–8 weeks)</td>
<td>14–20 weeks</td>
<td>γ-rays</td>
<td>0.8</td>
<td>Whole body</td>
</tr>
<tr>
<td>AItm3Sri mice, male and female (5–15 weeks)</td>
<td>~104 weeks</td>
<td>Iron</td>
<td>148</td>
<td>Whole body</td>
</tr>
<tr>
<td>Lck-Bax (Lck-Bax38R1 and Sint3-/-Lck-Bax38&amp;1) mice, male and female (4–9.2 weeks)</td>
<td>Life time or until euthanasia</td>
<td>γ-rays</td>
<td>0.8</td>
<td>Whole body</td>
</tr>
<tr>
<td>Apemin/+ mice, female (6–8 weeks)</td>
<td>~14–16 weeks</td>
<td>Iron</td>
<td>148</td>
<td>Whole body</td>
</tr>
<tr>
<td>C3H/HeNoNrl mice, male (8–10 weeks)</td>
<td>~114 weeks</td>
<td>γ-rays</td>
<td>0.8</td>
<td>Whole body</td>
</tr>
</tbody>
</table>

30
There have been several recent reports of increased metastatic potential of tumors in models of HZE carcinogenesis (Trani et al, 2010, 2014; Datta et al. 2013, Weil et al. 2014; Illa-Bocacha et al. 2014); and reports of acceleration of cancer progression with dose fractionation (Delgado et al. 2014). However, the risk and detriment of cancer will not be fully characterized until the relationship between radiation quality, dose, and latency, where tumors appear earlier after high-LET irradiation, is adequately described. The earlier latency and increasing effectiveness found with HZE ions similar to earlier studies with neutrons (Ullrich 1984; Fry and Storer 1987), along with the lack of response of gamma-rays seen in many low dose studies, suggests that the scaling concepts using in current risk assessment approaches are unable to describe important qualitative effects and that relative biological effectiveness factors may in principle be indefinable or a faulty concept.

Further studies employing additional animal models will be necessary to understand the risk of radiation carcinogenesis due to HZE ions along with studies to protect against initiation or progression of the disease. Some key questions still unanswered are outlined in the recent review by Barcellos-Hoff et al. (2015).

It is clear that ionizing radiation acts not only as a cancer initiator, where it directly imparts DNA damage and mutation, but it acts also as a cancer promoter, where it influences cancer development through a variety of indirect mechanisms that foster growth of pre-initiated cells through changes in the tissue microenvironment. Several studies have debated the relative importance of these effects comparing low and high-LET radiation impacts on direct DNA damage and mutation or on microenvironment effects such as extracellular matrix remodeling, intercellular communication and other non-targeted effects as contributors to carcinogenesis (Barcellos-Hoff et al. 2005, 2013; Illa-Bochaca et al. 2014). Tissue effects independent of DNA damage that have

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**Table 6 Extended.**

<table>
<thead>
<tr>
<th>Type of cancer</th>
<th>Summary of findings</th>
<th>Refs.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Skin</td>
<td>Skin tumor prevalence increased for Neon and Argon ions, and was similar for Argon and Iron ions. High LET more effective than low LET in inducing carcinogenesis. Dose fractionation lowers tumor incidence with electron exposure; no dose reduction or increased risk found after fractionation effect with high LET.</td>
<td>Burns et al.</td>
</tr>
<tr>
<td>Ovarian tumors, lung and mammary adenocarcinomas</td>
<td>Ovarian tumor induction similar for neutrons and γ rays up to ~0.5 Gy. For lung and mammary tumors dose-response curve after neutrons showed “bending-over” in dose range from 10–20 rad.</td>
<td>Ullrich</td>
</tr>
<tr>
<td>Harderian gland</td>
<td>Tumorigenic effects increase with LET up to 193 KeV/μ. Fractionated exposure 1.5x more effective than acute exposures.</td>
<td>Alpen et al.</td>
</tr>
<tr>
<td>Mammary</td>
<td>Increased incidence up to 1.5 Gy for protons and photons and saturation for higher doses; up to 0.6 Gy for Fe, with bending over at higher doses. Tumor sparing with fractionated high-LET exposure.</td>
<td>Fry et al.</td>
</tr>
<tr>
<td>AML and HCC</td>
<td>AML-linear increase with dose, no difference between high and low LET. HCC-linear increase in 0-0.4 Gy range and “bending-over” at higher doses. High LET more effective inducer of solid tumors than leukemias than low LET.</td>
<td>Weil et al.</td>
</tr>
<tr>
<td>Intestinal tumors</td>
<td>Significant increase in tumor burden post iron exposure in comparison to γ-rays in ApcMin/+ mice, no significant LET effect in Fli(AKR)/XApcMin/+ mice.</td>
<td>Trani et al.</td>
</tr>
<tr>
<td>Lymphomas and HCC</td>
<td>No increase for lymphomas. Incidence of HCC doubled after Fe in both wt and AtnASIS mice, but difference was not significant.</td>
<td>Yamamoto et al.</td>
</tr>
<tr>
<td>Thymic lymphomas</td>
<td>Dose-dependent acceleration of lymphomagenesis after iron ions, no increase after silicon ions. Lymphomagenesis due to Bax overexpression is enhanced in a LET and gender dependent manner. Mitochondrial dysfunction and increased superoxide may play a role in this phenotype.</td>
<td>Jacobus et al.</td>
</tr>
<tr>
<td>Intestinal tumors, and invasive carcinomas</td>
<td>Significant increase in intestinal tumor frequency and larger and higher grade tumors with iron ions in comparison to γ-rays.</td>
<td>Datta et al.</td>
</tr>
<tr>
<td>AML and HCC</td>
<td>No increase AMI incidence after silicon or iron ions compared to γ-rays or protons. Increased incidence for HCC after silicon or iron when compared to γ-rays or protons. Increase in metastatic HCC after silicon or iron when compared to γ-rays or protons.</td>
<td>Weil et al.</td>
</tr>
</tbody>
</table>
been associated with cancer initiation or progression include genomic instability (Park et al. 2003), extracellular matrix remodeling, persistent inflammation, oxidative damage (Mothersill et al. 2004; Werner et al. 2014) and cell migration and invasion (Patel et al. 2012). Other studies are exploring possible relationships between radiation and the activation of dormant tumors and modulation of angiogenesis (Folkman et al. 1989). Barcellos-Hoff et al. (2015) summarize the current status of the field of space radiation carcinogenesis and highlight the importance of elucidating the impact of radiation quality in experimental systems at the mechanistic level.

5. Bystander Effects

Bystander or non-targeted effects are defined as “effects manifesting in non-irradiated cells that received a signal(s) communicated from an irradiated cell” (Morgan and Sowa 2015), including induction of DNA damage responses in un-hit bystander cells (Klammer et al. 2015). These non-targeted effects, illustrated in Figure 11 from Li et al. (2014), may lead to supra-linear dose-response curves at low doses, perhaps reducing the effectiveness of spacecraft shielding, but they may also be protective by removing damaged cells from the organism. Both effects challenge the conventional linear no-threshold risk model assumption, which is currently adopted for radioprotection on Earth and in space. They also suggest important targets for biological countermeasures that are likely to be more effective than countermeasures targeting DNA damage.

To date, at least two modes of bystander signal transmission have been identified, namely, via gap junctions between cells in direct contact with each other and medium-mediated diffusion (Nelson 2003). Recent studies have focused on elucidating the nature of the transmittal agent and have identified several molecules as potential mediators of the response, such as calcium (Lyng et al. 2006), nitric oxide (Shao et al. 2008), reactive oxygen species (ROS) (Yang et al. 2005; Narayanan et al. 1997; Autsavapromporn et al. 2013), cytokines such as interleukin-8 (Narayanan et al. 1999) and transforming growth factor-β (Shao et al. 2008), and enzymes such as Cox-2 (Zhou et al. 2005), NADPH oxidase (Azzam et al. 2002) and DNA damage response (Yang et al. 2011). The varied results from these studies suggest that the transmission mode is likely dependent on several factors, such as cell density, cell type, radiation dose, and the biological endpoint assessed. Cells have been shown to exhibit several responses to bystander signals, including genomic instability or delayed death, induction of apoptosis, enhanced cell growth, and mutations (Mothersill and Seymour 1997; Lorimore et al. 1998; Wu et al. 1999; Ponnaiya et al. 2011). Additionally, a generalized stress response and alterations in protein levels have also been detected (Lyng et al. 2000; Mothersill et al. 2001; Azzam et al. 1998).
Figure 11 (from Li et al. 2014) HZE-particle irradiation induces targeted and nontargeted (bystander) effects. Communication of stress-inducing molecules from cells exposed to an HZE particle (cells in red) and its fragmentation products (cells in orange or green) propagate stressful effects that lead to induction of oxidative stress in bystander cells (yellow). The progeny of the targeted and bystander cells may also experience oxidative stress. Cells in white are nonaffected bystander cells.

