Described herein are particular embodiments relating to a microfluidic device that may be utilized for cell sensing, counting, and/or sorting. Particular aspects relate to a microfabricated device that is capable of differentiating single cell types from dense cell populations. One particular embodiment relates a device and methods of using the same for sensing, counting, and/or sorting leukocytes from whole, undiluted blood samples.

22 Claims, 10 Drawing Sheets
References Cited

OTHER PUBLICATIONS


U.S. Appl. No. 60/922,296, filed Apr. 6, 2007, Tai et al.


References Cited

OTHER PUBLICATIONS


* cited by examiner
FIG. 12A
MICROFLUIDIC DEVICE

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of priority under 35 U.S.C. §120 as a continuation of U.S. patent application Ser. No. 14/685,480, filed Apr. 13, 2015, and now U.S. Pat. No. 9,234,884, which is a continuation of U.S. patent application Ser. No. 14/296,199, filed Jun. 4, 2014, and issued as U.S. Pat. No. 9,029,158 on May 12, 2015, which is a divisional of U.S. patent application Ser. No. 12/062,808, filed Apr. 4, 2008, now abandoned, which claims priority under 35 U.S.C. §119(e) to U.S. provisional patent application No. 60/922,296, filed Apr. 6, 2007, now expired, the contents of each of which are herein incorporated by reference in their entirety.

GOVERNMENT RIGHTS

This invention was made with government support under grant number NCC 9-58-317 awarded by National Space Biomedical Research Institute through NASA. The government has certain rights in this invention.

FIELD OF THE INVENTION

The present disclosure relates to fabricated microfluidic devices that can be utilized as cell sensors and/or actuators. In certain embodiments, the microfluidic device may be used for labeling, sensing, differentiating, and/or sorting targets, particularly cell populations.

BACKGROUND OF THE INVENTION

Standard cell sensors or actuators are generally based on flow cytometry and employ one or a combination of electrical impedance sensing, light scattering measurement, and chemical or immunostaining followed by optical sensing.


Alternative optical methods are based on light scattering and fluorescence staining of organelles, granules, and nuclei. Generally, low-angle scattered light contains information on cell size and high-angle scattered light can be used to probe internal composition of the cell. To achieve 5-part differential, certain leukocyte populations, such as eosinophils, require special stain to change its scattering characteristics from other granulocytes, and basophils typically need to be counted separately following the differential lysis of other leukocytes. McKenzie, Clinical Laboratory Hematology, Prentice Hall, 2004; Fujimoto, Sysmex J. Int. 9 (1999).

In general, conventional automated cell analyzers are bulky, expensive, and mechanically complex, which restricts their locations to hospitals or central laboratories. Conventional cell analyzers require larger sample volumes and generate more waste than the systems developed using microdevices. Furthermore, for analysis of certain cell types, such as leukocytes, accuracy and speed of counting, differentiation, and/or sorting is important for determining disease state and treatment.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS(S)

FIG. 1A shows the molecular structure of acridine orange.
FIG. 1B shows leukocyte staining results with acridine orange.
FIG. 2 shows the top view of one embodiment of a novel fabricated microfluidic apparatus.
FIG. 3 shows one embodiment of the optical system setup.
FIG. 4A shows erythrocyte concentration in whole blood.
FIG. 4B shows leukocyte staining in whole blood with acridine orange at a concentration of 100 ng/mL.
FIG. 4C shows leukocyte staining in whole blood with acridine orange at a concentration of 1 µg/mL.
FIG. 4D shows leukocyte staining in whole blood with acridine orange at a concentration of 10 µg/mL.
FIG. 4E shows leukocyte staining in whole blood with acridine orange at a concentration of 100 µg/mL.
FIG. 4F shows leukocyte staining in whole blood with acridine orange at a concentration of 1 mg/mL.
FIG. 5 shows fluorescent signal bleaching from a single leukocyte in one embodiment.
FIG. 6A shows an image of background and a 5 µm bead with focused laser illumination flow taken by CCD camera with long pass emission filter, according to one embodiment.
FIG. 6B shows an image of background and a 5 µm bead with laser illumination flow taken by CCD camera with long pass emission filter, according to one embodiment.
FIG. 6C shows an image of background and a 5 µm bead with diffused laser illumination flow taken by CCD camera with long pass emission filter, according to one embodiment.
FIG. 6D shows an image of background and the trace of a 5 µm bead with diffused laser illumination flow taken by CCD camera with long pass emission filter, according to one embodiment.
FIG. 7A shows a graph of detection of 5 µm fluorescent beads detection with photo-diode detector with long pass emission filter.
FIG. 8A shows background image of focused laser beam from video taken by CCD camera with long pass emission filter, according to one embodiment.
FIG. 8B shows a signal from a leukocyte from diluted whole blood testing from video taken by CCD camera with long pass emission filter, according to one embodiment.
FIG. 9 shows the lime trace of amplified photodiode signal of acridine orange stained undiluted whole blood with green emission filter centered at 525 nm, and peaks labeled, according to one embodiment.
FIG. 10 shows a histogram of signal intensity from photodiode detector with green emission filter centered at 525 nm.
In certain embodiments, the microfluidic apparatus further comprises a detection zone, and/or a filter array, each in fluid communication with said channel and said fluid outlet.