Results in tissues suggest that biological responses differ between high and low LET radiation depending on the model context considered (2D vs. 3D vs. animal). Because of the many types of particles, energies, and doses of interest in space, extensive animal experimentation has been prohibited by costs in the past. Studies involving 3D human co-culture have been an effective method to study cancer risks in a more realistic context (Barcellos-Hoff et al. 2005; Riballo et al. 2006; Yang et al. 2007). However, the results obtained from 2D and 3D cell culture models have been conflicting. For example, some studies have demonstrated decreased radiosensitivity and bystander effects in 3D compared with 2D models (Olive et al. 1994; Su et al. 2010), some have demonstrated no significant differences between models (Lin et al., 2009), and still other studies have revealed increased radiosensitivity in 3D models (Roig et al. 2009). The conflicting results from these studies, which employed a wide range of cell types and model parameters, highlight not only the importance of tissue architecture when evaluating radiation effects including bystander responses, but also the importance of investigating these responses in different cellular and tissue systems.

Several investigations have explored bystander effects in animal models. In these studies (Koturbash et al. 2006; Mancuso et al. 2008), shielding was placed over a portion of the bodies of mice prior to irradiation, and bystander effects (e.g., DNA damage, DNA methylation, and apoptosis) were noted in this shielded region. Jain et al. (2011) observed late changes in molecular signaling pathways in liver mitochondria from head only irradiated animals, further solidifying the importance of non-targeted responses in vivo. Further studies in animal models are needed to better elucidate the contribution of non-targeted effects to cancer risk.

Although many unknowns remain regarding the bystander effect and its implications, an important characteristic of this effect is that it is triggered by low radiation doses, irrespective of radiation quality, and saturates with increasing dose, usually by 10-30 cGy (Yang et al. 2005; Schettino et al. 2005; Yang et al. 2011). Thus, bystander effects are particularly important at low doses and low particle fluences and may have important consequences for astronauts on long-
duration missions due to the low particle fluences that characterize the space radiation field. However, generalizations regarding the potential risk of bystander effects cannot yet be made due to the challenges of combining bystander study results from the literature into a cohesive framework and uncertainties in extrapolating results from *in vitro* studies to predict human cancer risk.

6. **Adaptive Response**

There is general agreement that significant differences exist in cellular responses to low-dose compared to high-dose radiation exposure, which may indicate distinct underlying mechanisms. There have been several studies performed that indicate an adaptive response to low-dose ionizing radiation can provide a level of protection against future exposures (Bhattacharjee and Ito 2001; Mortazavi et al. 2003; Elmore et al. 2008; Rithidech et al. 2012). This may be particularly important for understanding risks in the space environment because the GCR environment is comprised predominantly of protons, and it is realistic to expect that cells will be exposed to multiple hits of protons prior to being traversed by an HZE particle. Several studies have begun examining whether prior exposure to protons provides any level of adaptive protection prior to the HZE exposure with diverse results that are dependent on the model system, the order of ions, time between exposures, doses used, and the endpoints being measured. For example a recent study by Buonanno et al. (2015) shows a low dose exposure to protons (20 cGy) provided protection against chromosomal damage induced by a subsequent exposure to iron ions (50 cGy). However, Elmore et al. (2011) reported no evidence for a adaptive effects on cell transformation when they examined combinations of low dose iron (10 cGy) followed by 1 Gy proton or low dose proton (10 cGy) followed by 1 Gy iron. Other studies examined the dependence on LET (Sowa et al. 2011); the impact of order or particle delivery (Sutherland et al. 2005) and impact of time intervals (Zhou et al. 2006; Bennett et al. 2007). These studies highlight the difficulties and as well as importance of development of realistic GCR simulator capabilities, described at the end of this report, to accurately assess biological impact of the mixed field environment in space.

7. **Oxidative stress and inflammation**

Oxidative stress is an imbalance between the production of reactive oxygen species (ROS) and the ability of the body to mitigate their harmful effects. It is a well-known byproduct of radiation exposure and impacts all components of the cell including proteins, DNA and lipids through the production of free radicals or peroxides, illustrated in Figure 12. It has been linked to numerous diseases, premature aging, and persistent ROS are thought to play a critical role in DNA damage, telomere dysfunction and fuel inflammatory processes which can lead to the development of cancer (Azzam et al. 2012; Sridharan et al. 2015; Li et al. 2014). Genomic instability has been
Figure 12. Effects of high atomic number (Z) and high-energy (E) (HZE) ion irradiation in mammalian cells. Traversal of a cell by a densely ionizing HZE particle results in direct and indirect cellular effects on macromolecules. Absorption of ionizing radiation by living cells directly disrupts atomic structures, producing chemical and biological changes and indirectly through radiolysis of cellular water and generation of reactive chemical species by stimulation of oxidases and nitric oxide synthases. Ionizing radiation may also disrupt mitochondrial functions significantly contributing to short- and long-term effects leading to persistent alterations in lipids, proteins, nDNA, and mtDNA (From Li et al. 2014).

highlighted as an evolving hallmark of cancer (Coleman and Tsongalis 1999; Negrini et al. 2010), and has been reported in a number of studies as a result of persistent ROS (Werner et al. 2014; Kim et al. 2006; Azzam et al. 2012). Datta et al. (2012) reported the LET-dependent presence of markers of chronic oxidative stress in intestinal tissue of mice up to one year post exposure, with higher elevation of intracellular ROS and mitochondrial superoxide in cells exposed to high LET compared to low LET. A similar study by Cheema et al. (2014) revealed distinct LET-dependent metabolomics changes related to inflammatory signaling in mouse intestinal tissue at 2 months post exposure. Additional evidence indicates that other factors associated with spaceflight, such as microgravity, may be associated with an increase in ROS production, cellular antioxidants and tissue remodeling due to a shift in biological and metabolic homeostasis which may have important implications in the context of space radiation exposure (Mao et al. 2014). A summary of ROS related studies using high-LET radiation is provided in Table 7 from Sridharan et al. 2015.
Table 7. Summary of ROS Related High-LET Studies (from Sridharan et al. 2015)

<table>
<thead>
<tr>
<th>Model System/Co-type</th>
<th>Time after exposure</th>
<th>Radiation Quality (Dose)</th>
<th>Energy MeV/u</th>
<th>INCAPSE MN</th>
<th>FOLD REDUCTION</th>
<th>~FOLD INCREASE in ROS</th>
<th>MESC Expos</th>
<th>High vs Low LET Difference</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human AG1512 fibroblasts. Co-cultured with direct exposed cells</td>
<td>1-24 h</td>
<td>Krays 0.5, 20 Gy</td>
<td>250 kV</td>
<td>3</td>
<td>~2 fold</td>
<td>~2 fold</td>
<td>∼1.5</td>
<td>MN=micronuclei, CE¼cloning efficiency. Folds not provided in article are estimated based on graphs to make comparisons possible.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>250</td>
<td>250</td>
<td>1.3@12h</td>
<td>1.5@6h</td>
<td></td>
<td>∼2 fold</td>
<td>~2 fold increase at doses</td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td>Rat Hippocampus neural precursor c.e.</td>
<td>∼0 h</td>
<td>Protons 20 Gy</td>
<td>250 kV</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.1 fold</td>
<td>MN=micronuclei, CE¼cloning efficiency. Folds not provided in article are estimated based on graphs to make comparisons possible.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>250</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.1 fold</td>
<td></td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td>C57BL/6 mouse and ENstem-A human neural stem cells</td>
<td>∼0 h</td>
<td>Protons 20 Gy</td>
<td>250 kV</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.1 fold</td>
<td>Protons more effective in inducing ROS in human vs mouse</td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>250</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.3 fold</td>
<td>1.1 fold</td>
<td></td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td>C57BL/6 mouse brain cortex</td>
<td>1 &amp; 2 m</td>
<td>y-rays (20 Gy) Fe</td>
<td>1.006</td>
<td>1.1@2 m</td>
<td>1.4@2 m</td>
<td>1.1@2 m</td>
<td>1.1@2 m</td>
<td>Protons more effective in inducing ROS than x-ray</td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>1.006</td>
<td>1.1@1.1</td>
<td>1.25@1.1</td>
<td>1.1@1.1</td>
<td>1.1@1.1</td>
<td></td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td>C57BL/6 mouse intestine</td>
<td>1 y</td>
<td>y-rays (20 Gy) Fe</td>
<td>1.006</td>
<td>1.1@1y</td>
<td>1.25@1r</td>
<td>1.1@1y</td>
<td>1.1@1y</td>
<td>Protons more effective in inducing ROS than x-ray</td>
<td>Yang et al. (2007)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>1.006</td>
<td>1.1@1y</td>
<td>1.25@1r</td>
<td>1.1@1y</td>
<td>1.1@1y</td>
<td></td>
<td>Yang et al. (2007)</td>
</tr>
</tbody>
</table>

Notes. MN¼micronuclei, CE¼cloning efficiency. Folds not provided in article are estimated based on graphs to make comparisons possible.

* Energy is MeV/u except where indicated as kVp.

** Bystander experiment indicates paper is looking at indirect effects in cells not directly exposed.

Along with chronic oxidative stress, inflammation has been identified as a critical pathway in the development of cancer and other chronic diseases. Inflammatory responses triggered after radiation exposure can act directly, or indirectly via the tissue microenvironment, to promote sustained oxidative damage, alterations in cellular gene expression and signaling pathways that enhance cell proliferation and cell survival and support other abnormal changes associated with cancer development (Hayashi et al. 2003; Colotta et al. 2009; Del Prete et al. 2011; Barcellos-Hoff et al. 2014). Several studies provide evidence for chronic inflammatory responses following HZE ion exposure at relatively long times post exposure. Lorimore and Wright (2003) exposed hematopoietic stem cells to ionizing radiation and reported genomic instability as a result of...
inflammatory responses. In a more recent study by Jangiam et al. (2015), mice exposed to $^{48}$Ti ions exhibited a dose dependent increase in both oxidative stress and inflammation in liver tissue that persisted up to 6 months following exposure, and Tungjai et al. (2013) reported similar evidence for chronic inflammation in both the heart and bone marrow of mice at 6 months post exposure to $^{28}$Si ions. A summary of high-LET inflammation related studies is Table 8 from Sridharan et al. 2015.

**Table 8. Summary of high-LET inflammation related studies (from Sridharan et al. 2015).**

<table>
<thead>
<tr>
<th>Model system/cell type</th>
<th>Time after exposure</th>
<th>Radiation quality/dose (Gy)</th>
<th>Energy Mev/n$^a$</th>
<th>Biological end points</th>
<th>Radiation quality effects</th>
<th>Refs.</th>
</tr>
</thead>
<tbody>
<tr>
<td>CS7BL/6 mice female (9–10 weeks), spleen and lymphocytes</td>
<td>113 days</td>
<td>Si (2 Gy), Fe (0.1, 0.3, 2 Gy)</td>
<td>1200</td>
<td>High percentage of B cells, low percentage of T cell and monocytes after 2 Gy Fe low NK cells, decreased basal DNA synthesis, increased mitogenes response after 2 Gy Si</td>
<td>Radiation quality affects the number and distribution of lymphocytes</td>
<td>Gridley et al., 2009</td>
</tr>
<tr>
<td>Mouse bone marrow and heart (10–12 weeks)</td>
<td>1 week, 1 month, 6 months</td>
<td>Si (0.1, 0.25, 0.5 Gy)</td>
<td>Whole-body IR</td>
<td>Increased cleaved PARP</td>
<td>Levels of apoptosis and inflammation higher in irradiated samples compared to sham,</td>
<td>Tungjai et al., 2013</td>
</tr>
<tr>
<td>Human A-bomb survivor lymphocytes</td>
<td>40 years</td>
<td>γ rays and neutrons (0.005–1.5 Gy)</td>
<td>300</td>
<td>Increased activated NFκB</td>
<td>γ-rays and neutron radiation significant effect on inflammatory responses</td>
<td>Hayashi et al., 2003</td>
</tr>
<tr>
<td>CS7BL/6 mice female (9–10 weeks), spleen and lymphocytes</td>
<td>21 days, 56 days</td>
<td>1. LDR$^{**}$ γ rays (0.01 Gy, 0.03 eGy/h); 2. γ rays (2 Gy, 0.9 GY/min); 3. LDR + gamma; 4. protons (1 Gy); 5. LDR + protons</td>
<td>γ rays (1.17 and 1.33), proton 250</td>
<td>21 days lymphocyte populations (3 = 5 &lt; 1); 56 days CD4/CD25 Foxp3 T cells (4 &gt; 1 = 3 &lt; 5); 56 days IL-2 (1 = 5 &gt; 0 Gy); 456 days IL-10 (3 &gt; 0 Gy); 21 days TGF-β (5 &lt; 3); 21 days INK 4 (5 &gt; 3); Note. Numbers indicate radiation regimen described in Radiation Quality column. = is approximate.</td>
<td>Immune responses to acute 2 Gy radiation dependent on radiation quality; time of assessment, pre-exposure to γ rays</td>
<td>Gridley et al., 2013</td>
</tr>
</tbody>
</table>

$^a$ Energy is noted where provided in this article.
$^{**}$ LDR = Low dose rate.

Of critical interest for space radiation risk assessment are studies that suggest that the complex DNA damage produced by heavy ion radiation is associated with persistent oxidative stress, chronic inflammation, and genomic instability at levels not observed following exposure to similar doses of low-LET radiation. These biological responses may underlie the observations of decreased latency and the potential for increased aggression of tumors induced by heavy ion exposure. Also, their potential contribution to the etiology of cardiovascular and CNS diseases, the other major radiation risk areas of concern for spaceflight, make them potentially relevant targets for cross-risk biological countermeasure development.
8. Animal Models for Carcinogenesis

A significant challenge for studying the development of cancer due to space radiation is in observing the effect in relevant experimental models. Many studies have been performed using traditional cell culture in both two dimensional and more recently, three dimensional forms to study the effect of various doses, qualities and dose rates due to space radiation. Data from exposure to ionizing radiation from nuclear disasters or radiotherapy aids in understanding some effects however, it is not possible to recreate an exposure to the space radiation environment in humans on the ground. Therefore, it is critical to use relevant animal models to help further understand the risk of carcinogenesis and develop suitable biological countermeasures to help mitigate the initiation or promotion of carcinogenesis. A recent review by Rivina et al. (2013) discussed the value and relevance of using common laboratory mice to study cancer. They argue that due to its molecular and physiological similarities to man, small size, ease of breeding in captivity and a fully sequenced genome the *Mus musculus* remains one of the best animal model systems for cancer research. Studies on radiation carcinogenesis due to HZE ions have been performed with rat strains and stocks, inbred mouse strains and their F1 hybrids, genetically diverse stock, and genetically engineered mouse models (Barcellos-Hoff et al. 2015, Sridharan et al. 2015). Genetically engineered models have provided insight into the development of lung cancer and colon cancer (Delgado et al. 2014; Moding and Kirsch 2012; Trani et al 2014) while the development of a collaborative cross inbred mouse has provided a model to better represent the heterogeneous human population (Threadgill et al. 2011).

VI. Models of Cancer Risks and Uncertainties

A. Track-structure based Risk Model

This section describes the revisions to the NASA cancer risk model and associated uncertainties. NASA’s previous estimates of radiation risk were based on cancer mortality, background mortality, and models of excess relative risk (ERR) and excess absolute risk (EAR) from the Japanese LSS cohort (Cucinotta et al. 2006). More recently, the BEIR VII report (BEIR 2006) has suggested that cancer mortality risk should be estimated by utilizing incident cancer rates, transferring that risk to the cohort of interest, and scaling the transferred risk by the ratio of the cancer morality and incident rates of the host population. The discussion that follows is taken from Cucinotta et al. (2013).