A microfluidic apparatus comprising a substrate having at least one first channel having a defined physical feature; at least one first fluid comprising sheath fluid and said fluid biological sample; at least one second channel having a defined physical feature, wherein said second channel is in fluid communication with at least one fluid flow outlet; and a fluid biological sample. In certain embodiments, said fluid biological sample comprises blood.

In certain embodiments, the microfluidic apparatus further comprises a detection zone, and/or a filter array, each in fluid communication with said channel and said fluid outlet.

In certain embodiments, said filter structure comprises a filter array, said first fluid comprises sheath fluid and said second fluid comprises blood.

In certain embodiments disclosed herein relate to a detection system comprising a microfluidic apparatus and further comprising a light source; a lens assembly; a filter assembly; and an image capture device. In some embodiments, the detection system further comprises at least one display unit or at least one recording unit. In certain particular embodiments, said excitation source comprises a laser, particularly an argon laser. In particular embodiments, said filter assembly comprises an excitation filter, and at least one emission filter, and said display unit comprises a personal computer.

In particular embodiments, the microfluidic device may be used for labeling, sensing, differentiating, and/or sorting cell populations.


In the area of optical sensing, microfabrication has allowed development of microdevices to replace glass cap-
filary-based flow chambers, and to integrate compact optics and provide on-chip sample transport.

Cell sensing and counting, particularly of leukocytes, is cumbersome due in part to the cell population numbers. For leukocyte differential in microdevices based on optical sensing, a Y-groove micro-channel was fabricated by anisotropic wet etching of a silicon substrate and 3-part leukocyte differential was demonstrated for diluted blood without sheath flow by two-parameter light scattering. Altenendorf, Proceedings of the Int’l Conference on Solid State Sensors and Actuators, v. 1, p. 531 (1997).

However, until the instant embodied disclosure, it was necessary to dilute cell samples for cell sensors and actuators for many reasons. One reason dilution has been necessary is in order to prevent the coincidence effect in which multiple cells appear in the detection zone simultaneously. In human blood, the ratio of erythrocytes, or red blood cells, to leukocytes is on the order of about a thousand to one, a dilution factor of from about one hundred to several tens of thousands is typically required to avoid erythrocyte interference for electrical impedance or light scattering detection. Furthermore, for counting leukocytes in samples where leukocytes are specifically fluorescently labeled, a dilution of at least ten times is usually required. Sheenan and Storey, J. Pathol. Bacteriol. 59; 336 (1947); Kass, J. Clin. Pathol. 76; 810-12 (1981); Weigl et al., Biomed. Microdev. 3; 267-274 (2001).

Dilution is also often required in order to avoid clogging sample chambers, and also in order to remove erythrocytes that are lysed prior to running the sample, particularly for electrical impedance or light scattering detection. Some of these protocols also require an additional fixation buffer.