The risk of radiation exposure induced cancer (REIC) is calculated by folding the instantaneous hazard rate for cancer incidence, $\lambda_t$, with the probability of surviving to age $t$ free of cancer:

$$
\text{REIC}(a_E, H_T) = \hat{\lambda} \int_0^{a_{max}} dt \int_0^{H_T} \frac{\hat{S}_0(t)}{S_0(t)} \exp \left[ \int_a^{a_{max}} d\tau \int_0^{H_T} \frac{\lambda_M(z)}{\hat{\lambda}} dz \right],
$$

where $a$ is the age, $a_E$ is the age at exposure, $t$ is the time, $a_{max}$ is the maximum attained age, $S_0(t)$ is the survival function of the background population, $H_T$ is the organ dose equivalent for each tissue, $T$, and $\lambda_M$ is the cancer mortality rate. The tissue specific instantaneous cancer incidence rate is given by
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\[
\begin{align*}
(2) \quad & l_I(H_T, a_E, a) = [n_T \text{ ERR}(a_E, a) l_{I0}(a) + (1-n_T) \text{ EAR}(a_E, a)] \frac{H_T}{\text{DDREF}}, \\
\end{align*}
\]

where \( \nu_T \) is a parameter used to allow contribution for multiplicative (ERR) and additive risk models (EAR), \( \lambda_{I0} \) is the cancer incidence rate of the background population, and DDREF is the dose-rate and dose reduction factor.

Likewise, the risk of radiation exposure induced death (REID) is determined by folding the instantaneous cancer mortality hazard rate, \( \lambda_M \), with the probability of surviving to age \( t \) free of cancer (Cucinotta et al. 2013)

\[
\begin{align*}
(3) \quad & \text{REID}(a_E, H_T) = \int_{0}^{\infty} dt l_M(a, a_E, H_T) \mathcal{S}_0(t) \exp \left[ \int_{a_E}^{a_{\text{max}}} dz l_M(z, a_E, H_T) \right] \\
\end{align*}
\]

with

\[
\begin{align*}
(4) \quad & l_M(H_T, a_E, a) = [n_T \text{ ERR}(a_E, a) l_{M0}(a) + (1-n_T) l_{I0} \text{ EAR}(a_E, a)] \frac{H_T}{\text{DDREF}}. \\
\end{align*}
\]

In equation (4), \( \lambda_{M0} \) is the cancer mortality hazard rate for the background population.

Generalized ERR and EAR models were fitted to LSS cancer mortality and incidence data with Poisson likelihood methods and include latency (UNSCEAR 2008). The ERR is linear-quadratic in dose response and is given as

\[
\begin{align*}
(5) \quad & \text{ERR}(a, a_E, D) = (aD + bD^2)e^{\gamma D} \exp[k_1 a_E^2 + k_2 \ln(a - a_E) + k_3 \ln(a) + k_4 \ln(a_E)], \\
\end{align*}
\]

where \( a, \beta, \gamma, \kappa_1, \kappa_2, \kappa_3, \kappa_4, \) and \( l_5 \) are cancer specific parameters (UNSCEAR 2008). A similar generalized EAR model is used with a different set of parameters. Although the above ERR and EAR models are used for most tissues, there are some exceptions. The BEIR VII, 2006 models are used for breast and thyroid cancer risks and the Preston et al. (2007) models are used for the prostate, uterus, ovary, oral cavity, non-melanoma skin risks, as well as for other tissues that were not included in the BEIR VII and UNSCEAR models.

1. Biological Effects Related to Track Structure

The tissue dose equivalents appearing in equations (1) - (4) may be obtained by folding the physical tissue fluence with a radiation quality factor, \( Q \), representing the increased effectiveness of HZE particles, as compared to \( \gamma \)-rays, for the same biological endpoint. These factors are based on subjective assessment of maximum RBE values for relevant endpoints from radiobiology experiments. In the past, NASA has utilized the LET-dependent quality factor, \( Q(L) \), recommended by the ICRP (ICRP 1991).
Animal studies generally demonstrate that HZE nuclei have higher carcinogenic effectiveness than low-LET radiation. However, the number of studies of animal carcinogenesis made with HZE nuclei is extremely limited. These studies used one or only a few ion types, providing little information on the possible radiation quality dependence of RBE. Both the ICRP and NCRP have noted limitations in radiobiology data to assess radiation quality factors (ICRP 2003; NCRP 2006).

Despite the limitations in available data, an improved description of the quality factor for the particles found in space was provided by Cucinotta et al. (2013) by considering track-structure effects. Track structure descriptions are used in theoretical models of biological response to understand and extrapolate limited radiobiology data to other radiation qualities and doses. More recently, Borak et al. (2014), have presented strong arguments for revised quality factors that are new functions of LET for solid cancers and leukemia. This revisits the method NASA used previously where quality factor was dependent on LET under the ICRP (ICRP 1991) recommendations. A significant difference in the work by Borak et al. (2014) and the ICRP (ICRP 1991) is the ability to distinguish between solid cancers and leukemia.

Observations by Goodhead et al. (1980) and earlier arguments from Katz (1970) predict that biological effects would be highly influenced by δ-ray effects rather than by LET alone. The number of δ-rays created by an ion with charge \( Z \) traversing a biological target is proportional to the track structure parameter \( X_{TR} = (Z^*/\beta)^2 \), where \( Z^* \) is the effective charge number that adjusts \( Z \) by atomic screening effects important at low kinetic energies and high \( Z \), and \( \beta \) is the ion velocity relative to the speed of light. Figure 13 shows such a description comparing the frequency of energy deposition above 300 eV in a volume the size of the nucleosome. The comparisons illustrate that the parameter \( (Z^*/\beta)^2 \) provides an improved descriptor of energy deposition in small volumes compared to LET. Deviations from a unique \( (Z^*/\beta)^2 \) dependence occur at low energy where the curves branch for distinct charge numbers.

Figure 13. Number of nucleosomes per cell receiving 300 eV or more as a function of LET (left panel) or \( (Z^*/\beta)^2 \) (right panel). Calculations are shown for H, He, Si, and Fe nuclei using methods of Cucinotta et al. (2000).
Figure 14. Comparison of quality factor based on track structure to quality factor based on LET propagated through aluminum and water for A) solid cancers and B) leukemia (Borak et al. 2014)

For identical LET values, the ion with the lowest charge is predicted to be more effective at energies above a few MeV/u (Cucinotta and Kim 2013). The recent work by Borak et al. (2014) indicates that quality factor can be determined as a function of LET without taking into consideration charge or energy of the incident radiation. A comparison of NASA’s quality factor and the quality factor calculated using the method of Borak et al. (2014) propagated through aluminum and water for solid cancers and leukemia demonstrate good agreement (Figure 14).

Research at the NSRL is making new estimates of radiation quality effects for a variety of endpoints with the focus on approaches to mechanistic understanding of biological effectiveness. However, very few comprehensive studies have been completed at this time. Here we note that, in the past, very detailed studies of radiation quality were made for DNA breaks, as well as for cell inactivation and mutation for a large number of ion types. Such extensive studies would be difficult to repeat today because of the higher costs of many current experimental approaches, and certainly would take many years to complete. These older studies are useful to consider in terms of track-structure models.