Dyes

In the present disclosure, a dye, such as Acridine orange (3,6-dimethylaminocarboxymethyl, FIG. 1), can be used to differentiate a target, such as cells, organelles, granules, nuclei, molecules (including double or single stranded nucleic acids, such as DNA, or RNA, chromosomes, and also including synthetic forms). In one particular embodiment, leukocytes may be detected, counted, or sorted without need for lysing erythrocytes or fixing the cell sample. Certain dyes, such as Acridine orange, are also desirable due to the fast diffusion into cells, easy commercial availability, and excitation and emission wavelength compatibility with common light sources (i.e. argon laser and other broad spectrum light sources in visible range) and optical filters. Kosenow, Acta Haematol. 7, 217 (1952); Schilcher, Blood, 19, 200 (1962); Jackson, Blood, 17, 643 (1961); Hallermann et al., Verh Deutsch Ges Inn Med. 70, 217 (1964).

Acridine orange is a pH-sensitive fluorescent cationic dye that binds to double-stranded DNA by electrostatic attraction and intercalation of the Acridine orange between base pairs. Upon binding, the excitation maximum becomes 502 nm and the emission maximum becomes 525 nm (green). Acridine orange also binds to RNA and single-stranded DNA, with a shifted excitation maximum of 460 nm and an emission maximum of 650 nm (red). Adams and Kamentsky, Acta Cytol. 15, 289 (1971); Adams and Kamentsky Acta Cytol. 18, 389-91 (1974); Steinbek et al., Acta Cytol. 17, 113-17 (1973). Acridine orange is also desirable in that it is hydrophobic in neutral pH, and can easily diffuse through the cell membrane and cell nuclear membrane to bind to RNA and DNA. In living cells, Acridine orange is protonated in the acidic environment of lysosomes, which makes it cationic and prevents the dye from leaking out of lysosome membranes. Moriyama et al., J. Biochem. 92; 1333-36 (1982). When Acridine orange is used for leukocyte analysis, the cell nucleus is stained green with slightly mixed red, a result of double-stranded DNA and single-stranded RNA, while the cell cytoplasm is stained red due to the RNA and lysosomes. Thus, leukocyte counting can be achieved easily by using the strong signal from the green fluorescent channel. Leukocyte differentiation can be achieved by analyzing the signal from the red fluorescence channel.

For fresh-stained leukocytes, a 3-part differential (lymphocytes, monocytes, and granulocytes) can be achieved by studying the red fluorescent signal of an Acridine orange stained diluted blood sample, whereas a 5-part differential leukocytes (lymphocytes, monocytes, neutrophils, eosinophils, and basophils) has been demonstrated with hypotonic dilution and fresh Acridine orange-stained leukocyte samples. Adams and Kamentsky, Acta Cytol. 15, 289 (1971); Adams and Kamentsky, Acta Cytol. 18, 389-391 (1974); Steinbek et al., Acta Cytol. 17, 113-17 (1973).

Other dyes can be utilized with certain embodiments described in the instant disclosure, such as ethidium bromide, three-dye combinations (ethidium bromide, brilliant green, and, and/or a polymethine dye). Shapiro et al., J. Histochem. Cytochem. 24, 396-411 (1976); Shapiro et al., J. Histochem. Cytochem. 25, 836-844 (1977); U.S. Pat. No. 4,376,820; U.S. Pat. No. 4,882,284; Tibbe, et al., Nat. Biotechnol. 17, 1210-1213 (1999); U.S. Pat. No. 4,400,370; Kass, J. Histochem. Cytochem. 36, 711-715 (1988).

Apparatus

One embodiment of the instant disclosure relates to a device or apparatus for cell counting or differentiating. In particular embodiments, the device or apparatus comprises a substrate formed from a material, such as silicon, glass, plastic, metal, or other material. One particular embodiment of the instant disclosure was fabricated using soft lithography. Quake, Science 290, 1536-40 (2000). Other photolithographic or etching techniques could also be used, according to specific embodiments.

One embodiment of the device was microfabricated using two parts of PDMS (polydimethylsiloxane) (Sylgard 184, Dow Corning, MI, USA) mixed vigorously in 10:1 ratio. After degassing in vacuum for about 30 minutes, the mixture was poured onto DRIE-etched silicon mold, that had been pretreated with HMDS (hexamethyldisilazane) for easy separation after baking. The molds were baked at 80°C for 30 minutes. The hardened PDMS was separated from the silicon mold, and PDMS sheet was cut into pieces and fluidic access holes were punched on each piece with a Luer stub adapter (Beecton Dickinson, NJ, USA). Each PDMS piece was carefully placed on a cleaned glass slide and baked overnight at 80°C. In some cases, oxygen plasma treatment (300 m Torr, 25 W, 30 s) was used for PDMS and glass slides in order to improve adhesion between them, particularly with devices that were intended to be reused. Bhattacharyya et al., J. Microelectromechan. Syst. 14, 590-97 (2005).

In one particular embodiment, the channel structure was molded on a 1 cm x 1 cm PDMS block, with the thickness of the PDMS block at less than 3 mm. In one particular embodiment the channel depth was 16 μm in order to accommodate large leukocyte sizes.

One exemplary embodiment of the device is shown in FIG. 2. For this particular embodiment, a first fluid flow inlet allows for deposition of, for example sheath flow fluid, and is in fluid communication with a bifurcated channel with a first channel arm and a second channel arm that both converge at a junction of a reservoir and the
In one particular embodiment, the apparatus further comprises a second fluid flow inlet that allows for deposition of, for example, a sample fluid, such as blood, that is in fluid communication with a filter array structure, by way of a branched sample flow zone channel and a fluid flow outlet. In this particular exemplary embodiment, 2-D hydrodynamic focusing was adopted to control the particle position of the cell sample in the detection zone. According to the embodiment shown in FIG. 2, the ratio of cross-sectional area of sheath flow to core sample flow was 10:1, and the channel width of the detection zone was 50 µm, with the width of the focused sample flow preferably 5 µm or less. In particular embodiments, the channels comprise a physical feature, such as a depression or a protrusion.

One other exemplary embodiment of the device is shown in FIG. 13A. For this particular embodiment the fluid flow inlet allows for deposition of a sample fluid, such as a biological sample, or other fluid sample containing a target. In one particular embodiment, the biological sample includes a cell sample, such as blood. In this exemplary embodiment, the fluid inlet is in fluid communication with a first channel that is juxtaposed to a second channel which comprises the detection zone which is also in fluid communication with the fluid flow outlet. In certain embodiments, the height of the first and/or second channels is approximately 5 µm, approximately 8 µm, approximately 10 µm, approximately 12 µm, approximately 15 µm, approximately 20 µm, approximately 25 µm, approximately 30 µm, approximately 35 µm, approximately 40 µm, or any value therebetween. In certain embodiments, the width of the second channel is approximately 5 µm, 10 µm, approximately 15 µm, approximately 20 µm, approximately 25 µm, approximately 30 µm, approximately 35 µm, approximately 40 µm, or any value therebetween. In the exemplary embodiment shown in FIG. 13A, the second channel width was approximately 20 µm in size.

One other exemplary embodiment of the device is shown in FIG. 13B. For this particular embodiment, the fluid flow inlet allows for deposition of a sample fluid, such as a biological sample, or other fluid sample containing a target. In one particular embodiment, the biological sample includes a cell sample, such as blood. In this exemplary embodiment, the fluid inlet is in fluid communication with a first channel that is juxtaposed to a second channel which comprises the detection zone which is also in fluid communication with the fluid flow outlet. In certain embodiments, the height of the first and second channels is approximately 5 µm, approximately 8 µm, approximately 10 µm, approximately 12 µm, approximately 15 µm, approximately 20 µm, approximately 25 µm, approximately 30 µm, approximately 35 µm, approximately 40 µm, or any value therebetween. In certain embodiments, the width of the second channel is approximately 5 µm, 10 µm, approximately 15 µm, approximately 20 µm, approximately 25 µm, approximately 30 µm, approximately 35 µm, approximately 40 µm, or any value therebetween. In the exemplary embodiment shown in FIG. 13B, the second channel width was approximately 30 µm in size.

In one particular embodiment, the channel length of the detection zone is 1000 µm. A filter structure upstream of the sample flow zone 220 may also be included in certain embodiments, which filtered out contaminants, including erythrocyte rouleaux, and other large particle aggregates to prevent clogging in the detection zone 240. In certain embodiments, the size of the rectangular pillar structure components of the filter structure was 200 µm x 40 µm. The spacing between the pillars in each of the three rows was 40 µm, 30 µm, and 20 µm respectively, which allows for even the largest leukocytes to pass through the filter region 230.