2. Risk Cross Section and NASA Quality Factor

The biological action cross section, $\sigma$, is the probability for a given endpoint - such as mutation or induction of tumors - per unit fluence and is useful when exponential and linear dose response curves are suitable. The following parametric model of the risk cross section is recommended for low doses (Wilson et al. 1993; Cucinotta and Wilson 1995; Cucinotta et al. 2013)

$$S(Z,E) = S_0P(Z,E) + \frac{a \cdot LET}{6.24} [1 - P(Z,E)],$$

where
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(7) \[ P(Z, E) = [1 - \exp(-X_{TR} / \kappa)]^m. \]

In equation (7), \( \beta \) is the ratio of the particle speed to the speed of light, \( X_{TR} \) is the track structure parameter. The parameters \( \Sigma_0, \kappa, \) and \( m \) are estimated from radiobiology experiments. Note that \( \Sigma_0 \) is the maximum value of the risk cross section and is related to \( \text{RBE}_{\text{max}} \) (Cucinotta et al. 2013). The recommended NASA radiation quality factor is defined by using the risk cross section in equation (6), and is given by (Cucinotta et al. 2013, Cucinotta 2015)

(8) \[ Q_{\text{NASA}} = \frac{6.24 S_0 a_E}{L} P(Z, E) \left(1 - e^{-E/E_{\text{TH}}}ight). \]

where \( a_\gamma \) is the low LET slope parameter, which is estimated from human epidemiology data from \( \gamma \) radiation. The extra term in parentheses on the far right of equation (8) modifies the quality factor (or action cross section) at low energies to account for thindown. The value of \( E_{\text{TH}} \) has been set at 0.2 based on experimental data for H and He. Thindown results when the spatial distribution of \( \delta \)-rays from an ion becomes limited by kinematics to a size smaller than the biological target.

The revised radiation quality factor may be used to compute tissue dose equivalents for space radiation environments, allowing the cancer incidence and mortality rates to be computed in terms of particle-specific charge and kinetic energy, instead of LET alone. (Cucinotta et al. 2013). Borak et al. (2014), however, have demonstrated good agreement between NASA’s quality factor and an LET dependent quality factor for solid cancers and leukemia suggesting this may be a revised method to consider for determining quality factor in general.

3. Uncertainties in the Projection Model

There are various sources of uncertainty in assessing the risk associated with radiation exposure. In the current risk model, these are classified as uncertainties in the low-LET risk model, risk cross section (quality factor), and physics (fluence). The following section discusses various probability distribution functions that are used for propagating uncertainty in the estimation of REIC and REID.

Uncertainties in Low-LET Epidemiology Data

Low-LET risk uncertainties are incorporated into the hazard rate by sampling quantiles (random variables) from probability distribution functions (PDF) that represent dosimetric, statistical, bias, risk transfer, and risk coefficient uncertainties (Cucinotta et al. 2013):

(9) \[ l_L(E, a_E, a) = \frac{l_0(a_E, a) x_D x_a x_T x_B}{\text{DDREF}}. \]

where \( \lambda_0 \) is the baseline hazard rate per Sv, and \( x_a \) are the quantiles whose values are sampled from the associated PDF. Note that the DDREF applies only to the solid cancer risk and not the leukemia.
risk under the stated assumptions. Cucinotta et al. (2013) defines the following subjective PDFs, \( P_a(x_a) \), for each factor that contributes to the acute low LET-risk projection:

1. \( P_D \) represents the random and systematic errors in the estimation of the doses received by atomic-bomb blast survivors. It is represented by a log-normal distribution with a geometric mean of 0.9 and a geometric standard deviation of 1.3;
2. \( P_s \) represents the distribution in uncertainty in the risk coefficient that is associated with the increase in risk with increasing radiation exposure. It is assumed as a normally distributed PDF with a mean of one and a tissue specific standard deviation;
3. \( P_B \) represents any bias resulting for over- or under-reporting cancer incidents. It is assumed as a normal distribution with a mean of 1.0 and a standard deviation of 0.05;
4. \( P_T \) represents the uncertainty in the transfer of cancer risk following radiation exposure from the Japanese population to the US population. Both additive and relative risk models were considered by NCRP 126 in assessing the uncertainties in such transfer. \( P_T \) is a uniform distribution about the preferred weight (Cucinotta et al. 2013)
5. \( P_{Dr} \) represents the uncertainty in the knowledge of the extrapolation of risks to low dose and dose-rates, embodied in the dose and dose-rate reduction factor (DDREF). Based on a Baysian analysis, Cucinotta et al. (2013) have recommended a t-distribution with a central estimate of 1.5 for the \( P_{Dr} \).

**Uncertainties due to Dose-Rate and Protraction Effects for Ions**

It is not feasible to perform long duration (months) exposure to space relevant radiation doses and dose-rates. For low dose-rate and protracted proton and HZE radiation exposure of more than a few months, new biological factors may influence risk assessments, including redistribution in the cell cycle, repopulation, or promotional effects, especially when particle fluences are large enough to lead to multiple hits of target cells or surrounding cells and tissue environments. With the increase in proton and carbon ion therapy treatment centers, there is a limited amount of human data on high-dose, high dose-rate protons and carbon ions, however, no human data exists on other HZE ions and there is very little experimental data at space relevant doses and dose-rates for these particle types. Confidence in using radio-epidemiological data for acute (A-bomb survivors) or fractionated (patient) data is decreased when applied to protracted exposure. Experimental data for protracted proton or heavy ion irradiation in experimental models of carcinogenesis is almost non-existent. Burns et al. (1994) found split doses of argon ions separated by a few hours up to one-day increased the risk of skin cancer in rats. Alpen et al. (1994) found using seven two-week fractions of 0.07 Gy of iron an increase in risk of 50% compared to a single acute dose of 0.4 Gy for Harderian gland tumors in mice. A study of chromosomal aberrations in human lymphocytes (George et al. 2001) for acute and low dose-rates (0.08 Gy/hr) with 250 MeV protons, showed less sparing than found for gamma-rays. The Skyhook study of Ainsworth et al. (1986) considered life-shortening in mice comparing single acute with weekly fractions of several ions; however, the results were unclear with regards to any increase or decrease in risk.

For gamma-rays and neutrons, a good number of studies for cancer induction or life-shortening in mice exists, showing sparing effects for gamma-rays, and that neutron effects may be increased due to protraction under certain conditions in some tissues (Ullrich 1984; NCRP 1990). Contrary to these reports, Daniels and Schubauer-Berigan (2011) performed a meta-analysis across multiple studies involving human workers exposed to protracted low-dose gamma radiation with doses
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ranging between 5.6-810mGy and found it to be significantly associated with the incidence of leukemia. Other studies did not find an increase in tumor incidence low doses and dose-rates of 0.05 mGy/d (20 mGy total dose) or 1.1 mGy/d (400 mGy total dose), but noted an increased incidence of several tumor types affecting both sexes at 21 mGy/d (8000 mGy total dose) (Tanaka et al. 2007). Important questions related to the differences in life-span, cell turn-over rates, or mechanisms of initiation or promotion in humans and mice, make estimates of the effects of protraction on risk difficult. If protraction effects do increase the risk from high LET radiation, then such effects would be more important for a Mars mission than the shorter lunar missions. In space, each cell will be traversed about every two to three days by a proton or delta-ray produced by ions in adjacent cells, with a decreasing frequency from weeks to months as the charge of the HZE nuclei increases (Cucinotta et al. 1998). Studies of mixed-fields of protons and HZE ions are needed to understand uncertainties in dose-rate and protraction effects from space radiation. Efforts are underway to incorporate a mixed field GCR simulated environment at the NSRL in order to address these questions (Norbury et al. 2016). Uncertainties related to radiation quality, dose-rate, and protraction could lead to correlations that will be difficult to describe when based on limited experimental data. Methods to treat correlation effects will be needed when additional data on protraction effects become available.

Radiation Quality and Latency or Temporal Patterns of Risk

There is an additional radiation quality uncertainty introduced assuming the time dependence for low and high LET radiation is identical. Data on tumors or genomic instability in mice with neutrons (Ullrich et al. 1984, 1998; NCRP 1990) and the studies of rat or mammary carcinogenesis with HZE nuclei (Burns et al. 1994; Dicello et al. 2004; Barcellos-Hoff et al. 2015), suggest that the latency time is appreciably reduced for high LET compared to low LET radiation. There is sparse data available to estimate the impact of these differences on uncertainties. A radiation quality dependent latency is more important in the additive transfer model than the multiplicative transfer model, especially at younger ages of exposure. We ignore these uncertainties; however, we replace the 10 year minima latency assumption made for low LET by the step-in latency model (Pierce et al. 1996) used for the leukemia risk. The effects of these assumptions will need to be addressed when data and knowledge on underlying mechanisms become available.

Uncertainties in Quality Factor (Risk Cross Section)

Uncertainties in the quality factor are incorporated by Monte Carlo samples of quantiles from PDF associated with m, κ, and Σ0 / αγ from equations (7) and (8). There is an additional PDF that multiplies QNASA and is used as a correction to the quality factor for high energy protons. The PDF for each of the uncertainties are described below, and parameter estimates have been provided elsewhere (Cucinotta et al. 2013).

1. Slope parameter m: A discrete distribution for the slope parameter is used, with slope m = [1, 2.5, 3, 3.5, 4] and corresponding weights [0.15, 0.20, 0.40, 0.20, 0.05].
2. Track structure parameter κ: A normal distribution with mean of 1 and standard deviation of 1/3 is used.
3. PDF for $Q_{\text{max}} = \Sigma_0 / \alpha$: A log normal distribution with geometric mean (GM) of 1 and geometric standard deviation (GSD) of 1.4 is used for solid cancer; for leukemia, a normal distribution with mean of 1 and standard deviation of 1.6 is used.

3. High energy proton correction: For protons with energies greater than 150 MeV, a normal distribution with mean of 1 and standard deviation of 0.15 modifies $Q_{\text{NASA}}$.

Uncertainties in Fluence (Physics)

Uncertainty in determining the radiation environment within radiosensitive tissues includes uncertainties in estimating the ambient radiation field in space (boundary condition), shielding geometry and human phantom model, nuclear and atomic physics, and particle transport. In the current NASA cancer risk projection model, these combined uncertainties have been subjectively represented with a normal distribution with mean 1.05 and standard deviation of 1/3 is used light ions ($Z \leq 4$). For heavy ions ($Z > 4$), a normal distribution with mean 1 and standard deviation of 1/4 is used. Ongoing verification and validation efforts are focused on more rigorously quantifying combined physics uncertainties used in risk projection models.

B. Systems Biology Modeling

Systems biology is a rapidly growing field due to the advances in the various -omics techniques over the last decade. Along with it, computational biology has emerged as a useful path to reconcile the various -omics relationships and draw conclusions from the vast data sets generated. This area has become a focus for radiation carcinogenesis and several groups have begun implementing systems biology to understand different biological organizations along different time-scales to uncover relationships of the key processes involved (Barcellos-Hoff et al. 2014). Gene and molecular networks focus on mapping mechanistic and structural properties of the system (Conesa and Mortazavi 2014). Using these techniques, Del Prete et al. (2011) revealed two molecular pathways involved in cancer development due to inflammation. In a study involving low dose radiation (10cGy X-rays), researchers applied a comprehensive -omics analysis including proteomic, phosphoproteomic and metabolomic platforms to determine the temporal impact on human tissue using in vitro 3D full thickness human skin models (Tilton et al. 2015). Their research uncovered signaling mechanisms and common molecular and pathway responses to low dose ionizing radiation. Comprehensive -omics characterization of cancers that arise due to heavy ion exposure will support systems biology risk modeling efforts, extrapolation of results to human cancers, and also help to drive biomarker identification for disease monitoring and future countermeasure development and testing.

Computational and theoretical models are currently being developed to describe mechanistic or structural behavior of systems and networks as well. Several researchers have employed agent based modeling (ABM) to better understand stasis of specific diseases. Mukhopadhyay et al. (2010) used ABM to simulate cancer processes while von Neubeck et al. (2013) developed an ABM to determine the effects of heavy ion radiation on skin homeostasis. Others have developed mathematical models to extrapolate risks from high-dose to low-dose based on radiation induced
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foci (Neumaier et al. 2012), analyze the NHEJ repair pathway (Li et al. 2014) and to study radiation induced base excision repair (Rahmanian et al. 2014).

Multi-scale systems approaches that relate molecular damage and modifications of signal transduction pathways to cellular and tissue effects are important to achieve the required levels of accuracy in risk estimation required for long duration space flight. An integrated systems biology based risk assessment model using multiscale modeling approaches will provide the accuracy to predict individual based risk in support of long duration space travel. The integration of high throughput genomics and proteomics data sets for critical cancer development processes identified through mechanistic studies into a systems biology understanding of biomedical and fundamental biological processes is a critical aspect in achieving this overall goal. It is anticipated that this risk assessment model or computational framework will ultimately result from the assembly of distinct modular components, or building blocks, that connect across multiple levels of biological organization. The ultimate goal is to develop these models to provide information on the individual level. This will be important for future exploration missions since the size of the crew will be limited and individual response will be critical to their health and the mission. Personalized medicine is on the forefront of research. Schmidt and Goodwin (2013) describe an -omics based systems analysis to facilitate development of personalized countermeasures for astronauts. It is likely that this area will be advanced enough to be of benefit to NASA and the crew by the time a deep exploration mission occurs involving humans.

VII. Risk for Exploration Mission Operational Scenarios

The accuracy of GCR environmental models, transport codes, and nuclear interaction cross sections described above allow NASA to make predictions of space environments and organ exposures to be encountered for missions to the moon or Mars. However, there are major questions that arise due to the lack of knowledge on biological effects. For cancer risk projections, propagating individual uncertainties in factors that enter risk model calculations is used to place reasonable bounds on cancer risks to be encountered.

A. Risk Estimates for Space Exploration Missions

In implementing a numerical procedure for propagating the uncertainties discussed in section 3, equation (9) is inserted into the expression for REID (or REIC), and a large number of Monte Carlo trials (~10^5) are performed. In each trial, deviates are sampled from the uncertainty distributions and applied to the radiation quality factor or hazard rate. Results for the REID estimates are binned and the median values and confidence intervals found. Table 9 gives estimates for the number of safe days in space for various age groups and populations. The number of safe days is defined as the maximum number of days in space to be below 3% REID at a 95% confidence level (CL).
Table 9. Safe days in space, which is defined as the maximum number of days to be below 3% REID at a 95% CL. Calculations are for average solar minimum with 20 g/cm² aluminum shielding. Values in parentheses are for the case of the deep solar minimum of 2009 (Cucinotta et al. 2013).

<table>
<thead>
<tr>
<th>$a_E$, y</th>
<th>Females</th>
<th>Males</th>
</tr>
</thead>
<tbody>
<tr>
<td>35</td>
<td>106 (95)</td>
<td>187 (180)</td>
</tr>
<tr>
<td>45</td>
<td>139 (125)</td>
<td>227 (212)</td>
</tr>
<tr>
<td>55</td>
<td>161 (159)</td>
<td>277 (246)</td>
</tr>
</tbody>
</table>

B. Biological and Physical Countermeasures

Identifying effective countermeasures to reduce the biological damage produced by radiation remains a long-term goal of space research. As noted by Durante and Cucinotta (2008), such countermeasures may not be needed for a lunar base, but probably for the Mars mission, and definitely for exploring Jupiter or Saturn’s moon Titan or the nearby satellites. In all basic radioprotection textbooks, it is stated that there are three means to reduce exposure to ionizing radiation: increasing the distance from the radiation source, reducing the exposure time, and by shielding. Distance plays no role in space, as space radiation is omnipresent. Time in space is likely to be increased rather than decreased given plans for exploration and colonization. Shielding remains a plausible countermeasure, albeit a prohibitively costly one in light of current launch mass capabilities and may actually increase exposures beyond certain thicknesses due to neutron build-up and electromagnetic cascades (Slaba et al. 2013a).

Other strategies can be effective in reducing exposure, or the effects of the irradiation, in space. These strategies include the choice of an appropriate time of flight, administration of drugs or dietary supplements to reduce the radiation effects, and crew selection.

Biomarkers will aid in determining when and if a biological countermeasure is required based on individual exposures and sensitivities. This is a rapidly expanding area of research. Biomarkers have currently been developed in support of mass triage for potential nuclear disasters or radiation terrorism. The primary method to assess radiation exposure previously was by analyzing chromosome aberrations: dicentric for short-lived damage and translocations for long-lived. This is a time consuming method and is being replaced by biomarkers that are based on genetic damage identified from peripheral blood mononuclear cells (Dressman et al. 2007; Paul and Amundson 2008; Lucas et al. 2014), breath (Phillips et al. 2015), protein (Bazan et al. 2014) and more recently metabolites (Laiakis et al. 2015). These biomarker methods offer rapid assessment, providing information quickly that will facilitate treatment options.