The optical system was set up on an optical bench as shown in FIG. 3 (transmitted laser-induced fluorescent detection system or LIF). In one particular embodiment, the system setup comprises an excitation or laser source, a lens assembly, a microfluidic apparatus, an optional additional lens assembly, a filter assembly, and an image capture device. In certain embodiments, one or more emission filters comprise, the filter assemblies, and an image capture device. In particular embodiments, the image capture device may be coupled to communicate with a display unit or computing device such as a personal computer. One of skill in the art would recognize that multiple and various computer software programs are available that allow for integration, compilation, analysis, reconfiguration, and other manipulation of data received from the system, particularly by way of the computing device.

In one particular exemplary embodiment, an argon laser (National Laser NLC210BL, 483 nm, and 15-30 mW adjustable, Salt Lake City, Utah, USA) is used as the excitation source. An aperture 310 of 50 µm diameter is put in front of the laser output to facilitate the alignment process and lower the illumination intensity. In certain embodiments, a long-working-distance microscope objective (USMCO M Plan Apo, 10x, 0.28 NA, Dayton, Nev., USA) is used as a condenser lens.

Another long-working-distance microscope objective (Bausch & Lomb, 50x, 0.45 NA, Rochester, N.Y., USA) is used as an objective lens. In the same embodiment, three emission filters are used in one particular test: 488 nm long pass filter (Chroma H1500 LP, transition width=4.9 nm, edge steepness=2.5 nm, Rockingham, Vt., USA), a green bandpass filter with central wavelength 525 nm and a bandwidth 50 nm (Chroma DS525_50 m), and a red bandpass filter with central wavelength 650 nm and bandwidth 50 nm (Chroma DS650_50 m). A broadband non-polarizing hybrid cube beamsplitter 370 (Newport 05BC17MB.1, 400-700 nm, R/T=45%/45%, Irvine, Calif., USA) is used to direct light to the photodiode detector and CCD camera simultaneously.

The signal is electrically amplified and detected either with a silicon photodiode receiver module or a photon multiplier tube (PMT, Hamamatsu H5784-20, 400-700 nm, R/T=45%/45%, Irvine, Calif., USA) is used to direct light to the photodiode detector and CCD camera simultaneously.
In one exemplary embodiment, fresh human blood was obtained from healthy donors and used within 3 days of collection. EDTA was added to the blood samples in order to prevent coagulation. For Acridine orange staining, the stock solution was added to obtain a final dye-concentration of 10 µg/mL in Ficoll-Paque Plus. Ficoll-Paque Plus was also used as the sheath flow solution. Fluorescent polystyrene beads (5 µm green fluorescent beads) were purchased from Duke Scientific Corporations, Fremont, Calif., USA.

Cell nucleus stain Acridine orange was obtained from Molecular Probes, Eugene, Oreg., USA, and dissolved in water to achieve a 10 mg/mL stock solution. Blood diluted Ficoll-Paque Plus was purchased from Amersham Biosciences, Sweden. Phosphate buffered saline (10xPBS) was obtained from Ambion (9625), Austin, Tex., USA.

Staining results were observed under a fluorescent microscope (Nikon E800, Japan) with a triple band filter block DAPI-FITC-TRITC, which has excitation filter wavelengths of 385-400 nm, 475-490 nm, and 545-565 nm, and emission filter wavelengths of 450-465 nm, 505-535 nm and 580-620 nm. Images were taken with a cooled CCD camera (RT-KE color 3-shot, Diagnostic Instruments, Sterling Heights, Mich., USA). Rough count of leukocytes was made with a hemacytometer (Hauser Scientific, Horsham, Pa., USA). When necessary, blood or fluorescent beads were diluted with Ficoll-Paque Plus (specific gravity 1.077 g/mL) to match the specific gravity of the solvent to leukocytes. All fluids were pumped into the devices using syringe pumps (Harvard Apparatus Pico Plus, Holliston, Mass., USA).

In this particular embodiment, an analog CCD camera was used for video recording at a matched camera frame rate of 30 frames per second and then converted to digital format and stored in a computer. Other imaging capture devices, such as CMOS, PMT, or still other devices may also be used with particular embodiments described herein. In certain exemplary test runs, the system set up utilizing a photodiode detector and PMT are more sensitive than the CCD camera and have a faster time response. During one exemplary test run, the optical system was first roughly aligned on a dummy device with the aid of images from CCD camera. A 10 µm diameter illumination spot on the detection zone is easily achieved with proper alignment.