1. Radioprotectors and Mitigators

The search for efficient radioprotectors and mitigators is a major goal of research in radiation protection and therapy. Both radiation injury and oxygen poisoning occur through the formation of reactive oxygen species (ROS); therefore, antioxidants have been heavily studied as a way to mitigate the damage resulting from ROS (Weiss and Landauer 2003).
The ideal biological countermeasure will provide cross-risk mitigation or protection through antioxidants that provide free radical scavenging, common pathways (eg., inflammatory) or other biological pathway that intersects multiple risks. Several biological countermeasures have been developed and studied in support of acute radiation exposure. WR-2721 (amifostine or Ethyol®, MedImmune Oncology, Inc.) is a phosphorothioate that confers radioprotection and is approved by the FDA. It has been investigated extensively as an antioxidant that scavenges free radicals (Xiao and Whitnall 2009; Langell et al. 2008). Although it has demonstrated positive results as a radioprotector, it has shown toxicity at the levels required to induce a radioprotective effect with undesirable side effects (Bogo et al. 1985; Whitnall 2012). An analog to WR-2721 is an aminothiol, PrC-210 (ProCertus BioPharm, Inc.), offering the same benefits of WR-2721 without the adverse side effects and toxicity (Peebles et al. 2012; Soref et al. 2011; Whitnall 2012). Natural occurring antioxidants are less effective than phosphorothioate agents in protection against high-dose acute radiation burden. However, nutritional antioxidants have a low toxicity, can be used for prolonged time, and they seem to play a key role in the prevention of cancer (Halliwell 2000; Bingham and Riboli 2004). A diet rich in fruit and vegetables significantly reduced the risk of cancer in the A-bomb survivor cohort (Sauvaget et al. 2004). Retinoids and vitamins (A, C, and E) are probably the most well-known and studied natural radioprotectors, but hormones (e.g. melatonin), glutathione, superoxide dismutase, phytochemicals from plant extracts (including green tea and cruciferous vegetables), and metals (especially selenium, zinc, and copper salts) are also under study as dietary supplements for individuals overexposed to radiation (Weiss and Landauer 2000), including astronauts. In addition, there is evidence of a reduced antioxidant capacity during spaceflight, as shown by reduced superoxide dismutase (SOD) and total antioxidant activity in some astronauts returning from missions on the International Space Station (Smith et al. 2005).

Understanding the effectiveness of antioxidants in space is complicated by the presence of HZE particles. In principle, antioxidants should provide reduced or no protection against the initial damage from densely ionizing radiation, because the direct effect is more important than free radical-mediated indirect radiation damage at high LET. However, there is an expectation that some benefits should occur for persistent oxidative damage related to inflammation and immune responses. Recent experiments suggest that an efficient radioprotection by dietary supplements can be achieved even in case of exposure to high LET radiation. Ascorbate reduces the frequency of mutations in human-hamster hybrid cells exposed to high LET C-ions (Waldren et al. 2004). Vitamin A strongly reduces the induction of fibroma in rats exposed to swift $^{56}$Fe ions (Burns et al. 2007). Dietary supplementation with Bowan-Birk protease inhibitors (BBI) (Guan et al. 2006; Kennedy 2014), L-selenomethionine or a combination of selected antioxidant agents (Kennedy et al. 2007) could partially or completely prevent the decrease in the total antioxidant status in the plasma of mice exposed to proton or HZE particle radiation, and neoplastic transformation of human thyroid cells in vitro. Eskiocak et al. (2010) reported that CDDO-Me (Methyl-2-cyano-3,12 dioxyolean-1,9 diene-28-oate), an antioxidative, anti-inflammatory modulator, protected human colon epithelial cells against radiation induced neoplastic transformation after exposure to 2 Gy protons followed by 0.5 Gy $^{56}$Fe ions or the ions delivered individually. The latest in vivo studies using space relevant radiation dose levels of 0.5 Gy, 1-GeV/n $^{56}$Fe ions, showed treatment with antioxidant combination or BBI decreased the levels of malignant lymphoma (Kennedy et al. 2008) and decreased the incidence of rare tumors (such as Harderian gland) (Kennedy et al. 2011). However, because the mechanisms of biological effects may be different for low dose-rate
compared to acute irradiation, new studies for protracted exposures will be needed to understand the potential benefits of biological countermeasures.

Even if antioxidants can act as radioprotectors or mitigators, this does not necessarily translate as an advantage for cancer risk. If antioxidants protect cells by rescuing them from apoptosis, then this may allow the survival of damaged cells, which eventually can initiate tumor progression. Concern about this possibility is sustained by a recent meta-study of the effects antioxidant supplements in the diet of normal subjects (Bjelakovic et al. 2007). The authors did not find statistically significant evidence that antioxidant supplements have beneficial effects on mortality. On the contrary, they concluded that β-carotene, vitamin A, and vitamin E seem to increase the risk of death. Concerns not only include rescuing cells that still sustain DNA mutations, but also the altered methylation patterns that can result in genomic instability (Kovalchuk et al. 2004). An approach to target damaged cells for apoptosis may be advantageous for chronic exposures to galactic cosmic radiation (GCR). Radioprotectors and mitigators tested for acute exposures at high doses should be used with care – rescuing cells may make the problem worse in the long term.

Non-steroidal anti-inflammatory drugs (NSAIDS) have been shown to alter tumors and the tumor microenvironment by blocking cell proliferation and promoting apoptosis. (Rayburn et al. 2009; Balkwill et al. 2012). Epidemiological evidence has indicated the incidence of breast, colon and lung cancers is inversely related to the use of aspirin or NSAIDS (Rayburn et al. 2009). NSAIDS are commonly used to reduce cardiovascular events potentially offering cross-risk mitigation, however, dose and duration are important factors to consider when employing the use of these drugs. Other anti-inflammatory targets include chemokine-receptor antagonists, cytokine-receptor antagonists, and COX inhibitors with several clinical trials currently underway to investigate the therapeutic effect of agents on these targets (Mantovani et al. 2008).

2. Shielding

For terrestrial radiation workers, additional protection against radiation exposure is usually provided through increased shielding. Unfortunately, shielding in space is problematic, especially when galactic cosmic rays (GCR) are considered. High-energy radiation is very penetrating: a thin or moderate shielding is generally efficient in reducing the exposure, but as the thickness increases, shield effectiveness drops. This is the result of the production of a large number of secondary particles, including neutrons, caused by nuclear interactions of the GCR with the shield. These particles have generally lower energy, but can have higher quality factors than incident cosmic primary particle. Radiation shielding effectiveness depends on the atomic constituents of the material used. Shielding effectiveness per unit mass is the highest for hydrogen, and decreases with increasing atomic number (Wilson et al. 1995; Wernerth et al. 2013). Liquid hydrogen would display the maximum performance as shield material but is not practical, since it is a low temperature liquid. Instead, polyethylene (CH₂) could be a good compromise. Secondary neutron production increases with the mass number of the atomic constituents of the material and can grow to be large values for materials such as aluminum or the regolith on the Martian surface, or for heavier materials such as lead. For SPE shielding, the situation is much better and the majority of events on record can be reduced to reasonable dose levels (< 100 mSv) with localized shielding of polyethylene inside a lightly shielded vehicle or habitat (Figure 15).
Risk of Radiation Carcinogenesis

**Figure 15.** Effective doses versus depth in several materials for GCR at solar minimum and the 1972 SPE (Slaba et al. 2013).

VIII. Gaps

There are 15 gaps associated with the risk of radiation carcinogenesis and several gaps related to this risk including acute radiation exposure, degenerative tissue effects and impact on the central nervous system.

**Risk:** Given that crewmembers are exposed to radiation from the space environment, there is the possibility for increased cancer morbidity or mortality.

**Gaps:**

Cancer - 1: How can experimental models of tumor development for the major tissues (lung, colon, stomach, breast, liver, and leukemias) be developed to represent the major processes in radiation carcinogenesis and extrapolated to human risk and clinical outcome projections?

Cancer - 2: How can experimental models of tumor development for the other tissues (bladder, ovary, brain, esophagus, skin, etc) be developed to represent the major processes in radiation carcinogenesis and extrapolated to human risk and clinical outcome projections?

Cancer - 3: How can experimental models of carcinogenesis be applied to reduce the uncertainties in radiation quality effects from SPE’s and GCR, including effects on tumor spectrum, burden, latency and progression (e.g., tumor aggression and metastatic potential)?
Risk of Radiation Carcinogenesis

Cancer - 4: How can models of cancer risk be applied to reduce the uncertainties in dose-rate dependence of risks from SPE's and GCR?

Cancer - 5: How can models of cancer risk be applied to reduce the uncertainties in individual radiation sensitivity including genetic and epigenetic factors from SPE and GCR?

Cancer - 6: How can models of cancer risk be applied to reduce the uncertainties in the age and sex dependence of cancer risks from SPE's and GCR?

Cancer - 7: How can systems biology approaches be used to integrate research on the molecular, cellular, and tissue mechanisms of radiation damage to improve the prediction of the risk of cancer and to evaluate the effectiveness of countermeasures? How can epidemiology data and scaling factors support this approach?

Cancer - 8: What are the most effective biomedical or dietary countermeasures to mitigate cancer risks from exposure to SPE and GCR? What side effects should be tolerated versus mission risks?

Cancer - 9: Are there significant effects from other spaceflight factors (microgravity, stress, altered circadian rhythms, changes in immune responses, depressed nutrition, bone loss, etc.) that modify the carcinogenic risk from space radiation?

Cancer - 10: Are space validation experiments needed for verifying knowledge of carcinogenic or other risks prior to long-term deep space missions, and if so what experiments should be undertaken?

Cancer - 11: What are the most effective shielding approaches to mitigate cancer risks?

Cancer - 12: What quantitative models, numerical methods, and experimental data are needed to accurately describe the primary space radiation environment and transport through spacecraft materials and tissue to evaluate dose composition in critical organs for mission relevant radiation environments (ISS, Free-space, Lunar, or Mars)?

Cancer - 13: What are the most effective approaches to integrate radiation shielding analysis codes with collaborative engineering design environments used by spacecraft and planetary habitat design efforts?

Cancer - 14: What biodosimetry methods are required for exploration missions and how can biomarker approaches be used for outcome prediction and surveillance?

Cancer - 15: Given that the majority of astronauts are never smokers, are there research approaches that can elucidate the potential confounding effects of tobacco use inherent in population-based epidemiology data on space radiation cancer risk estimates?
The SRPE overlaps with several of the gaps within other HRP Elements as outlined in the HRP Integrated Research Plan (IRP). SRPE works with the other HRP Elements to integrate gaps as necessary in accordance with the IRP.

IX. Conclusion

The evidence for cancer risks from humans exposed to low LET radiation is extensive for doses above 100 mSv. There are important uncertainties for low LET radiation at lower doses (<100 mSv), for low dose-rates, and in transferring risks between populations with different genetic, dietary, etc, attributes. Human epidemiology can be applied to space exposures; however, there are additional uncertainties related to the quality of radiation in space that is known to produce both qualitative and quantitative differences compared to low LET radiation in experimental models. The doses to be expected on space missions, and the nuclear type and energies are well understood. NASA has existing models that quite accurately determine radiation physics parameters in space. Reducing the uncertainties in risk assessment is required before a mission to Mars can be undertaken and has led to a number of investigations guided by molecular and genetic research on carcinogenesis and degenerative diseases. The large uncertainties in risk projection models will only be reduced by improving basic understanding of the underlying biological processes and their disruption by space radiation. There are unique aspects involved in this approach due to the specific challenges to biological systems presented by space radiation, especially HZE ions. It is unlikely that the radiation risk problem for space exploration will be solved by a simple countermeasure, such as shielding or radioprotective drugs. The risk will be understood and controlled only with more basic research in the field of cancer induction by charged particles (Cucinotta and Durante 2006).

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XII. List of Acronyms

2D Two-Dimensional
3D Three-Dimensional
AB Atomic Bomb
ABM Agent Based Modeling
ALARA As Low as Reasonably Achievable
AT Ataxia-Telangiectasia
ATM Ataxia-Telangiectasia-Mutated
BEIR Biological Effects of Ionizing Radiation
BRCA Breast Cancer Gene
CI Confidence Intervals
CML Chronic Myeloid Leukemia
DDREF Dose- and Dose-Rate Effectiveness Factor
DNA DeoxyriboNucleic Acid
<table>
<thead>
<tr>
<th>Acronym</th>
<th>Description</th>
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<tbody>
<tr>
<td>DNA-PK</td>
<td>DNA-dependent Protein Kinase</td>
</tr>
<tr>
<td>DSB</td>
<td>Double Strand Break</td>
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<tr>
<td>EAR</td>
<td>Excess Absolute Risk</td>
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<tr>
<td>ERR</td>
<td>Excess Relative Risk</td>
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<tr>
<td>GCR</td>
<td>Galactic Cosmic Rays</td>
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<tr>
<td>GSD</td>
<td>Geometric Standard Deviation</td>
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<td>Gy</td>
<td>Gray</td>
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<tr>
<td>H2AXP</td>
<td>Histone H2AX Phosphorylated</td>
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<tr>
<td>HF</td>
<td>Human Fibroblasts</td>
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<tr>
<td>HPBL</td>
<td>Human Peripheral Blood Lymphocytes</td>
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<tr>
<td>HZE</td>
<td>High Charge and Energy</td>
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<tr>
<td>HZETRN</td>
<td>High-Charge-and Energy TRaNsport</td>
</tr>
<tr>
<td>ICRP</td>
<td>International Commission on Radiological Protection</td>
</tr>
<tr>
<td>IOM</td>
<td>Institute of Medicine</td>
</tr>
<tr>
<td>ISS</td>
<td>International Space Station</td>
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<tr>
<td>keV/μm</td>
<td>kilo-electron Volt per micrometer</td>
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<tr>
<td>LEO</td>
<td>Low Earth Orbit</td>
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<tr>
<td>LET</td>
<td>Linear Energy Transfer</td>
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<tr>
<td>LLE</td>
<td>Loss of Life-Expectancy</td>
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<tr>
<td>LLE-REID</td>
<td>Loss of Life-Expectancy amongst Exposure Induced-Death</td>
</tr>
<tr>
<td>LSS</td>
<td>Life-Span Study</td>
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<tr>
<td>MeV</td>
<td>Megaelectron Volt</td>
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<tr>
<td>mGy</td>
<td>milliGray</td>
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<tr>
<td>mSv</td>
<td>milliSievert</td>
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<tr>
<td>NAS</td>
<td>National Academy of Sciences</td>
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<tr>
<td>NCRP</td>
<td>National Council on Radiation Protection and Measurements</td>
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<td>NSRL</td>
<td>NASA Space Radiation Laboratory</td>
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<tr>
<td>PDF</td>
<td>Probability Distribution Function</td>
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<tr>
<td>PEL</td>
<td>Permissible Exposure Limit</td>
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<tr>
<td>Q</td>
<td>radiation Quality factor</td>
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<tr>
<td>QMSFRG</td>
<td>Quantum Multiple Scattering FRaGmentation</td>
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<tr>
<td>RBE</td>
<td>Relative Biological Effectiveness</td>
</tr>
<tr>
<td>REID</td>
<td>Risk of Exposure Induced Death</td>
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<tr>
<td>REM</td>
<td>Röntgen Equivalent Man</td>
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<tr>
<td>SEER</td>
<td>Surveillance, Epidemiology and End Results Program</td>
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<tr>
<td>siRNA</td>
<td>silencing RNA</td>
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<tr>
<td>SPE</td>
<td>Solar Particle Event</td>
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<td>SSB</td>
<td>single strand break</td>
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<td>STS</td>
<td>Space Transportation System</td>
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<tr>
<td>Sv</td>
<td>Sievert</td>
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<tr>
<td>TEPC</td>
<td>Tissue Equivalent Proportional Counter</td>
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<tr>
<td>TLD</td>
<td>ThermoLuminescent Dosimeters</td>
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<tr>
<td>UNSCEAR</td>
<td>United National Scientific Committee on the Effects of Atomic Radiation</td>
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