As shown in FIG. 12A and FIG. 12B, the instant apparatus may be incorporated into a hand-held unit comprising a laser source (such as a laser emitting diode or LED), at least one lens, at least one filter assembly with optional beamsplitter, a microfluidic apparatus as described herein on a microchip or other substrate, an input/output port, at least one image capture device, which may be a photomultiplier tube. In certain embodiments, the hand-held unit may be assembled and enclosed by an outer casing or casings, and rivets or bolts.

Cell Detection

One aspect of the instant disclosure relates to methods of counting and/or differentiating cells, particularly leukocytes, from undiluted cell samples, such as human or other animal blood, by utilizing microfabricated devices. In one exemplary embodiment, cell detection was conducted utilizing Acridine orange and fresh whole human blood.

In one exemplary embodiment, fresh human blood was obtained from healthy donors and used within 3 days of collection. EDTA was added to the blood samples in order to prevent coagulation. For Acridine orange staining, the stock solution was added to obtain a final dye-concentration of 10 µg/mL in Ficoll-Paque Plus. Ficoll-Paque Plus was also used as the sheath flow solution. Fluorescent polystyrene beads (5 µm green fluorescent beads) were purchased from Duke Scientific Corporations, Fremont, Calif., USA.

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Staining results were observed under a fluorescent microscope (Nikon E800, Japan) with a triple band filter block DAPI-FITC-TRITC, which has excitation filter wavelengths of 385-400 nm, 475-490 nm, and 545-565 nm, and emission filter wavelengths of 450-465 nm, 505-535 nm and 580-620 nm. Images were taken with a cooled CCD camera (RT-KE color 3-shot, Diagnostic Instruments, Sterling Heights, Mich., USA). Rough count of leukocytes was made with a hemacytometer (Hauser Scientific, Horsham, Pa., USA). When necessary, blood or fluorescent beads were diluted with Ficoll-Paque Plus (specific gravity 1.077 g/mL) to match the specific gravity of the solvent to leukocytes. All fluids were pumped into the devices using syringe pumps (Harvard Apparatus Pico Plus, Holliston, Mass., USA).

In this particular embodiment, an analog CCD camera was used for video recording at a matched camera frame rate of 30 frames per second and 30 nL/minute sheath flow rate. For photodiode detection, a 0.1 µL/minute sample flow rate and a 1 µL/minute sheath flow rate were used. A 1 µL/minute sample flow and a 10 µL/minute sheath flow were used with the photon multiplier tube instrument.

In order to achieve a high signal-to-noise ratio, the maximal concentration for cell staining was established using routine methods in the art. Adams and Kamentsky, Acta Cytol. 15: 289 (1971). As shown in FIG. 4, whole blood samples were analyzed with different Acridine orange concentrations. The optimal concentration for leukocyte staining was determined to be approximately in the range of 0.01 µg/mL. In the particular exemplary embodiment utilized in FIG. 4, the distance between the coverslip and the grid surface was approximately 100 µm. As can be seen in FIG. 4A, an abundance of erythrocytes were present under the field of view, yet these cells did not interfere with the fluorescent signal from the leukocytes, as shown in FIG. 4B-F.

As can be seen in FIG. 5, the exemplary embodiment utilized in cell detection did not experience any significant photobleaching. The signal was fitted as a first-order exponential decay with time constant of 6.4/±0.7 seconds. Two more tests confirmed that the photobleaching time constant for one particular embodiment was between 1 second and 10 seconds. The photobleaching time constant for one particular embodiment was characterized by filling the device with Acridine orange-stained whole blood. The channel was scanned by the laser spot and the illumination was set to be the same as that used in testing. The entire process was recorded with a CCD camera. Whenever a fluorescing leukocyte was observed with fluorescent emission clearly distinct from the background, we stopped moving the laser spot and waited until the leukocyte was photographed above background level. The images were extracted from the video, converted to 8-bit gray scale images, and analyzed with a Matlab program. The data was fitted to a single time-constant exponential decay.

Additionally, green fluorescent beads were tested at a concentration of about 2 x 107/µL, as observed by CCD camera, and shown in FIG. 6. Sample flow rate was set at about 3 nL/min, and sheath flow was about 30 nL/min. In one exemplary test run, a hydrodynamic focused laser beam, as shown in FIG. 6A, created an enlarged light circle as shown in FIG. 6B. Only a single bead normally appeared in each image. With diffused laser illumination, as shown in FIG. 6C, the trace of the bead could be identified, as shown in FIG. 6D. Hydrodynamic focusing limits the cross-sectional area of the detection zone without shrinking the channel diameter, thus the signal-to-noise ratio may be improved without increasing the risk of clogging the channel. Also, the reduction of the cross-section of the core flow reduces the coincidence effect. Finally, enclosing the core sample flow with sheath flow minimizes fluorescent dye absorption in the device walls, thus reducing background noise. As indicated in FIG. 7, bead signals from the photodiode detector could easily be identified.

As shown in FIG. 8, using both red and green emission filters, images extracted from video taken by the CCD camera show the signal identified from a leukocyte stained with Acridine orange, as well as the signal obtained from the fluorescent control bead. For photodiode detection, the expected leukocyte detection rate would average about 4-11 cells per second for a normal individual.

In one exemplary embodiment, a time trace over 50 seconds of an undiluted blood sample stained with Acridine orange using a green emission filter, and a throughput of up to about 1000 leukocytes per second was attained. Maxima signal Intensity (peak height as in FIG. 9) from the green
fluorescent channel with 525 nm emission filter was studied by plotting its histogram, as shown in FIG. 10. As expected, the lower-intensity portion is likely contributed mainly by lymphocytes, while the higher-intensity portion is likely mainly from monocytes, with the center-region is likely mostly from granulocytes. Steinikam et al., Acta Cytol. 17; 113-117 (1973).

In one exemplary embodiment, a time trace over 50 seconds of an undiluted blood sample stained with Acridine orange using a red fluorescent channel with 650 nm emission filter was conducted. As shown in FIG. 11, two peaks were identified. The lower intensity is dominated by lymphocytes and the higher-intensity peak is largely monocytes and granulocytes. The time between the start of staining the cells to photodiode recording was typically greater than 15 minutes.

In both exemplary studies, the maximal throughput was about 1000 leukocytes per second utilizing one embodiment of the PMT detector. By using undiluted blood, minimal sample volume was maintained, which increases the throughput. Since sample throughput is proportional to volume flow rate, but is limited by the maximal pumping rate and response time of the sensing system, a 3 nL/minute core flow rate was used with the CCD camera detection. Under this flow rate, a typical leukocyte traveled through the detection zone in approximately 30 milliseconds, which roughly equals the CCD frame acquisition time.

Flow rates for varying embodiments may be suitable for a range from approximately 1 nL/minute, approximately 2 nL/minute, approximately 3 nL/minute, approximately 4 nL/minute, approximately 5 nL/minute, approximately 6 nL/minute, approximately 7 nL/minute, approximately 8 nL/minute, approximately 9 nL/minute, approximately 10 nL/minute, approximately 20 nL/minute, approximately 30 nL/minute, approximately 40 nL/minute, approximately 50 nL/minute, approximately 60 nL/minute, approximately 70 nL/minute, approximately 80 nL/minute, approximately 90 nL/minute, approximately 100 nL/minute, approximately 110 nL/minute, approximately 120 nL/minute, approximately 130 nL/minute, approximately 140 nL/minute, approximately 150 nL/minute, or any value therebetween for photodiode detection. Likewise, for PMT detection, flow rates for varying embodiments may be suitable for a range from approximately 200 nL/minute, approximately 300 nL/minute, approximately 400 nL/minute, approximately 500 nL/minute, approximately 600 nL/minute, approximately 700 nL/minute, approximately 800 nL/minute, approximately 900 nL/minute, approximately 1 µL/minute, approximately 2 µL/minute, approximately 3 µL/minute, approximately 4 µL/minute, or any value therebetween.

In one exemplary embodiment, the time response of the photodiode receiver module under low sensitivity setting was 0.16 milliseconds, and 0.6 milliseconds under high sensitivity, while the time response of the PMT detector in one exemplary run was about 16 microseconds.

Furthermore, by decreasing the cross-sectional area, the linear flow velocity of the core flow is increased, which requires faster sensing, and reduces the coincidence effect by increasing the average distance between cells in the detection zone.

Thus, by utilizing particular embodiments disclosed herein relating to a microfluidic device, leukocyte sensing, counting, and sorting can be achieved one-by-one in a micro flow cytometer system. Furthermore, dense cell suspensions, such as whole, undiluted blood may be utilized in certain embodiments described herein, which provides for reduced sample and waste volume, reduced processing time, and completely eliminates on-chip mixing and buffer storage. In particular aspects, leukocytes can be sensed one-by-one in a micro flow cytometer system.

As described herein, certain embodiments of the device can be implemented in various sizes and conformations, including but not limited to a bench-top device, a handheld device (such as is shown in FIG. 12), an implantable device, a nanotechnology device, or other size or conformation. In the smaller exemplary conformations, high-illumination LED is used for excitation and a minipump is used to manipulate the sample in suction mode, while fluorescent signals from green and red channels can be detected simultaneously.

INCORPORATION BY REFERENCE

All of the above U.S. patents, U.S. patent application publications, U.S. patent applications, foreign patents, foreign patent applications and non-patent publications referred to in this specification and/or listed in the Application Data Sheet, are incorporated herein by reference, in their entirety.

The invention claimed is:
1. A method for analyzing cells, the method comprising:
   (a) introducing a fluid sample comprising fluorescently labeled cells and fluorescently labeled control beads to an inlet of a first microfluidic channel, wherein the fluorescently labeled cells are labeled with a fluorophore different than a fluorophore label of the control beads;
   (b) flowing the fluid sample from the inlet of the first microfluidic channel to an outlet of a second microfluidic channel, wherein an outlet of the first microfluidic channel is physically and fluidically coupled to an inlet of the second microfluidic channel and wherein the flow of the fluid sample is aided with a pump;
   (c) exciting the fluorophores of the fluorescently labeled cells and of the fluorescently labeled control beads using an excitation source;
   (d) detecting fluorescence of the fluorescently labeled cells and fluorescence of the fluorescently labeled control beads as they pass through a detection zone within the second microfluidic channel, wherein the detection is performed on individual cells;
   (e) analyzing the detected fluorescence of the fluorescently labeled cells and fluorescence of the fluorescently labeled control beads using a computer software inscribed on a computing device; and
   (f) providing a result concerning one or more properties of the cells being analyzed.
2. The method of claim 1, wherein the cells are leukocytes.
3. The method of claim 1, wherein the fluid sample is a treated blood sample.
4. The method of claim 1, wherein the second microfluidic channel has fixed dimensions.
5. The method of claim 4, wherein the fixed dimensions are of approximately 40 microns by approximately 50 microns.
6. The method of claim 1, wherein the first channel and the second channel are obstacle-free.
7. The method of claim 1, wherein the flowing of the fluid sample from the inlet of the first microfluidic channel to the outlet of the second microfluidic channel increases a core flow rate of the fluid sample to form an increased flow rate.
8. The method of claim 1, wherein the method further comprises identifying the fluorescently labeled cells.
9. The method of claim 1, wherein the method further comprises sorting the fluorescently labeled cells.

10. The method of claim 1, wherein the method further comprises counting the fluorescently labeled cells.

11. The method of claim 1, further comprising video recording a measurement on a recording unit.

12. The method of claim 1, wherein the method can detect cells at a rate of up to about 1000 cells per second.

13. The method of claim 1, wherein the excitation source is a laser or a LED.

14. The method of claim 1, wherein the excitation source comprises a lens assembly.

15. The method of claim 1, further comprising manipulating the detected fluorescence using a computer software program.

16. The method of claim 15, further comprising displaying a result of the manipulation on a display unit.

17. The method of claim 1, wherein the fluorescently labeled beads are polymeric beads.

18. The method of claim 1, wherein the fluorescently labeled beads are polystyrene beads.

19. The method of claim 1, wherein the step of detecting fluorescence comprises using an image capture device.

20. The method of claim 1, wherein flowing the fluid sample comprises pumping the fluid sample through a microfluidic device with the pump.

21. The method of claim 1, wherein the fluid sample does not comprise a sheath fluid.

22. The method of claim 1, wherein the fluid sample is an undiluted blood sample.

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